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Balyasnikova et al.

(54) CAR T CELLS RECOGNIZING **CANCER-SPECIFIC IL 13RA2**

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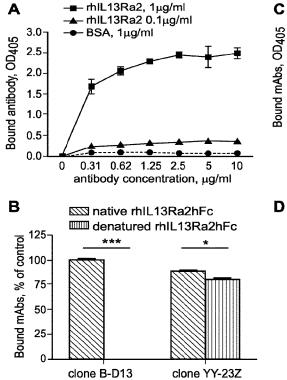
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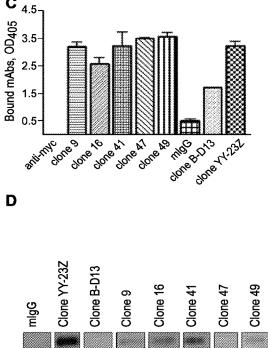
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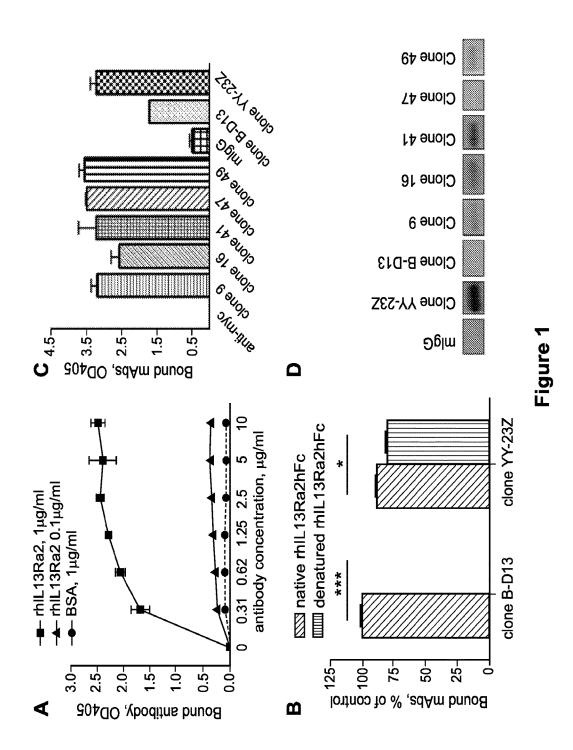
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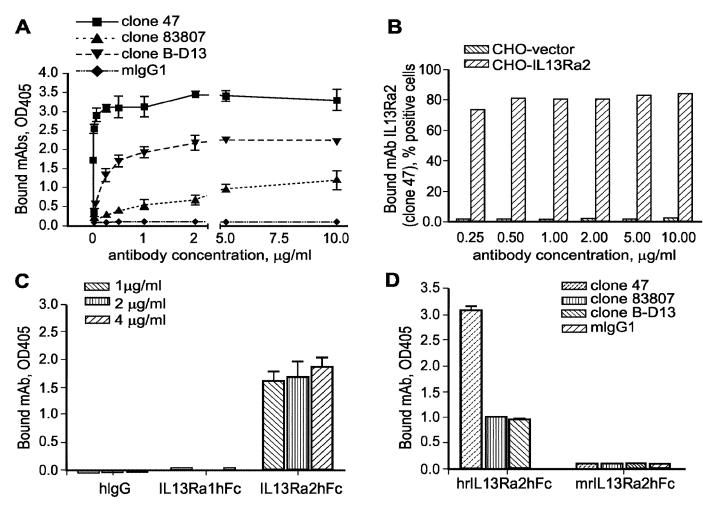
ABSTRACT (57)

Provided are specific binding molecules, or fragments thereof, that bind to an epitope of IL13R α 2, a receptor polypeptide preferentially found on the surface of cancer cells rather than healthy cells. Exemplary specific binding molecules are bispecific binding molecules that comprise a fragment of an IL13R α 2 binding molecule and a peptide providing a second function providing a signaling function of the signaling domain of a T cell signaling protein, a peptide modulator of T cell activation, or an enzymatic component of a labeling system. Also provided are polynucleotides encoding such a specific binding molecule (e.g., bispecific binding molecule), vectors, host cells, pharmaceutical compositions and methods of preventing, treating or ameliorating a symptom associated with a cancer disease such as a solid tumor disease (e.g., glioblastoma multiforme).

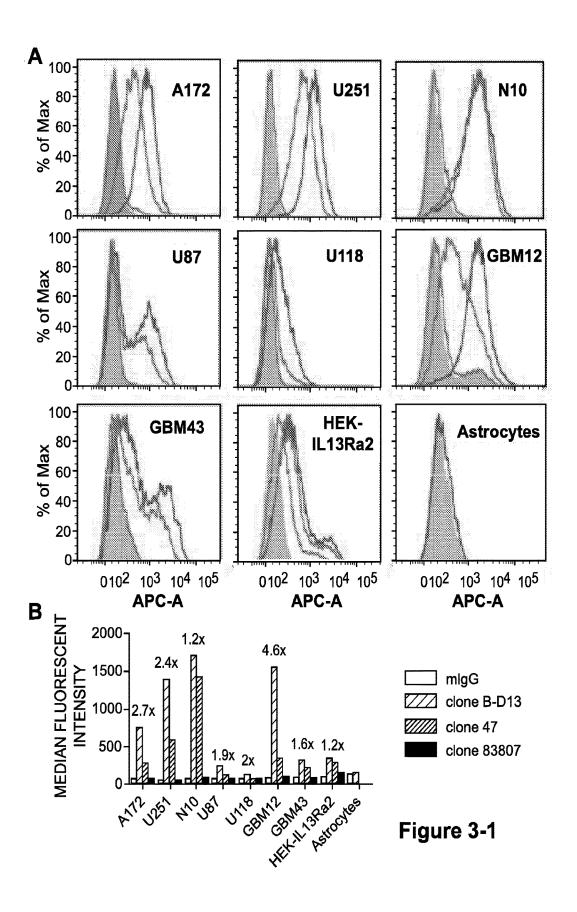


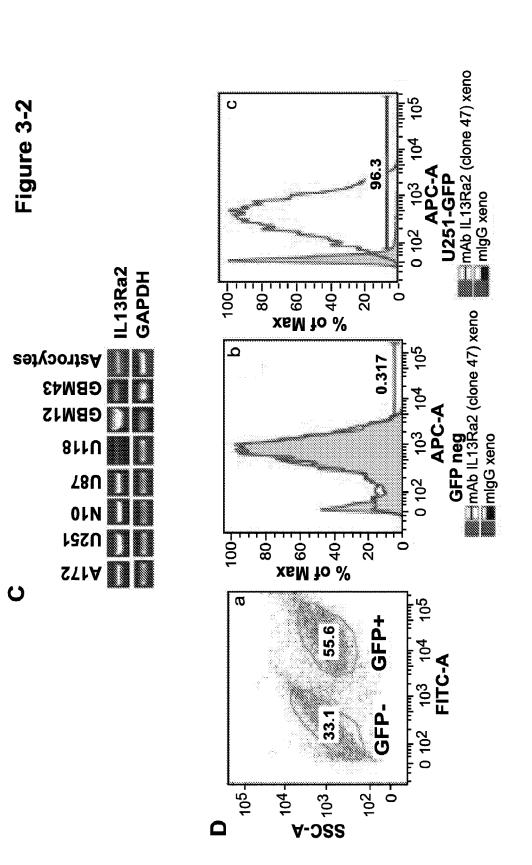


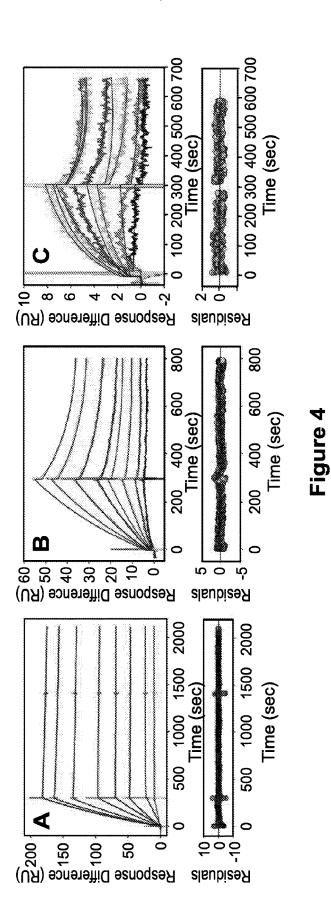


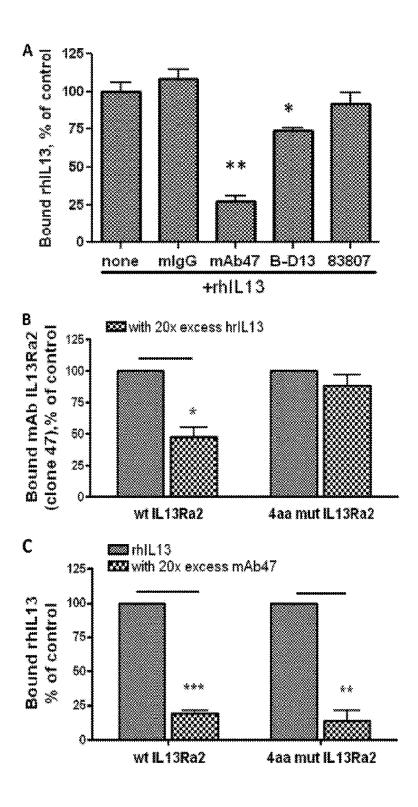












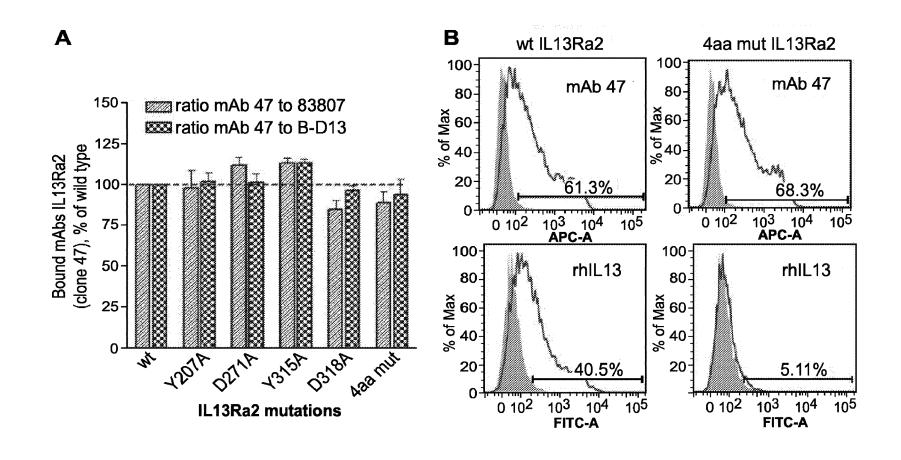


Figure 6

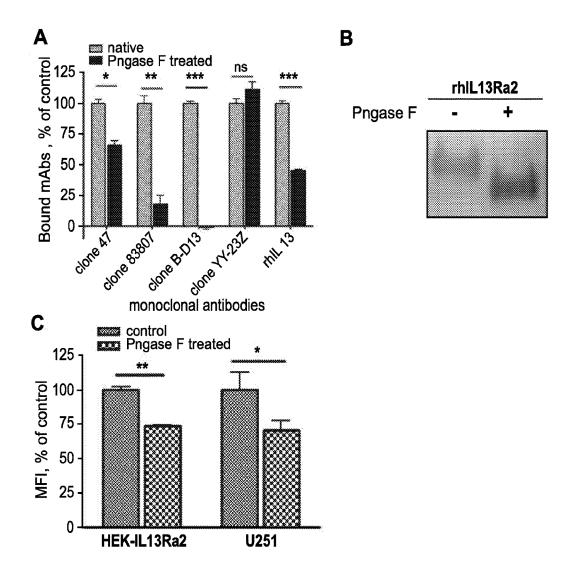
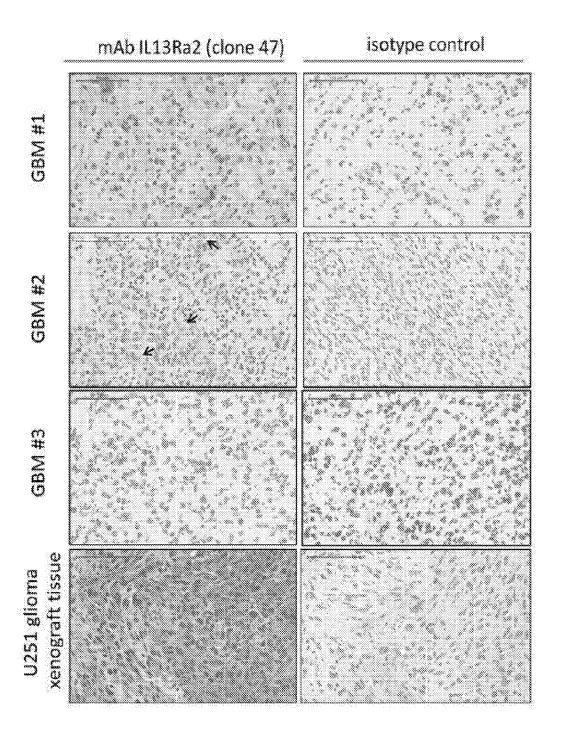
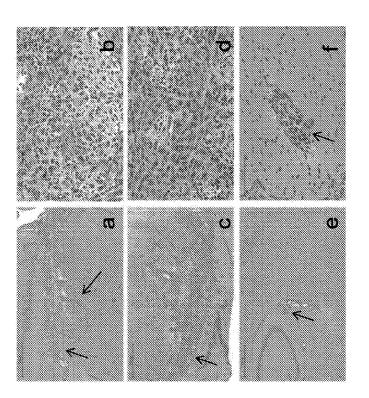
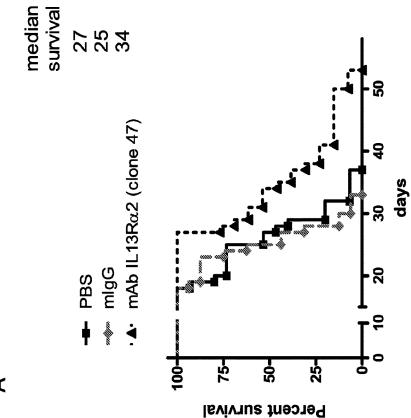


Figure 7



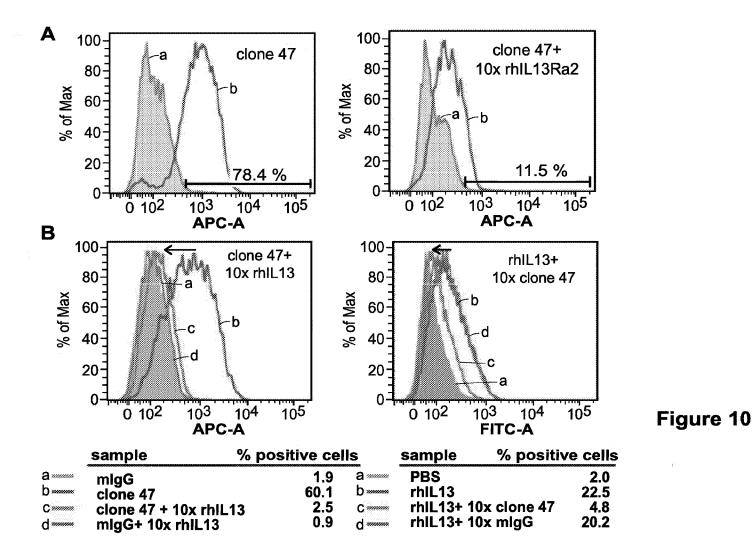




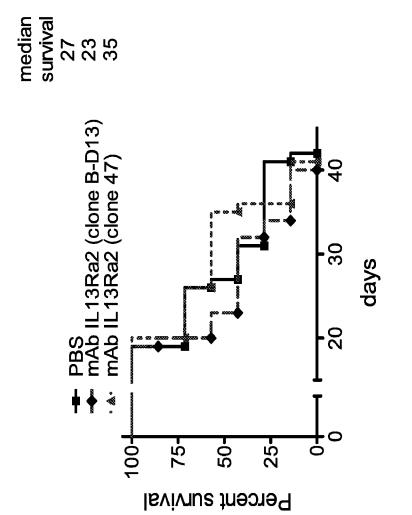


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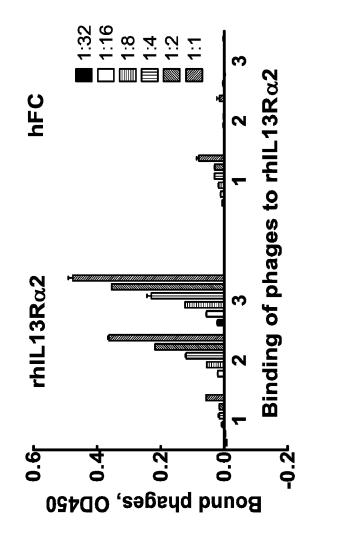
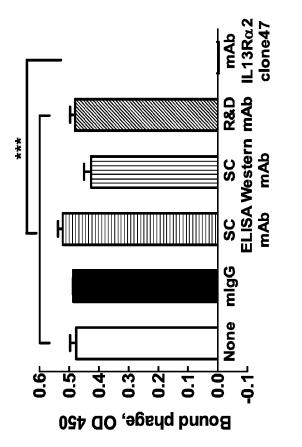
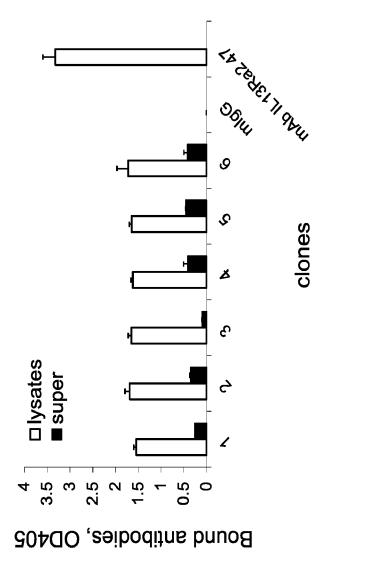
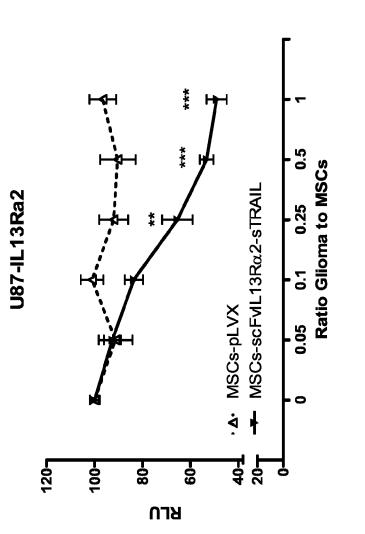


Figure 12











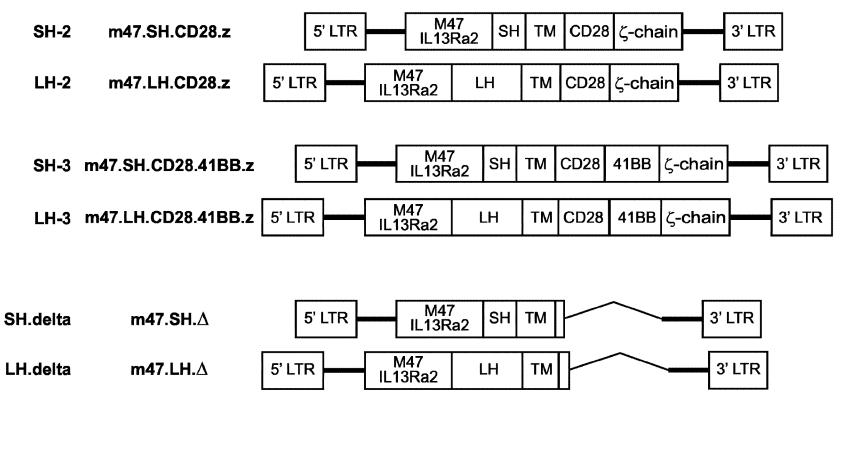


Figure 16

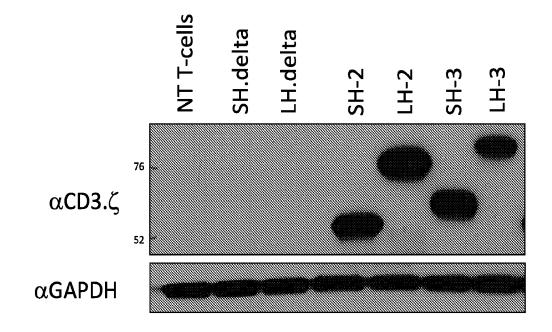


Figure 17

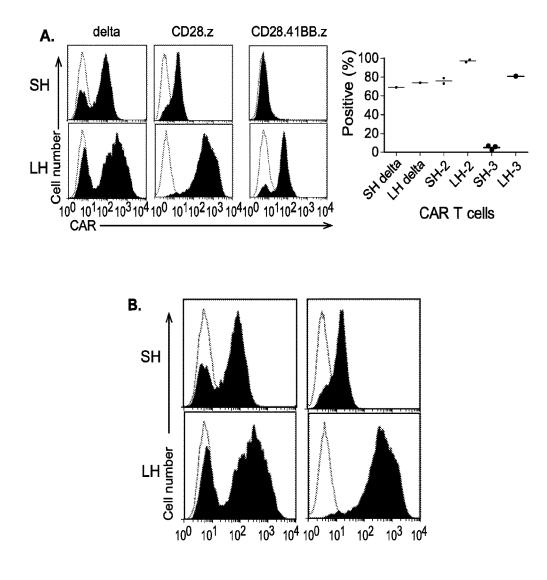


Figure 18

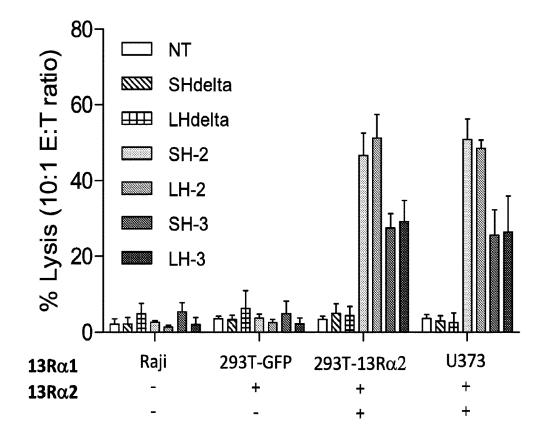


Figure 19

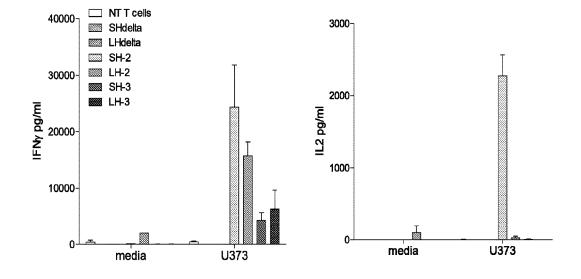


Figure 20

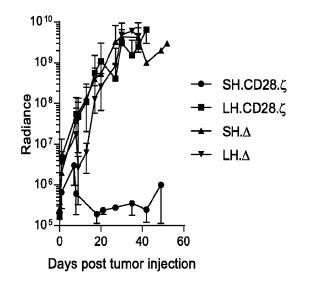


Figure 21

	Expression (WB)	Cell surface expression	Cyto	IFNγ	IL2	Anti-glioma activity in vivo
SH2A	n/a	+	-	•		-
SH3∆	n/a	+	-	-	-	-
SH2	+	+	+	++	+	+
LH2	+	+	+	++	-	bk.
SH3	+	+/-	+	+	-	ND
LH3	+	+	+	+	-	ND



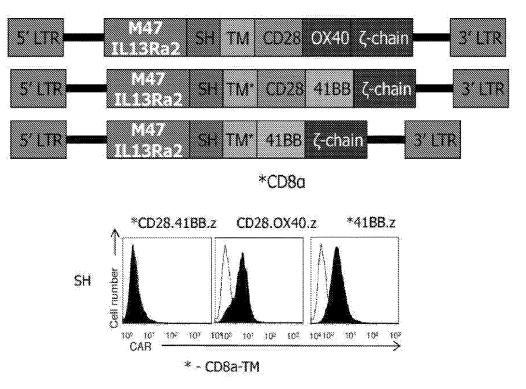


Figure 23

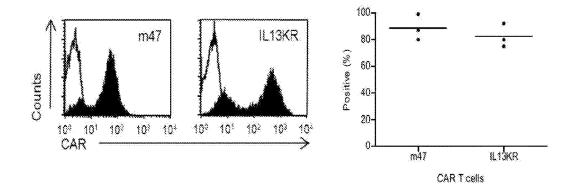


Figure 24

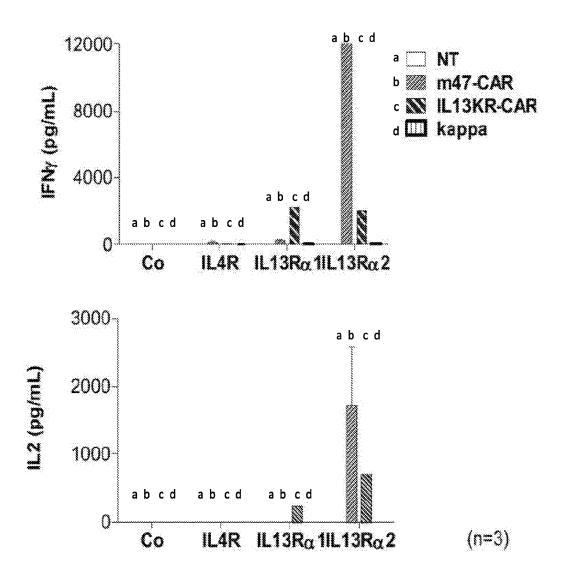
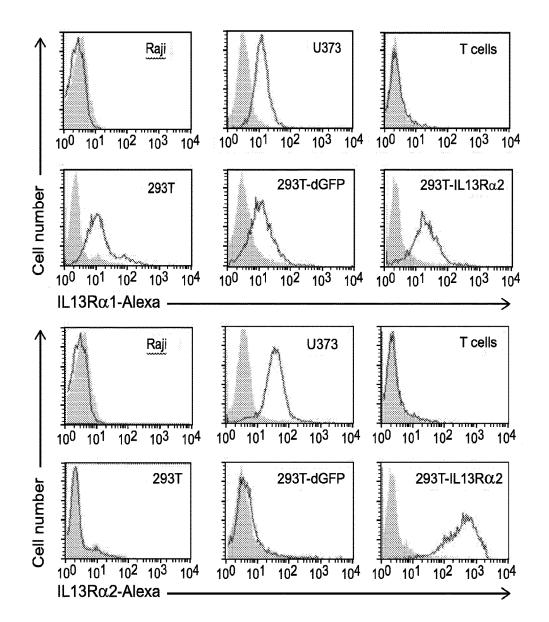


Figure 25



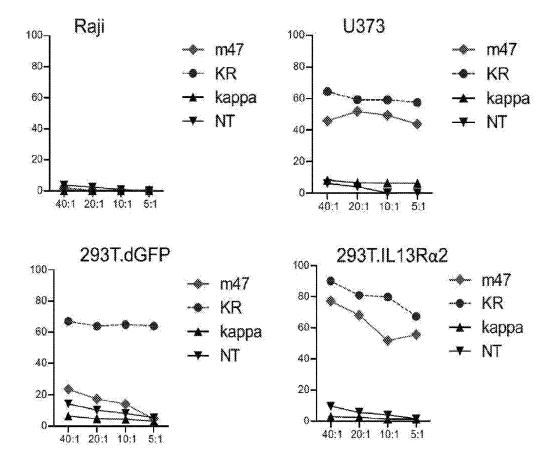
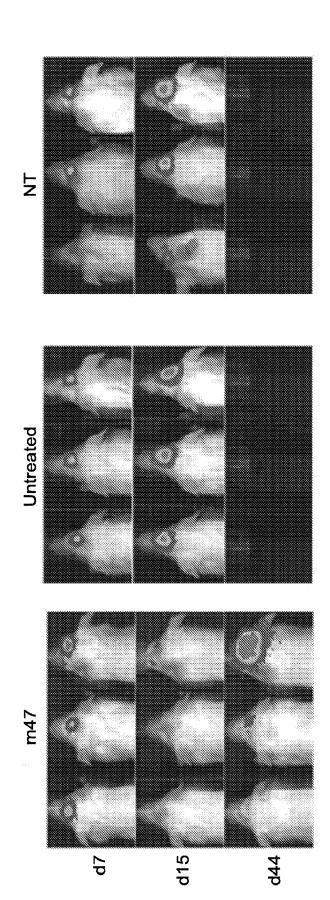


Figure 27





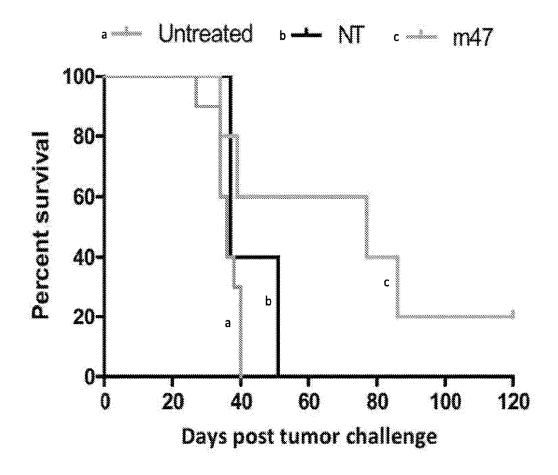
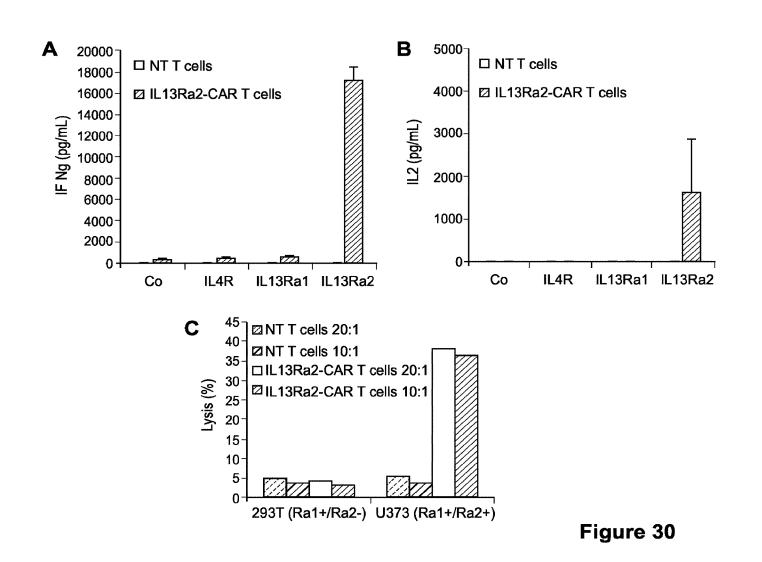


Figure 29





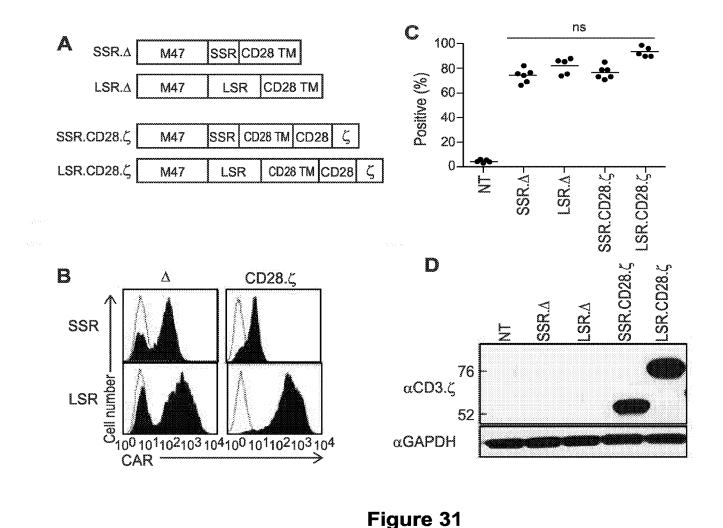
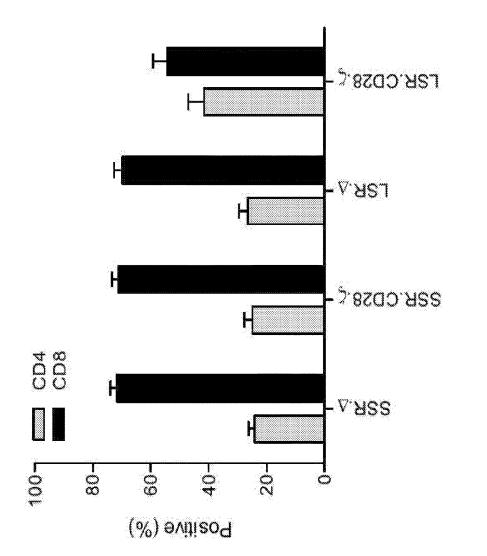
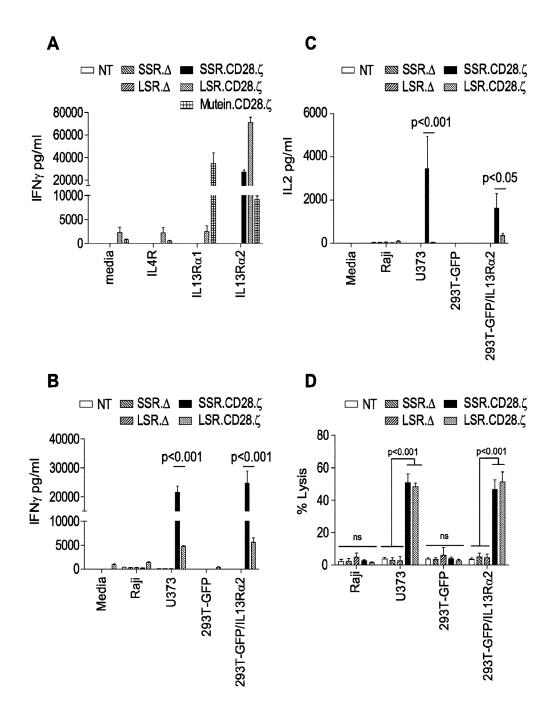


Figure 31







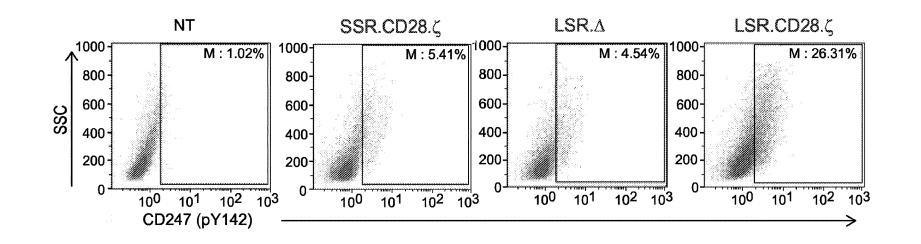
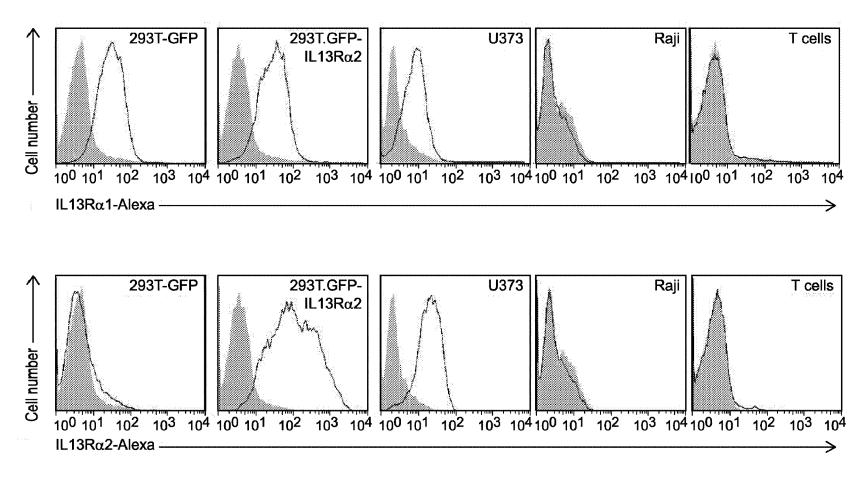


Figure 34





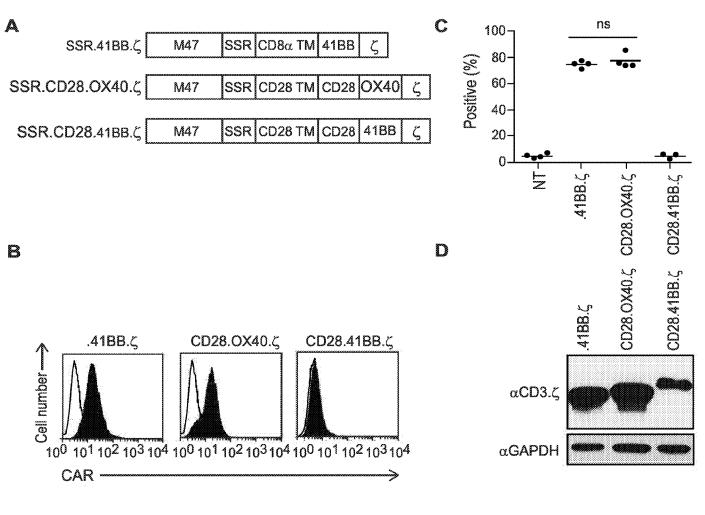


Figure 36

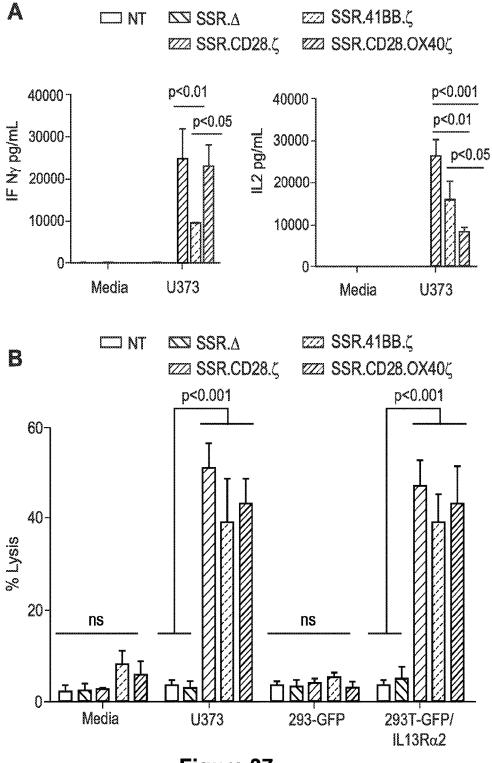


Figure 37

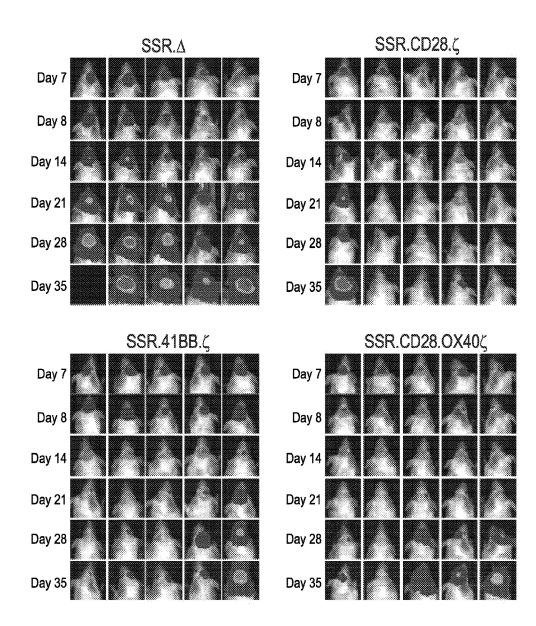


Figure 38A

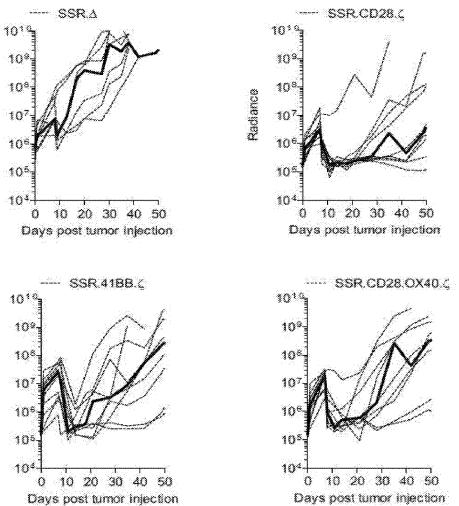




Figure 38B

B

Radiance

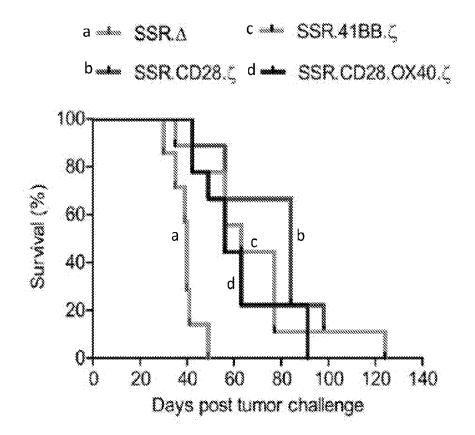
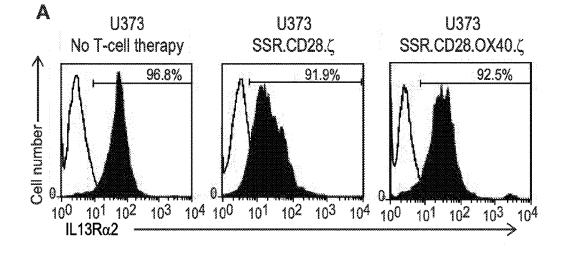


Figure 38C



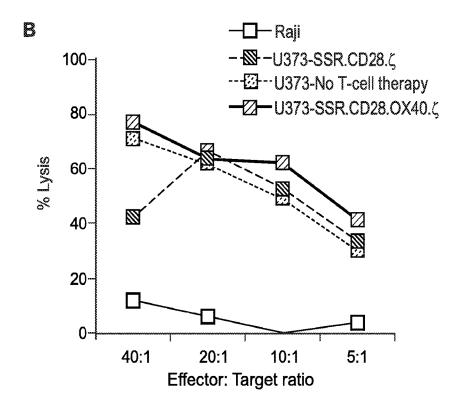
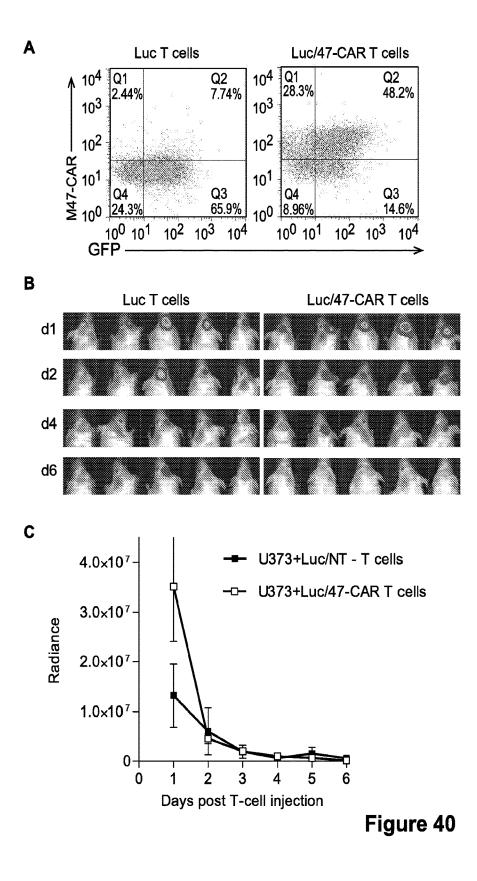
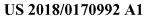


Figure 39





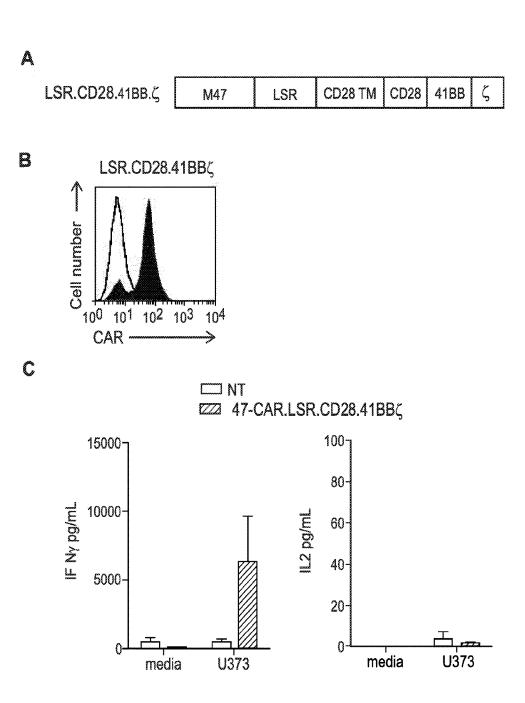
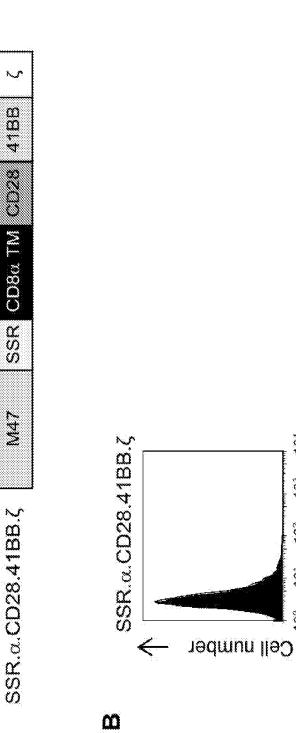


Figure 41





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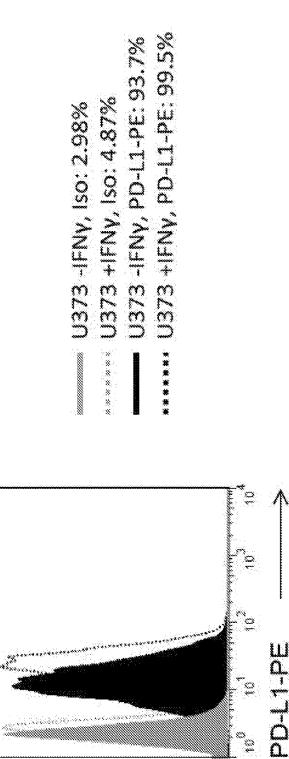
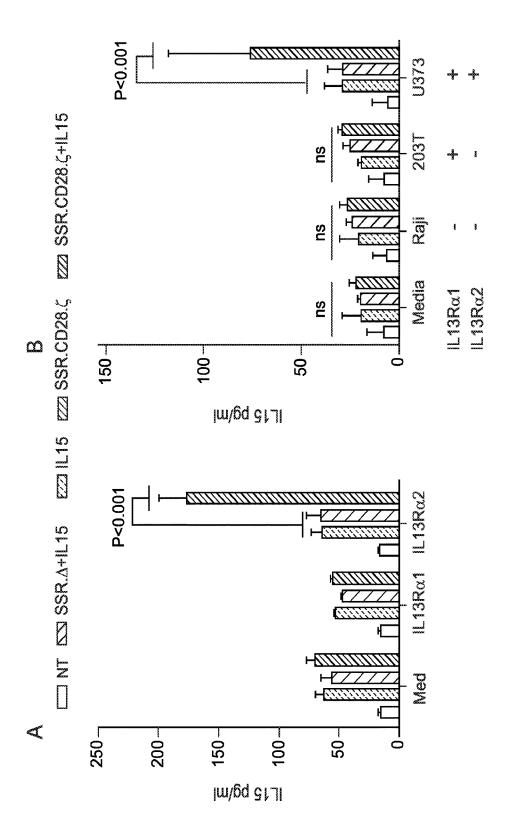




Figure 44



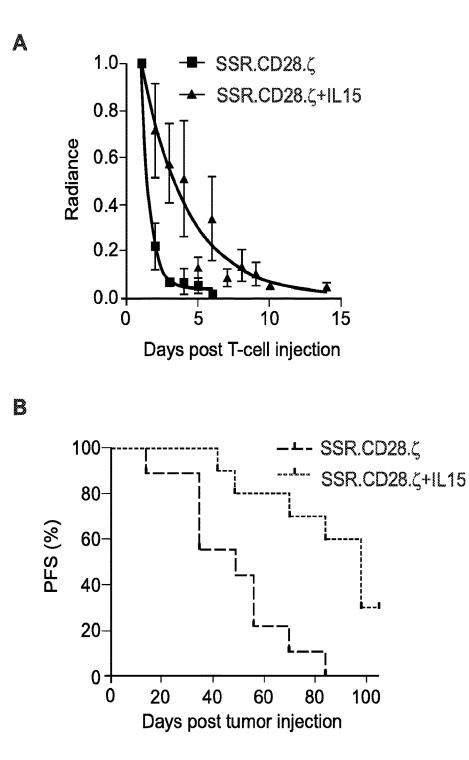


Figure 45

CAR T CELLS RECOGNIZING CANCER-SPECIFIC IL 13RA2

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application claims the priority benefit under 35 U.S.C. § 119(e) of Provisional U.S. Patent Application No. 62/107,980, filed Jan. 26, 2015 and Provisional U.S. Patent Application No. 62/245,771, filed Oct. 23, 2015, the disclosures of which are incorporated herein by reference in their entireties.

INCORPORATION BY REFERENCE OF MATERIAL SUBMITTED ELECTRONICALLY

[0002] This application contains, as a separate part of the disclosure, a Sequence Listing in computer-readable form which is incorporated by reference in its entirety and identified as follows: Filename: 49923A_Seqlisting.txt; 176,301 bytes, created Jan. 22, 2016.

FIELD OF THE DISCLOSURE

[0003] The disclosure relates generally to the fields of cancer biology and to molecular antibody-receptor technology.

BACKGROUND

[0004] Cancer is a major threat to human and non-human animal health, leading to reduced quality of life and, in too many cases, death. The burden placed on national, regional and local healthcare organizations to treat and prevent the various forms of cancer is significant in terms of the resources and manpower required. One of the main weapons vertebrates, including humans, have to combat disease is a functioning immune system. A brief consideration of immunotherapies to treat or prevent cancer might lead one to conclude that the effort held out little hope of success because immune systems guard against foreign, or non-self, materials and cancer cells arise from within, i.e., they are self materials. Continued progress in our understanding of cancer and immunology is modifying that view, however. [0005] Mutant antigens are powerful targets for tumor destruction, e.g., in mice, and tumor-infiltrating lymphocytes targeting these mutations cause durable tumor regression in patients. Nevertheless, non-mutant antigens have been presumed by many scientists to be cancer-specific or "relatively cancer-specific" and safe antigens for vaccine approaches. However, adoptively transferred T cells can be orders of magnitude more effective and destructive than vaccinations. As a result, targeting MAGE-A3, HER-2 or CEA with T cells has caused death or serious toxicity in clinical trials now halted (8-11). As was shown in 2002,

cancer cells with extremely high or very low expression levels of a target antigen differ only in the induction of immune responses, but not at the effector phase (15).

[0006] The high affinity interleukin-13 receptor $\alpha 2$ (IL13R $\alpha 2$) is selectively expressed at a high frequency by glioblastoma multiforme (GBM) as well as several other tumor types. One approach for targeting this tumor-specific receptor utilizes the cognate ligand, IL-13, conjugated to cytotoxic molecules. This approach, however, lacks specificity because the lower affinity receptor for IL-13, IL13R α 1, is widely expressed by normal tissues.

[0007] Most human cancers lack specific antigens that are predictably present and serve as effective targets for eradication by T cells. Every cancer cell type harbors a unique set of mutations causing different tumor-specific antigens. Identifying an effective unique antigen and isolating an appropriate TCR for transduction of autologous T cells for adoptive immunotherapy is still difficult despite the enormous technological progress being made. Adoptive immunotherapy using antibodies or T cells is clinically as well as experimentally the most effective immunotherapy, at least when clinically relevant cancers are considered (22). The remarkable success of adoptive immunotherapy with chimeric antibody receptors (CARs) and bispecific T cell engaging proteins (BiTEs) is, however, largely restricted to those specific for CD19/CD20-eradicating B cell malignancies and normal B cells in patients, i.e., hematopoietic cancers. Thus, there is a need to identify shared, yet tumorspecific, antigens on a wide range of solid tumors, and a concomitant need to develop prophylactics and therapeutics that can diagnose, prevent, treat or ameliorate a symptom of these cancers, along with methods for diagnosing, preventing and treating various cancers.

SUMMARY

[0008] Disclosed herein are T cells expressing a chimeric antigen receptor (i.e., CAR) that specifically recognizes and binds to the $\alpha 2$ Interleukin 13 Receptor (i.e., IL13R $\alpha 2$). The IL13Ra2-specific CARs, generally referred to herein as 47-CARs, when expressed in T cells effectively target and kill IL13Ra2-positive target cells. Also disclosed is evidence establishing that 47-CARs with a short spacer region, or SSR (i.e., 47-CAR.SSR), exhibit greater capacity to induce IL2-production in an antigen-dependent manner. Further disclosed herein is experimental evidence that 47-CAR.SSR T cells have potent anti-tumor activity in vivo. [0009] The disclosure provides (i) the sequences of heavy (SEQ ID NO:7) and light (SEQ ID NO:8) chain variable regions of a monoclonal antibody (i.e., the clone 47 antibody) specifically targeting human tumor-associated antigen, IL13R α 2, and (ii) data demonstrating the functionality of the protein encoded by the heavy and light chain cDNAs in the format of an scFv antibody or fusion to other functional moieties. The sequences of the heavy and light chain constant regions were also determined and were found to be identical to the corresponding sequences in Genbank Acc. No. DQ381544.1. In particular, the CH1 sequence of the clone 47 antibody is set forth in SEQ ID NO:104, CH2 in SEQ ID NO:105 and CH3 in SEQ ID NO:106; the light chain constant region sequence of the clone 47 antibody is set forth in SEQ ID NO:107; and the hinge region of the clone 47 antibody in SEQ ID NO:108. The heavy and light chain can be arranged in different formats, such as singlechain antibody, diabodies, bi- and tri-specific antibodies, fusions with therapeutic proteins and other moieties, human or humanized whole antibodies as well as human or humanized Fab fragments and other functional derivatives. The single-chain antibody or other arrangements of the protein encoded by the heavy and light chains, e.g., a bispecific binding molecule, may be expressed and conjugated to therapeutic carriers (e.g., viruses, cells, nanomaterials) for specific delivery of therapeutic to IL13Ra2-overexpressing tumors or for imaging tumor burden.

[0010] Proteins expressed by tumor cells but not by normal cells are attractive molecules for the selective delivery of cytotoxic molecules. Accordingly, interleukin-13 receptor $\alpha 2$ (IL13R $\alpha 2$), the high affinity receptor for interleukin-13 (IL-13), is a promising candidate. IL13R $\alpha 2$ is expressed at a high frequency in the aggressive and incurable form of primary brain tumor known as glioblastoma multiforme (GBM) (1-3), as well as by other solid tumors (4). In contrast, normal tissues express little to no IL13R $\alpha 2$, with the exception of the testes (6). Notably, IL13R $\alpha 1$, a different receptor with low affinity for IL-13, is expressed ubiquitously by many tissues (7-9), making it a poor candidate for selective targeting of tumor-specific immunotherapeutic applications.

[0011] Several studies have investigated the therapeutic properties of an IL-13 fusion protein conjugated to a recombinant cytotoxin derived from *Pseudomonas* exotoxin A (IL-13PE) that induces apoptosis in IL13R α 2-expressing glioma cells in vitro, in preclinical animal models, and in patients tested in clinical trials (17-22). Such agents, however, lack a high specificity of interaction with IL13R α 2 because they alternatively bind to the ubiquitously expressed IL13R α 1. Therefore, developing highly selective antibody fragments that can be combined with effectors (e.g., T-cells, toxins) for specificity to IL13R α 2-expressing cells is expected to yield therapeutically beneficial results.

[0012] The disclosure captures the tumor specificity of IL13Ra2 by providing protein binding partners specific for IL13Ra2, rather than mimicking IL13 itself, which would result in a molecule exhibiting a capacity to bind to both IL13Ra1 and IL13Ra2. In addition, the disclosure provides a polynucleotide encoding one of these cancer-specific IL13Ra2 binding partners, including polynucleotides comprising codon-optimized coding regions for binding partners specific for an epitope of one of these IL13Ra2 binding partners. Expressly contemplated are fusion proteins or chimeras that comprise an IL13R α 2 binding partner as defined above in operable linkage to a peptide providing a second function, such as a signaling function of the signaling domain of a T cell signaling protein, a peptide modulator of T cell activation or an enzymatic component of a labeling system. Exemplary T cell signaling proteins include 4-1BB (CD137), CD3ζ, and fusion proteins, e.g., CD28-CD3ζ and 4-1BB-CD3ζ. 4-1BB (CD137) and CD28 are co-stimulatory molecules of T cells; CD3 c is a signal-transduction component of the T-cell antigen receptor. In certain embodiments, the IL13Ra2-specific CAR may be expressed in two fragments that are inactive without the addition of an exogenous substance. By way of non-limiting example, the CAR would consist of two molecules: 1) the first molecule would contain the IL13R α 2-specific scFc, a hinge, a transmembrane domain, a costimulatory domain, and a heterodimerizer domain (Exto-TM-HD), and 2) the first molecule would contain a transmembrane domain, a costimulatory domain, a heterodimerizer domain, a CD35 activating domain (Cyto-HD) (Wu et al; Science. 2015 Oct. 16; 350(6258):aab407). Expression of Exto-TM-HD and Cyto-HD in cells would result in an inactive IL13Ra2-CAR unless a small molecule, for example but not limited to, a rapalog A/C Heterodimerizer is added that links Exto-TM-HD and Cyto-HD, allowing for pharmacological control of IL13Ra2-CAR activity. The peptide or protein providing a second function may provide a modulator of T cell activation, such as IL15, IL15R α , of an IL15/IL15R α fusion, or it may encode a label or an enzymatic component of a labeling system useful in monitoring the extent and/or location of binding, in vivo or in vitro. Agent encoding these prophylactically and therapeutically active biomolecules placed in the context of T cells, such as autologous T cells, provide a powerful platform for recruiting adoptively transferred T cells to prevent or treat a variety of cancers in some embodiments of the disclosure. Codon optimization of the coding regions for binding partners specific for epitopes found on cancer cells provides an efficient approach to delivery of the diagnostic, prophylactic, and/or therapeutic proteins disclosed herein. [0013] In one aspect, the disclosure provides an Interleukin 13 Receptor $\alpha 2$ (IL13R $\alpha 2$) binding partner comprising the antibody heavy chain variable fragment (V_H) complementarity determining region 1 (CDR1) of SEQ ID NO:1, the V_H CDR2 of SEQ ID NO: 2, the V_H CDR3 of SEQ ID NO: 3, the light chain (V_L) complementarity determining region 1 (CDR1) of SEQ ID NO: 4, the V_L CDR2 of SEQ ID NO: 5, and the V_L CDR3 of SEQ ID NO: 6, wherein the IL13Ra2 binding partner specifically binds to an epitope of IL13Ra2. In some embodiments, the V_H sequence is set forth as SEQ ID NO: 7 and in some of the same and some different embodiments, the V_L sequence is set forth as SEQ ID NO: 8.

[0014] A related aspect of the disclosure provides a bispecific binding molecule comprising a fragment of the IL13R α 2 binding partner described herein that binds to the IL13R α 2 epitope covalently linked to a peptide providing a second function to form a bispecific binding molecule. In some embodiments, the second function of the peptide is selected from the group consisting of a signaling function of the signaling domain of a T cell signaling protein, a peptide modulator of T cell activation, and an enzymatic component of a labeling system. In some embodiments, the fragment is a single-chain variable fragment (scFv), which may be contained within a bi-specific T-cell engager (BiTE) or a chimeric antigen receptor (CAR). Some embodiments are provided wherein the bispecific binding molecule as described herein is conjugated to a therapeutic carrier.

[0015] Another aspect of the disclosure is drawn to a pharmaceutical composition comprising the IL13R α 2 binding partner as described herein and a pharmaceutically acceptable carrier, adjuvant or diluent.

[0016] A related aspect provides a kit comprising the pharmaceutical composition described herein and a protocol for administration of the composition. Also related is an aspect providing a polynucleotide encoding the IL13R α 2 binding partner as described herein and a vector comprising the polynucleotide as described herein. Yet another aspect is directed to a host cell comprising the polynucleotide described herein.

[0017] Yet another aspect of the disclosure provides a method of preventing, treating or ameliorating a symptom of a cancer disease comprising administering a therapeutically effective amount of the pharmaceutical composition as described herein. In some embodiments, the cancer is a solid tumor, such as a glioblastoma multiforme (GBM). In some embodiments, the cancer is treated by inhibiting the growth rate of the solid tumor. In some embodiments, the symptom ameliorated is pain.

[0018] More particularly, one aspect of the disclosure is drawn to an IL13R α 2-specific chimeric antigen receptor (CAR) comprising: (A) each of the amino acid sequences of: NYLMN (SEQ ID NO: 1); RIDPYDGDIDYNQNFKD (SEQ ID NO: 2); GYGTAYGVDY (SEQ ID NO: 3); RAS-ESVDNYGISFMN (SEQ ID NO: 4); AASRQGSG (SEQ ID

NO: 5); and QQSKEVPWT (SEQ ID NO: 6), (B) a hinge region, (C) a transmembrane domain, and (D) an endodomain comprising a signaling domain a CD3 zeta chain and a signaling domain of CD28. In some embodiments, the endodomain further comprises a signaling domain of one or more of: CD137, CD134, CD27, CD40, ICOS, and Myd88, optionally, wherein the endodomain comprises one or more of the amino acid sequences of SEQ ID NOs: 68, 70, 72, 74, 76, and 78. In some embodiments, the hinge region comprises the amino acid sequence of SEQ ID NO: 35 or SEQ ID NO: 37. In some embodiments, the IL13R α 2-specific CAR comprises a transmembrane domain of CD28. In some embodiments, the IL13Ra2-specific CAR comprises the amino acid sequence of SEQ ID NO: 39. In some embodiments, the CD3 zeta chain signaling domain comprises the amino acid sequence of SEQ ID NO: 41. In some embodiments, the IL13Ra2-specific CAR comprises the amino acid sequence of SEQ ID NO: 47. In some embodiments, the IL13R α 2-specific CAR comprises the amino acid sequence of SEQ ID NO 49 or 51. In some embodiments, the IL13Rα2-specific CAR comprises one or both of the amino acid sequences of SEQ ID NO: 7 and/or SEQ ID NO: 8. In some embodiments, the IL13R α 2-specific CAR of claim 9, wherein the amino acid sequence of SEQ ID NO: 7 is fused to the amino acid sequence of SEQ ID NO: 8 through a linker. In some embodiments, the linker comprises the amino acid sequence of EEGEFSEAR (SEQ ID NO 10). In some embodiments, the IL13R α 2-specific CAR comprises the amino acid sequence of SEQ ID NO: 13. In some embodiments, the IL13Ra2-specific CAR comprises the amino acid sequence of SEQ ID NO: 53 or SEQ ID NO: 55.

[0019] In a related aspect, the disclosure provides a nucleic acid encoding any of the IL13R α 2-specific CARs disclosed or described herein. In some embodiments, the nucleic acid comprises the sequence of SEQ ID NO: 34, 36, 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, or 65.

[0020] In yet another aspect, the disclosure provides a vector comprising a nucleic acid disclosed or described herein. In some embodiments, the vector is a retroviral vector.

[0021] In another aspect, the disclosure provides a host cell comprising a vector disclosed or described herein. In some embodiments, the host cell is a human host cell. In some embodiments, the host cell is a T-lymphocyte. In some embodiments, the host cell is a natural killer cell.

[0022] In a related aspect, the disclosure provides a cell population comprising a host cell disclosed or described herein. In some embodiments, the cell population comprises at least 10^7 host cells.

[0023] Another aspect is drawn to a pharmaceutical composition comprising an IL13R α 2-specific CAR as disclosed or described herein, a nucleic acid as disclosed or described herein, a vector as disclosed or described herein, a host cell as disclosed or described herein, or a cell population as disclosed or described herein, and a pharmaceutically acceptable carrier.

[0024] Another aspect of the disclosure provides a method of treating a cancer in a subject, comprising administering to the subject a cell population as disclosed or described herein, in an amount effective to treat the cancer in the subject. In some embodiments, the cancer is colon cancer. In some embodiments, the host cells of the cell population are cells obtained from the subject. In some embodiments, the curculation contents, the cells of the cell population are cells obtained from the subject.

obtained from the subject are T-lymphocytes. In some embodiments, the cells obtained from the subject are natural killer cells.

[0025] Another aspect of the disclosure provides an IL13Ra2-specific chimeric antigen receptor (CAR) comprising: (A) an ectodomain comprising each of the amino acid sequences of: (i) NYLMN (SEQ ID NO: 1); (ii) RIDPYDGDIDYNQNFKD (SEQ ID NO: 2); (III) GYG-TAYGVDY (SEQ ID NO: 3); (iv) RASESVDNYGISFMN (SEQ ID NO: 4); (v) AASRQGSG (SEQ ID NO: 5); and (vi) QQSKEVPWT (SEQ ID NO: 6); (B) a spacer region; (C) a transmembrane domain; and (D) an endodomain selected from the group consisting of CD3. ζ , CD28. ζ , CD28. OX40. ζ, CD28.41BB.ζ and 41BB.ζ. In some embodiments, the spacer region comprises no more than 100 amino acids, or no more than 50 amino acids, or no more than 25 amino acids, or the spacer region comprises SEQ ID NO:103 (PKSCDKTHTCPPCPAPEL) from the IgG1 hinge region. In some embodiments, the transmembrane domain comprises the transmembrane domain of CD28, such as a transmembrane domain comprising the amino acid sequence of SEQ ID NO:39, or CD8a. In some embodiments, the endodomain further comprises a signaling domain of one or more of: CD137, CD134, CD27, CD40, ICOS, and Myd88. In some embodiments, the endodomain comprises one or more of the amino acid sequences of SEQ ID NOs: 68, 70, 72, 74, 76, and 78. In some embodiments comprising the CD3 zeta chain signaling domain, the IL13Ra2-specific CAR comprises the amino acid sequence of SEQ ID NO: 41. In some embodiments, the IL13Ra2-specific CAR comprises the amino acid sequence of SEQ ID NO: 47. In some embodiments, the IL13Ra2-specific CAR comprises the amino acid sequence of SEQ ID NO 49 or 51. In some embodiments, the IL13Ra2-specific CAR comprises one or both of the amino acid sequences of SEQ ID NO: 7 and/or SEQ ID NO: 8.

[0026] The disclosure also contemplates embodiments wherein the amino acid sequence of SEQ ID NO: 7 is fused to the amino acid sequence of SEQ ID NO: 8 through a linker. In some embodiments, the linker comprises the amino acid sequence of EEGEFSEAR (SEQ ID NO 10). In some of these embodiments, the IL13R α 2-specific CAR comprises the amino acid sequence of SEQ ID NO: 13. In some embodiments, the IL13R α 2-specific CAR comprises the amino acid sequence of SEQ ID NO: 53 or SEQ ID NO: 55.

[0027] Another aspect of the disclosure is drawn to a nucleic acid encoding the IL13R α 2-specific CAR disclosed herein. In some embodiments, the nucleic acid comprises the sequence of SEQ ID NO: 34, 36, 38, 40, 42, 44, 46, 48, 50, 52, 54, 56, and 65.

[0028] Still another aspect of the disclosure is drawn to a vector comprising the nucleic acid disclosed herein. In some embodiments, the vector is a retroviral vector.

[0029] Yet another aspect of the disclosure is a host cell comprising the vector disclosed herein. In some embodiments, the host cell is a human host cell. In some embodiments, the host cell is a T-lymphocyte or a natural killer cell. In some embodiments, the cells obtained from the subject are T cells, and/or other lymphocytes including, but not limited to, NKT cells, $\gamma\delta$ T cells, mucosa associated invariant T cells or MAIT cells, and innate lymphoid cells. In addition, stem and/or progenitor cells may be obtained from

the subject that are subsequently differentiated into the aforementioned immune cells.

[0030] Another aspect of the disclosure is a cell population comprising the host cell disclosed herein. In some embodiments, the cell population comprises at least 10^7 host cells. [0031] In another aspect, the disclosure provides a pharmaceutical composition comprising an IL13R α 2-specific CAR as disclosed herein, a nucleic acid as disclosed herein, a vector as disclosed herein, a host cell as disclosed herein, or a cell population as disclosed herein, and a pharmaceutically acceptable carrier.

[0032] Yet another aspect of the disclosure is a method of treating a cancer in a subject, comprising administering to the subject a cell population as disclosed herein, in an amount effective to treat the cancer in the subject. In some embodiments, the cancer is colon cancer. In some embodiments, the host cells of the cell population are cells obtained from the subject. In some embodiments, the cells, and/or other lymphocytes including, but not limited to, NKT cells, $\gamma\delta$ T cells, mucosa associated invariant T cells or MAIT cells, and innate lymphoid cells. In addition, stem and/or progenitor cells may be obtained from the subject that are subsequently differentiated into the aforementioned immune cells.

[0033] In some embodiments, the immune or stem and/or progenitor cells that are genetically modified to be IL13R α 2-specific by expressing a CAR or BITE molecule may be further genetically modified to enhance their antitumor activity. Non-limiting examples of additional genetic modification include, but are not limited to: i) CARs or BITEs that are specific for other antigens expressed on tumor cells or within the tumor environment, ii) cytokines (e.g., various interleukins such as IL7, IL12, IL15, IL21), iii) chimeric cytokine receptors (e.g., IL7R, IL15R), iv) chemokine receptors (e.g., CCR2b, CXCR2), iv) chimeric activating receptors (e.g., IL4/IL7R, IL4/IL2R, TGFB/ TLR4R), v) silencing negative regulators (e.g., PD1, SHP1), vi) silencing endogenous TCR expression, and vii) inducible suicide genes (e.g., CD20, truncated EGFR, inducible caspase 9).

[0034] Other features and advantages of the disclosure will become apparent from the following detailed description, including the drawing. It should be understood, however, that the detailed description and the specific examples, while indicating preferred embodiments, are provided for illustration only, because various changes and modifications within the spirit and scope of the invention will become apparent to those skilled in the art from the detailed description.

BRIEF DESCRIPTION OF THE DRAWING

[0035] The patent or application file contains at least one drawing executed in color. Copies of this patent or patent application publication with color drawing(s) will be provided by the Office upon request and payment of the necessary fee.

[0036] FIG. 1. Characterization of antigen recognition and screening of hybridoma clones. A, binding of B-D13 mAb to ELISA plates coated with rhIL13R α 2hFc at 0.1 and 1 µg/ml. B, binding of IL13R α 2 mAb to native and denatured (at 95° C. in the presence of (3-mercaptoethanol) rhIL13R α 2hFc in a plate-bound ELISA. A paired t test was used to evaluate the difference between control groups (n=4). *, p<0.01; ***, p<0.001. Error bars represent S.D. These data are represen-

tative of two independent experiments. C, screening of selected hybridoma populations against rhIL13R α 2hFc in a plate-bound ELISA. D, screening of selected hybridoma populations against rhIL13R α 2hFC using a Western blot.

[0037] FIG. **2.** The IL13R α 2 (clone 47) mAb specifically binds to rhlL13R α 2 and IL13R α 2 expressed on the cell surface of CHO cells. A, binding of IL13R α 2 (clone 47, 83807, and B-D13) mAbs to rhlL13R α 2 in a plate-bound ELISA. B, binding of the IL13R α 2 (clone 47) mAb to human IL13R α 2 expressed on the surface of CHO cells. C, cross-reactivity of the IL13R α 2 (clone 47) mAb with hrIL13R α 1. D, cross-reactivity of IL13R α 2 (clones 47, 83807, and B-D13) mAbs with mouse rIL13R α 2. Error bars represent S.D.

[0038] FIG. 3. Binding of IL13R α 2 mAb to glioma cells. A, flow charts of IL13Ra2 (clones 47, 83807, and B-D13) mAbs binding to the surface of glioma cells, normal human primary astrocytes, and HEK cells transfected with IL13Ra2. B, data of the median fluorescence intensity of binding between the IL13R α 2 (clones 47, 83807, and B-D13) mAbs to various cell lines analyzed by flow cytometry. Numbers above the bars represent the difference in the binding of clone 47 when compared with clone B-D13 for each cell line. The color key is the same for A and B. C, mRNA expression for IL13Ra2 in glioma cells as well as normal human primary astrocytes. D, panels a-c, flow cytometry demonstrating the specific binding of the IL13R α 2 (clone 47) mAb to GFP-tagged U251 glioma cells from an intracranial xenograft (xeno). The curve with a clear area under the curve in sub-panel b depicts the binding of mAb IL13R α 2 (clone 47) to GFP negative cells; the curve with a clear area under the curve in sub-panel c depicts the binding of mAb IL13R α 2 (clone 47) to GFP positive cells. Curves in sub-panels b and c with gray areas under the curves show the results when exposing control IgG to GFP-negative (sub-panel b) or GFP-positive (sub-panel c) cells. neg, negative. A, area; SSC-A, side scatter area; APC-A, allophycocyanin area.

[0039] FIG. 4. The affinity between the IL13R α 2 (clone 47) mAb and rhIL13R α 2. The kinetics of interaction of IL13R α 2 (clone 47) mAb (A) and the commercially available mAb clones 83807 (B) and B-D13 (C) with rhIL13Ra2 as visualized by SPR in a Biacore 3000 are shown. The rhIL13Ra2 was injected at concentrations ranging from 1 to 100 nM (1 nM, 2.5 nM, 5 nM, 7.5 nM, 10 nM, 15 nM, 20 nM, 25 nM concentrations shown, lower to upper curves) at a constant flow rate of 20 µl/minute over immobilized antibodies and over a control dextran surface (these values were subtracted from the signal). The association and dissociation phases were monitored for 300 s by following the change in SPR signal (colored curves) given in RU. Black curves represent the fit of the data to a one-site binding model. For derived kinetic parameters, see Table 1. Lower panels show residuals from a one-site binding model, indicating an excellent fit.

[0040] FIG. **5**. The IL13R α 2 (clone 47) mAb competes with rhIL-13 for the binding site of IL13R α 2. A, using a competitive binding plate assay, the IL13R α 2 (clone 47) mAb but not control mIgG or antibody clones 83807, B-D13, and YY-23Z significantly abolished the binding of rhIL-13 to the rhIL13R α 2Fc chimera absorbed to plastic. One-way analysis of variance followed by Dunnett's post hoc test was performed. Data from a single representative experiment are shown. B, recombinant human IL-13 com-

petes with the IL13R α 2 (clone 47) mAb for the binding site of WT IL13R α 2 but not with the 4-amino acid (4aa) mutant IL13R α 2 expressed on the surface of HEK cells. C, the IL13R α 2 (clone 47) mAb competes with rhIL-13 for the binding site of the WT and 4-amino acid mutant form of IL13R α 2. A paired t test was performed. Data represent the summary of three independent experiments shown in B and C. *, p<0.05; **, p<0.01; ***, p<0.001. Error bars represent S.D.

[0041] FIG. 6. The contribution of Tyr207, Asp271, Tyr315, and Asp318 residues of IL13R α 2 to the binding of the IL13R α 2 (clone 47) mAb. A, variants of cDNA encoding individual mutations to Ala or a combinatorial 4-amino acid mutant (4aa mut) of IL13Ra2 was generated. HEK cells were transfected with a control vector or a vector encoding the IL13R α 2 variants. After 48 hours, binding of the IL13Ra2 (clone 47) mAb to the surface of transfected cells was analyzed by flow cytometry. Anti-IL13Ra2 antibody clones 83807 and B-D13 were used as reference antibodies in this assay. Binding of antibodies was determined as the percentage of positive cells. The ratio of bound clones was determined for each IL13R α 2 mutant and compared with that of the wild-type receptor. One-way analysis of variance followed by Dunnett's post hoc test was performed. Data represent a summary of four independent experiments. Error bars represent S.D. B, representative graphs of flow cytometry data demonstrating the binding of clone 47 or rhIL-13 to the WT and 4-amino acid-mutated variant of the IL13Ra2 receptor expressed on the surface of HEK cells. Filled curves: negative control, staining with isotype control IgG+ secondary antibody; Open curves: staining with the anti-IL13Rα2 (clone 47) monoclonal antibody+secondary antibody. A, area; APC-A, allophycocyanin area; FITC-A, fluorescein isothiocyanate area.

[0042] FIG. 7. Effect of N-linked glycosylation on the binding of IL13R α 2 to recombinant IL13R α 2. A, binding of IL13Rα2 to control and Pngase F-treated rhIL13Rα2. Plates were coated with hrIL13R α 2 at 1 µg/m1 and treated with native buffer or with 1 milliunit/well Pngase F in native buffer for 3 hours at 37° C. An ELISA for binding of the IL13R α 2 (clone 47) mAb in comparison with antibody clones B-D13, 83807, and YY-23Z and rhIL-13 was performed, and the data of one representative experiment from three independent experiments are shown. A paired t test was used to evaluate the difference between control and Pngase F-treated groups (n=4). *, p<0.5; **, p<0.01; ***, p<0.001. B, a Western blot shows the lower molecular weight of Pngase F-treated rhIL13Ra2 due to removal of N-linked glycosylation adducts from the molecule. C, flow cytometry shows the binding of IL13Rα2 mAbs to IL13Rα2-expressing U251 and HEK293 cells treated with 1 milliunit of Pngase F for 1 hour at 37° C. The data are representative of three independent experiments. A paired t test was used to evaluate the difference between control and Pngase F-treated groups. *, p<0.5. MFI, mean fluorescence intensity. Error bars represent S.D.

[0043] FIG. 8. The IL13R α 2 (clone 47) mAb recognizes IL13R α 2 in GBM tissues and in a human glioma xenograft. Immunohistochemistry on frozen tissue sections from three human GBM samples and a U251 xenograft was performed with the IL13R α 2 (clone 47) mAb or mIgG at a concentration of 3 µg/ml. Staining of GBM tissues demonstrates positive staining of the majority of cells in sample 1, positive reactivity in only a fraction of the cells in sample 2, and

negative staining in sample 3. Staining in all three samples was performed in the same experiment. Positive staining was also detected in U251 xenograft tissue. Arrows point to individual positive cells. Scale bars=100 µm.

[0044] FIG. 9. The IL13R α 2 (clone 47) mAb improves the survival of mice in an orthotopic human glioma xenograft model. A, the survival of animals injected with U251 glioma cells (2.5×10⁴) alone or in combination with either control IgG or the IL13R α 2 (clone 47) mAb. B, a representative photomicrograph of 10-µm-thick tissue sections stained with H&E from mice injected with U251 cells alone (panels a and b) or in combination with mIgG (panels c and d) or mAbIL13R α 2 (clone 47) (panels e and f). Arrows point to the tumor and invading cells. Scale bars (panels a, c, and e)=100 µm. Scale bars (panels b, d, and f)=100 µm.

[0045] FIG. **10**. A competitive binding assay for the IL13R α 2 (clone 47) mAb to the surface of N10 glioma cells. A. The IL13R α 2 (clone 47) mAb was pre-incubated with 10× excess rhIL13R α 2 for 30 minutes on ice. N10 cells were subsequently incubated with isotype control mIgG or IL13R α 2 (clone 47) mAb alone or in the presence of rhIL13R α 2 and bound antibodies were analyzed by flow cytometry. B. N10 glioma cells were pre-incubated either with 10× excess rhIL13 (left panel) or with 10× excess of IL13R α 2 (clone 47) mAb for 30 minutes on ice (right panel). N10 cells were subsequently incubated with isotype control mIgG, IL13R α 2 (clone 47) mAb for rhIL13. Bound antibodies or rhIL13 were detected with secondary antibodies and analyzed by flow cytometry. Data are presented as % of positive cells.

[0046] FIG. **11**. The effects of IL13R α 2 (clone 47) mAb on the survival of mice with an established human U251 glioma. Mice were intracranially-injected with 2.5×10⁴ U251 glioma cells and treated three days later with a single injection of PBS (n=7) or 10 µg IL13R α 2 (clone 47 or B-D13) mAb (n=7). The analysis of the animal's survival was performed using the Log-rank test. Median survival was determined to be 27 days in the PBS group, versus 23 and 35 days in the groups treated with B-D13 and 47 IL13R α 2 mAb, respectively (p>0.05).

[0047] FIG. 12. Binding of IL13R α 2 clone 47 phages with IL13R α 2hFc in plate ELISA. These data demonstrate that phages presenting scFv IL13R α 2 (clone 47) are positively selected against IL13R α 2Fc chimeric protein after 3 rounds of biopanning.

[0048] FIG. **13**. Specificity of binding scFv IL13R α 2 clone 47 with IL13R α 2hFc—competitive assay. These data show that binding of the scFvIL13R α 2 (clone 47) presented on the phage surface to recombinant IL13R α 2 is completely abolished by parental monoclonal antibody (clone 47), but not other antibodies against IL13R α 2. It indicates that scFvIL13R α 2 (clone 47) and parental monoclonal antibody (clone 47) share the epitope (i.e., recognition site) on the IL13R α 2 molecule. Each data point is an average of 3 independent replicates in all figures. Data presented as mean±SEM. *** p<0.001.

[0049] FIG. **14**. Binding of soluble scFv IL13R α 2 (clone 47) with IL13R α 2hFc chimera. These data show that soluble scFvIL13R α 2 (clone47) generated in a prokaryotic expression system (*E. coli*) binds specifically to IL13R α 2Fc recombinant protein. Parental antibody, mAb IL13R α 2 (clone 47), and control mouse IgG served as positive and negative controls, respectively

[0050] FIG. **15**. The effect of mesenchymal stem cells secreting scFvIL13R α 2-sTRAIL fusion protein on the U87-IL13R α 2 glioma cell line. These data show that mesenchymal stem cells modified to secrete a genetic fusion of scFvIL13R α 2(clone 47) with TRAIL protein exhibit a therapeutic effect in the IL13R α 2-expressing U87 glioma cell line. The results establish the efficacy of conjugating the scFV to a TRAIL cytokine. The amount of cancer cell killing is equivalent to the use of TRAIL alone without the scFV, but it is expected that the scFV-TRAIL would be less harmful to non-cancer tissues, given the specificity conferred by the scFV targeting IL13R α 2.

[0051] FIG. **16**. Schematic maps of retroviral vector encoding IL13R α 2-specific scFv CARs. The CAR consists of the immunoglobulin heavy-chain leader peptide, the IL13R α 2-specific scFv clone 47 (M47), a short hinge (SH) or long hinge (LH), a transmembrane domain (TM) derived from CD28, and a CD28. ζ endodomain. LTR: long terminal repeat (retroviral backbone). Domains are identified as block structures. Maps are not to scale.

[0052] FIG. 17. IL13R α 2-scFv CAR T cell agent: Expression of α CD3. ζ relative to α GAPDH of CAR agent in T cells. SH: short hinge. LH: long hinge.

[0053] FIG. **18**. IL13R α 2-scFv CARs are expressed on the surface of T cells. IL13R α 2-cAR T cells were generated by retroviral transduction and CAR expression was determined by FACS analysis. Short hinge CARs were detected with an antibody specific for murine scFV. Long hinge CARs were detected with an antibody specific for the long hinge. Isotype antibody control: open curve; Specific Antibody: filled curve.

[0054] FIG. 19. Functional characterization of IL13R α 2-CAR T cells-Cytotoxicity. Standard ⁵¹Chromium cytotoxicity assays were performed with Raji (IL13Ra1-/ IL13Rα2-), 293T (IL13Rα1+/IL13Rα2-), 293T genetically modified to express IL13Ra2 cells (293T-IL13Ra2; IL13R α 1+/IL13R α 2+), or U373 (IL13R α 1+/IL13R α 2+) cells as targets. As effectors nontransduced (NT) T cells, IL13Ra2-CAR.SH.CD28. CT cells, IL13Ra2-CAR.LH. CD28. ζ T cells, IL13R α 2-CAR.SH. Δ T cells, or IL13R α 2-CAR.LH. Δ T cells were used. Only IL13R α 2-CAR.SH. killed with IL13R α 2+ target cells (U373 and 293T-IL13Ra2; n=4). T cells expressing nonfunctional CARs (IL13R α 2-CAR.SH. Δ and IL13R α 2-CAR.LH. Δ) had not cytolytic activity, demonstrating that the killing activity depends on the expression of a functional IL13R α 2-CAR. NT T cells killed none of the targets, further confirming specificity.

[0055] FIG. 20. Functional characterization of IL13R α 2-CAR T cells—IFN γ and IL2 Cytokine secretions. A. NT T cells, IL13R α 2-CAR.SH.CD28. ζ T cells, IL13R α 2-CAR.LH. Δ T cells, IL13R α 2-CAR.SH. Δ T cells, or IL13R α 2-CAR.LH. Δ T cells were co-cultured with U373 cells for 24 to 48 hours (n=4). Only IL13R α 2-CAR.SH. CD28. ζ T cells and IL13R α 2-CAR.LH.CD28. ζ T cells secreted IFN γ demonstrating target cell recognition in contrast to IL13R α 2-CAR.SH. Δ T cells, IL13R α 2-CAR.LH. Δ T cells or NT T cells. B. NT T cells, IL13R α 2-CAR.SH. CD28. ζ T cells, IL13R α 2-CAR.SH. Δ T cells, IL13R α 2-CAR.SH.CD28. ζ T cells hours (n=4). Only IL13R α 2-CAR.SH.CD28. ζ T cells secreted IL2, demonstrating that IL13R α 2-CAR.SH.CD28. ζ induces

superior T cell activation in comparison to IL13R α 2-CAR. LH.CD28. ξ . IL13R α 2-CAR.SH. Δ T cells, IL13R α 2-CAR. LH. Δ T cells or NT T cells also did not induce IL2 production.

[0056] FIG. **21**. IL13R α 2-SH CARs have anti-glioma activity in vivo. Severe combined immunodeficient (SCID) mice were injected with 1×10^5 firefly luciferase expressing U373 cells intracranially. On day 7 mice were treated either with 1×10^6 IL13R α 2-CAR.SH.CD28. ζ T cells, IL13R α 2-CAR.LH.CD28. ζ T cells, IL13R α 2-CAR.LH. Δ T cells (5 mice per group). Tumor growth was monitored by bioluminescence imaging. Only IL13R α 2-CAR.SH.CD28. ζ T cells had significant anti-glioma effects with 4/5 mice having a complete response.

[0057] FIG. **22**. Properties of m47 CAR T cell agent. The m47-CAR T cells recognize IL13R α 2⁺, but not IL13R α 1⁺ targets. The data show that the short hinge CD28z-CAR (SH2) T cells perform better in terms of effector function than CD28z-CAR (SH3), CD28z-CAR (LH2), CD28z-CAR (LH3), CD28z-CAR (SH3 Δ).

[0058] FIG. 23. Functional comparison of m47 CAR T cell agents. Open curve: secondary antibody; Filled curve: $IL13R\alpha 2Fc+secondary$ antibody.

[0059] FIG. **24**. The m47 CAR T cell agent is highly expressed after transduction. Open curve: secondary antibody; Filled curve: IL13R α 2Fc+secondary antibody.

[0060] FIG. 25. The m47 CAR T cell produce interferon γ and interleukin 2, but only after IL13R α 2 stimulation.

[0061] FIG. **26**. IL13R α 2- and IL13R α 1-positive cell lines are made by genetic modification of HEK 293T cells. Filled curve: isotype antibody control; Open curve: specific antibody.

[0062] FIG. **27**. The m47 CAR T cells kill only IL13R α 2⁺ cell lines. The in vitro experiments provide data establishing that m47 CAR T cells present a recombinant CAR protein on the cell surface that does not recognize IL4R, IL13R α 1 or any receptor other than its specific recognition of IL13R α 2. The specificity of the recognition extends to a specificity for only those cell lines expressing IL13R α 2.

[0063] FIG. **28**. In vivo data comparing effect of m47 CAR T cell agent, untreated and NT-treated glioblastoma multiforme xenografts in nude mice. The U373 glioblastoma multiforme xenograft mouse model was used. At day 0, 1×10^5 GFP-fluc U373 cells were administered per mouse. On day 7, 2×10^6 m47 CAR T cells or NT cells were administered. Untreated samples did not receive treatment on day 7. No exogenous interleukin 2 was administered and results of the survival analysis were recorded by serial bioluminescence imaging. n=3.

[0064] FIG. **29**. The m47 CAR T cell agent prolonged the survival of nude mice with glioblastoma multiforme.

[0065] FIG. **30**. Characterization of IL13R α 2-CAR T cells. (A, B) Co-culture assay with recombinant protein demonstrated interferon γ and interleukin 2 production in an IL14R α 2-dependent fashion; (C) Cytolytic activity in standard chromium release assay.

[0066] FIG. **31.** Generation of 47 CAR T cells. (A) Scheme of M47 CARs. All CARs contained an N-terminal leader sequence, a codon-optimized synthetic gene encoding M47 in scFv format, a spacer region, a CD28 transmembrane domain, and signaling domains derived from CD28 and CD3- ζ . The spacer region was either the IgG1 hinge (16 amino acids; short spacer region; M47-CAR.SSR.CD28. ζ) or the IgG1-CH2CH3 domain. LSR. Δ and SSR. Δ M47-

CARs without signaling domains were constructed and served as controls. (C,B) CAR expression was confirmed using FACS analysis. Representative plots (B) and summary data (C) are shown (mean 74.1%-93.3%, n=5-6 per CAR construct). Open curve: secondary antibody; Filled curve: IL13R α 2Fc+secondary antibody. (D) Expression of full-length 47-CAR.SSR.CD28. ζ and 47-CAR.LSR.CD28. ζ by Western blot analysis using a CD3- ζ antibody.

[0067] FIG. 32. Phenotypic analysis of 47-CAR T cell lines. CAR T cells were analyzed for CD4 and CD8 surface expression using CD4-PacBlue and CD8-PerCP antibodies (BD Biosciences). The four CAR T cell lines analyzed for surface expression of CD4 and CD8 were SSR. Δ , SSR. CD28.ζ, LSR.Δ, and LSR.CD28.ζ. The histogram provides the results of the analysis, with light gray bars indicating CD4 expression and black bars indicating CD8 expression. [0068] FIG. 33. 47-CAR T cells release cytokines after stimulation with recombinant IL13Ra2 protein or IL13Ra2positive cells. 47-CAR or non-transduced (NT) T cells were stimulated with recombinant IL13Ra1, IL13Ra2, or IL4Ra proteins. After 24 hours, IFNy (A) was measured by ELISA (n=4). T cells expressing 47-CAR constructs, but not controls, expressed significant levels of IFNy (p<0.001) when stimulated with recombinant IL13Ra2 protein in comparison to IL13Ra1 and IL4Ra stimulated T cells. 47-CAR T cells were co-cultured with Raji, U373 cells, 293T-GFP, and 293T-GFP/IL13Rα2 at a 1:2 E:T ratio. NT and CAR.Δ, T cells served as controls. (B,C) After 24 hours, cytokines (IFNy, IL2) were measured by ELISA (n=3). (B) U373 and 293T-GFP-IL13Ra2 (IFN γ); SSR. Δ vs SSR.CD28. ζ : p<0. 001; LSR.Δ vs LSR.CD28.ζ: p<0.05. (C); U373 and 293T-GFP-IL13Rα2 (IL2); SSR.Δ vs SSR.CD28.ζ: p<0.01; LSR. Δ vs LSR. CD28. ζ: NS. (D) 4-hour cytotoxicity assay at an E:T ratio of 10:1 (n=4).

[0069] FIG. 34. LSR.CD28. ζ T cells show a self-activation phenotype during ex vivo expansion. T cells were analyzed for phosphor-CD3- ζ expression using CD247 (pY142)-AF647 antibody (BD Biosciences).

[0070] FIG. **35**. Cell surface expression of IL13R α 1 and IL13R α 2. Cell lines were analyzed for IL13R α 1 and IL13R α 2 are expression using primary goat anti-IL13R α 1 and anti-IL13R α 2 antibodies (AF152 and AF146, respectively; R&D) followed by secondary rabbit anti-goat IgG Alexa647 antibody (Life Technologies). Filled curve: isotype antibody control; Open curve: specific antibody.

[0071] FIG. **36**. Generation of SSR 47-CARs with CD28. OX40. ζ , CD28.41BB. ζ or 41BB. ζ endodomains. (A) Scheme of SSR 47-CARs. (B, C) CAR expression was confirmed using FACS analysis. Representative plots (B) and summary data (C) are shown. 47-CAR.SSR.CD28. OX40. ζ and 47-CAR.SSR.CD28.41BB. ζ : mean: 74.6%-77. 5% (n=4); 47-CAR.SSR.CD28.41BB. ζ : mean: 4.9% (n=3). Open curve: secondary antibody; Filled curve: IL13R α 2Fc+ secondary antibody. (D) Expression of 47-CAR.SSR.41BB. ζ , M47-CAR.SSR.OX40.CD28. ζ and M47-CAR.SSR. 41BB.CD28. ζ by Western blot analysis.

[0072] FIG. **37**. Comparison of 47-CAR.SSR.CD28. ζ , 47-CAR.SSR.41BB. ζ , and 47-CAR.SSR.CD28.OX40. ζ T cells. (A) 47-CAR T cells were co-cultured with U373 cells at a 1:2 E:T ratio. NT and CAR. Δ T cells served as controls. After 24 hours, IFN γ and IL2 were measured by ELISA (n=3); SSR. Δ vs SSR.CD28. ζ (U373; IFN γ): p<0.001; SSR. Δ vs SSR.41BB. ζ (U373; IFN γ): p<0.001; SSR. Δ vs SSR.CD28.OX40. ζ for (U373; IFN γ): p<0.001; SSR. Δ vs

SSR.CD28.ζ (U373; IL2): p<0.001; SSR.Δ vs SSR.41BB.ζ (U373; IL2): p<0.001; SSR.Δ vs SSR.CD28.OX40.ζ (U373; IL2): p<0.01. (B) 4-hour cytotoxicity assay at an E:T ratio of 10:1 (n=4).

[0073] FIG. **38**. Treatment of glioma xenograft with T cells expressing 47-CARs results in tumor regression and improved overall survival. U373 glioma bearing mice were treated on day 7 with SSR.CD28. ζ (n=9), SSR.41BB. ζ (n=9) or SSR.OX40.CD28. ζ (n=9) T cells. SSR. Δ CAR T cells (n=7) served as controls. (A) Representative images for each group and (B) quantitative bioluminescence (radiance=photons/sec/cm²/sr) imaging data for all mice are shown (dotted lines: individual mice; solid lines: median). (C) Kaplan-Meier survival analysis (SSR. Δ vs SSR.CD28. ζ ; p=0.0002; SSR. Δ vs SSR.41BB. ζ : p=0.0039; SSR. Δ vs SSR.OX40.CD28. ζ : p=0.0092; SSR.CD28. ζ vs SSR.41BB. ζ ; p=0.4723; SSR.CD28. ζ vs SSR.OX40.CD28. ζ ; p=0.3582; SSR.41BB. ζ vs SSR.OX40.CD28. ζ ; p=0.3582; SSR.41BB. ζ vs SSR.OX40.CD28. ζ ; p=0.8374).

[0074] FIG. **39.** Analysis of U373 cells isolated from recurrent tumors. U373 cells were isolated from recurrent tumors of mice that were treated with 47-CAR T cells. After short-term culture (2 to 7 days), FACS analysis and cytotoxicity assays were performed. (A) FACS analysis for IL13R α 2. (B) 47-CAR T cells killed U373 tumor cells isolated from recurrent tumors in contrast to Raji cells in a standard four-hour cytotoxicity assay for Cr release from labeled cells. Open curve: isotype antibody control; Filled curve: specific antibody.

[0075] FIG. **40**. Limited persistence of 47-CAR T cell in vivo. 47.SSR.CD28. ζ -CAR T cells were transduced to express eGFP.ffLuc. (A) FACS analysis confirmed the expression of the CAR and eGFP.ffLuc transgenes. (B, C) 1×10^5 unmodified U373 cells were injected intracranially into mice. On day 7, mice received 2×10^6 47.SSR.CD28. ζ eGFP.ffLuc CAR T cells intracranially using the same tumor coordinates. Bioluminescence imaging was used to monitor T cell persistence.

[0076] FIG. **41**. Generation and characterization of LSR-CD28.41BB. ζ CAR T cells. (A) Scheme of LSR.CD28. 41BB. ζ CAR construct. (B) CAR expression was confirmed using FACS analysis. Representative plot. Open curve: secondary antibody; Filled curve: IL13R α 2Fc+secondary antibody. (C) LSR.CD28.41BB. ζ CAR T cells were co-cultured with U373 cells at a 1:2 E:T ratio. NT T cells served as controls. After 24 hours, IFN γ or IL2 was measured by ELISA (n=3).

[0077] FIG. **42**. Generation of SSR. α .CD28.41BB. ζ CAR T cells. (A) Scheme of SSR. α .CD28.41BB. ζ CAR construct. (B) CAR expression was tested using FACS analysis (representative plot shown).

[0078] FIG. **43**. FACS analysis of PD-L1 expression on U373 cell surface with and without IFN γ stimulation. U373 cells were cultured with or without IFN γ (100 units/ml). After 24 hours, U373 cells were analyzed for PD-L1 expression using a CD271 PE antibody (BD Biosciences).

[0079] FIG. **44**. Transgenic expression of IL15 in SSR. CD28. ζ T cells results in enhanced antigen-dependent IL15 secretion. T cells were stimulated with (A) recombinant proteins or (B) cell lines.

[0080] FIG. **45**. Transgenic expression of IL15 results in (A) enhanced in vivo persistence of SSR.CD28. ζ T cells resulting in improved (B) progression-free survival (PFS).

DETAILED DESCRIPTION

[0081] The disclosure provides binding agents, or partners, that specifically recognize interleukin 13 receptor $\alpha 2$ (IL13R α 2) for use in diagnosing, preventing, treating or ameliorating a symptom of any of a wide range of cancers characterized by cells presenting IL13R α 2. More particularly, the disclosure provides (i) the sequences of the six complementarity determining regions of a monoclonal antibody (m47) that specifically targets human tumor-associated antigen, i.e., interleukin 13 receptor $\alpha 2$ (IL13R $\alpha 2$), and (ii) data demonstrating the functionality of the protein encoded by the heavy and light chain cDNAs in the format of an scFv antibody or conjugate (e.g., fusion) to other functional moieties. The six complementarity determining regions of the m47 monoclonal antibody confer binding specificity for IL13R α 2, consistent with the understanding in the immunological arts. In some embodiments, the scFv comprises the complete heavy and light chain variable regions of antibody m47, or the complete heavy and light chains of antibody m47. In some embodiments, the heavy and light chain fragments comprise, e.g., the m47 CDRs, or the m47 variable regions, and these domains can be arranged in different formats, such as a single-chain variable fragment of an antibody, i.e., a scFv, a diabody, a bi-specific antibody fragment, a tri-specific antibody fragment, a fusion protein with any of a wide variety of therapeutic proteins and/or other moieties, a humanized antibody fragment, a Fab fragment, a Fab' fragment, a F(ab)2' fragment and any other functional format for a bi-functional peptide providing a targeting function and an effector function. Moreover, the single-chain antibody or other arrangements of the protein encoded by the heavy and light chains could be expressed and conjugated to therapeutic carriers (e.g., viruses, cells, nanomaterials) for specific delivery of a therapeutic to an IL13R α 2-expressing tumor. The materials according to the disclosure are also useful in imaging tumor burden.

[0082] The technology addresses the most serious obstacle to progress in immunotherapy, i.e., the virtual absence of defined, tumor-specific antigens that can be predictably found on at least a larger subgroup of human cancers and that can serve as effective targets for cancer eradication. Finding such antigens would move the field beyond the methods for treating CD19/CD20-expressing B cell malignancies.

[0083] The terms used throughout this disclosure are given their ordinary and accustomed meanings in the art, unless a different meaning is made clear from the text when considered in the context of the disclosure as a whole.

[0084] The disclosure describes the development and characterization of a monoclonal antibody (mAb) fragment specific to IL13R α 2 for the therapeutic purpose of targeting IL13R α 2-expressing tumors. The high affinity IL13R α 2 is selectively expressed at a high frequency by glioblastoma multiforme (GBM) as well as several other tumor types. One approach for targeting this tumor-specific receptor utilizes the cognate ligand, IL-13, conjugated to cytotoxic molecules. This approach, however, lacks specificity because the lower affinity receptor for IL-13, IL13Ra1, is widely expressed by normal tissues. A monoclonal antibody (mAb) specific to IL13Ra2 was expected to overcome the lack of specificity afflicting methodologies that recognized both IL13 receptors, i.e., IL13R α 1 as well as IL13R α 2. Such a mAb would be therapeutically useful in targeting and treating IL13Ra2-expressing cancers, including tumors.

[0085] As disclosed herein, hybridoma cell lines were generated and compared for binding affinities to recombinant human IL13Ra2 (rhIL13Ra2). Clone 47 demonstrated binding to the native conformation of IL13R α 2 and was therefore chosen for further studies. Clone 47 bound specifically and with high affinity (KD= 1.39×10^{-9} M) to rhIL13R α 2 but not to rhIL13R α 1 or murine IL13R α 2. Furthermore, clone 47 specifically recognized wild-type IL13R α 2 expressed on the surface of CHO and HEK cells as well as several glioma cell lines. Competitive binding assays revealed that clone 47 also significantly inhibited the interaction between human soluble IL-13 and IL13Ra2 receptor. Moreover, N-linked glycosylation of IL13R α 2 contributes in part to the interaction of the antibody to IL13R α 2. In vivo, the IL13R α 2 mAb improved the survival of nude mice intracranially implanted with a human U251 glioma xenograft.

[0086] The IL13R α 2-specific, scFv-based CAR, 47-CAR, constructed as disclosed herein, provided the material used in exploring the influence of long and short spacer regions, as well as endodomains, on its function. While 47-CAR. SSR.CD28. ζ (i.e., the 47-CAR binding region provided as an scFv joined to a short spacer region as defined herein, in turn joined to an unmodified or chimeric endodomain or T cell cytoplasmic domain) and 47-CAR.LSR.CD28.ζ (similar construct substituting a long spacer region (LSR)) recognized target cells as judged by IFNy production, only 47-CAR.SSR.CD28.ζ induced IL2 production, indicating better T-cell activation. An additional LSR 47-CAR containing a CD28.41BB.5 endodomain (FIG. 41) was shown to lack the ability to induce IL2 expression. These observations are consistent with knowledge that scFvs that bind to an epitope in close proximity to the cancer cell membrane, requiring long spacer regions for optimal CAR function, in contrast to scFvs that bind to epitopes distal to the cell membrane. The data disclosed herein indicates that the IL13Ra2 epitope recognized by 47-CARs is located distal to the cell membrane.

[0087] In greater particularity, four SSR 47-CARs were constructed, each with a different endodomain, i.e., CD28.ζ, 41BB.ζ CD28.OX40.ζ, and CD28.41BB.ζ. While all four CARs were expressed, as judged by Western blot analysis, no significant cell-surface expression was observed for 47-CAR.SSR.CD28.41BB.ζ. We explored if changing the transmembrane domain from CD28 to CD8a in 47-CAR. SSR.CD28.41BB.ζ would result in better cell-surface expression, but no increase in expression was observed. Because 47-CARs.LSR.CD28.41BB.ζ are expressed on the cell surface (FIG. **42**), the result indicates that the interplay between spacer region and endodomain influences CAR cell-surface expression.

[0088] 47-CAR.SSR.CD28.ζ, 47-CAR.SSR.41BB.ζ, and 47-CAR.SSR.CD28.OX40.ζ T cells had potent antitumor effect in vivo, resulting in a significant survival advantage. While mice treated with 47-CAR.SSR.CD28.ζ T cells had the longest median survival in comparison to 47-CAR.SSR. 41BB.ζ or 47-CAR.SSR.CD28.OX40.ζ T-cell treated mice, this difference did not reach significance. The experimental results also showed that addition of a second costimulatory endodomain did not improve antitumor activity in vivo. Limited T-cell persistence in vivo was identified as the principal limitation on therapy. This limitation may be overcome by the transgenic expression of cytokines³⁶ or by blocking inhibitory molecules that are secreted or present on the surface of gliomal cells. For example, gliomas such as U373 express PD-L1, which is upregulated in the presence of IFN γ (FIG. 43), and could be targeted in future studies. [0089] The experimental results disclosed herein establish that T cells redirected to IL13R α 2 with 47-CARs have potent anti-tumor activity against glioma cells in vitro, and induce the regression of established GBM xenografts in vivo. 47-CARs are expected to be of value in the treatment of not only IL13R α 2-positive GBMs but also other malignancies in which IL13R α 2 is expressed.

[0090] The experimental results disclosed herein establish that T cells redirected to IL13R α 2 with 47-CARs and that also express IL15 have enhanced anti-tumor activity in the GBM xenografts in vivo.

[0091] The disclosure is based, at least in part, on the discovery that IL13Ra2 is found preferentially on cancer cells such as tumor cells. This receptor functions as a cancer-, or tumor-, specific antigen that has been used to elicit the high-affinity monoclonal antibody m47, along with antigen binding fragments of that antibody. The VL and VH variable regions of the m47 antibody have been engineered into a single chain (sc) variable fragment (scFv) to generate conjugates, such as chimeric antigen receptors (i.e., CARs), for introduction into T cells for adoptive transfer. Thus, CAR-transduced T cells are expected to target a tumorspecific IL13Ra2 epitope, leading to eradication of cancer cells presenting the receptor. It is believed that CARtransduced T cells recognizing IL13Ra2 will destroy large solid tumors. CAR-transduced T cells, however, target cancer cells only directly and antigen-negative cancer cells may escape. It is expected that CAR-transduced T cells also will be effective in eliminating antigen-negative cancer cells via the bystander effect.

[0092] Disclosed herein are experiments establishing the development of IL13R α 2-specific CARs with a scFv47-based antigen-binding domain (47-CARs). The data show that 47-CARs perform better with a short spacer region, which provides for optimal functionality, and that 47-CAR T cells are able to recognize and kill only IL13R α 2-positive and not IL13R α 1-positive target cells in vitro. In addition, 47-CAR T cells induce tumor regression in an orthotopic xenograft mouse model of GBM, which was associated with a significant survival advantage.

[0093] The protein conjugates according to the disclosure are specific for IL13R α 2, which is associated with cancers, e.g., tumors. In addition, the disclosure provides a polynucleotide encoding one of these cancer-specific binding partners, including polynucleotides comprising codon-optimized coding regions for binding partners specific for an epitope of IL13R α 2. The polynucleotides of the disclosure encode conjugates, or bi-functional polypeptides, useful in diagnosing, preventing, treating, or ameliorating a symptom of cancer, such as any of a variety of human cancers, including those forming solid tumors. Also contemplated are vectors comprising a polynucleotide as disclosed herein, a host cell comprising such a polynucleotide and/or a vector as described above, and methods of treating, preventing or ameliorating a symptom of, a cancer disease, e.g., a solid tumor, a primary cancer site or a metastasized cancer.

[0094] The various forms of conjugates known in the art are contemplated by the disclosure. These conjugates provide exquisitely cancer—as well as protein-specific antibody receptors that can be incorporated into a variety of backbones providing effector function, such as bispecific T

cell Engagers (BiTEs) or chimeric antigen receptors (CARs), as noted below. Exemplary conjugates of the disclosure include CARs, fusion proteins, including fusions comprising single-chain variable (antibody) fragment (scFv) multimers or scFv fusions to coding regions encoding products useful in treating cancer, e.g., IL15, IL15R α , or IL15/ IL15Ra agent, diabodies, tribodies, tetrabodies, and bispecific bivalent scFvs, including bispecific tandem bivalent scFvs, also known as bispecific T cell engagers, or BiTEs. Any of these conjugate forms, moreover, may exhibit any of various relative structures, as it is known in the art that different domain orders (e.g., H2N-VH-linker-VL-CO2H and H₂N-VL-linker-VH-CO₂H) are compatible with specific binding. Higher order forms of the conjugates described herein are also contemplated, such as peptibodies comprising at least one form of the conjugates disclosed herein. The conjugates of the disclosure specifically bind to a cancer-specific epitope (e.g., an IL13R α 2) and the polynucleotides encoding them may be codon-optimized, e.g., for maximal translation, for expression in the targeted cells (e.g., human or mouse cells). Codon optimization in the context of expressing the conjugates of the disclosure, such as CARs, is important to ensuring that production of the protein is both efficient and robust enough to be useful as a source of therapeutic.

[0095] The disclosure also contemplates conjugates in which a targeting moiety (an anti-IL13R α 2 antibody or fragment thereof) is linked to a peptide providing a second function, e.g., an effector function, such as a T cell signaling domain involved in T cell activation, a peptide that affects or modulates an immunological response to cancer cells, or an enzymatic component of a labeling system that results in a CAR encoded by a polynucleotide according to the disclosure, if the coding region for the conjugate is codon-optimized for expression in a target cell. Exemplary conjugates include an anti-IL13R α 2 scFv linked to a hinge, a transmembrane domain, and an effector compound or domain, e.g., CD28, CD35, CD134 (OX40), CD137 (41BB), ICOS, CD40, CD27, or Myd88, thereby yielding a CAR.

[0096] The polynucleotide aspect of the disclosure comprises embodiments in which an unexpected variation on codon optimization in slower-growing higher eukaryotes such as vertebrates, e.g., humans, is provided that is focused on translation optimization (maximizing high-fidelity translation rates) rather than the typical codon optimization used in such organisms, which is designed to accommodate mutational bias and thereby minimize mutation. Also disclosed are the methods of diagnosing, preventing, treating or ameliorating a symptom of a cancer. Schematically described, the polynucleotides comprise a codon-optimized coding region for an antigen receptor specifically recognizing an IL13R α 2 epitope linked to any one of the following: a coding region for a T cell signaling domain involved in T cell activation, a gene product that affects or modulates an immunological response to cancer cells such as an IL15/ IL15Ra fusion, or a labeling component such as an enzymatic component of a labeling system. The linked coding regions result in polynucleotides encoding conjugates according to the disclosure, such as BiTEs or chimeric antigen receptors (CARs).

[0097] In methods of diagnosing, preventing, treating or ameliorating a symptom of a cancer, the compositions of the disclosure are typically administered in the form of a conjugate-transduced cell, such as a T cell, an NK cell, or a lymphocyte including, but not limited to, NKT cells, $\gamma\delta$ T cells, mucosa associated invariant T cells or MAIT cells, or innate lymphoid cells, although administration of a vector comprising a polynucleotide of the disclosure or administration of a polynucleotide of the disclosure are also contemplated, depending on the functionalities of the conjugate. Combining a polynucleotide, vector or host cell of the disclosure with a physiologically suitable buffer, adjuvant or diluent yields a pharmaceutical composition according to the disclosure, and these pharmaceutical compositions are suitable for administration to diagnose, prevent, treat, or ameliorate a symptom of, a cancer.

[0098] In the course of experimental work described herein, hybridoma cell lines were generated and compared for binding affinities to recombinant human IL13Ra2 (rhIL13R α 2). Clone 47 demonstrated binding to the native conformation of IL13R α 2 and was therefore characterized further. Clone 47 bound specifically and with high affinity (KD 1.39×10^{-9} M) to rhIL13Ra2 but not to rhIL13Ra1 or murine IL13Ra2. Furthermore, clone 47 specifically recognized wild-type IL13Ra2 expressed on the surface of CHO and HEK cells as well as several glioma cell lines. Competitive binding assays revealed that clone 47 also significantly inhibited the interaction between human soluble IL-13 and IL13Ra2 receptor. Moreover, N-linked glycosylation of IL13Ra2 was found to contribute, in part, to the interaction of the antibody with IL13Ra2. In vivo, the IL13R α 2 monoclonal antibody improved the survival of nude mice intracranially implanted with a human U251 glioma xenograft. Collectively, these data establish the efficacy of the immunomodulatory treatment of cancer disclosed herein.

[0099] Overexpression of IL13R α 2 in glioblastoma multiforme (GBM) but not in normal brain tissue uniquely positions this receptor as a candidate for targeting tumor cells. GBM is a highly infiltrative tumor, often making complete surgical removal impossible. Moreover, GBM is highly resistant to radiation and chemotherapy (16), warranting further development of novel and targeted therapies for the treatment of patients.

[0100] A phage display library approach has been used to select small antibody fragments specific to human IL13R α 2, followed by their evaluation in vitro and in vivo (23). Despite the high specificity of interaction with IL13R α 2. conjugation with toxins has failed to increase cytotoxicity in IL13Ra2-expressing glioma and renal cell carcinoma cell lines when compared with the effects of IL-13PE38. The low affinity of generated antibody fragments is the most reasonable explanation for the lack of success. Antibody fragments derived from phage display libraries are known to be lower in affinity and avidity than antibodies generated by conventional hybridoma technology (24). Modifications of those small antibody fragments are often required to enhance their affinity and avidity to targeted proteins. In recent years, monoclonal antibodies have shown increasing success as targeted anticancer and diagnostic agents (25, 26), and a further search for high affinity reagents with restricted specificity to tumor-associated antigens is needed. The experiments disclosed herein were designed to discover, develop, and characterize a high affinity antibody that specifically recognizes IL13R α 2 expressed on the surface of cancer cells. Consistent with that design, disclosed herein are experiments establishing the generation of an antibody possessing the properties critical for immunotherapeutic targeting of IL13R α 2-expressing tumors in vivo, and potentially suitable for various other applications.

[0101] Monoclonal antibodies appear to be valuable research and diagnostic tools as well as therapeutic agents. Monoclonal antibodies specific for tumor-associated antigens have significant advantages over systemic chemotherapies due to the ability to specifically target cancer cells while avoiding interaction with untransformed tissue. Therefore, the search for novel "magic bullets" continues to grow, confirmed by a global market for therapeutic antibodies worth \$48 billion as of 2010. Therapeutic antibodies are products of traditional hybridoma technology or screening of libraries for antibody fragments and their subsequent engineering into humanized fragments or full size molecules. Prior to this study, the hybridoma cell line secreting a high affinity antibody to the tumor-specific antigen IL13Ra2 was unavailable to the scientific community. Here, we describe the generation and characterization of a high affinity antibody to the tumor-specific antigen IL13R α 2 and discuss its potential use in different applications.

[0102] The specificity of interaction of newly discovered antibodies to human IL13Ra2 was analyzed by ELISA using the rhIL13Ra2hFc fusion protein, recombinant human IL13R α 2 expressed on the surface of CHO and HEK cells, and several glioma cell lines expressing IL13R α 2 at various levels by flow cytometry. The antibody identified herein, and agent using the binding domain thereof, demonstrated a specificity of interaction to human $IL13R\alpha 2$ and did not cross-react with human IL13R α 1 or mouse IL13R α 2. Moreover, the specificity of binding to $IL13R\alpha 2$ was confirmed in competitive binding assays using rhIL13Ra2hFc fusion protein by ELISA or by flow cytometry for detection of IL13R α 2 expressed on the surface of HEK cells. In these assays, IL13Ra2 (clone 47) mAb competed with recombinant human IL-13 for its epitope and was able to block about 80% of the binding between IL-13 and IL13R α 2. Conversely, human recombinant IL-13 was able to block about 50% of antibody binding to IL13Ra2. Similarly, a significant decrease in the binding of IL13R α 2 (clone 47) mAb to N10 glioma cells was observed when rhIL13R2hFc chimera and rhIL-13 were used as competitors. The binding of rhIL-13 to N10 cells was also abolished by IL13R α 2 (clone 47) mAb. These data indicate that the two molecules have significant overlap in their recognition sites for IL13R α 2.

[0103] IL-13 is a small 10-kDa molecule (31), whereas an antibody is about 15 times greater in molecular mass. The ability of rhIL-13 to compete with an antibody for a binding site suggests that the inhibitory property of the antibody is likely due to the specific interaction with amino acid residues contributing to the binding of IL-13 to the cognate receptor rather than to steric hindrance, which can also prevent the interaction of IL-13 with its receptor. Previously, Tyr207, Asp271, Tyr315, and Asp318 were identified as critical residues of IL13R α 2 necessary for interaction with IL-13 (28). In the assays disclosed herein, the binding of IL-13 to a mutant IL13R α 2 carrying a combination of all 4 amino acid mutations to alanine was significantly abolished when compared with the wild-type receptor. Binding of the IL13R α 2 mAb to either the individual or the 4-amino acid mutant form of IL13R α 2, however, was not significantly affected. These findings indicate that Tyr207, Asp271, Tyr315, and Asp318 residues are not critical for the recognition of IL13R α 2 by the IL13R α 2 mAb. The human IL13Ra2 and murine IL13Ra2 are structurally conserved

and share 59% amino acid identity (32). Moreover, Tyr207, Asp271, Tyr315, and Asp318 residues are conserved in human and murine IL13R α 2. Absence of binding of the IL13R α 2 mAb to murine IL13R α 2hFc fusion further supports the expectation that these amino acid residues contribute to the binding of IL-13 to IL13R α 2 and are not critical for the interaction of this antibody with the receptor.

[0104] To further characterize the interaction of IL13R α 2 with the antibody and antibody agent disclosed herein, the affinity of the IL13Ra2 mAb was measured and compared with the binding properties of two commercially available antibodies using the surface plasmon resonance method. The affinity of the IL13R α 2 mAb was determined to be equal to 1.39×10^{-9} M, greatly exceeding the affinity of comparable commercially available antibodies by up to 75-fold. In agreement with the affinity studies, the IL13Ra2 mAb (clone 47) demonstrated superiority to two commercial antibodies in binding to the IL13Ra2 expressed on the surface of various glioma cells and in ELISA. Although many properties of antibodies, including the affinity and avidity, in vivo stability, rate of clearance and internalization, tumor penetration, and retention, should be considered prior to specific usage, it has been reported that higher affinity antibodies are better for immunotherapeutic tumortargeting applications (33). The single chain antibody fragment (scFv) MR1-1 against epidermal growth factor receptor variant III demonstrates about 15-fold higher affinity than the parental scFvMR1 and also showed on average a 244% higher tumor uptake than that for the scFvMR1 (34). It is likely that the high affinity properties of the IL13R α 2 mAb and agent thereof that are disclosed herein will be advantageous for applications utilizing antibodies or associated derivatives for targeting tumor cells expressing IL13Rα2.

[0105] The N-linked glycosylation of IL13R α 2 has been identified as a necessary requirement for efficient binding to IL-13 (30). Taking into consideration that the IL13R α 2 mAb disclosed herein inhibits about 80% of IL-13 binding to the cognate receptor, IL13R α 2, it is reasonable to expect that the binding of this antibody, or an agent containing its binding domain, with the deglycosylated form of IL13R α 2 could also be affected. The IL13R α 2 molecule has four potential sites of N-linked glycosylation. The binding of the antibody to rhIL13Ra2 or to IL13Ra2 expressed on the surface of HEK or U251 cells treated with Pngase F was decreased by 35 and 30%, respectively, when compared with non-treated control. A partial change in binding activity for the clone 47 when compared with clones 83807 and B-D13 suggests that removal of carbohydrate adducts from IL13Ra2 with Pngase F causes conformational changes of the receptor, indirectly affecting the binding of both IL-13 (30) and the IL13R α 2 mAb to IL13R α 2. This also supports the expectation that the antibody binds directly to the IL13R α 2 amino acid backbone rather than interacting with carbohydrate moieties added post-translationally. Supporting this expectation, several studies have previously demonstrated that the conformational profile and structural rigidity of proteins depends on N-linked glycosylation (22, 35-38).

[0106] To investigate the therapeutic properties of the IL13R α 2 mAb and its agent, an in vivo study was performed whereby glioma cells and the IL13R α 2 (clone 47) mAb were intracranially co-injected into brain, or antibody was injected into established tumor-bearing mice. Interestingly, the IL13R α 2 mAb was able to delay tumor progression and

improve survival of animals with intracranial U251 glioma xenografts most significantly in the co-injected model, demonstrating a trend in the improvement of median survival in animals with established glioma. Although the underlying mechanism for this antitumor effect remains unclear, the result establishes the therapeutic applicability of this antibody, or its agent (containing the IL13R α 2 binding domain in the form of the six CDR regions, or in the form of the two variable domains of the clone 47 anti-IL13R α 2), alone or in combination with a pharmaceutical carrier, thereby providing therapies for the treatment of IL13R α 2-expressing glial and other lineage tumors. Several antibodies have been shown to mediate a cytotoxic effect in tumors through Fc-mediated activation of complement (39). Antibody-dependent cell-mediated cytotoxicity-induced activation of effector cells can also contribute to the cytotoxic effect of antibodies against targeted cells (40, 41). Anti-IL13Ra2 activity derived from the sera of animals challenged with D5 melanoma cells expressing human IL13R α 2 demonstrates the ability to inhibit cellular growth in vitro (4).

[0107] Cancers amenable to the described treatments include cancers in which IL13Ra2 has been found to be expressed, including glioblastoma; medulloblastoma; Kaposi sarcoma; and head and neck, ovarian, pancreatic, kidney, and colorectal cancers (2, 43-47). Although the role of IL13R α 2 in some cancers is not yet defined, recent reports have demonstrated that $IL13R\alpha^2$ contributes to the invasive phenotype of ovarian, pancreatic, and colorectal cancers (5, 13). Moreover, Minn et al. (42) have suggested a relationship between IL13Ra2 expression and breast cancer metastasis to the lung. Additionally, Fichtner-Feigl et al. (11) demonstrated that the interaction of IL-13 with IL13R α 2 upregulates TGF- β 1, mediating fibrosis in a bleomycin-induced model of lung fibrosis. In light of this finding, it is expected that the anti-IL13R α 2 antibody (clone 47) and binding agents thereof, will be able to attenuate TGF-β1-induced pulmonary fibrosis.

[0108] As disclosed herein, the described experiments led to the generation of an anti-IL13R α 2 antibody and binding agents thereof, all of which are specific to human IL13R α 2. The antibody and its agent possess a high affinity for IL13R α 2 and compete with IL-13 for the binding site on IL13R α 2. The antibody recognizes antigen expressed on the cell surface of glioma cells as well as other IL13R α 2 expressing cells, establishing the suitability for targeting IL13R α 2-expressing tumor cells in vivo. The anti-IL13R α 2 antibody and binding agents thereof are also expected to be efficacious and cost effective in diagnostic imaging, delivery of antibody radionuclide conjugates, bioassays for the detection of IL13R α 2, and as a carrier for therapeutic agents in various types of IL13R α 2-overexpressing tumors.

[0109] In methods of diagnosing, preventing, treating or ameliorating a symptom of a cancer, the compositions of the disclosure are typically administered in the form of conjugate-transduced T cells, although administration of a vector comprising a polynucleotide of the disclosure or administration of a polynucleotide of the disclosure are also contemplated, depending on the functionalities of the conjugate. Combining a polynucleotide, vector or host cell of the disclosure with a physiologically suitable buffer, adjuvant or diluent yields a pharmaceutical composition according to the disclosure, and these pharmaceutical compositions are suitable for administration to diagnose, prevent, treat, or ameliorate a symptom of, a cancer.

[0110] A conjugate according to the disclosure, such as a fusion protein composed of an scFv-receptor for an IL13R α 2 epitope fused to IL15/IL15R α , is also contemplated. It is expected that the fusion protein will eliminate clinical size tumors or only incipient and microdisseminated cancer cells. The disclosure further contemplates the simultaneous targeting of two independent IL13R α 2 epitopes on a human cancer, which may be essential for preventing escape from treatment, such as CAR treatment.

[0111] Simultaneous targeting of different epitopes of IL13R α 2 by CARs should reduce the chance of escape of a cancer subpopulation, which provides a strong reason for identifying additional IL13R α 2 antibody products and/or epitopes.

[0112] The disclosure provides materials and methods that are adaptable and can serve as the basis for a platform technology with considerable growth potential. The cancerspecific nature of IL13R α 2 is expected to provide targets for cancer diagnostics, prophylactics and therapeutics that offer major advantages over previously and presently used targets.

[0113] Consistent with the spirit of the foregoing, the following provides a description of the materials and methods provided herein.

[0114] Disclosed herein are IL13R α 2 binding agents comprising each of the amino acid sequences of NYLMN (SEQ ID NO: 1); RIDPYDGDIDYNQNFKD (SEQ ID NO: 2); GYGTAYGVDY (SEQ ID NO: 3); RASESVDNYGISFMN (SEQ ID NO: 4); AASRQGSG (SEQ ID NO: 5); and QQSKEVPWT (SEQ ID NO: 6). In exemplary aspects, the binding agent comprises each of the foregoing six amino acid sequences in addition to further sequences which provide a framework to support a three-dimensional conformation that binds to IL13R α 2. In exemplary aspects, the IL13Rα2 binding agent comprises one or both of the amino acid sequences of SEQ ID NO: 7 and/or SEQ ID NO: 8. In exemplary aspects, the IL13Ro2 binding agent comprises the amino acid sequence of SEQ ID NO: 7. In exemplary aspects, the IL13Ra2 binding agent comprises the amino acid sequence of SEQ ID NO: 8. In exemplary aspects, the IL13Ra2 binding agent comprises both the amino acid sequences of SEQ ID NO: 7 and SEQ ID NO: 8. In exemplary aspects wherein both the amino acid sequences of SEQ ID NO: 7 and SEQ ID NO: 8 are present in the binding agent, the amino acid sequence of SEQ ID NO: 7 is fused to the amino acid sequence of SEQ ID NO: 8 through a linker. Suitable linkers are known in the art. In exemplary aspects, the linker comprises a short amino acid sequence of about 5 to about 25 amino acids, e.g., about 10 to about 20 amino acids. In exemplary aspects, the linker comprises the amino acid sequence of EEGEFSEAR (SEQ ID NO 10). In exemplary aspects, the linker comprises the amino acid sequence of AKTTPPKLEEGEFSEARV (SEQ ID NO: 80). In exemplary aspects, IL13R α 2 binding agent comprises the amino acid sequence of SEQ ID NO: 13.

[0115] In exemplary embodiments, the binding agent provided herein further comprises additional amino acid sequences. In exemplary aspects, the binding agent further comprises a constant region of a heavy chain and/or a constant region of a light chain. Sequences for heavy and light chain constant regions are publically available. For example, the National Center of Biotechnology Information (NCBI) nucleotide database provides a sequence of the constant region of the IgG1 kappa light chain. See GenBank

Accession No. DQ381549.1, incorporated herein by reference. In exemplary aspects, the binding agent comprises an amino acid sequence of SEQ ID NO: 28. In exemplary aspects, the binding agent comprises a modified amino acid sequence of SEQ ID NO: 28. In exemplary aspects, the binding agent comprises an amino acid sequence which is at least 90%, at least 93%, at least 95%, or at least 98% identical to SEQ ID NO: 28. Also, for example, the NCBI nucleotide database provides a sequence of the constant region of the Mus musculus IgG1. See GenBank Accession No. DQ381544.1. In exemplary aspects, the binding agent comprises an amino acid sequence of SEQ ID NO: 29. In exemplary aspects, the binding agent comprises a modified amino acid sequence of SEQ ID NO: 29. In exemplary aspects, the binding agent comprises an amino acid sequence which is at least 90%, at least 93%, at least 95%, or at least 98% identical to SEQ ID NO: 29.

[0116] In exemplary aspects, the IL13R α 2 binding agent is an antibody, or an antigen-binding fragment thereof. In exemplary aspects, the antibody comprises each of the amino acid sequences of SEQ ID NOs: 1-6. In exemplary aspects, the antibody comprises the amino acid sequence of SEQ ID NO: 7 and/or SEQ ID NO: 8. In exemplary aspects, the antibody comprises the amino acid sequences of SEQ ID NO: 7 and SEQ ID NO: 8. In exemplary aspects, the antibody comprises the amino acid sequences of SEQ ID NO: 7 and SEQ ID NO: 8 and the amino acid sequence of SEQ ID NO: 7 is fused to the amino acid sequence of SEQ ID NO: 8 through a linker. In exemplary aspects, the linker comprises a short amino acid sequence of about 5 to about 25 amino acids, e.g., about 10 to about 20 amino acids. In exemplary aspects, the linker comprises the amino acid sequence of EEGEFSEAR (SEQ ID NO 10). In exemplary aspects, the linker comprises the amino acid sequence of AKTTPPKLEEGEFSEARV (SEQ ID NO: 80). In exemplary aspects, the antibody comprises the amino acid sequence of SEQ ID NO: 13.

[0117] In exemplary aspects, the antibody can be any type of immunoglobulin that is known in the art. For instance, the antibody can be of any isotype, e.g., IgA, IgD, IgE, IgG, or IgM. The antibody can be monoclonal or polyclonal. The antibody can be a naturally-occurring antibody, i.e., an antibody isolated and/or purified from a mammal, e.g., mouse, rabbit, goat, horse, chicken, hamster, human, and the like. In this regard, the antibody may be considered to be a mammalian antibody, e.g., a mouse antibody, rabbit antibody, goat antibody, horse antibody, chicken antibody, hamster antibody, human antibody, and the like. The term "isolated" as used herein means having been removed from its natural environment. The term "purified," as used herein relates to the isolation of a molecule or compound in a form that is substantially free of contaminants normally associated with the molecule or compound in a native or natural environment and means having been increased in purity as a result of being separated from other components of the original composition. It is recognized that "purity" is a relative term, and not to be necessarily construed as absolute purity or absolute enrichment or absolute selection. In some aspects, the purity is at least or about 50%, is at least or about 60%, at least or about 70%, at least or about 80%, or at least or about 90% (e.g., at least or about 91%, at least or about 92%, at least or about 93%, at least or about 94%, at least

or about 95%, at least or about 96%, at least or about 97%, at least or about 98%, at least or about 99% or is approximately 100%.

[0118] In exemplary aspects, the antibody comprises a constant region of an IgG. In exemplary aspects, the antibody comprises a constant region of an IgG_1 . In exemplary aspects, the antibody comprises a constant region of an IgG kappa light chain. For instance, the antibody may comprise the amino acid sequence of SEQ ID NO: 28. In exemplary aspects, the antibody comprises an amino acid sequence that is highly similar to SEQ ID NO: 28. For instance, the antibody may comprise an amino acid sequence having at least 85% sequence identity to SEQ ID NO: 28, or an amino acid sequence having at least 90% sequence identity to SEQ ID NO: 28, or an amino acid sequence having at least 93% sequence identity to SEQ ID NO: 28, or an amino acid sequence having at least 95% sequence identity to SEQ ID NO: 28, or an amino acid sequence having at least 98% sequence identity to SEQ ID NO: 28.

[0119] In exemplary aspects, the antibody comprises a constant region of a *Mus musculus* IgG_1 . For instance, the antibody may comprise the amino acid sequence of SEQ ID NO: 30. In exemplary aspects, the antibody comprises an amino acid sequence which is highly similar to SEQ ID NO: 30. For instance, the antibody may comprise an amino acid sequence having at least 85% sequence identity to SEQ ID NO: 30, or an amino acid sequence having at least 90% sequence identity to SEQ ID NO: 30, or an amino acid sequence having at least 93% sequence identity to SEQ ID NO: 30, or an amino acid sequence having at least 95% sequence identity to SEQ ID NO: 30, or an amino acid sequence having at least 95% sequence identity to SEQ ID NO: 30, or an amino acid sequence having at least 98% sequence identity to SEQ ID NO: 30.

[0120] The anti-IL13R α 2 antibodies and fragments thereof of the disclosure can have any level of affinity or avidity for IL13Ra2. The dissociation constant (IQ may be any of those exemplary dissociation constants described herein with regard to binding units. Binding constants, including dissociation constants, are determined by methods known in the art, including, for example, methods that utilize the principles of surface plasmon resonance, e.g., methods utilizing a BiacoreTM system. In accordance with the foregoing, in some embodiments, the antibody is in monomeric form, while in other embodiments, the antibody is in polymeric form. In certain embodiments in which the antibody comprises two or more distinct antigen binding regions or fragments, the antibody is considered bispecific, trispecific, or multi-specific, or bivalent, trivalent, or multivalent, depending on the number of distinct epitopes that are recognized and bound by the binding agent.

[0121] Because the binding agent of the disclosures can compete with IL13 for binding to IL13R α 2, the antibody in exemplary aspects is considered to be a blocking antibody or neutralizing antibody. In some aspects, the K_D of the binding agent is about the same as the K_D of the native ligand, IL13, for IL13R α 2. In some aspects, the K_D of the binding agent is lower (e.g., at least 0.5-fold lower, at least 1-fold lower, at least 2-fold lower, at least 50-fold lower, at least 10-fold lower, at least 25-fold lower, at least 50-fold lower, at least 75-fold lower, at least 100-fold lower) than the K_D of IL13R α 2. In exemplary aspects, the K_D is between about 0.0001 nM and about 100 nM. In some embodiments, the K_D is at least or about 0.001 nM, at least or about 0.001 nM, at least or about 0.001 nM, at least or about 0.1 nM, at least 0.5 nM and about 100 nM.

or about 1 nM, or at least or about 10 nM. In some embodiments, the K_D is no more than or about 100 nM, no more than or about 75 nM, no more than or about 50 nM, or no more than or about 25 nM. In exemplary aspects, the antibody has a K_D for human IL13R α 2 that is no greater than about 1.39×10⁻⁹ M.

[0122] In exemplary aspects, the binding agent, e.g., antibody, or antigen binding fragment thereof, does not bind to human IL13R α 1.

[0123] In exemplary embodiments, the antibody is a genetically engineered antibody, e.g., a single chain antibody, a humanized antibody, a chimeric antibody, a CDR-grafted antibody, an antibody that includes portions of CDR sequences specific for IL13R α 2 (e.g., an antibody that includes CDR sequences of SEQ ID NOs: 1-6), a humaneered or humanized antibody, a bispecific antibody, a trispecific antibody, and the like, as defined in greater detail herein. Genetic engineering techniques also provide the ability to make fully human antibodies in a non-human.

[0124] In some aspects, the antibody is a chimeric antibody. The term "chimeric antibody" is used herein to refer to an antibody containing constant domains from one species and the variable domains from a second, or more generally, containing stretches of amino acid sequence from at least two species.

[0125] In some aspects, the antibody is a humanized antibody. The term "humanized" when used in relation to antibodies is used to refer to antibodies having at least CDR regions from a nonhuman source that are engineered to have a structure and immunological function more similar to true human antibodies than the original source antibodies. For example, humanizing can involve grafting CDR from a non-human antibody, such as a mouse antibody, into a human antibody. Humanizing also can involve select amino acid substitutions to make a non-human sequence look more like a human sequence, as would be known in the art.

[0126] Use of the terms "chimeric or humanized" herein is not meant to be mutually exclusive; rather, is meant to encompass chimeric antibodies, humanized antibodies, and chimeric antibodies that have been further humanized. Except where context otherwise indicates, statements about (properties of, uses of, testing, and so on) chimeric antibodies apply to humanized antibodies, and statements about humanized antibodies pertain also to chimeric antibodies. Likewise, except where context dictates, such statements also should be understood to be applicable to antibodies and antigen binding fragments of such antibodies.

[0127] In some aspects of the disclosure, the binding agent is an antigen binding fragment of an antibody that specifically binds to an IL13R α 2 in accordance with the disclosure. The antigen binding fragment (also referred to herein as "antigen binding portion") may be an antigen binding fragment of any of the antibodies described herein. The antigen binding fragment can be any part of an antibody that has at least one antigen binding site, including, but not limited to, Fab, F(ab')₂, dsFv, sFv, scFv, diabodies, triabodies, bisscFvs, fragments expressed by a Fab expression library, domain antibodies, VhH domains, V-NAR domains, VH domains, VL domains, and the like. Antibody fragments of the invention, however, are not limited to these exemplary types of antibody fragments.

[0128] In exemplary aspects, the IL13R α 2 binding agent is an antigen binding fragment. In exemplary aspects, the antigen binding fragment comprises each of the amino acid

sequences of SEQ ID NOs: 1-6. In exemplary aspects, the antigen binding fragment comprises the amino acid sequence of SEQ ID NO: 7 and/or SEQ ID NO: 8. In exemplary aspects, the antigen binding fragment comprises the amino acid sequences of SEQ ID NO: 7 and SEQ ID NO: 8. In exemplary aspects, the antigen binding fragment comprises the amino acid sequences of SEQ ID NO: 7 and SEQ ID NO: 8 and the amino acid sequence of SEQ ID NO: 7 is fused to the amino acid sequence of SEQ ID NO: 8 through a linker. In exemplary aspects, the linker comprises a short amino acid sequence of about 5 to about 25 amino acids, e.g., about 10 to about 20 amino acids. In exemplary aspects, the linker comprises the amino acid sequence of EEGEF-SEAR (SEQ ID NO 10). In exemplary aspects, the linker comprises the amino acid sequence of AKTTPPKLEEGEF-SEARV (SEQ ID NO: 80). In exemplary aspects, the antigen binding fragment provided herein comprises the amino acid sequence of SEQ ID NO: 13.

[0129] In exemplary aspects, the antigen binding fragment comprises a leader sequence. Optionally, the leader sequence, in some aspects, is located N-terminal to the heavy chain variable region. In exemplary aspects, the antigen binding fragment comprises an Ig kappa leader sequence. Suitable leader sequences are known in the art, and include, for example, an Ig kappa leader sequence of METDTLLLWVLLLWVPGSTGD (SEQ ID NO: 9).

[0130] In exemplary aspects, an antigen binding fragment comprises one more tag sequences. Tag sequences may assist in the production and characterization of the manufactured antigen binding fragment. In exemplary aspects, the antigen binding fragment comprises one or more tag sequences C-terminal to the light chain variable region. Suitable tag sequences are known in the art and include, but are not limited to, Myc tags, His tags, and the like. In exemplary aspects, an antigen binding fragment comprises a Myc tag of GGPEQKLISEEDLN (SEQ ID NO: 11). In exemplary aspects, an antigen binding fragment comprises a His tag sequence of HHHHHH (SEQ ID NO: 12).

[0131] In exemplary aspects, the antigen binding fragment of the disclosures comprises, from the N- to the C-terminus, a leader sequence, a heavy chain variable region, a linker sequence, a light chain variable region, a Myc tag (e.g., SEQ ID NO: 11), and a His tag (e.g., SEQ ID NO: 12). In exemplary aspects, the antigen binding fragment of the disclosure comprises the amino acid sequence of SEQ ID NO: 14.

[0132] In exemplary aspects, the antigen binding fragment is a domain antibody. A domain antibody comprises a functional binding unit of an antibody, and can correspond to the variable regions of either the heavy (V_H) or light (V_L) chains of antibodies. A domain antibody can have a molecular weight of approximately 13 kDa, or approximately one-tenth the weight of a full antibody. Domain antibodies may be derived from full antibodies, such as those described herein. The antigen binding fragments in some embodiments are monomeric or polymeric, bispecific or trispecific, and bivalent or trivalent.

[0133] Antibody fragments that contain the antigen binding, or idiotope, of the antibody molecule share a common idiotype and are contemplated by the disclosure. Such antibody fragments may be generated by techniques known in the art and include, but are not limited to, the $F(ab')_2$ fragment which may be produced by pepsin digestion of the antibody molecule; the Fab' fragments which may be generated by reducing the disulfide bridges of the $F(ab')_2$ fragment, and the two Fab' fragments which may be generated by treating the antibody molecule with papain and a reducing agent.

[0134] In exemplary aspects, the binding agent provided herein is a single-chain variable region fragment (scFv) antibody fragment. An scFv may consist of a truncated Fab fragment comprising the variable (V) domain of an antibody heavy chain linked to a V domain of an antibody light chain via a synthetic peptide, and it can be generated using routine recombinant DNA technology techniques (see, e.g., Janeway et al., *Immunobiology*, 2^{nd} Edition, Garland Publishing, New York, (1996)). Similarly, disulfide-stabilized variable region fragments (dsFv) can be prepared by recombinant DNA technology (see, e.g., Reiter et al., Protein Engineering, 7, 697-704 (1994)).

[0135] In exemplary aspects, the IL13R α 2 binding agent provided herein is an scFv. In exemplary aspects, the scFv comprises each of the amino acid sequences of SEQ ID NOs: 1-6. In exemplary aspects, the scFv comprises the amino acid sequence of SEQ ID NO: 7 or SEQ ID NO: 8. In exemplary aspects, the scFv comprises the amino acid sequences of SEQ ID NO: 7 and SEQ ID NO: 8. In exemplary aspects, the scFv comprises the amino acid sequences of SEQ ID NO: 7 and SEQ ID NO: 8 and the amino acid sequence of SEQ ID NO: 7 is fused to the amino acid sequence of SEQ ID NO: 8 through a linker. In exemplary aspects, the linker comprises a short amino acid sequence of about 5 to about 25 amino acids, e.g., about 10 to about 20 amino acids. In exemplary aspects, the linker comprises the amino acid sequence of EEGEFSEAR (SEQ ID NO 10). In exemplary aspects, the linker comprises the amino acid sequence of AKTTPPKLEEGEFSEARV (SEQ ID NO: 80). In exemplary aspects, the scFv provided herein comprises the amino acid sequence of SEQ ID NO: 13.

[0136] Recombinant antibody fragments, e.g., scFvs of the disclosure, can also be engineered to assemble into stable multimeric oligomers of high binding avidity and specificity to different target antigens. Such diabodies (dimers), triabodies (trimers) or tetrabodies (tetramers) are well known in the art. See e.g., Kortt et al., *Biomol Eng.* 2001 18:95-108, (2001) and Todorovska et al., *J Immunol Methods.* 248:47-66, (2001).

[0137] In exemplary aspects, the binding agent is a bispecific antibody (bscAb). Bispecific antibodies are molecules comprising two single-chain Fv fragments joined via a glycine-serine linker using recombinant methods. The V light-chain (V_I) and V heavy-chain (V_H) domains of two antibodies of interest in exemplary embodiments are isolated using standard PCR methods. The V_L and V_H cDNAs obtained from each hybridoma are then joined to form a single-chain fragment in a two-step fusion PCR. Bispecific fusion proteins are prepared in a similar manner. Bispecific single-chain antibodies and bispecific fusion proteins are antibody substances included within the scope of the present invention. Exemplary bispecific antibodies are taught in U.S. Patent Application Publication No. 2005-0282233A1 and International Patent Application Publication No. WO 2005/087812, both applications of which are incorporated herein by reference in their entireties.

[0138] In exemplary aspects, the binding agent is a bispecific T-cell engaging antibody (BiTE) containing two scFvs produced as a single polypeptide chain. In exemplary aspects, the binding agent is a BiTE comprising two scFvs,

wherein at least one comprises each of the amino acid sequences of SEQ ID NOs: 1-6 or comprises SEQ ID NO: 7 and/or SEQ ID NO: 8. Methods of making and using BiTE antibodies are described in the art. See, e.g., Cioffi et al., *Clin Cancer Res* 18: 465, Brischwein et al., *Mol Immunol* 43:1129-43 (2006); Amann M et al., *Cancer Res* 68:143-51 (2008); Schlereth et al., *Cancer Res* 65: 2882-2889 (2005); and Schlereth et al., *Cancer Immunol Immunother* 55:785-796 (2006).

[0139] In exemplary aspects, the binding agent is a dual affinity re-targeting antibody (DART). DARTs are produced as separate polypeptides joined by a stabilizing interchain disulphide bond. In exemplary aspects, the binding agent is a DART comprising an scFv comprising each of the amino acid sequences of SEQ ID NOs: 1-6 or comprises SEQ ID NO: 7 and/or SEQ ID NO: 8. Methods of making and using DART antibodies are described in the art. See, e.g., Rossi et al., *MAbs* 6: 381-91 (2014); Fournier and Schirrmacher, BioDrugs 27:35-53 (2013); Johnson et al., *J Mol Biol* 399:436-449 (2010); Brien et al., *J Virol* 87: 7747-7753 (2013); and Moore et al., *Blood* 117:4542 (2011).

[0140] In exemplary aspects, the binding agent is a tetravalent tandem diabody (TandAbs) in which an antibody fragment is produced as a non-covalent homodimer folder in a head-to-tail arrangement. In exemplary aspects, the binding agent is a TandAbs comprising an scFv comprising each of the amino acid sequences of SEQ ID NOs: 1-6 or comprises SEQ ID NO: 7 and/or SEQ ID NO: 8. TandAbs are known in the art. See, e.g., McAleese et al., *Future Oncol* 8: 687-695 (2012); Portner et al., *Cancer Immunol Immunother* 61:1869-1875 (2012); and Reusch et al., *MAbs* 6:728 (2014).

[0141] In exemplary aspects, the BiTE, DART, or Tand-Abs comprises the CDRs of SEQ ID NOs: 1-6. In exemplary aspects, the BiTE, DART, or TandAbs comprises the amino acid sequence of SEQ ID NOs: 7 and 8. In exemplary aspects, the BiTE, DART, or TandAbs comprises SEQ ID NOs: 13.

[0142] Suitable methods of making antibodies are known in the art. For instance, standard hybridoma methods are described in, e.g., Harlow and Lane (eds.), Antibodies: A Laboratory Manual, CSH Press (1988), and CA. Janeway et al. (eds.), Immunobiology, 5th Ed., Garland Publishing, New York, N.Y. (2001)).

[0143] Monoclonal antibodies for use in the invention may be prepared using any technique that provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique originally described by Koehler and Milstein (Nature 256: 495-497, 1975), the human B-cell hybridoma technique (Kosbor et al., Immunol Today 4:72, 1983; Cote et al., Proc Natl Acad Sci 80: 2026-2030, 1983) and the EBV-hybridoma technique (Cole et al., Monoclonal Antibodies and Cancer Therapy, Alan R Liss Inc, New York N.Y., pp 77-96, (1985).

[0144] Briefly, a polyclonal antibody is prepared by immunizing an animal with an immunogen comprising a polypeptide of the present invention and collecting antisera from that immunized animal. A wide range of animal species can be used for the production of antisera. In some aspects, an animal used for production of anti-antisera is a non-human animal including rabbits, mice, rats, hamsters, goat, sheep, pigs or horses. Because of the relatively large blood volume of rabbits, a rabbit, in some exemplary aspects, is a

preferred choice for production of polyclonal antibodies. In an exemplary method for generating a polyclonal antisera immunoreactive with the chosen IL13R α 2 epitope, 50 µg of IL13R α 2 antigen is emulsified in Freund's Complete Adjuvant for immunization of rabbits. At intervals of, for example, 21 days, 50 µg of epitope are emulsified in Freund's Incomplete Adjuvant for boosts. Polyclonal antisera may be obtained, after allowing time for antibody generation, simply by bleeding the animal and preparing serum samples from the whole blood.

[0145] Briefly, in exemplary embodiments, to generate monoclonal antibodies, a mouse is injected periodically with recombinant IL13R α 2 against which the antibody is to be raised (e.g., 10-20 µg IL13Ra2 emulsified in Freund's Complete Adjuvant). The mouse is given a final pre-fusion boost of a IL13R α 2 polypeptide containing the epitope that allows specific recognition of lymphatic endothelial cells in PBS, and four days later the mouse is sacrificed and its spleen removed. The spleen is placed in 10 ml serum-free RPMI 1640, and a single cell suspension is formed by grinding the spleen between the frosted ends of two glass microscope slides submerged in serum-free RPMI 1640, supplemented with 2 mM L-glutamine, 1 mM sodium pyruvate, 100 units/ml penicillin, and 100 µg/ml streptomycin (RPMI) (Gibco, Canada). The cell suspension is filtered through sterile 70-mesh Nitex cell strainer (Becton Dickinson, Parsippany, N.J.), and is washed twice by centrifuging at 200 g for 5 minutes and resuspending the pellet in 20 ml serum-free RPMI. Splenocytes taken from three naive Balb/c mice are prepared in a similar manner and used as a control. NS-1 myeloma cells, kept in log phase in RPMI with 11% fetal bovine serum (FBS) (Hyclone Laboratories, Inc., Logan, Utah) for three days prior to fusion, are centrifuged at 200 g for 5 minutes, and the pellet is washed twice.

[0146] Spleen cells (1×10^8) are combined with 2.0×10^7 NS-1 cells and centrifuged, and the supernatant is aspirated. The cell pellet is dislodged by tapping the tube, and 1 ml of 37° C. PEG 1500 (50% in 75 mM Hepes, pH 8.0) (Boehringer Mannheim) is added with stirring over the course of 1 minute, followed by the addition of 7 ml of serum-free RPMI over 7 minutes. An additional 8 ml RPMI is added and the cells are centrifuged at 200 g for 10 minutes. After discarding the supernatant, the pellet is resuspended in 200 ml RPMI containing 15% FBS, 100 μ M sodium hypoxanthine, 0.4 μ M aminopterin, 16 μ M thymidine (HAT) (Gibco), 25 units/ml IL-6 (Boehringer Mannheim) and 1.5×10⁶ splenocytes/ml and plated into 10 Corning flat-bottom 96-well tissue culture plates (Corning, Corning N.Y.).

[0147] On days 2, 4, and 6, after the fusion, 100 µl of medium is removed from the wells of the fusion plates and replaced with fresh medium. On day 8, the fusion is screened by ELISA, testing for the presence of mouse IgG binding to IL13R α 2 as follows. Immulon 4 plates (Dynatech, Cambridge, Mass.) are coated for 2 hours at 37° C. with 100 ng/well of IL13R α 2 diluted in 25 mM Tris, pH 7.5. The coating solution is aspirated and 200 µl/well of blocking solution (0.5% fish skin gelatin (Sigma) diluted in CMF-PBS) is added and incubated for 30 minutes at 37° C. Plates are washed three times with PBS containing 0.05% Tween 20 (PBST) and 50 µl culture supernatant is added. After incubation at 37° C. for 30 minutes, and washing as above, 50 µl of horseradish peroxidase-conjugated goat anti-mouse IgG(Fc) (Jackson ImmunoResearch, West Grove, Pa.)

diluted 1:3500 in PBST is added. Plates are incubated as above, washed four times with PBST, and 100 μ l substrate, consisting of 1 mg/ml o-phenylene diamine (Sigma) and 0.1 μ l/ml 30% H₂O₂ in 100 mM citrate, pH 4.5, are added. The color reaction is stopped after 5 minutes with the addition of 50 μ l of 15% H₂SO₄. The A₄₉₀ absorbance is determined using a plate reader (Dynatech).

[0148] Selected fusion wells are cloned twice by dilution into 96-well plates and visual scoring of the number of colonies/well after 5 days. The monoclonal antibodies produced by hybridomas are isotyped using the Isostrip system (Boehringer Mannheim, Indianapolis, Ind.).

[0149] When the hybridoma technique is employed, myeloma cell lines may be used. Such cell lines suited for use in hybridoma-producing fusion procedures preferably are non-antibody-producing, have high fusion efficiency, and enzyme deficiencies that render them incapable of growing in certain selective media that support the growth of only the desired fused cells (hybridomas). For example, where the immunized animal is a mouse, one may use P3-X63/Ag8, P3-X63-Ag8.653, NS1/1.Ag 4 1, Sp210-Ag14, FO, NSO/U, MPC-11, MPC11-X45-GTG 1.7 and S194/15XX0 Bul; for rats, one may use R210.RCY3, Y3-Ag 1.2.3, IR983F and 4B210; and U-266, GM1500-GRG2, LICR-LON-HMy2 and UC729-6 are all useful in connection with cell fusions. It should be noted that the hybridomas and cell lines produced by such techniques for producing the monoclonal antibodies are contemplated to be compositions of the disclosure.

[0150] Depending on the host species, various adjuvants may be used to increase an immunological response. Such adjuvants include, but are not limited to, Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanin, and dinitrophenol. BCG (bacilli Calmette-Guerin) and Corynebacterium parvum are potentially useful human adjuvants. [0151] Alternatively, other methods, such as EBV-hybridoma methods (Haskard and Archer, J. Immunol. Methods, 74(2), 361-67 (1984), and Roder et al., Methods Enzymol., 121, 140-67 (1986)), and bacteriophage vector expression systems (see, e.g., Huse et al., Science, 246, 1275-81 (1989)) that are known in the art may be used. Further, methods of producing antibodies in non-human animals are described in, e.g., U.S. Pat. Nos. 5,545,806, 5,569,825, and 5,714,352, and U.S. Patent Application Publication No. 2002/0197266 A1).

[0152] Antibodies may also be produced by inducing in vivo production in the lymphocyte population or by screening recombinant immunoglobulin libraries or panels of highly specific binding reagents as disclosed in Orlandi et al. (Proc. Natl. Acad. Sci. 86: 3833-3837; 1989), and Winter and Milstein (Nature 349: 293-299, 1991).

[0153] Furthermore, phage display can be used to generate an antibody of the disclosure. In this regard, phage libraries encoding antigen-binding variable (V) domains of antibodies can be generated using standard molecular biology and recombinant DNA techniques (see, e.g., Sambrook et al. (eds.), Molecular Cloning, A Laboratory Manual, 3^{rd} Edition, Cold Spring Harbor Laboratory Press, New York (2001)). Phage encoding a variable region with the desired specificity are selected for specific binding to the desired antigen, and a complete or partial antibody is reconstituted comprising the selected variable domain. Nucleic acid sequences encoding the reconstituted antibody are introduced into a suitable cell line, such as a myeloma cell used for hybridoma production, such that antibodies having the characteristics of monoclonal antibodies are secreted by the cell (see, e.g., Janeway et al., supra, Huse et al., supra, and U.S. Pat. No. 6,265,150). Related methods also are described in U.S. Pat. Nos. 5,403,484; 5,571,698; 5,837, 500; and 5,702,892. The techniques described in U.S. Pat. Nos. 5,780,279; 5,821,047; 5,824,520; 5,855,885; 5,858, 657; 5,871,907; 5,969,108; 6,057,098; and 6,225,447, are also contemplated as useful in preparing antibodies according to the disclosure.

[0154] Antibodies can be produced by transgenic mice that are transgenic for specific heavy and light chain immunoglobulin genes. Such methods are known in the art and described in, for example U.S. Pat. Nos. 5,545,806 and 5,569,825, and Janeway et al., supra.

[0155] Methods for generating humanized antibodies are well known in the art and are described in detail in, for example, Janeway et al., supra, U.S. Pat. Nos. 5,225,539; 5,585,089; and 5,693,761; European Patent No. 0239400 B1; and United Kingdom Patent No. 2188638. Humanized antibodies can also be generated using the antibody resurfacing technology described in U.S. Pat. No. 5,639,641 and Pedersen et al., J. Mol. Biol., 235:959-973 (1994).

[0156] Techniques developed for the production of "chimeric antibodies," the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used (Morrison et al., Proc. Natl. Acad. Sci. 81: 6851-6855, 1984; Neuberger et al., Nature 312: 604-608, 1984; and Takeda et al., Nature 314: 452-454; 1985). Alternatively, techniques described for the production of single-chain antibodies (U.S. Pat. No. 4,946,778) can be adapted to produce IL13R α 2-specific single chain antibodies.

[0157] A preferred chimeric or humanized antibody has a human constant region, while the variable region, or at least a CDR, of the antibody is derived from a non-human species. Methods for humanizing non-human antibodies are well known in the art. (see U.S. Pat. Nos. 5,585,089, and 5,693,762). Generally, a humanized antibody has one or more amino acid residues introduced into a CDR region and/or into its framework region from a source which is non-human. Humanization can be performed, for example, using methods described in Jones et al. (Nature 321: 522-525, 1986), Riechmann et al., (Nature, 332: 323-327, 1988) and Verhoeyen et al. (Science 239:1534-1536, 1988), by substituting at least a portion of a rodent complementaritydetermining region (CDR) for the corresponding region of a human antibody. Numerous techniques for preparing engineered antibodies are described, e.g., in Owens and Young, J. Immunol. Meth., 168:149-165 (1994). Further changes can then be introduced into the antibody framework to modulate affinity or immunogenicity.

[0158] Consistent with the foregoing description, compositions comprising CDRs may be generated using, at least in part, techniques known in the art to isolate CDRs. Complementarity-determining regions are characterized by six polypeptide loops, three loops for each of the heavy or light chain variable regions. The amino acid position in a CDR is defined by Kabat et al., "Sequences of Proteins of Immunological Interest," U.S. Department of Health and Human Services, (1983), which is incorporated herein by reference. For example, hypervariable regions of human antibodies are roughly defined to be found at residues 28 to 35, from 49-59 and from residues 92-103 of the heavy and light chain variable regions [Janeway et al., supra]. The murine CDRs also are found at approximately these amino acid residues. It is understood in the art that CDR regions may be found within several amino acids of the approximated amino acid positions set forth above. An immunoglobulin variable region also consists of four "framework" regions surrounding the CDRs (FR1-4). The sequences of the framework regions of different light or heavy chains are highly conserved within a species, and are also conserved between human and murine sequences.

[0159] Compositions comprising one, two, and/or three CDRs of a heavy chain variable region or a light chain variable region of a monoclonal antibody are generated. For example, using antibody of hybridoma clone 47 comprising the CDRs having the sequences of SEO ID NOs: 1-6, polypeptide compositions comprising these CDRs are generated. Polypeptide compositions comprising one, two, three, four, five and/or six complementarity-determining regions of an antibody are also contemplated. Using the conserved framework sequences surrounding the CDRs, PCR primers complementary to these consensus framework sequences are generated to amplify the CDR sequence located between the primer regions. Techniques for cloning and expressing nucleotide and polypeptide sequences are well-established in the art [see e.g., Sambrook et al., Molecular Cloning: A Laboratory Manual, 2nd Edition, Cold Spring Harbor, N.Y. (1989)]. The amplified CDR sequences are ligated into an appropriate plasmid. The plasmid comprising one, two, three, four, five and/or six cloned CDRs optionally contains additional polypeptide encoding regions linked to the CDR.

[0160] It is contemplated that modified polypeptide compositions comprising one, two, three, four, five, or six CDRs of a heavy or light chain of SEQ ID NOs: 1-6 are generated, wherein a CDR is altered to provide increased specificity or affinity or avidity to the target IL13R α 2. Sites at locations in the CDRs are typically modified in series, e.g., by substituting first with conservative choices (e.g., hydrophobic amino acid substituted for a non-identical hydrophobic amino acid substituted for a charged amino acid), and then with more dissimilar choices (e.g., hydrophobic amino acid, and then deletions or insertions may be made at the target site.

[0161] Framework regions (FR) of a murine antibody are humanized by substituting compatible human framework regions chosen from a large database of human antibody variable sequences, including over twelve hundred human V_H sequences and over one thousand V_L sequences. The database of antibody sequences used for comparison is downloaded from Andrew C. R. Martin's KabatMan web page (http://www.rubic.rdg.ac.uk/abs/). The Kabat method for identifying CDRs provides a means for delineating the approximate CDR and framework regions of any human antibody and comparing the sequence of a murine antibody for similarity to determine the CDRs and FRs. Best matched human V_H and V_L sequences are chosen on the basis of high overall framework matching, similar CDR length, and minimal mismatching of canonical and V_H/V_L contact residues. Human framework regions most similar to the murine sequence are inserted between the murine CDRs. Alternatively, the murine framework region may be modified by making amino acid substitutions of all or part of the native framework region that more closely resemble a framework region of a human antibody.

[0162] "Conservative" amino acid substitutions are made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues involved. For example, nonpolar (hydrophobic) amino acids include alanine (Ala, A), leucine (Leu, L), isoleucine (Ile, I), valine (Val, V), proline (Pro, P), phenylalanine (Phe, F), tryptophan (Trp, W), and methionine (Met, M); polar neutral amino acids include glycine (Gly, G), serine (Ser, S), threonine (Thr, T), cysteine (Cys, C), tyrosine (Tyr, Y), asparagine (Asn, N), and glutamine (Gln, Q); positively charged (basic) amino acids include arginine (Arg, R), lysine (Lys, K), and histidine (His, H); and negatively charged (acidic) amino acids include aspartic acid (Asp, D) and glutamic acid (Glu, E). "Insertions" or "deletions" are preferably in the range of about 1 to 20 amino acids, more preferably 1 to 10 amino acids. The variation may be introduced by systematically making substitutions of amino acids in a polypeptide molecule using recombinant DNA techniques and assaying the resulting recombinant variants for activity. Nucleic acid alterations can be made at sites that differ in the nucleic acids from different species (variable positions) or in highly conserved regions (constant regions). Methods for expressing polypeptide compositions useful in the invention are described in greater detail below.

[0163] Additionally, another useful technique for generating antibodies for use in the methods of the disclosure may be one which uses a rational design-type approach. The goal of rational design is to produce structural analogs of biologically active polypeptides or compounds with which they interact (agonists, antagonists, inhibitors, peptidomimetics, binding partners, and the like). In this case, the active polypeptides comprise the sequences of SEQ ID NOs: 1-6 disclosed herein. By creating such analogs, it is possible to fashion additional antibodies which are more immunoreactive than the native or natural molecule. In one approach, one would generate a three-dimensional structure for the antibodies or an epitope binding fragment thereof. This could be accomplished by x-ray crystallography, computer modeling or by a combination of both approaches. An alternative approach, "alanine scan," involves the random replacement of residues throughout a molecule with alanine, and the resulting effect on function is determined.

[0164] It also is possible to solve the crystal structure of the specific antibodies. In principle, this approach yields a pharmacore upon which subsequent drug design can be based. It is possible to bypass protein crystallography altogether by generating anti-idiotypic antibodies to a functional, pharmacologically active antibody. As a mirror image of a mirror image, the binding site of anti-idiotype antibody is expected to be an analog of the original antigen. The anti-idiotype antibody is then be used to identify and isolate additional antibodies from banks of chemically- or biologically-produced peptides.

[0165] Chemically synthesized bispecific antibodies may be prepared by chemically crosslinking heterologous Fab or $F(ab')_2$ fragments by means of chemicals such as heterobifunctional reagent succinimidyl-3-(2-pyridyldithiol)-propionate (SPDP, Pierce Chemicals, Rockford, Ill.). The Fab and $F(ab')_2$ fragments can be obtained from intact antibody by digesting it with papain or pepsin, respectively (Karpovsky et al., J. Exp. Med. 160:1686-701, 1984; Titus et al., J. Immunol., 138:4018-22, 1987).

[0166] Methods of testing antibodies for the ability to bind to the epitope of the IL13R α 2, regardless of how the antibodies are produced, are known in the art and include any antibody-antigen binding assay such as, for example, radioimmunoassay (RIA), ELISA, Western blot, immuno-precipitation, and competitive inhibition assays (see, e.g., Janeway et al., infra, and U.S. Patent Application Publication No. 2002/0197266 A1).

[0167] Selection of antibodies from an antibody population for purposes herein also include using blood vessel endothelial cells to "subtract" those antibodies that crossreact with epitopes on such cells other than IL13R α 2 epitopes. The remaining antibody population is enriched in antibodies preferential for IL13R α 2 epitopes.

[0168] Aptamers

[0169] Recent advances in the field of combinatorial sciences have identified short polymer sequences (e.g., oligonucleic acid or peptide molecules) with high affinity and specificity to a given target. For example, SELEX technology has been used to identify DNA and RNA aptamers with binding properties that rival mammalian antibodies, the field of immunology has generated and isolated antibodies or antibody fragments which bind to a myriad of compounds, and phage display has been utilized to discover new peptide sequences with very favorable binding properties. Based on the success of these molecular evolution techniques, it is certain that molecules can be created which bind to any target molecule. A loop structure is often involved with providing the desired binding attributes as in the case of aptamers, which often utilize hairpin loops created from short regions without complementary base pairing, naturally derived antibodies that utilize combinatorial arrangement of looped hyper-variable regions and new phage-display libraries utilizing cyclic peptides that have shown improved results when compared to linear peptide phage display results. Thus, sufficient evidence has been generated to indicate that high affinity ligands can be created and identified by combinatorial molecular evolution techniques. For the present disclosure, molecular evolution techniques can be used to isolate binding agents specific for the IL13R α 2 disclosed herein. For more on aptamers, see generally, Gold, L., Singer, B., He, Y. Y., Brody. E., "Aptamers As Therapeutic And Diagnostic Agents," J. Biotechnol. 74:5-13 (2000). Relevant techniques for generating aptamers are found in U.S. Pat. No. 6,699,843, which is incorporated herein by reference in its entirety.

[0170] In some embodiments, the aptamer is generated by preparing a library of nucleic acids; contacting the library of nucleic acids with a growth factor, wherein nucleic acids having greater binding affinity for the growth factor (relative to other library nucleic acids) are selected and amplified to yield a mixture of nucleic acids enriched for nucleic acids with relatively higher affinity and specificity for binding to the growth factor. The processes may be repeated, and the selected nucleic acids mutated and rescreened, whereby a growth factor aptamer is identified. Nucleic acids may be screened to select for molecules that bind to more than target. Binding more than one target can refer to binding more than one simultaneously or competitively. In some embodiments, a binding agent comprises at least one

aptamer, wherein a first binding unit binds a first epitope of an IL13R α 2 and a second binding unit binds a second epitope of the IL13R α 2.

[0171] With regard to the binding agents of the compositions of the disclosure, ligand-induced activation of the IL13R α 2 is reduced upon binding of the binding agent to the IL13R α 2. As used herein, the term "reduce" as well as like terms, e.g., "inhibit," do not necessarily imply 100% or a complete reduction or inhibition. Rather, there are varying degrees of reduction or inhibition of which one of ordinary skill in the art recognizes as having a potential benefit or therapeutic effect. Accordingly, in some embodiments, ligand-induced activation of the IL13R α 2 is completely abolished. In some embodiments, ligand-induced activation is substantially reduced, e.g., reduced by about 10% (e.g., by about 20%, by about 30%, by about 40%, by about 50%, by about 60%, by about 70%, by about 80%, by about 90%) or more, as compared to ligand-induced activation of the IL13R α 2 when the binding agent is absent or not bound to the IL13Ra2. Methods of measuring ligand-induced activation of an IL13R α 2 are known in the art, and include, for example, the assays described in the Examples, below.

[0172] Conjugates

[0173] Conjugates comprising a targeting domain and an effector domain are disclosed herein. In exemplary embodiments, the conjugate comprises any one of the binding agents disclosed herein as the targeting domain to localize the conjugate to a cell expressing IL13R α 2, e.g., a tumor cell expressing the same, and an effector domain. In exemplary aspects, the conjugate is a fusion protein. In exemplary aspects, the conjugate is a chimeric protein. As used herein, the term "chimeric" refers to a molecule composed of parts of different origins. A chimeric molecule, as a whole, is non-naturally occurring, e.g., synthetic or recombinant, although the parts which comprise the chimeric molecule may be naturally occurring.

[0174] Exemplary Effector Domains

[0175] As used herein, the term "effector domain" refers to a portion of a conjugate that effects a desired biological function. In exemplary aspects, the effector domain identifies or locates IL13R α 2-expressing cells. For example, the effector domain may be a diagnostic agent, e.g., a radiolabel, a fluorescent label, an enzyme (e.g., that catalyzes a calorimetric or fluorometric reaction), a substrate, a solid matrix, or a carrier (e.g., biotin or avidin). The diagnostic agent in some aspects is an imaging agent. Many appropriate imaging agents are known in the art, as are methods of attaching the labeling agents to the peptides of the invention (see, e.g., U.S. Pat. Nos. 4,965,392; 4,472,509; 5,021,236; and 5,037, 630; each incorporated herein by reference). The imaging agents are administered to a subject in a pharmaceutically acceptable carrier, and allowed to accumulate at a target site having the lymphatic endothelial cells. This imaging agent then serves as a contrast reagent for X-ray, magnetic resonance, positron emission tomography, single photon emission computed tomography (SPECT), or sonographic or scintigraphic imaging of the target site. Of course, it should be understood that the imaging may be performed in vitro where tissue from the subject is obtained through a biopsy, and the presence of lymphatic endothelial cells is determined with the aid of the imaging agents described herein in combination with histochemical techniques for preparing and fixing tissues. Paramagnetic ions useful in the imaging agents of the invention include for example chromium (III),

manganese (II), iron (III), iron (II), cobalt (II), nickel (II) copper (II), neodymium (III), samarium (III), ytterbium(III), gadolinium (III), vanadium (II), terbium (III), dysprosium (III), holmium (III) and erbium (III). Ions useful for X-ray imaging include, but are not limited to, lanthanum (III), gold (III), lead (II) and particularly bismuth (III). Radioisotopes for diagnostic applications include for example, ²¹¹astatine, ¹⁴carbon, ⁵¹chromium, ³⁶chlorine, ⁵⁷cobalt, ⁶⁷copper, ¹⁵²europium, ⁶⁷gallium, ³hydrogen, ¹²³iodine, ¹²⁵iodine, ¹¹¹indium, ⁵⁹iron, ³²phosphorus, ¹⁸⁶rhenium, ⁷⁵selenium, ³⁵sulphur, ⁹⁹mtechnicium, ⁹⁰yttrium, and ⁹⁹zirconium.

[0176] The effector domain may be one which alters the physico-chemical characteristics of the conjugate, e.g., an effector which confers increased solubility and/or stability and/or half-life, resistance to proteolytic cleavage, modulation of clearance. In exemplary aspects, the effector domain is a polymer, a carbohydrate, or a lipid.

[0177] The polymer may be branched or unbranched. The polymer may be of any molecular weight. The polymer in some embodiments has an average molecular weight of between about 2 kDa to about 100 kDa (the term "about" indicating that in preparations of a water-soluble polymer, some molecules will weigh more, some less, than the stated molecular weight). The average molecular weight of the polymer is in some aspects between about 5 kDa and about 50 kDa, between about 12 kDa to about 40 kDa or between about 20 kDa to about 35 kDa. In some embodiments, the polymer is modified to have a single reactive group, such as an active ester for acylation or an aldehyde for alkylation, so that the degree of polymerization may be controlled. The polymer in some embodiments is water soluble so that the protein to which it is attached does not precipitate in an aqueous environment, such as a physiological environment. In some embodiments when, for example, the composition is used for therapeutic use, the polymer is pharmaceutically acceptable. Additionally, in some aspects, the polymer is a mixture of polymers, e.g., a co-polymer, a block co-polymer. In some embodiments, the polymer is selected from the group consisting of: polyamides, polycarbonates, polyalkylenes and derivatives thereof, including polyalkylene glycols, polyalkylene oxides, polyalkylene terepthalates, polymers of acrylic and methacrylic esters, including poly (methyl methacrylate), poly(ethyl methacrylate), poly (butylmethacrylate), poly(isobutyl methacrylate), poly (hexylmethacrylate), poly(isodecyl methacrylate), poly (lauryl methacrylate), poly(phenyl methacrylate), poly (methyl acrylate), poly(isopropyl acrylate), poly(isobutyl acrylate), and poly(octadecyl acrylate), polyvinyl polymers including polyvinyl alcohols, polyvinyl ethers, polyvinyl esters, polyvinyl halides, poly(vinyl acetate), and polyvinylpyrrolidone, polyglycolides, polysiloxanes, polyurethanes and co-polymers thereof, celluloses including alkyl cellulose, hydroxyalkyl celluloses, cellulose ethers, cellulose esters, nitro celluloses, methyl cellulose, ethyl cellulose, hydroxypropyl cellulose, hydroxy-propyl methyl cellulose, hydroxybutyl methyl cellulose, cellulose acetate, cellulose propionate, cellulose acetate butyrate, cellulose acetate phthalate, carboxylethyl cellulose, cellulose triacetate, and cellulose sulphate sodium salt, polypropylene, polyethylenes including poly(ethylene glycol), poly(ethylene oxide), and poly(ethylene terephthalate), and polystyrene. In some aspects, the polymer is a biodegradable polymer, including a synthetic biodegradable polymer (e.g., polymers of lactic acid and glycolic acid, polyanhydrides, poly(ortho)esters, polyurethanes, poly(butic acid), poly(valeric acid), and poly (lactide-cocaprolactone)), and a natural biodegradable polymer (e.g., alginate and other polysaccharides including dextran and cellulose, collagen, chemical derivatives thereof (substitutions, additions of chemical groups, for example, alkyl, alkylene, hydroxylations, oxidations, and other modifications routinely made by those skilled in the art), albumin and other hydrophilic proteins (e.g., zein and other prolamines and hydrophobic proteins)), as well as any copolymer or mixture thereof. In general, these materials degrade either by enzymatic hydrolysis or exposure to water in vivo, by surface or bulk erosion. In some aspects, the polymer is a bioadhesive polymer, such as a bioerodible hydrogel described by H. S. Sawhney, C. P. Pathak and J. A. Hubbell in Macromolecules, 1993, 26, 581-587, the teachings of which are incorporated herein, polyhyaluronic acids, casein, gelatin, glutin, polyanhydrides, polyacrylic acid, alginate, chitosan, poly(methyl methacrylates), poly(ethyl methacrylates), poly(butylmethacrylate), poly(isobutyl methacrylate), poly(hexylmethacrylate), poly(isodecyl methacrylate), poly(lauryl methacrylate), poly(phenyl methacrylate), poly (methyl acrylate), poly(isopropyl acrylate), poly(isobutyl acrylate), and poly(octadecyl acrylate). In some embodiments, the polymer is a water-soluble polymer or a hydrophilic polymer. Suitable water-soluble polymers are known in the art and include, for example, polyvinylpyrrolidone, hydroxypropyl cellulose (HPC; Klucel), hydroxypropyl methylcellulose (HPMC; Methocel), nitrocellulose, hydroxypropyl ethylcellulose, hydroxypropyl butylcellulose, hydroxypropyl pentylcellulose, methyl cellulose, ethvlcellulose (Ethocel), hydroxyethyl cellulose, various alkyl celluloses and hydroxyalkyl celluloses, various cellulose ethers, cellulose acetate, carboxymethyl cellulose, sodium carboxymethyl cellulose, calcium carboxymethyl cellulose, vinyl acetate/crotonic acid copolymers, poly-hydroxyalkyl methacrylate, hydroxymethyl methacrylate, methacrylic acid copolymers, polymethacrylic acid, polymethylmethacrylate, maleic anhydride/methyl vinyl ether copolymers, poly vinyl alcohol, sodium and calcium polyacrylic acid, polyacrylic acid, acidic carboxy polymers, carboxypolymethylene, carboxyvinyl polymers, polyoxyethylene polyoxypropylene copolymer, polymethylvinylether co-maleic anhydride, carboxymethylamide, potassium methacrylate divinylbenzene co-polymer, polyoxyethyleneglycols, polyethylene oxide, and derivatives, salts, and combinations thereof. In some aspects, the water-soluble polymers or mixtures thereof include, but are not limited to, N-linked or O-linked carbohydrates, sugars, phosphates, carbohydrates; sugars; phosphates; polyethylene glycol (PEG) (including the forms of PEG that have been used to derivatize proteins, including mono-(C1-C 10) alkoxy- or aryloxy-polyethylene glycol); monomethoxy-polyethylene glycol; dextran (such as low molecular weight dextran of, for example, about 6 kD), cellulose; other carbohydrate-based polymers, poly-(Nvinyl pyrrolidone), polyethylene glycol, propylene glycol homopolymers, a polypropylene oxide/ethylene oxide copolymer, polyoxyethylated polyols (e.g., glycerol) and polyvinyl alcohol. Also encompassed by the disclosure are bifunctional crosslinking molecules which may be used to prepare covalently attached multimers. A particularly preferred water-soluble polymer for use herein is polyethylene glycol (PEG). As used herein, polyethylene glycol is meant to encompass any of the forms of PEG that can be used to derivatize other proteins, such as mono-(C1-C10) alkoxy- or

aryloxy-polyethylene glycol. PEG is a linear or branched neutral polyether, available in a broad range of molecular weights, and is soluble in water and most organic solvents. PEG is effective at excluding other polymers or peptides when present in water, primarily through its high dynamic chain mobility and hydrophobic nature, thus creating a water shell or hydration sphere when attached to other proteins or polymer surfaces. PEG is nontoxic, non-immunogenic, and approved by the Food and Drug Administration for internal consumption. Proteins or enzymes when conjugated to PEG have demonstrated bioactivity, non-antigenic properties, and decreased clearance rates when administered in animals. F. M. Veronese et al., Preparation and Properties of Monomethoxypoly(ethylene glycol)-modified Enzymes for Therapeutic Applications, in J. M. Harris ed., Poly(Ethylene Glycol) Chemistry-Biotechnical and Biomedical Applications, 127-36, 1992, incorporated herein by reference. Without wishing to be bound by theory, these phenomena may be due to the exclusion properties of PEG in preventing recognition by the immune system. In addition, PEG has been widely used in surface modification procedures to decrease protein adsorption and improve blood compatibility. S. W. Kim et al., Ann. N.Y. Acad. Sci. 516: 116-30 1987; Jacobs et al., Artif. Organs 12: 500-501, 1988; Park et al., J. Poly. Sci, Part A 29:1725-31, 1991, each incorporated herein by reference in its entirety. Hydrophobic polymer surfaces, such as polyurethanes and polystyrene, can be modified by the grafting of PEG (MW 3,400) and employed as nonthrombogenic surfaces. Surface properties (contact angle) can be more consistent with hydrophilic surfaces, due to the hydrating effect of PEG. More importantly, protein (albumin and other plasma proteins) adsorption can be greatly reduced, resulting from the high chain motility, hydration sphere, and protein exclusion properties of PEG. PEG (MW 3,400) was determined as an optimal size in surface immobilization studies, Park et al., J. Biomed. Mat. Res. 26:739-45, 1992, while PEG (MW 5,000) was most beneficial in decreasing protein antigenicity. F. M. Veronese et al., In J. M. Harris, et al., Poly(Ethylene Glycol) Chemistry-Biotechnical and Biomedical Applications, 127-36. Methods for preparing pegylated binding agent polypeptides may comprise the steps of (a) reacting the polypeptide with polyethvlene glycol (such as a reactive ester or aldehyde derivative of PEG) under conditions whereby the binding agent polypeptide becomes attached to one or more PEG groups, and (b) obtaining the reaction product(s). In general, the optimal reaction conditions for the acylation reactions will be determined based on known parameters and the desired result. For example, the larger the ratio of PEG:protein, the greater the percentage of poly-pegylated product. In some embodiments, the binding agent will have a single PEG moiety at the N-terminus. See U.S. Pat. No. 8,234,784, incorporated by reference herein.

[0178] In some embodiments, the effector domain is a carbohydrate. In some embodiments, the carbohydrate is a monosaccharide (e.g., glucose, galactose, fructose), a disaccharide (e.g., sucrose, lactose, maltose), an oligosaccharide (e.g., raffinose, stachyose), a polysaccharide (e.g., a starch, amylase, amylopectin, cellulose, chitin, callose, laminarin, xylan, mannan, fucoidan, or galactomannan).

[0179] In some embodiments, the effector domain is a lipid. The lipid, in some embodiments, is a fatty acid, eicosanoid, prostaglandin, leukotriene, thromboxane, N-acyl ethanolamine, glycerolipid (e.g., mono-, di-, tri-

substituted glycerols), glycerophospholipid (e.g., phosphatidylcholine, phosphatidylinositol, phosphatidylethanolamine, phosphatidylserine), sphingolipid (e.g., sphingosine, ceramide), sterol lipid (e.g., steroid, cholesterol), prenol lipid, saccharolipid, or a polyketide, oil, wax, cholesterol, sterol, fat-soluble vitamin, monoglyceride, diglyceride, triglyceride, or a phospholipid.

[0180] Lethal Domains

[0181] In exemplary aspects, the effector domain is a lethal domain that confers lethality, such that when the conjugate is localized to a cell expressing IL13R α 2, e.g., a tumor cell expressing the same. The effector domain confers upon the conjugate the ability to kill an IL13R α 2-expressing cell once the binding agent has found and bound to its IL13R α 2 target.

[0182] In exemplary aspects, the effector domain is a cytotoxin (also referred to herein as a "cytotoxic agent"). The cytotoxic agent is any molecule (chemical or biochemical) which is toxic to a cell. In some embodiments, the cytotoxic agent is a chemotherapeutic agent. Chemotherapeutic agents are known in the art and include, but are not limited to, platinum coordination compounds, topoisomerase inhibitors, antibiotics, antimitotic alkaloids and difluoronucleosides, as described in U.S. Pat. No. 6,630,124. In some embodiments, the chemotherapeutic agent is a platinum coordination compound. The term "platinum coordination compound" refers to any tumor cell growth-inhibiting platinum coordination compound that provides the platinum in the form of an ion. In some embodiments, the platinum coordination compound is cis-diamminediaquoplatinum (II)-ion; chloro(diethylenetriamine)-platinum(II) chloride; dichloro(ethylenediamine)-platinum(II), diammine(1,1-cyclobutanedicarboxylato) platinum(II) (carboplatin); spiroplatin; iproplatin; diammine(2-ethylmaethylenediaminemalonatoplatinum lonato)-platinum(II); aqua(1,2-diaminodyclohexane)-sulfatoplatinum(II); (II): (1,2-diaminocyclohexane)malonatoplatinum(II); (4-caroxyphthalato)(1,2-diaminocyclohexane)platinum(II); (1,2diaminocyclohexane)-(isocitrato)platinum(II); (1,2-diaminocyclohexane)cis(pyruvato)platinum(II); (1,2diaminocyclohexane)oxalatoplatinum(II); ormaplatin; or tetraplatin. In some embodiments, cisplatin is the platinum coordination compound employed in the compositions and methods of the present invention. Cisplatin is commercially available under the name PLATINOLTM from Bristol Myers-Squibb Corporation and is available as a powder for constitution with water, sterile saline or other suitable vehicle. Other platinum coordination compounds suitable for use in the present invention are known and are available commercially and/or can be prepared by conventional techniques. Cisplatin, or cis-dichlorodiammineplatinum II, has been used successfully for many years as a chemotherapeutic agent in the treatment of various human solid malignant tumors. More recently, other diamino-platinum complexes have also shown efficacy as chemotherapeutic agents in the treatment of various human, solid, malignant tumors. Such diamino-platinum complexes include, but are not limited to, spiroplatinum and carboplatinum. Although cisplatin and other diamino-platinum complexes have been widely used as chemotherapeutic agents in humans, they have had to be delivered at high dosage levels that can lead to toxicity problems such as kidney damage.

[0183] In some embodiments, the chemotherapeutic agent is a topoisomerase inhibitor. Topoisomerases are enzymes

that are capable of altering DNA topology in eukaryotic cells. They are critical for cellular functions and cell proliferation. Generally, there are two classes of topoisomerases in eukaryotic cells, type I and type II. Topoisomerase I is a monomeric enzyme of approximately 100,000 molecular weight. The enzyme binds to DNA and introduces a transient single-strand break, unwinds the double helix (or allows it to unwind), and subsequently reseals the break before dissociating from the DNA strand. Various topoisomerase inhibitors have recently shown clinical efficacy in the treatment of humans afflicted with ovarian cancer, esophageal cancer or non-small cell lung carcinoma. In some aspects, the topoisomerase inhibitor is camptothecin or a camptothecin analog. Camptothecin is a water-insoluble, cytotoxic alkaloid produced by Camptotheca accuminata trees indigenous to China and Nothapodytes foetida trees indigenous to India. Camptothecin exhibits tumor cell growth-inhibiting activity against a number of tumor cells. Compounds of the camptothecin analog class are typically specific inhibitors of DNA topoisomerase I. By the term "inhibitor of topoisomerase" is meant any tumor cell growth-inhibiting compound that is structurally related to camptothecin. Compounds of the camptothecin analog class include, but are not limited to; topotecan, irinotecan and 9-amino-camptothecin. In additional embodiments, the cytotoxic agent is any tumor cell growth-inhibiting camptothecin analog claimed or described in U.S. Pat. No. 5,004,758; European Patent Application Number 88311366.4 (Publication Number EP 0 321 122); U.S. Pat. No. 4,604,463; European Patent Application Publication Number EP 0 137 145; U.S. Pat. No. 4,473,692; European Patent Application Publication Number EP 0 074 256; U.S. Pat. No. 4,545,880; European Patent Application Publication Number EP 0 074 256; European Patent Application Publication Number EP 0 088 642; Wani et al., J. Med. Chem., 29, 2358-2363 (1986); and Nitta et al., Proc. 14th International Congr. Chemotherapy, Kyoto, 1985, Tokyo Press, Anticancer Section 1, p. 28-30. In particular, the disclosure contemplates a compound called CPT-11. CPT-11 is a camptothecin analog with a 4-(piperidino)piperidine side chain joined through a carbamate linkage at C-10 of 10-hydroxy-7-ethyl camptothecin. CPT-11 is currently undergoing human clinical trials and is also referred to as irinotecan; Wani et al, J. Med. Chem., 23, 554 (1980); Wani et al., J. Med. Chem., 30, 1774 (1987); U.S. Pat. No. 4,342,776; European Patent Application Publication Number EP 418 099; U.S. Pat. No. 4,513,138; European Patent Application Publication Number EP 0 074 770; U.S. Pat. No. 4,399,276; European Patent Application Publication Number 0 056 692; the entire disclosure of each of which is hereby incorporated by reference. All of the above-listed compounds of the camptothecin analog class are available commercially and/or can be prepared by conventional techniques including those described in the above-listed references. The topoisomerase inhibitor may be selected from the group consisting of topotecan, irinotecan and 9-aminocamptothecin.

[0184] The preparation of numerous compounds of the camptothecin analog class (including pharmaceutically acceptable salts, hydrates and solvates thereof) as well as the preparation of oral and parenteral pharmaceutical compositions comprising such a compound of the camptothecin analog class and an inert, pharmaceutically acceptable carrier or diluent, is extensively described in U.S. Pat. No. 5,004,758; and European Patent Application Number

88311366.4(Publication Number EP 0 321 122), the teachings of each of which are incorporated herein by reference in its entirety.

[0185] In still another embodiment of the invention, the chemotherapeutic agent is an antibiotic compound. Suitable antibiotics include, but are not limited to, doxorubicin, mitomycin, bleomycin, daunorubicin and streptozocin. In some embodiments, the chemotherapeutic agent is an antimitotic alkaloid. In general, antimitotic alkaloids can be extracted from Cantharanthus roseus, and have been shown to be efficacious as anticancer chemotherapy agents. A great number of semi-synthetic derivatives have been studied both chemically and pharmacologically (see, O. Van Tellingen et al, Anticancer Research, 12, 1699-1716 (1992)). The antimitotic alkaloids of the present invention include, but are not limited to, vinblastine, vincristine, vindesine, Taxol and vinorelbine. The latter two antimitotic alkaloids are commercially available from Eli Lilly and Company, and Pierre Fabre Laboratories, respectively (see, U.S. Pat. No. 5,620, 985). In one aspect of the disclosure, the antimitotic alkaloid is vinorelbine.

[0186] In another embodiment of the invention, the chemotherapeutic agent is a diffuoronucleoside. 2'-deoxy-2',2'diffuoronucleosides are known in the art as having antiviral activity. Such compounds are disclosed and taught in U.S. Pat. Nos. 4,526,988 and 4,808,614. European Patent Application Publication 184,365 discloses that these same diffuoronucleosides have oncolytic activity. In certain specific aspects, the 2'-deoxy-2',2'-diffuoronucleoside used in the compositions and methods of the disclosure is 2'-deoxy-2', 2'-diffuorocytidine hydrochloride, also known as gemcitabine hydrochloride. Gemcitabine is commercially available or can be synthesized in a multi-step process as disclosed in U.S. Pat. Nos. 4,526,988, 4,808,614 and 5,223,608, the teachings of each of which are incorporated herein by reference in its entirety.

[0187] In exemplary aspects, the effector domain is an apoptosis tag which causes the IL13R α 2-expressing cell to apoptose. In exemplary aspects, the apoptosis tag is a TRAIL protein, or a portion thereof. In exemplary aspects, the apoptosis tag comprises the amino acid sequence of SEQ ID NO: 27. In exemplary aspects, the conjugate comprises the amino acid sequence of SEQ ID NO: 25.

[0188] In exemplary embodiments, the effector domain is an Fc domain of IgG or other immunoglobulin. For substituents such as an Fc region of human IgG, the fusion can be fused directly to a binding agent or fused through an intervening sequence. For example, a human IgG hinge, CH2 and CH3 region may be fused at either the N-terminus or C-terminus of a binding agent to attach the Fc region. The resulting Fc-fusion agent enables purification via a Protein A affinity column (Pierce, Rockford, Ill.). Peptide and proteins fused to an Fc region can exhibit a substantially greater half-life in vivo than the unfused counterpart. A fusion to an Fc region allows for dimerization/multimerization of the fusion polypeptide. The Fc region may be a naturally occurring Fc region, or may be modified for superior characteristics, e.g., therapeutic qualities, circulation time, reduced aggregation. As noted above, in some embodiments, the binding agent are conjugated, e.g., fused to an immunoglobulin or portion thereof (e.g., variable region, CDR, or Fc region). Known types of immunoglobulins (Ig) include IgG, IgA, IgE, IgD or IgM. The Fc region is a C-terminal region of an Ig heavy chain, which is responsible for binding

to Fc receptors that carry out activities such as recycling (which results in prolonged half-life), antibody dependent cell-mediated cytotoxicity (ADCC), and complement dependent cytotoxicity (CDC).

[0189] For example, according to some definitions the human IgG heavy chain Fc region stretches from Cys226 to the C-terminus of the heavy chain. The "hinge region" generally extends from Glu216 to Pro230 of human IgG1 (hinge regions of other IgG isotypes may be aligned with the IgG1 sequence by aligning the cysteines involved in cysteine bonding). The Fc region of an IgG includes two constant domains, CH2 and CH3. The CH2 domain of a human IgG Fc region usually extends from amino acids 231 to amino acid 341. The CH3 domain of a human IgG Fc region usually extends from amino acids 342 to 447. References made to amino acid numbering of immunoglobulins or immunoglobulin fragments, or regions, are all based on Kabat et al. 1991. Sequences of Proteins of Immunological Interest, U.S. Department of Public Health, Bethesda, Md., incorporated herein by reference. In related embodiments, the Fc region may comprise one or more native or modified constant regions from an immunoglobulin heavy chain, other than CH1, for example, the CH2 and CH3 regions of IgG and IgA, or the CH3 and CH4 regions of IgE.

[0190] Suitable conjugate moieties include portions of immunoglobulin sequence that include the FcRn binding site. FcRn, a salvage receptor, is responsible for recycling immunoglobulins and returning them to circulation in the blood. The region of the Fc portion of IgG that binds to the FcRn receptor has been described based on X-ray crystallography (Burmeister et al. 1994, Nature 372:379). The major contact area of the Fc with the FcRn is near the junction of the CH2 and CH3 domains. Fc-FcRn contacts are all within a single Ig heavy chain. The major contact sites include amino acid residues 248, 250-257, 272, 285, 288, 290-291, 308-311, and 314 of the CH2 domain and amino acid residues 385-387, 428, and 433-436 of the CH3 domain.

[0191] Some conjugate moieties may or may not include Fc γ R binding site(s). Fc γ R are responsible for antibodydependent cell-mediated cytotoxicity (ADCC) and complement-dependent cytotoxicity (CDC). Examples of positions within the Fc region that make a direct contact with Fc γ R are amino acids 234-239 (lower hinge region), amino acids 265-269 (B/C loop), amino acids 297-299 (C/E loop), and amino acids 327-332 (F/G) loop (Sondermann et al., Nature 406: 267-273, 2000). The lower hinge region of IgE has also been implicated in the FcRI binding (Henry, et al., Biochemistry 36, 15568-15578, 1997). Residues involved in IgA receptor binding are described in Lewis et al., (J Immunol. 175:6694-701, 2005). Amino acid residues involved in IgE receptor binding are described in Sayers et al. (J Biol Chem. 279(34):35320-5, 2004).

[0192] Amino acid modifications may be made to the Fc region of an immunoglobulin. Such variant Fc regions comprise at least one amino acid modification in the CH3 domain of the Fc region (residues 342-447) and/or at least one amino acid modification in the CH2 domain of the Fc region (residues 231-341). Mutations believed to impart an increased affinity for FcRn include T256A, T307A, E380A, and N434A (Shields et al. 2001, J. Biol. Chem. 276:6591). Other mutations may reduce binding of the Fc region to FcγRI, FcγRIIA, FcγRIIB, and/or FcγRIIA without significantly reducing affinity for FcRn. For example, substitution

of the Asn at position 297 of the Fc region with Ala or another amino acid removes a highly conserved N-glycosylation site and may result in reduced immunogenicity with concomitant prolonged half-life of the Fc region, as well as reduced binding to Fc γ Rs (Routledge et al. 1995, Transplantation 60:847; Friend et al. 1999, Transplantation 68:1632; Shields et al. 1995, J. Biol. Chem. 276:6591). Amino acid modifications at positions 233-236 of IgG1 have been made that reduce binding to Fc γ Rs (Ward and Ghetie 1995, Therapeutic Immunology 2:77 and Armour et al. 1999, Eur. J. Immunol. 29:2613). Some exemplary amino acid substitutions are described in U.S. Pat. Nos. 7,355,008 and 7,381, 408, each of which is incorporated by reference herein in its entirety.

[0193] In some embodiments, the binding agent is fused to alkaline phosphatase (AP). Methods for making Fc or AP fusion agents are provided in WO 02/060950.

[0194] Chimeric Antigen Receptors (CARs)

[0195] In exemplary aspects, the effector domain is a T-cell signaling domain. In exemplary aspects, the conjugate is a chimeric antigen receptor (CAR). Chimeric antigen receptors (CARs) are engineered transmembrane proteins that combine the specificity of an antigen-specific antibody with a T-cell receptor's function. In general, CARs comprise an ectodomain, a spacer region, a transmembrane domain, and an endodomain. The ectodomain of a CAR in exemplary aspects comprises an antigen recognition region, which may be an scFV of an antigen-specific antibody. The ectodomain also in some embodiments comprises a signal peptide which directs the nascent protein into the endoplasmic reticulum. In exemplary aspects, the ectodomain comprises a spacer which links the antigen recognition region to the transmembrane domain. The transmembrane (TM) domain is the portion of the CAR which traverses the cell membrane. In exemplary aspects, the TM domain comprises a hydrophobic alpha helix. In exemplary aspects, the TM domain comprises all or a portion of the TM domain of CD28. In exemplary aspects, the TM domain comprises all or a portion of the TM domain of $CD8\alpha$. The endodomain of a CAR comprises one or more signaling domains. In exemplary aspects, the endodomain comprises the zeta chain of CD3, which comprises three copies of the Immunoreceptor Tyrosine-based Activation Motif (ITAM). An ITAM generally comprises a Tyr residue separated by two amino acids from a Leu or Ile. In the case of immune cell receptors, e.g., the T cell receptor and the B cell receptor, the ITAMs occur in multiples (at least two) and each ITAM is separated from another by 6-8 amino acids. The endodomain of CARs may also comprises additional signaling domains, e.g., portions of proteins that are important for downstream signal transduction. In exemplary aspects, the endodomain comprises signaling domains from one or more of CD28, 41BB or 4-1BB (CD137), ICOS, CD27, CD40, OX40 (CD134), or Myd88. Sequences encoding signaling domains of such proteins are provided herein as SEQ ID NOs: 39-42, 68-79, 81, and 83. Methods of making CARs, expressing them in cells, e.g., T-cells, and utilizing the CAR-expressing T-cells in therapy, are known in the art. See, e.g., International Patent Application Publication Nos. WO2014/208760, WO2014/190273, WO2014/186469, WO2014/184143, WO2014180306, WO2014/179759, WO2014/153270, U.S. Application Publication Nos. US20140369977, US20140322212, US20140322275, US20140322183, US20140301993, US20140286973, US20140271582,

US20140271635, US20140274909, European Application Publication No. 2814846, each of which are incorporated by reference in their entirety.

[0196] In exemplary aspects, the conjugate of the disclosure is an IL13Ra2-specific chimeric antigen receptor (CAR) comprising a binding agent described herein, a hinge region, and an endodomain comprising a signaling domain of a CD3 zeta chain and a signaling domain of CD28, CD134, and/or CD137. In exemplary aspects, the CAR comprises (A) each of the amino acid sequence of: NYLMN (SEQ ID NO: 1); RIDPYDGDIDYNQNFKD (SEQ ID NO: 2); GYGTAYGVDY (SEQ ID NO: 3); RASESVDNYG-ISFMN (SEQ ID NO: 4); AASRQGSG (SEQ ID NO: 5); and QQSKEVPWT (SEQ ID NO: 6), (B) a hinge region; and (C) an endodomain comprising a signaling domain of a CD3 zeta chain and a signaling domain of CD28, CD134, and/or CD137. In exemplary aspects, the CD3 zeta chain signaling domain comprises the amino acid sequence of SEQ ID NO: 41. In exemplary aspects, the CAR further comprises a transmembrane (TM) domain based on the TM domain of CD28. In exemplary aspects, the CAR comprises the amino acid sequence of SEQ ID NO: 47. In exemplary aspects, the CAR further comprises a transmembrane (TM) domain based on the TM domain of CD8 α . In exemplary aspects, the CAR comprises the amino acid sequence of SEQ ID NO: 85. In exemplary aspects, the hinge region comprises the amino acid sequence of SEQ ID NO: 35 or SEQ ID NO: 37. In exemplary aspects, the CAR comprises the amino acid sequence of SEQ ID NO: 49 or SEQ ID NO: 51. In exemplary aspects, the endodomain of the CAR of the disclosures comprises the amino acid sequence of SEQ ID NO: 53 or SEQ ID NO: 55. In exemplary aspects, the endodomain of the CAR of the disclosure comprises the amino acid sequence of SEQ ID NO: 87 or SEQ ID NO: 89. In exemplary aspects, the endodomain of the CAR of the disclosure comprises the amino acid sequence of SEQ ID NO: 91, SEQ ID NO: 93 or SEQ ID NO: 95. In exemplary aspects, the endodomain of the CAR of the disclosure comprises the amino acid sequence of SEQ ID NO: 97, SEQ ID NO: 99 or SEQ ID NO: 101.

[0197] In exemplary aspects, the endodomain further comprises a signaling domain of one or more of: CD137, CD134, CD27, CD40, ICOS, and Myd88. In exemplary aspects, the endodomain comprises one or more of the amino acid sequences of SEQ ID NOs: 68, 70, 72, 74, 76, and 78, which provide a sequence comprising a CD27 signaling domain, a sequence comprising a CD40 signaling domain, a sequence comprising a CD134 signaling domain, a sequence comprising a CD137 signaling domain, a sequence comprising a ICOS signaling domain, and a sequence comprising a Myd88 signaling domain, respectively.

[0198] In exemplary aspects, the CAR comprises (A) each of the amino acid sequence of: NYLMN (SEQ ID NO: 1); RIDPYDGDIDYNQNFKD (SEQ ID NO: 2); GYG-TAYGVDY (SEQ ID NO: 3); RASESVDNYGISFMN (SEQ ID NO: 4); AASRQGSG (SEQ ID NO: 5); and QQSKEVPWT (SEQ ID NO: 6), (B) a hinge region; (C) an endodomain comprising a signaling domain of a CD3 zeta chain and a signaling domain of CD28 and at least one other signaling domain. In exemplary aspects, the CAR comprises an endodomain comprising a signaling domain of 41BB (CD137). In exemplary aspects the CAR comprises an endodomain comprising an amino acid sequence of SEQ ID NO: 81. In exemplary aspects, the CD137 signaling is

N-terminal to a CD3 zeta chain signaling chain. In exemplary aspects, the endodomain comprises the amino acid sequence of SEQ ID NO: 87. In exemplary aspects, the CAR comprises the amino acid sequence of SEQ ID NO: 91. In exemplary aspects, the CAR comprises the amino acid sequence of SEQ ID NO: 97.

[0199] In exemplary aspects, the CAR comprises an endodomain comprising a signaling domain of OX40 (CD134). In exemplary aspects the CAR comprises an endodomain comprising an amino acid sequence of SEQ ID NO: 83. In exemplary aspects, the CD137 signaling is N-terminal to a CD3 zeta chain signaling chain. In exemplary aspects, the endodomain comprises the amino acid sequence of SEQ ID NO: 89. In exemplary aspects, the CAR comprises the amino acid sequence of SEQ ID NO: 95. In exemplary aspects, the CAR comprises the amino acid sequence of SEQ ID NO: 99.

[0200] In exemplary aspects, the CAR comprises (A) each of the amino acid sequence of: NYLMN (SEQ ID NO: 1); RIDPYDGDIDYNQNFKD (SEQ ID NO: 2); GYG-TAYGVDY (SEQ ID NO: 3); RASESVDNYGISFMN (SEQ ID NO: 4); AASRQGSG (SEQ ID NO: 5); and QQSKEVPWT (SEQ ID NO: 6), (B) a hinge region; (C) a transmembrane domain of CD8a chain, and (D) an endodomain comprising a signaling domain of a CD3 zeta chain, and, optionally, at least one other signaling domain. In exemplary aspects, the transmembrane domain comprises the amino acid sequence of SEQ ID NO: 85. In exemplary aspects, the CAR further comprises a CD137 signaling domain and a CD3 zeta chain signaling domain. In exemplary aspects, the CAR comprises the amino acid sequence of SEQ ID NO: 93. In exemplary aspects, the CAR comprises the amino acid sequence of SEQ ID NO: 101.

[0201] As an example, sequences of three additional IL13R α 2-specific CARs are provided. One CAR contains a CD8 α TM domain, and a 41BB.zeta signaling domain (SEQ ID NO:93 encoded by SEQ ID NO:94). The other two CARs contain a CD28 TM domain and either a CD28.CD134.zeta (SEQ ID NO:99 encoded by SEQ ID NO:100) or CD28. CD137.zeta (SEQ ID NO:101 encoded by SEQ ID NO:102) signaling domain.

[0202] Nucleic Acids, Vectors, Host Cells

[0203] Further provided by the disclosures is a nucleic acid comprising a nucleotide sequence encoding any of the binding agents and conjugates (e.g., chimeric proteins, fusion proteins, CARs) described herein. The nucleic acid may comprise any nucleotide sequence which encodes any of the binding agents and conjugates described herein. In exemplary aspects, the nucleic acid comprises a nucleotide sequence encoding each of the CDRs of SEQ ID NOs: 1-6. In exemplary aspects, the nucleic acid of the disclosures comprises a nucleic acid sequence which encodes a SEQ ID NO: 7 and/or SEQ ID NO: 8. In exemplary aspects, the nucleic acid of the disclosures comprises a nucleic acid sequence which encodes SEQ ID NO: 13 or SEQ ID NO: 14. In exemplary aspects, the nucleic acid provided herein comprises the sequence of SEQ ID NO: 15 and/or SEQ ID NO: 16. In exemplary aspects, the nucleic acid comprises a nucleotide sequence of SEQ ID NO: 66 or 67. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes the sequence of SEQ ID NO: 25. In exemplary aspects, the nucleic acid comprises a nucleotide sequence of SEQ ID NO: 26. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes

each of SEQ ID NOs: 1-6 and encodes SEQ ID NO: 28 or 30. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes each of SEQ ID NOs: 1-6 and encodes an amino acid sequence which is at least 90% identical to SEQ ID NO: 28 or 30. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes each of SEQ ID NOs: 1-6 and comprises SEQ ID NO: 29 or 31. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes SEQ ID NO: 33. In exemplary aspects, the nucleic acid comprises a nucleotide sequence of SEQ ID NO: 34. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes each of SEQ ID NOs: 1-6 and encodes SEQ ID NO: 35 or 37. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes each of SEQ ID NOs: 1-6 and comprises SEQ ID NO: 36 or 38. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes each of SEQ ID NOs: 1-6 and encodes SEQ ID NO: 39 or 41. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes each of SEQ ID NOs: 1-6 and comprises SEQ ID NO: 40 or 42. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes each of SEQ ID NOs: 1-6 and encodes SEQ ID NO: 47. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes each of SEQ ID NOs: 1-6 and comprises SEQ ID NO: 48. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes each of SEQ ID NOs: 1-6 and encodes SEQ ID NO: 49 or 51. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes each of SEQ ID NOs: 1-6 and comprises SEQ ID NO: 50 or 52. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes SEQ ID NO: 53 or 55. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes each of SEQ ID NOs: 1-6 and comprises SEQ ID NO: 54 or 56. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes each of SEO ID NOs: 1-6 and encodes one or more of SEO ID NOs: 68, 70, 72, 74, 76, and 78. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes each of SEQ ID NOs: 1-6 and comprises one or more of SEQ ID NOs: 69, 71, 73, 75, 77, and 79. In exemplary aspects, the nucleic acid comprises a nucleotide sequence which encodes each of SEQ ID NOs: 1-6 and comprises one or more of SEQ ID NOs: 82, 84, 86, 88, 90, 92, 94, 96. In exemplary aspects, the nucleic acid comprises a nucleotide sequence comprising one of SEQ ID NOs: 98, 100, and 102.

[0204] By "nucleic acid" as used herein includes "polynucleotide," "oligonucleotide," and "nucleic acid molecule," and generally means a polymer of DNA or RNA, which may be single-stranded or double-stranded, synthesized or obtained (e.g., isolated and/or purified) from natural sources, which may contain natural, non-natural or altered nucleotides, and which may contain a natural, non-natural or altered internucleotide linkage, such as a phosphoroamidate linkage or a phosphorothioate linkage, instead of the phosphodiester found between the nucleotides of an unmodified oligonucleotide. It is generally preferred that the nucleic acid does not comprise any insertions, deletions, inversions, and/or substitutions. However, it may be suitable in some instances, as discussed herein, for the nucleic acid to comprise one or more insertions, deletions, inversions, and/or substitutions.

[0205] In exemplary aspects, the nucleic acids of the disclosures are recombinant. As used herein, the term "recombinant" refers to (i) molecules that are constructed outside living cells by joining natural or synthetic nucleic acid segments to nucleic acid molecules that may replicate in a living cell, or (ii) molecules that result from the replication of those described in (i) above. For purposes herein, the replication may be in vitro replication or in vivo replication.

[0206] The nucleic acids in exemplary aspects are constructed based on chemical synthesis and/or enzymatic ligation reactions using procedures known in the art. See, for example, Sambrook et al., supra, and Ausubel et al., supra. For example, a nucleic acid may be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed upon hybridization (e.g., phosphorothioate derivatives and acridine substituted nucleotides). Examples of modified nucleotides that may be used to generate the nucleic acids include, but are not limited to, 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xanthine, 4-acetylcytosine, 5-(carboxyhydroxymethyl) uracil, 5-carboxymethylaminomethyl-2-thiouridme, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N6-isopentenyladenine, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N-substituted adenine, 7-methylguanine, 5-methylammomethyluracil, 5-methoxyaminomethyl-2thiouracil. beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5-oxyacetic acid methylester, 3-(3-amino-3-N-2-carboxypropyl) uracil, and 2,6-diaminopurine. Alternatively, one or more of the nucleic acids of the disclosures may be purchased from companies, such as Macromolecular Resources (Fort Collins, Colo.) and Synthegen (Houston, Tex.).

[0207] The nucleic acids of the disclosures in exemplary aspects are incorporated into a recombinant expression vector. In this regard, the disclosures provides recombinant expression vectors comprising any of the nucleic acids of the disclosures. For purposes herein, the term "recombinant expression vector" means a genetically-modified oligonucleotide or polynucleotide construct that permits the expression of an mRNA, protein, polypeptide, or peptide by a host cell, when the construct comprises a nucleotide sequence encoding the mRNA, protein, polypeptide, or peptide, and the vector is contacted with the cell under conditions sufficient to have the mRNA, protein, polypeptide, or peptide expressed within the cell. The vectors of the disclosures are not naturally-occurring as a whole. However, parts of the vectors may be naturally-occurring. The inventive recombinant expression vectors may comprise any type of nucleotides, including, but not limited to DNA and RNA, which may be single-stranded or double-stranded, synthesized or obtained in part from natural sources, and which may contain natural, non-natural or altered nucleotides. The recombinant expression vectors may comprise naturallyoccurring or non-naturally occurring internucleotide linkages, or both types of linkages. In exemplary aspects, the altered nucleotides or non-naturally occurring internucleotide linkages do not hinder the transcription or replication of the vector.

[0208] The recombinant expression vector of the disclosures may be any suitable recombinant expression vector, and may be used to transform or transfect any suitable host. Suitable vectors include those designed for propagation and expansion or for expression or both, such as plasmids and viruses. The vector may be selected from the group consisting of the pUC series (Fermentas Life Sciences), the pBluescript series (Stratagene, LaJolla, Calif.), the pET series (Novagen, Madison, Wis.), the pGEX series (Pharmacia Biotech, Uppsala, Sweden), and the pEX series (Clontech, Palo Alto, Calif.). Bacteriophage vectors, such as λ GTIO, λ GT1 1, λ ZapII (Stratagene), λ EMBL4, and λ NM1 149, also may be used. Examples of plant expression vectors include pBIO1, pBI101.2, pBI101.3, pBI121 and pBIN19 (Clontech). Examples of animal expression vectors include pEUK-Cl, pMAM and pMAMneo (Clontech). Preferably, the recombinant expression vector is a viral vector, e.g., a retroviral vector.

[0209] The recombinant expression vectors of the disclosures may be prepared using standard recombinant DNA techniques described in, for example, Sambrook et al., supra, and Ausubel et al., supra. Constructs of expression vectors, which are circular or linear, may be prepared to contain a replication system functional in a prokaryotic or eukaryotic host cell. Replication systems may be derived, e.g., from CoIE1, 2μ plasmid, λ , SV40, bovine papilloma virus, and the like.

[0210] In exemplary aspects, the recombinant expression vector comprises regulatory sequences, such as transcription and translation initiation and termination codons, which are specific to the type of host (e.g., bacterium, fungus, plant, or animal) into which the vector is to be introduced, as appropriate and taking into consideration whether the vector is DNA- or RNA-based.

[0211] The recombinant expression vector may include one or more marker genes, which allow for selection of transformed or transfected hosts. Marker genes include biocide resistance, e.g., resistance to antibiotics, heavy metals, etc., complementation in an auxotrophic host to provide prototrophy, and the like. Suitable marker genes for the inventive expression vectors include, for instance, neomycin/G418 resistance genes, hygromycin resistance genes, histidinol resistance genes, tetracycline resistance genes, and ampicillin resistance genes.

[0212] The recombinant expression vector may comprise a native or normative promoter operably linked to the nucleotide sequence encoding the binding agent or conjugate or to the nucleotide sequence which is complementary to or which hybridizes to the nucleotide sequence encoding the binding agent or conjugate. The selection of promoters, e.g., strong, weak, inducible, tissue-specific and developmental-specific, is within the ordinary skill of the artisan.

[0213] Similarly, the combining of a nucleotide sequence with a promoter is also within the skill of the artisan. The promoter may be a non-viral promoter or a viral promoter, e.g., a cytomegalovirus (CMV) promoter, an SV40 promoter, an RSV promoter, and a promoter found in the long-terminal repeat of the murine stem cell virus.

[0214] The inventive recombinant expression vectors may be designed for either transient expression, for stable expression, or for both. Also, the recombinant expression vectors may be made for constitutive expression or for inducible expression. Further, the recombinant expression vectors may be made to include a suicide gene.

[0215] As used herein, the term "suicide gene" refers to a gene that causes the cell expressing the suicide gene to die. The suicide gene may be a gene that confers sensitivity to an agent, e.g., a drug, upon the cell in which the gene is expressed, and causes the cell to die when the cell is contacted with or exposed to the agent. Suicide genes are known in the art (see, for example, Suicide Gene Therapy: Methods and Reviews. Springer, Caroline J. (Maycer Research UK Centre for Maycer Therapeutics at the Institute of Maycer Research, Sutton, Surrey, UK), Humana Press, 2004) and include, for example, the Herpes Simplex Virus (HSV) thymidine kinase (TK) gene, cytosine deaminase, purine nucleoside phosphorylase, and nitroreductase.

[0216] The disclosures further provides a host cell comprising any of the nucleic acids or vectors described herein. As used herein, the term "host cell" refers to any type of cell that may contain the nucleic acid or vector described herein. In exemplary aspects, the host cell is a eukaryotic cell, e.g., plant, animal, fungi, or algae, or may be a prokaryotic cell, e.g., bacteria or protozoa. In exemplary aspects, the host cells is a cell originating or obtained from a subject, as described herein. In exemplary aspects, the host cell originates from or is obtained from a mammal. As used herein, the term "mammal" refers to any mammal, including, but not limited to, mammals of the order Rodentia, such as mice and hamsters, and mammals of the order Lagomorpha, such as rabbits. It is preferred that the mammals are from the order Carnivora, including Felines (cats) and Canines (dogs). It is more preferred that the mammals are from the order Artiodactyla, including bovines (cows) and swines (pigs) or of the order Perssodactyla, including equines (horses). It is most preferred that the mammals are of the order Primates, Ceboids, or Simoids (monkeys) or of the order Anthropoids (humans and apes). An especially preferred mammal is the human.

[0217] In exemplary aspects, the host cell is a cultured cell or a primary cell, i.e., isolated directly from an organism, e.g., a human. The host cell in exemplary aspects is an adherent cell or a suspended cell, i.e., a cell that grows in suspension. Suitable host cells are known in the art and include, for instance, DH5a E. coli cells, Chinese hamster ovarian (CHO) cells, monkey VERO cells, T293 cells, COS cells, HEK293 cells, and the like. For purposes of amplifying or replicating the recombinant expression vector, the host cell is preferably a prokaryotic cell, e.g., a DH5 α cell. For purposes of producing a binding agent or a conjugate, the host cell is in some aspects a mammalian cell. In exemplary aspects, the host cell is a human cell. While the host cell may be of any cell type, the host cell may originate from any type of tissue, and may be of any developmental stage. In exemplary aspects, the host cell is a hematopoietic stem cell or progenitor cell. See, e.g, Nakamura De Oliveira et al., Human Gene Therapy 24:824-839 (2013). The host cell in exemplary aspects is a peripheral blood lymphocyte (PBL). In exemplary aspects, the host cell is a natural killer cell. In exemplary aspects, the host cell is a T cell.

[0218] For purposes herein, the T cell may be any T cell, such as a cultured T cell, e.g., a primary T cell, or a T cell from a cultured T cell line, e.g., Jurkat, SupTl, etc., or a T cell obtained from a mammal. If obtained from a mammal, the T cell may be obtained from numerous sources, includ-

ing but not limited to blood, bone marrow, lymph node, the thymus, or other tissues or fluids. T cells may also be enriched for or purified. The T cell may be obtained by maturing hematopoietic stem cells, either in vitro or in vivo, into T cells. In exemplary aspects, the T cell is a human T cell. In exemplary aspects, the T cell is a T cell isolated from a human. The T cell may be any type of T cell, including NKT cell, and may be of any developmental stage, including but not limited to, CD4+/CD8+ double positive T cells, CDA+ helper T cells, e.g., Th1 and Th2 cells, CD8+ T cells (e.g., cytotoxic T cells), peripheral blood mononuclear cells (PBMCs), peripheral blood leukocytes (PBLs), tumor infiltrating cells (TILs), memory T cells, naive T cells, and the like. Preferably, the T cell is a CD8+ T cell or a CD4+ T cell. [0219] Also provided by the disclosures is a population of cells comprising at least one host cell described herein. The population of cells may be a heterogeneous population comprising the host cell comprising any of the recombinant expression vectors described, in addition to at least one other cell, e.g., a host cell (e.g., a T cell), which does not comprise any of the recombinant expression vectors, or a cell other than a T cell, e.g., a B cell, a macrophage, a neutrophil, an erythrocyte, a hepatocyte, an endothelial cell, an epithelial cells, a muscle cell, a brain cell, etc. Alternatively, the population of cells may be a substantially homogeneous population, in which the population comprises mainly of host cells (e.g., consisting essentially of) comprising the recombinant expression vector. The population also may be a clonal population of cells, in which all cells of the population are clones of a single host cell comprising a recombinant expression vector, such that all cells of the population comprise the recombinant expression vector. In exemplary embodiments of the disclosures, the population of cells is a clonal population comprising host cells expressing a nucleic acid or a vector described herein.

[0220] Pharmaceutical Compositions and Routes of Administration

[0221] In some embodiments of the disclosures, the binding agents, conjugates, nucleic acids, vectors, host cells, or populations of cells, are admixed with a pharmaceutically acceptable carrier. Accordingly, pharmaceutical compositions comprising any of the binding agents, conjugates, nucleic acids, vectors, host cells, or populations of cells described herein and comprising a pharmaceutically acceptable carrier, diluent, or excipient are contemplated.

[0222] The pharmaceutically acceptable carrier is any of those conventionally used and is limited only by physicochemical considerations, such as solubility and lack of reactivity with the active binding agent(s), and by the route of administration. The pharmaceutically acceptable carriers described herein, for example, vehicles, adjuvants, excipients, and diluents, are well-known to those skilled in the art and are readily available to the public. In one aspect the pharmaceutically acceptable carrier is one that is chemically inert to the active ingredient(s) of the pharmaceutical composition, e.g., the first binding agent and the second binding agent, and one which has no detrimental side effects or toxicity under the conditions of use. The carrier in some embodiments does not produce adverse, allergic, or other untoward reactions when administered to an animal or a human. The pharmaceutical composition in some aspects is free of pyrogens, as well as other impurities that could be harmful to humans or animals. Pharmaceutically acceptable carriers include any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents and the like; the use of which are well known in the art.

[0223] Acceptable carriers, excipients or stabilizers are nontoxic to recipients and are preferably inert at the dosages and concentrations employed, and include buffers such as phosphate, citrate, or other organic acids; antioxidants such as ascorbic acid; low molecular weight polypeptides; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, arginine or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrins; chelating agents such as EDTA; sugar alcohols such as sodium; and/or nonionic surfactants such as Tween, Pluronics or polyethylene glycol (PEG).

[0224] Therapeutic formulations of the compositions useful for practicing the methods disclosed herein, such as polypeptides, polynucleotides, or antibodies, may be prepared for storage by mixing the selected composition having the desired degree of purity with optional physiologically pharmaceutically-acceptable carriers, excipients, or stabilizers (Remington's Pharmaceutical Sciences, 18th edition, A. R. Gennaro, ed., Mack Publishing Company (1990)) in the form of a lyophilized cake or an aqueous solution. Pharmaceutical compositions may be produced by admixing with one or more suitable carriers or adjuvants such as water, mineral oil, polyethylene glycol, starch, talcum, lactose, thickeners, stabilizers, suspending agents, and the like. Such compositions may be in the form of solutions, suspensions, tablets, capsules, creams, salves, ointments, or other conventional forms.

[0225] The composition to be used for in vivo administration should be sterile. This is readily accomplished by filtration through sterile filtration membranes, prior to or following lyophilization and reconstitution. Therapeutic compositions generally are placed into a container having a sterile access port, for example, an intravenous solution bag or vial having a stopper pierceable by a hypodermic injection needle. The pharmaceutical forms suitable for injectable use include sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions. In some cases the form should be sterile and should be fluid to the extent that easy syringability exists. It should be stable under the conditions of manufacture and storage and should be preserved against the contaminating action of microorganisms, such as bacteria and fungi. The composition for parenteral administration ordinarily will be stored in lyophilized form or in solution.

[0226] The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and vegetable oils. The proper fluidity can be maintained, for example, by the use of a coating, such as lecithin, by the maintenance of the required particle size in the case of dispersion, and by the use of surfactants. The prevention of the action of microorganisms can be brought about by various antibacterial and/or antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the inclu-

sion in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

[0227] The choice of carrier will be determined in part by the particular type of binding agents of the pharmaceutical composition, as well as by the particular route used to administer the pharmaceutical composition. Accordingly, there are a variety of suitable formulations of the pharmaceutical composition.

[0228] The pharmaceutical composition of the present disclosures can comprise any pharmaceutically acceptable ingredient including, for example, acidifying agents, additives, adsorbents, aerosol propellants, air displacement agents, alkalizing agents, anticaking agents, anticoagulants, antimicrobial preservatives, antioxidants, antiseptics, bases, binders, buffering agents, chelating agents, coating agents, coloring agents, desiccants, detergents, diluents, disinfectants, disintegrants, dispersing agents, dissolution-enhancing agents, dyes, emollients, emulsifying agents, emulsion stabilizers, fillers, film-forming agents, flavor enhancers, flavoring agents, flow enhancers, gelling agents, granulating agents, humectants, lubricants, mucoadhesives, ointment bases, ointments, oleaginous vehicles, organic bases, pastille bases, pigments, plasticizers, polishing agents, preservatives, sequestering agents, skin penetrants, solubilizing agents, solvents, stabilizing agents, suppository bases, surface active agents, surfactants, suspending agents, sweetening agents, therapeutic agents, thickening agents, tonicity agents, toxicity agents, viscosity-increasing agents, waterabsorbing agents, water-miscible cosolvents, water softeners, or wetting agents.

[0229] The pharmaceutical compositions may be formulated to achieve a physiologically compatible pH. In some embodiments, the pH of the pharmaceutical composition may be at least 5, at least 5.5, at least 6, at least 6.5, at least 7, at least 7.5, at least 8, at least 8.5, at least 9, at least 9.5, at least 10, or at least 10.5 up to and including pH 11, depending on the formulation and route of administration. In certain embodiments, the pharmaceutical compositions may comprise buffering agents to achieve a physiologically compatible pH. The buffering at the desired pH such as, for example, phosphate buffers (e.g., PBS), triethanolamine, Tris, bicine, TAPS, tricine, HEPES, TES, MOPS, PIPES, cacodylate, MES, and others known in the art.

[0230] In some embodiments, the pharmaceutical composition comprising the binding agents described herein is formulated for parenteral administration, subcutaneous administration, intravenous administration, intramuscular administration, intraarterial administration, intrathecal administration, or interperitoneal administration. In other embodiments, the pharmaceutical composition is administered via nasal, spray, oral, aerosol, rectal, or vaginal administration. The compositions may be administered by infusion, bolus injection or by implantation device.

[0231] The following discussion on routes of administration is merely provided to illustrate exemplary embodiments and should not be construed as limiting the scope of the disclosed subject matter in any way.

[0232] Formulations suitable for oral administration can consist of (a) liquid solutions, such as an effective amount of the composition of the present disclosure dissolved in diluents, such as water, saline, or orange juice; (b) capsules, sachets, tablets, lozenges, and troches, each containing a predetermined amount of the active ingredient, as solids or

granules; (c) powders; (d) suspensions in an appropriate liquid; and (e) suitable emulsions. Liquid formulations may include diluents, such as water and alcohols, for example, ethanol, benzyl alcohol, and the polyethylene alcohols, either with or without the addition of a pharmaceutically acceptable surfactant. Capsule forms can be of the ordinary hard- or soft-shelled gelatin type containing, for example, surfactants, lubricants, and inert fillers, such as lactose, sucrose, calcium phosphate, and corn starch. Tablet forms can include one or more of lactose, sucrose, mannitol, corn starch, potato starch, alginic acid, microcrystalline cellulose, acacia, gelatin, guar gum, colloidal silicon dioxide, croscarmellose sodium, talc, magnesium stearate, calcium stearate, zinc stearate, stearic acid, and other excipients, colorants, diluents, buffering agents, disintegrating agents, moistening agents, preservatives, flavoring agents, and other pharmacologically compatible excipients. Lozenge forms can comprise a composition of the disclosure in a flavor, usually sucrose and acacia or tragacanth, as well as pastilles comprising a composition of the disclosure in an inert base, such as gelatin and glycerin, or sucrose and acacia, emulsions, gels, and the like, optionally also containing such excipients as are known in the art.

[0233] The compositions of the disclosure, alone or in combination with other suitable components, can be delivered via pulmonary administration and can be made into aerosol formulations to be administered via inhalation. These aerosol formulations can be placed into pressurized acceptable propellants, such as dichlorodifluoromethane, propane, nitrogen, and the like. They also may be formulated as pharmaceuticals for non-pressured preparations, such as in a nebulizer or an atomizer. Such spray formulations also may be used to spray mucosa. In some embodiments, the composition is formulated into a powder blend or into microparticles or nanoparticles. Suitable pulmonary formulations are known in the art. See, e.g., Qian et al., Int J Pharm 366: 218-220 (2009); Adjei and Garren, Pharmaceutical Research, 7(6): 565-569 (1990); Kawashima et al., J Controlled Release 62(1-2): 279-287 (1999); Liu et al., Pharm Res 10(2): 228-232 (1993); International Patent Application Publication Nos. WO 2007/133747 and WO 2007/141411. [0234] Topical formulations are well-known to those of

[0234] Topical formulations are well-known to those of skill in the art. Such formulations are particularly suitable in the context of the invention for application to the skin.

[0235] In some embodiments, the pharmaceutical composition described herein is formulated for parenteral administration. For purposes herein, parenteral administration includes, but is not limited to, intravenous, intraarterial, intramuscular, intracerebral, intracerebroventricular, intracardiac, subcutaneous, intraosseous, intradermal, intrathecal, intraperitoneal, retrobulbar, intrapulmonary, intravesical, and intracavernosal injections or infusions. Administration by surgical implantation at a particular site is contemplated as well.

[0236] Formulations suitable for parenteral administration include aqueous and non-aqueous, isotonic sterile injection solutions, which can contain anti-oxidants, buffers, bacteriostats, and solutes that render the formulation isotonic with the blood of the intended recipient, and aqueous and non-aqueous sterile suspensions that can include suspending agents, solubilizers, thickening agents, stabilizers, and preservatives. The term, "parenteral" means not through the alimentary canal but by some other route such as subcutaneous, intramuscular, intraspinal, or intravenous. The com-

position of the present disclosure can be administered with a physiologically acceptable diluent in a pharmaceutical carrier, such as a sterile liquid or mixture of liquids, including water, saline, aqueous dextrose and related sugar solutions, an alcohol, such as ethanol or hexadecyl alcohol, a glycol, such as propylene glycol or polyethylene glycol, dimethylsulfoxide, glycerol, ketals such as 2,2-dimethyl-1, 5,3-dioxolane-4-methanol, ethers, poly(ethyleneglycol) 400, oils, fatty acids, fatty acid esters or glycerides, or acetylated fatty acid glycerides with or without the addition of a pharmaceutically acceptable surfactant, such as a soap or a detergent, a suspending agent, such as pectin, carbomers, methylcellulose, hydroxypropylmethylcellulose, or carboxymethylcellulose, or emulsifying agents and other pharmaceutical adjuvants.

[0237] Oils, which can be used in parenteral formulations include petroleum, animal, vegetable, or synthetic oils. Specific examples of oils include peanut, soybean, sesame, cottonseed, corn, olive, petrolatum, and mineral. Suitable fatty acids for use in parenteral formulations include oleic acid, stearic acid, and isostearic acid. Ethyl oleate and isopropyl myristate are examples of suitable fatty acid esters.

[0238] The parenteral formulations in some embodiments contain preservatives or buffers. In order to minimize or eliminate irritation at the site of injection, such compositions optionally contain one or more nonionic surfactants having a hydrophile-lipophile balance (HLB) of from about 12 to about 17. The quantity of surfactant in such formulations will typically range from about 5% to about 15% by weight. Suitable surfactants include polyethylene glycol sorbitan fatty acid esters, such as sorbitan monooleate and the high molecular weight adducts of ethylene oxide with a hydrophobic base, formed by the condensation of propylene oxide with propylene glycol. The parenteral formulations can be presented in unit-dose or multi-dose sealed containers, such as ampoules and vials, and can be stored in a freeze-dried (lyophilized) condition requiring only the addition of the sterile liquid excipient, for example, water, for injections, immediately prior to use. Extemporaneous injection solutions and suspensions can be prepared from sterile powders, granules, and tablets of the kind previously described and known in the art.

[0239] Injectable formulations are in accordance with the invention. The requirements for effective pharmaceutical carriers for injectable compositions are well-known to those of ordinary skill in the art (see, e.g., *Pharmaceutics and Pharmacy Practice*, J. B. Lippincott Company, Philadelphia, Pa., Banker and Chalmers, eds., pages 238-250 (1982), and *ASHP Handbook on Injectable Drugs*, Toissel, 4th ed., pages 622-630 (1986)).

[0240] It will be appreciated by one of skill in the art that, in addition to the above-described pharmaceutical compositions, the composition of the disclosure can be formulated as inclusion complexes, such as cyclodextrin inclusion complexes, or liposomes.

[0241] Dose

[0242] For purposes herein, the amount or dose of the pharmaceutical composition administered is sufficient to effect, e.g., a therapeutic or prophylactic response, in the subject or animal over a reasonable time frame. For example, the dose of the pharmaceutical composition is sufficient to treat or prevent a disease or medical condition in a period of from about 12 hours, about 18 hours, about 1

to 4 days or longer, e.g., 5 days, 6 days, 1 week, 10 days, 2 weeks, 16 to 20 days, or more, from the time of administration. In certain embodiments, the time period is even longer. The dose is determined by the efficacy of the particular pharmaceutical composition and the condition of the animal (e.g., human), as well as the body weight of the animal (e.g., human) to be treated.

[0243] Many assays for determining an administered dose are known in the art. In some embodiments, an assay which comprises comparing the extent to which the binding agents block IL13R α 2-mediated cell growth upon administration of a given dose to a mammal among a set of mammals each of which is given a different dose of binding agents is used to determine a starting dose to be administered to a mammal. The extent to which the binding agents block IL13R α 2 mediated cell growth upon administration of a certain dose can be assayed by methods known in the art.

[0244] The dose of the pharmaceutical composition also will be determined by the existence, nature and extent of any adverse side effects that might accompany the administration of a particular pharmaceutical composition. Typically, the attending physician will decide the dosage of the pharmaceutical composition with which to treat each individual patient, taking into consideration a variety of factors, such as age, body weight, general health, diet, sex, binding agents of the pharmaceutical composition to be administered, route of administration, and the severity of the condition being treated.

[0245] By way of example and not intending to limit the invention, the dose of the binding agent of the present disclosure can be about 0.0001 to about 1 g/kg body weight of the subject being treated/day, from about 0.0001 to about 0.001 g/kg body weight/day, or about 0.01 mg to about 1 g/kg body weight/day. The pharmaceutical composition in some aspects comprise the binding agent of the present disclosure at a concentration of at least A, wherein A is about 0.001 mg/ml, about 0.01 mg/ml, 0 about 1 mg/ml, about 0.5 mg/ml, about 1 mg/ml, about 2 mg/ml, about 3 mg/ml, about 4 mg/ml, about 5 mg/ml, about 6 mg/ml, about 7 mg/ml, about 8 mg/ml, about 9 mg/ml, about 10 mg/ml, about 11 mg/ml, about 12 mg/ml, about 13 mg/ml, about 14 mg/ml, about 15 mg/ml, about 16 mg/ml, about 17 mg/ml, about 18 mg/ml, about 19 mg/ml, about 20 mg/ml, about 21 mg/ml, about 22 mg/ml, about 23 mg/ml, about 24 mg/ml, about 25 mg/ml or higher. In some embodiments, the pharmaceutical composition comprises the binding agent at a concentration of at most B, wherein B is about 30 mg/ml, about 25 mg/ml, about 24 mg/ml, about 23, mg/ml, about 22 mg/ml, about 21 mg/ml, about 20 mg/ml, about 19 mg/ml, about 18 mg/ml, about 17 mg/ml, about 16 mg/ml, about 15 mg/ml, about 14 mg/ml, about 13 mg/ml, about 12 mg/ml, about 11 mg/ml, about 10 mg/ml, about 9 mg/ml, about 8 mg/ml, about 7 mg/ml, about 6 mg/ml, about 5 mg/ml, about 4 mg/ml, about 3 mg/ml, about 2 mg/ml, about 1 mg/ml, or about 0.1 mg/ml. In some embodiments, the compositions may contain an analog at a concentration range of A to B mg/ml, for example, about 0.001 to about 30.0 mg/ml.

[0246] Additional dosing guidance can be gauged from other antibody therapeutics, such as bevacizumab (AvastinTM Genentech); Cetuximab (ExbituxTM Imclone), Panitumumab (VectibixTM Amgen), and Trastuzumab (HerceptinTM Genentech).

[0247] Timing of Administration

[0248] The disclosed pharmaceutical formulations may be administered according to any regimen including, for example, daily (1 time per day, 2 times per day, 3 times per day, 4 times per day, 5 times per day, 6 times per day), every two days, every three days, every four days, every five days, every six days, weekly, bi-weekly, every three weeks, monthly, or bi-monthly. Timing, like dosing can be fine-tuned based on dose-response studies, efficacy, and toxicity data, and initially gauged based on timing used for other antibody therapeutics.

[0249] Controlled Release Formulations

[0250] The pharmaceutical composition is in certain aspects modified into a depot form, such that the manner in which the active ingredients of the pharmaceutical composition (e.g., the binding agents) is released into the body to which it is administered is controlled with respect to time and location within the body (see, for example, U.S. Pat. No. 4,450,150). Depot forms in various aspects, include, for example, an implantable composition comprising a porous or non-porous material, such as a polymer, wherein the binding agents are encapsulated by or diffused throughout the material and/or degradation of the non-porous material. The depot is then implanted into the desired location within the body and the binding agents are released from the implant at a predetermined rate.

[0251] Accordingly, the pharmaceutical composition in certain aspects is modified to have any type of in vivo release profile. In some aspects, the pharmaceutical composition is an immediate release, controlled release, sustained release, extended release, delayed release, or bi-phasic release formulation. Methods of formulating peptides (e.g., peptide binding agents) for controlled release are known in the art. See, for example, Qian et al., J Pharm 374: 46-52 (2009) and International Patent Application Publication Nos. WO 2008/130158, WO2004/033036; WO2000/032218; and WO 1999/040942. Suitable examples of sustained-release preparations include semipermeable polymer matrices in the form of shaped articles, e.g., films, or microcapsules. Sustained release matrices include polyesters, hydrogels, polylactides (U.S. Pat. No. 3,773,919, EP 58,481), copolymers of L-glutamic acid and gamma ethyl-L-glutamate (Sidman, et al., Biopolymers, 22: 547-556 (1983)), poly (2-hydroxyethyl-methacrylate) (Langer, et al., J. Biomed. Mater. Res., 15:167-277 (1981) and Langer, Chem. Tech., 12:98-105 (1982)), ethylene vinyl acetate (Langer, et al, supra) or poly-D(-)-3-hydroxybutyric acid (EP 133,988). Sustainedrelease compositions also may include liposomes, which can be prepared by any of several methods known in the art (e.g., DE 3,218,121; Epstein, et al., Proc. Natl. Acad. Sci. USA, 82:3688-3692 (1985); Hwang, et al., Proc. Natl. Acad. Sci. USA, 77:4030-4034 (1980); EP 52,322; EP 36,676; EP 88,046; EP 143,949).

[0252] Combinations

[0253] The compositions of the disclosures may be employed alone, or in combination with other agents. In some embodiments, more than one type of binding agent are administered. For example, the administered composition, e.g., pharmaceutical composition, may comprise an antibody as well as an scFv. In some embodiments, the compositions of the disclosure are administered together with another therapeutic agent or diagnostic agent, including any of those described herein. Certain diseases, e.g., cancers, or patients may lend themselves to a treatment of combined agents to achieve an additive or even a synergistic effect compared to the use of any one therapy alone.

[0254] Uses

[0255] Based in part on the data provided herein, the binding agents, conjugates, host cells, populations of cells, and pharmaceutical compositions are useful for treating a neoplasm, tumor, or a cancer.

[0256] For purposes of the present disclosure, the term "treat" and "prevent" as well as words stemming therefrom, as used herein, do not necessarily imply 100% or complete treatment (e.g., cure) or prevention. Rather, there are varying degrees of treatment or prevention of which one of ordinary skill hi the art recognizes as having a potential benefit or therapeutic effect. In this respect, the methods of the present disclosures can provide any amount or any level of treatment or prevention of a cancer in a patient, e.g., a human. Furthermore, the treatment or prevention provided by the method disclosed herein can include treatment or prevention of one or more conditions or symptoms of the disease, e.g., cancer, being treated or prevented. Also, for purposes herein, "prevention" can encompass delaying the onset of the disease, or a symptom or condition thereof.

[0257] The materials and methods described herein are especially useful for inhibiting neoplastic cell growth or spread; particularly neoplastic cell growth for which the IL13R α 2 targeted by the binding agents plays a role.

[0258] Neoplasms treatable by the binding agents, conjugates, host cells, populations of cells, and pharmaceutical compositions of the disclosures include solid tumors, for example, carcinomas and sarcomas. Carcinomas include malignant neoplasms derived from epithelial cells which infiltrate, for example, invade, surrounding tissues and give rise to metastases. Adenocarcinomas are carcinomas derived from glandular tissue, or from tissues that form recognizable glandular structures. Another broad category of cancers includes sarcomas and fibrosarcomas, which are tumors whose cells are embedded in a fibrillar or homogeneous substance, such as embryonic connective tissue. The invention also provides methods of treatment of cancers of myeloid or lymphoid systems, including leukemias, lymphomas, and other cancers that typically are not present as a tumor mass, but are distributed in the vascular or lvmphoreticular systems. Further contemplated are methods for treatment of adult and pediatric oncology, growth of solid tumors/malignancies, myxoid and round cell carcinoma, locally advanced tumors, cancer metastases, including lymphatic metastases. The cancers listed herein are not intended to be limiting. Both age (child and adult), sex (male and female), primary and secondary, pre- and post-metastatic, acute and chronic, benign and malignant, anatomical location cancer embodiments and variations are contemplated targets. Cancers are grouped by embryonic origin (e.g., carcinoma, lymphomas, and sarcomas), by organ or physiological system, and by miscellaneous grouping. Particular cancers may overlap in their classification, and their listing in one group does not exclude them from another.

[0259] Carcinomas that may be targeted include adrenocortical, acinar, acinic cell, acinous, adenocystic, adenoid cystic, adenoid squamous cell, cancer adenomatosum, adenosquamous, adnexel, cancer of adrenal cortex, adrenocortical, aldosterone-producing, aldosterone-secreting, alveolar, alveolar cell, ameloblastic, ampullary, anaplastic cancer of thyroid gland, apocrine, basal cell, basal cell, alveolar, comedo basal cell, cystic basal cell, morphea-like basal cell, multicentric basal cell, nodulo-ulcerative basal cell, pigmented basal cell, sclerosing basal cell, superficial basal cell, basaloid, basosquamous cell, bile duct, extrahepatic bile duct, intrahepatic bile duct, bronchioalveolar, bronchiolar, bronchioloalveolar, bronchoalveolar, bronchoalveolar cell, bronchogenic, cerebriform, cholangiocellular, chorionic, choroids plexus, clear cell, cloacogenic anal, colloid, comedo, corpus, cancer of corpus uteri, cortisolproducing, cribriform, cylindrical, cylindrical cell, duct, ductal, ductal cancer of the prostate, ductal cancer in situ (DCIS), eccrine, embryonal, cancer en cuirasse, endometrial, cancer of endometrium, endometroid, epidermoid, cancer ex mixed tumor, cancer ex pleomorphic adenoma, exophytic, fibrolamellar, cancer fibrosum, follicular cancer of thyroid gland, gastric, gelatinform, gelatinous, giant cell, giant cell cancer of thyroid gland, cancer gigantocellulare, glandular, granulose cell, hepatocellular, Hurthle cell, hypernephroid, infantile embryonal, islet cell carcinoma, inflammatory cancer of the breast, cancer in situ, intraductal, intraepithelial, juvenile embryonal, intraepidermal. Kulchitsky-cell, large cell, leptomeningeal, lobular, infiltrating lobular, invasive lobular, lobular cancer in situ (LCIS), lymphoepithelial, cancer medullare, medullary, medullary cancer of thyroid gland, medullary thyroid, melanotic, meningeal, Merkel cell, metatypical cell, micropapillary, cancer molle, mucinous, cancer muciparum, cancer mucocellulare, mucoepidermoid, cancer mucosum, mucous, nasopharyngeal, neuroendocrine cancer of the skin, noninfiltrating, non-small cell, non-small cell lung cancer (NSCLC), oat cell, cancer ossificans, osteoid, Paget's disease of the bone or breast, papillary, papillary cancer of thyroid gland, periampullary, preinvasive, prickle cell, primary intrasseous, renal cell, scar, schistosomal bladder, Schneiderian, scirrhous, sebaceous, signet-ring cell, cancer simplex, small cell, small cell lung cancer (SCLC), spindle cell, cancer spongiosum, squamous, squamous cell, terminal duct, anaplastic thyroid, follicular thyroid, medullary thyroid, papillary thyroid, trabecular cancer of the skin, transitional cell. tubular, undifferentiated cancer of thyroid gland, uterine corpus, verrucous, villous, cancer villosum, yolk sac, squamous cell particularly of the head and neck, esophageal squamous cell, and oral cancers and carcinomas.

[0260] Sarcomas that may be targeted include adipose, alveolar soft part, ameloblastic, avian, botryoid, sarcoma botryoides, chicken, chloromatous, chondroblastic, clear cell sarcoma of kidney, embryonal, endometrial stromal, epithelioid, Ewing's, fascial, fibroblastic, fowl, giant cell, granulocytic, hemangioendothelial, Hodgkin's, idiopathic multiple pigmented hemorrhagic, immunoblastic sarcoma of B cells, immunoblastic sarcoma of T cells, Jensen's, Kaposi's, Kupffer cell, leukocytic, lymphatic, melanotic, mixed cell, multiple, lymphangio, idiopathic hemorrhagic, multipotential primary sarcoma of bone, osteoblastic, osteogenic, parosteal, polymorphous, pseudo-Kaposi, reticulum cell, reticulum cell sarcoma of the brain, rhabdomyosarcoma, Rous, soft tissue, spindle cell, synovial, telangiectatic, sarcoma (osteosarcoma)/malignant fibrous histiocytoma of bone, and soft tissue sarcomas.

[0261] Lymphomas that may targeted include AIDS-related, non-Hodgkin's, Hodgkin's, T-cell, T-cell leukemia/ lymphoma, African, B-cell, B-cell monocytoid, bovine malignant, Burkitt's, centrocytic, lymphoma cutis, diffuse, diffuse, large cell, diffuse, mixed small and large cell, diffuse, small cleaved cell, follicular, follicular center cell, follicular, mixed small cleaved and large cell, follicular, predominantly large cell, follicular, predominantly small cleaved cell, giant follicle, giant follicular, granulomatous, histiocytic, large cell, immunoblastic, large cleaved cell, large noncleaved cell, Lennert's, lymphoblastic, lymphocytic, intermediate; lymphocytic, intermediately differentiated, plasmacytoid; poorly differentiated lymphocytic, small lymphocytic, well differentiated lymphocytic, lymphoma of cattle; MALT, mantle cell, mantle zone, marginal zone, Mediterranean lymphoma mixed lymphocytic-histiocytic, nodular, plasmacytoid, pleomorphic, primary central nervous system, primary effusion, small b-cell, small cleaved cell, small noncleaved cell, T-cell lymphomas; convoluted T-cell, cutaneous t-cell, small lymphocytic T-cell, undefined lymphoma, u-cell, undifferentiated, aids-related, central nervous system, cutaneous T-cell, effusion (body cavity-based), thymic lymphoma, and cutaneous T cell lymphomas.

[0262] Leukemias and other blood cell malignancies that may be targeted include acute lymphoblastic, acute myeloid, lymphocytic, chronic myelogenous, hairy cell, lymphoblastic, myeloid, lymphocytic, myelogenous, leukemia, hairy cell, T-cell, monocytic, myeloblastic, granulocytic, gross, hand mirror-cell, basophilic, hemoblastic, histiocytic, leukopenic, lymphatic, Schilling's, stem cell, myelomonocytic, prolymphocytic, micromyeloblastic, megakaryoblastic, megakaryocytic, Rieder cell, bovine, aleukemic, mast cell, myelocytic, plasma cell, subleukemic, multiple myeloma, nonlymphocytic, and chronic myelocytic leukemias.

[0263] Brain and central nervous system (CNS) cancers and tumors that may be targeted include astrocytomas (including cerebellar and cerebral), gliomas (including malignant gliomas, glioblastomas, brain stem gliomas, visual pathway and hypothalamic gliomas), brain tumors, ependymoma, medulloblastoma, supratentorial primitive neuroectodermal tumors, primary central nervous system lymphoma, extracranial germ cell tumor, myelodysplastic syndromes, oligodendroglioma, myelodysplastic/myeloproliferative diseases, myelogenous leukemia, myeloid leukemia, multiple myeloma, myeloproliferative disorders, neuroblastoma, plasma cell neoplasm/multiple myeloma, central nervous system lymphoma, intrinsic brain tumors, astrocytic brain tumors, and metastatic tumor cell invasion in the central nervous system.

[0264] Gastrointestinal cancers that may be targeted include extrahepatic bile duct cancer, colon cancer, colon and rectum cancer, colorectal cancer, gallbladder cancer, gastrointestinal carcinoid tumor, gastrointestinal carcinoid tumors, gastrointestinal stromal tumors, bladder cancer, islet cell carcinoma (endocrine pancreas), pancreatic cancer, islet cell pancreatic cancer, prostate cancer rectal cancer, and polyps associated with colorectal neoplasia. A discussion of colorectal cancer is described in Barderas et al., *Cancer Research* 72: 2780-2790 (2012).

[0265] Bone cancers that may be targeted include osteosarcoma and malignant fibrous histiocytomas, bone marrow cancers, bone metastases, osteosarcoma/malignant fibrous histiocytoma of bone, and osteomas and osteosarcomas. Breast cancers that may be targeted include small cell carcinoma and ductal carcinoma.

[0266] Lung and respiratory cancers that may be targeted include bronchial adenomas/carcinoids, esophagus cancer esophageal cancer, esophageal cancer, hypopharyngeal can-

cer, laryngeal cancer, hypopharyngeal cancer, lung carcinoid tumor, non-small cell lung cancer, small cell lung cancer, small cell carcinoma of the lungs, mesothelioma, nasal cavity and paranasal sinus cancer, nasopharyngeal cancer, nasopharyngeal cancer, oral cavity and lip cancer, oropharyngeal cancer; paranasal sinus and nasal cavity cancer, and pleuropulmonary blastoma.

[0267] Urinary tract and reproductive cancers that may be targeted include cervical cancer, endometrial cancer, ovarian epithelial cancer, extragonadal germ cell tumor, extracranial germ cell tumor, extragonadal germ cell tumor, ovarian germ cell tumor, gestational trophoblastic tumor, spleen, kidney cancer, ovarian cancer, ovarian epithelial cancer, ovarian germ cell tumor, ovarian low malignant potential tumor, penile cancer, renal cell cancer (including carcinomas), renal cell cancer, renal pelvis and ureter (transitional cell cancer), transitional cell cancer of the renal pelvis, and ureter, gestational trophoblastic tumor, testicular cancer, ureter and renal pelvis, transitional cell cancer, urethral cancer, endometrial uterine cancer, uterine sarcoma, vaginal cancer, vulvar cancer, ovarian carcinoma, primary peritoneal epithelial neoplasms, cervical carcinoma, uterine cancer and solid tumors in the ovarian follicle), superficial bladder tumors, invasive transitional cell carcinoma of the bladder, and muscle-invasive bladder cancer.

[0268] Skin cancers and melanomas (as well as nonmelanomas) that may be targeted include cutaneous t-cell lymphoma, intraocular melanoma, tumor progression of human skin keratinocytes, basal cell carcinoma, and squamous cell cancer. Liver cancers that may be targeted include extrahepatic bile duct cancer, and hepatocellular cancers. Eye cancers that may be targeted include intraocular melanoma, retinoblastoma, and intraocular melanoma Hormonal cancers that may be targeted include: parathyroid cancer, pineal and supratentorial primitive neuroectodermal tumors, pituitary tumor, thymoma and thymic carcinoma, thymoma, thymus cancer, thyroid cancer, cancer of the adrenal cortex, and ACTH-producing tumors.

[0269] Miscellaneous other cancers that may be targeted include advanced cancers, AIDS-related, anal cancer, adrenal, cortical, aplastic anemia, aniline, betel or buyo cheek, cerebriform, chimney-sweeps, clay pipe, colloid, contact, cystic, dendritic, cancer a deux, duct, dye workers, encephaloid, cancer en cuirasse, endometrial, endothelial, epithelial, glandular, cancer in situ, kang, kangri, latent, medullary, melanotic, mule-spinners', non-small cell lung, occult cancer, paraffin, pitch workers', scar, schistosomal bladder, scirrhous, lymph node, small cell lung, soft, soot, spindle cell, swamp, tar, and tubular cancers.

[0270] Miscellaneous other cancers that may be targeted also include carcinoid (gastrointestinal and bronchial) Castleman's disease chronic myeloproliferative disorders, clear cell sarcoma of tendon sheaths, Ewing's family of tumors, head and neck cancer, lip and oral cavity cancer, Waldenstrom's macroglobulinemia, metastatic squamous neck cancer with occult primary, multiple endocrine neoplasia syndrome, multiple myeloma/plasma cell neoplasm, Wilms' tumor, mycosis fungoides, pheochromocytoma, sezary syndrome, supratentorial primitive neuroectodermal tumors, unknown primary site, peritoneal effusion, malignant pleural effusion, trophoblastic neo-plasms, and hemangiopericytoma.

[0271] In exemplary aspects, the cancer is any one of the foregoing described in which $IL13R\alpha 2$ is expressed on the

cells of the cancer. In exemplary aspects, the cancer is colon cancer. In exemplary aspects, the cancer is Glioblastoma Multiforme. In exemplary aspects, the method of treating cancer in a subject in need thereof comprises administering to the subject any of the binding agents, conjugates, nucleic acids, vectors, host cells, cell populations, or pharmaceutical compositions described herein, in an amount effective to treat the cancer. In exemplary aspects, the method comprises administering a conjugate described herein. In exemplary aspects, the method comprises administering host cells of the disclosures and the host cells are autologous cells in relation to the subject being treated. In exemplary aspects, the method comprises administering host cells of the disclosures and the host cells are cells obtained from the subject being treated. In exemplary aspects, the cells are T-lymphocytes. In alternative aspects, the cells are natural killer cells. [0272] The disclosure will be more fully understood by reference to the following examples, which detail exemplary embodiments of the disclosure. The examples should not, however, be construed as limiting the scope of the disclosure.

Example 1

[0273] Materials

[0274] Lipofectamine 2000 and the pEF6/Myc-His vector were obtained from Invitrogen. Monoclonal antibodies to IL13Ra2 (clones YY-23Z and B-D13) and the IsoStrip mouse monoclonal antibody isotyping kit were purchased from Santa Cruz Biotechnology (Santa Cruz, Calif.). The mAb to IL13R α 2 (clone 83807) and recombinant human and mouse IL13Ra2hFc and IL13Ra1hFc chimeras were purchased from R&D Systems (Minneapolis, Minn.). Biotinylated horse anti-mouse antibodies and the Elite kit were obtained from Vector Laboratories (Burlingame, Calif.). 3,3'-Diaminobenzidine substrate was purchased from Dako (Carpinteria, Calif.). Goat anti-mouse antibody conjugated with peroxidase was purchased from Chemicon International (Temicula, Calif.), and Pngase F was purchased from New England Biolabs (Ipswich, Mass.). The QuikChange LightningTM site-directed mutagenesis kit was purchased from Agilent Technologies, Inc. (Santa Clara, Calif.), and the RNeasy Plus™ kit was received from Qiagen (Valencia, Calif.). The cDNA iScript[™] kit, 7.5% Tris-HCl gel, and ImmunStarTM WesternCTM developing reagent and protein marker were purchased from Bio-Rad. The human IL-13 ELISA kit was purchased from eBioscience (San Diego, Calif.). GBM12 and GBM43 were kindly provided by Dr. David C. James (University of California-San Francisco), and the cDNA encoding human wild-type IL13R α 2 was obtained from Dr. Waldemar Debinski (Wake Forest University). Obtaining the cDNA encoding the human wild-type IL13R α 2 or most other proteins involves the use of wellknown techniques and readily available reagents.

[0275] Cell Lines

[0276] U373 (GBM), 293T (human embryonic kidney), and Raji (Burkitt's lymphoma) cell line were purchased from the American Type Culture Collection (ATCC; Manassas, Va.). The generation of U373 cells expressing enhanced green fluorescent protein and firefly luciferase (U373.eGFP. ffLuc), 293T cells expressing green fluorescent protein (293T.GFP) or IL13R α 2 and GFP (293T.IL13R α 2.GFP) were previously reported. See Chow et al., Mol. Ther. 21:629-637 (2013); Krebs et al., Cytotherapy 16:1121-1131 (2014). Cell lines were grown in RPMI or DMEM (Thermo Scientific HyClone, Waltham, Mass.; Lonza, Basel, Switzerland) with 10% fetal calf serum (FCS; HyClone, Logan, Utah) and 2 mM GlutaMAX-ITM (Invitrogen, Carlsbad, Calif.). The Characterized Cell Line Core Facility at MD Anderson Cancer Center, Houston, Tex., performed cell line validation.

[0277] Immunization

[0278] To obtain monoclonal antibodies with specificity to native IL13R α 2, the human recombinant IL13R α 2hFc fusion was used for immunization of animals and in all screening assays. Two 6-week-old female BALB/c mice were immunized with intraperitoneal injection of 10 µg of rhIL13Ra2hFc protein in complete Freund's adjuvant followed by intraperitoneal injection of 10 µg of rhIL13Ra2hFc protein in incomplete Freund's adjuvant at a 2-week interval for 2 months. Two weeks after the last intraperitoneal injection and 3 days before the fusion, a boost was performed by the combination of intravenous and intraperitoneal injection of 10 µg of antigen without Freund's adjuvant. The fusion of mouse spleen cells with the mouse myeloma cell line X63.Ag8.653 subclone P3O1 was performed by using a procedure described by Köhler and Milstein (27). Hybridoma supernatants were assayed for the presence of IL13Ra2 antibodies using an enzyme-linked immunosorbent assay (ELISA). Selected populations were cloned, and supernatants were assayed to identify the clones with strongest binding.

[0279] Generation of CHO Cell Line Expressing Human IL13R α 2

[0280] The cDNA encoding human wild-type IL13R α 2 was amplified with the following primer pair: forward, 5'-GCTTGGTACCGAATGGCTTTCGTTTGCTTGGC-3' (SEQ ID NO: 17) and reverse, 5'-GTTTTTGTTCGAATG-TATCACAGAAAAATTCTGG-3' (SEQ ID NO: 18). The purified PCR product was restricted with KpnI and BstBI enzymes, agarose gel-purified, and subsequently cloned into the pEF6/Myc-His vector in a reading frame with Myc and His6 tags. CHO cells were plated at 80% confluence and transfected with a plasmid encoding the IL13Ra2 using Lipofectamine 2000. The following day, 4 µg/ml blasticidin was added for selection of cells that had stably incorporated and expressed the IL13Ra2 transcript. A stable population of cells was further subcloned in 96-well plates at a density of one cell/well. Ten days later, single clones were screened by flow cytometry for cell surface expression of IL13Ra2 using an antibody to IL13Ra2 (clone B-D13). The clone with the highest level of IL13R α 2 expression was selected and expanded for subsequent screening of hybridomas secreting IL13Ra2 antibodies.

[0281] ELISA

[0282] 96-well plates were coated with 50 μ l of human or mouse recombinant IL13R α 2hFc or IL13R α 1hFc or human control IgG at a concentration of 1 μ g/ml overnight at 4° C. Following washes with TBS-Tween 20 buffer and blocking with 1% nonfat dry milk, 50 μ l of purified antibodies, serum, or hybridoma supernatants at various dilutions were applied to the plate and incubated for 1 hour at room temperature. Bound antibodies were detected with goat anti-mouse antibodies conjugated to alkaline phosphatase following the development with alkaline phosphatase substrate. Plates were read at A405 using a UniRead 800 plate reader (BioTek). [0283] Flow Cytometry

[0284] CHO or HEK cells expressing IL13R α 2; the glioma cell lines A172, N10, U251, U87, and U118; patientderived GBM12 and GBM43, and primary human astrocytes were stained with IL13R α 2 (clone 47) monoclonal antibody at 1 µg/ml followed by goat anti-mouse Alexa Fluor 647 (1:500). All staining procedures were performed on ice. Samples were analyzed using the BD FACSCanto flow cytometer and FACSDiVaTM software.

[0285] For the experiments disclosed in Examples 13-16, a FACSCalibur instrument (BD Bioscience, Mountain View, Calif.) was used to acquire immunofluorescence data that were analyzed with CellQuest (BD) or FCS Express software (De Novo Software, Los Angeles, Calif.). Isotype controls were immunoglobulin G1-fluorescein isothiocyanate (IgG1-FITC; BD Bioscience) and IgG1-phycoerythrin (IgG1-PE; BD Bioscience). SSR 47-CAR expression was detected by staining T cells with an IL13R α 2 chimera followed by Fc-FITC (Milipore) or Fc-PE (SouthernBiotech). LSR 47-CARs were detected using Fc-FITC or Fc-PE. U373 cells were analyzed for PD-L1 expression using a CD271 PE antibody (BD Bioscience). Forward- and sidescatter gating were used to discriminate live cells from dead cells. Cells were collected and washed once with phosphatebuffered saline (PBS) containing 1% FBS (Sigma; FACS buffer) prior to the addition of antibodies. Cell were incubated for 30 minutes on ice in the dark, washed once, and fixed in 0.5% paraformaldehyde/FACS buffer prior to analysis.

[0286] PCR

[0287] To determine the expression of IL13R α 2 in various glioma cells and astrocytes, total RNA was generated from the cell pellets using the RNeasy Plus kit. 200 ng of total RNA was then converted into cDNA using the cDNA iScript kit. The cDNA was further amplified by PCR for IL13R α 2 and GAPDH for 30 cycles using IL13R α 2 and GAPDH primers and visualized on a 1% agarose gel.

[0288] Surface Plasmon Resonance

[0289] The affinity and rates of interaction between IL13Ra2 (clone 47) monoclonal antibody, commercially available IL13Ra2 monoclonal antibodies (clones 83807 and B-D13), and target (rhIL13R α 2) were measured with a Biacore 3000 biosensor through surface plasmon resonance (SPR). The monoclonal antibodies were immobilized (covalently) to the dextran matrix of the sensor chip (CMS) using the amino coupling kit. The carboxyl groups on the sensor surfaces were activated with an injection of a solution containing 0.2M N-ethyl-N'-(3-diethylamino-propyl)-carbodiimide and 0.05M N-hydroxysuccinimide. The immobilization procedure was completed by the injection of 1Methanolamine hydrochloride to block the remaining ester groups. All steps of the immobilization process were carried out at a flow rate of 10 µl/minute. The control surface was prepared similarly with the exception that running buffer was injected rather than monoclonal antibodies. Binding reactions were performed at 25° C. in HBS-P buffer (20 mM HEPES, pH 7.4, 150 mM NaCl, and 0.005% (v/v) surfactant P20) using a flow rate of 20 µl/minute. Target (rhIL13Ra2) was added at various concentrations in the flow during the binding phase. The amount of protein bound to the sensor chip was monitored by the change in refractive index (represented by response units (RU)). The instrument was programmed to perform a series of binding measurements with increasing concentrations of target over the same surface. Triplicate injections of each concentration of target were performed. Sensorgrams (plots of changes in RU on the surface as a function of time) were analyzed using BIAevaluation v4.1. Affinity constants were estimated by curve fitting using a 1:1 binding model.

[0290] Data Preparation and Kinetic Analysis

[0291] The estimation of kinetic parameters was performed by repetitive injections of a range of target concentrations over the immobilized mAbs. Data were prepared by the method of "double referencing." This method utilizes parallel injections of each target sample over a control dextran surface as well as running buffer injections over both the immobilized mAbs and control dextran surfaces. Subtraction of these sensorgrams yielded the control; this was subtracted from the experimental sensorgram. Each data set (consisting of sensorgrams of increasing target concentrations over the same level of immobilized mAbs) was analyzed using various kinetic models. The BIAevaluation v 4.1 software was then used for data analysis. Affinity constants were estimated by curve fitting using a 1:1 binding model. Sensorgram association and dissociation curves were fit locally or globally. The rate of complex formation during the sample injection is described by an equation of the following type: dR/dt=k_aC(R_{max}-R)-k_dR (for a 1:1 interaction) where R is the SPR signal in RU, C is the concentration of analyte, R_{max} is the maximum analyte binding capacity in RU, and dR/dt is the rate of change of SPR signal. The early binding phase (300 s) was used to determine the association constant (k_a) between mAb and target. The dissociation phase (k_a) was measured using the rate of decline in RU on introduction of free buffer at the end of target injections. Data were simultaneously fit by the software program (global fitting algorithm), and the dissociation constant (K_D) of the complexes was determined as the ratio k_a/k_d . For quantitative analysis, three independent replicates were performed for each sample. Data are expressed as mean±S.E.

[0292] Competitive Binding Assay

[0293] For the competitive binding plate assay, a 96-well plate was coated with 50 µl of affinity-purified hrIL13R α 2hFc at 1 µg/ml in carbonate buffer, pH 9.6 and stored overnight at 4° C. After washing with PBS containing 0.05% Tween 20, mAbs to IL13R α 2 (10 µg/ml) or control mIgG were added for 30 minutes at room temperature. After washing, 50 µl of purified rhIL-13 in PBS and 0.1% BSA at 10 ng/ml were added for a 1-hour incubation at room temperature and assayed for bound rhIL-13 using detection reagents from a human IL-13 ELISA kit. Separately, HEK cells expressing wild-type IL13Ra2 or 4-amino-acid mutants (see Example 10) in the IL13R α 2 sequence were pretreated with either rhIL-13 or mAb IL13R α 2 (clone 47) at 2 µg/ml for 30 minutes on ice followed by a 1-hour incubation with IL13R α 2 (clone 47) mAb or rhIL-13 at 100 ng/ml, respectively. Binding of rhIL-13 to IL13R α 2 alone or in the presence of competitor was detected with human IL-13 mAb-FITC. Binding of IL13Ra2 (clone 47) mAb to rhIL13Ra2 alone or in the presence of competitor was detected with anti-mouse antibody conjugated to Alexa Fluor 649 and analyzed by flow cytometry.

[0294] Mutagenesis of IL13R α

[0295] Previously, Tyr²⁰⁷, Asp²⁷¹, Tyr³¹⁵, and Asp³¹⁸ of the human IL13R α 2 were identified as residues crucial for interaction with human IL-13 (28). To determine whether those residues were important for binding of $IL13R\alpha 2$ (clone 47) mAb to IL13R α 2, the Tyr²⁰⁷, Asp²⁷¹, Tyr³¹⁵, and Asp³¹⁸ residues were mutated to Ala separately or at the same time (4-amino-acid mutant) using the QuikChange Lightning site-directed mutagenesis kit according to the manufacturer's recommendations. Sequencing of selected clones was performed using conventional techniques, which confirmed the presence of the selected mutation. HEK cells were transfected with wild-type or mutated variants of IL13Ra2 cDNA in the pEF6 Myc-His vector using Lipofectamine Plus transfection reagent. 48 hours after transfection, the cells were collected and analyzed for binding to IL13Rα2 (clone 47) mAb via flow cytometry.

[0296] Western Blot [0297] The rhIL13R α 2 was applied to a 7.5% Tris-HCl gel (Bio-Rad) at 200 ng/lane and resolved under reducing conditions. After the transfer of proteins to a PVDF membrane (Bio-Rad) and blocking with 2% nonfat dry milk, the membrane was stained with anti-IL13Ra2 mAb (clones YY-23Z and B-D13) at 2 µg/ml or with supernatant collected from hybridoma clones (diluted 10 times), followed by goat anti-mouse antibody conjugated to peroxidase. Immun-StarTM WesternCTM was used to develop reactions. Images were captured using a Bio-Rad ChemiDoc imaging system. [0298] For experiments disclosed in Examples 13-16, cells were dissociated with PBS+3 mM EDTA and lysed in a buffer containing 50 mM Tris, 150 mM NaCl, 5 mM EDTA, 1% Triton X-100 (all from Sigma, St. Louis, Mo.), and protease inhibitors (Thermo Scientific, Waltham, Mass.). Protein concentrations were determined using a Bio-Rad protein assay (Bio-Rad, Hercules, Calif.) with bovine serum albumin (BSA) as the standard. Samples were denatured in Laemmli buffer (Bio-Rad) at 95° C. for 5 minutes. 5 µg of protein were loaded per well and run on a 10% polyacrylamide gel. Proteins were transferred to nitrocellulose membranes (BioRad). Membranes were blocked with 5% milk powder (MP) in Tris-buffered saline (TBS)+0.1% Tween-20 (Sigma) and then probed with anti-CD3.5 (sc-1239, Santa Cruz Biotechnology, Inc., Santa Cruz, Calif.) or GAPDH (sc-47724, Santa Cruz Biotechnology, Inc., Santa Cruz, Calif.) mouse monoclonal antibodies followed by a horseradish peroxidase (HRP) conjugated goat anti-mouse IgG antibody (sc-2005, Santa Cruz Biotechnology, Inc.). Blots were developed using SuperSignal West Dura Extended Duration Substrate (Thermo Scientific) and exposed to GeneMate Blue Basic Autoradiography Film (BioExpress, Kaysville, Utah).

[0299] Immunohistochemistry

[0300] The GBM tissues were collected in accordance with a protocol approved by the Institutional Review Board at the University of Chicago. Flash-frozen brain-tumor tissues were cut to a thickness of 10 µm. Tissue sections were fixed with -20° C. methanol and stained for human IL13R α 2 using mouse IL13R α 2 (clone 47) mAb at a concentration of 3 µg/ml or isotype control mIgG1. The bound antibodies were detected with biotinylated horse anti-mouse antibodies (1:100). The antigen-antibody binding was detected by the Elite kit with 3,3'-diaminobenzidine substrate. Slides were analyzed using the CRI Panoramic Scan Whole Slide Scanner and Panoramic Viewer software. [0301] Animal Study

[0302] All animals were maintained and cared for in accordance with the Institutional Animal Care and Use Committee protocol and according to National Institutes of Health guidelines. The animals used in the experiments were 6- to 7-week-old male athymic nu/nu mice. Mice were

anesthetized with an intraperitoneal injection of ketamine hydrochloride/xylazine (25 mg/ml/2.5 mg/ml) mixture. To establish intracranial tumors, a midline cranial incision was made, and a right-sided burr hole was placed 2 mm lateral to the sagittal suture and about 2 mm superior to λ . Animals were positioned in a stereotactic frame, and a Hamilton needle was inserted through the burr hole and advanced 3 mm. Intracranial penetration was followed by (i) injection of 2.5×10⁴ U251 glioma cells in 2.5 µl of sterile PBS in combination with 200 ng of mIgG or IL13Ra2 (clone 47) mAb or (ii) 3 days post-intracranial injection of glioma cells with PBS or 10 µg of IL13Ra2 (clone 47 or B-D13) mAb as described previously (29, incorporated herein by reference). All mice were monitored for survival. Three animals from each group were sacrificed at day 17, and brains were harvested and frozen for sectioning, hematoxylin and eosin (H&E) staining, and microscopic analysis.

[0303] Animal experiments disclosed in Examples 13-16 followed a protocol approved by the Baylor College of Medicine Institutional Animal Care and Use Committee. Experiments were performed as described in Ahmed et al., Clin. Cancer Res. 16:474-485 (2010) (incorporated herein by reference) with a few modifications. ICR-SCID mice were purchased from Taconic (IcrTac:ICR-Prkdcscid; Fox Chase C.B-17 SCID™ ICR; Taconic, Hudson, N.Y.). Male 7- to 9-week-old mice were anesthetized, head were shaved and the mice were immobilized in a Cunningham[™] Mouse/ Neonatal Rat Adaptor (Stoelting, Wood Dale, Ill.) stereotactic apparatus fitted into an E15600 Lab Standard Stereotaxic Instrument (Stoelting), and then scrubbed with 1% povidone-iodine. A 10 mm skin incision was made along the midline. The tip of a 30G 1/2 inch needle mounted on a Hamilton syringe (Hamilton, Reno, Nev.) served as the reference point. A 1 mm burr-hole was drilled into the skull 1 mm anterior and 2 mm to the right of the bregma. 1×10^5 U373.eGFP.ffLuc cells in 2.0 µL were injected 3 mm deep to the bregma, corresponding to the center of the right caudate nucleus over 5 minutes. The needle was left in place for 3 minutes to avoid tumor cell extrusion, and then withdrawn over 5 minutes. Seven days after tumor cell injection, animals were treated with 2×10^6 effector cells in 2 µL to the same tumor coordinates. The incision was closed with 2-3 interrupted 7.0 Ethilon sutures (Ethicon, Inc., Somerville, N.J.). A subcutaneous injection of 0.03-0.1 mg/kg buprenorphine (Buprenex® RBH, Hull, England) was given for pain control.

[0304] Generation of Retroviral Vectors Encoding IL13R α 2-scFv-Specific CARs

[0305] A codon-optimized gene was synthesized by GeneArt (Invitrogen, Carlsbad, Calif.) containing the immunoglobulin heavy-chain leader peptide37, and scFv47 flanked by 5' NcoI and 3' BamHI sites. This mini gene was subcloned into SFG retroviral vector containing IL13Ra2specific CARs (47-CARs) with short or long spacer regions (SSRs, LSRs) and CD28. \, CD28. OX40. \, CD28.41BB. \, or 41BB. contained a contained a CD28 transmembrane domain except for 47.SSR.CAR. 41BB.ζ, which had a CD8α transmembrane domain. 47.SSR.CAR and 47.LSR.CAR without an endodomain (47.SSR.CAR. Δ and 47.LSR.CAR. Δ) were generated by PCR cloning. All cloning of the CARs were verified by sequencing (Seqwright, Houston, Tex.). RD114pseudotyped retroviral particles were generated by transient transfection of 293T cells as previously described in Johnson et al., Sci. Transl. Med. 7:275ra22 (2015), incorporated herein by reference.

[0306] Generation of CAR T Cells

[0307] Human peripheral blood mononuclear cells (PB-MCs) from healthy donors were obtained under a Baylor College of Medicine IRB-approved protocol, after informed consent was obtained in accordance with the Declaration of Helsinki. To generate 47-CAR T cells, PBMCs were isolated by Lymphoprep (Greiner Bio-One, Monroe, N.C.) gradient centrifugation and then stimulated on non-tissue culture treated 24-well plates, which were precoated with OKT3 (CRL-8001, ATCC) and CD28 (BD Bioscience, Mountain View, Calif.) antibodies. Recombinant human interleukin-7 (IL7) and IL15 (IL7, 10 ng/mL; IL15, 5 ng/mL; Proleukin; Chiron, Emeryville, Calif.) were added to cultures on day 2 (Xu et al., Blood 123:3750-3759 (2014), incorporated herein by reference). On day 3, OKT3/CD28-stimulated T cells $(2.5 \times 10^5 \text{ cells/well})$ were transduced on RetroNectin® (Clontech, Mountainview, Calif.)-coated plates in the presence of IL7 and IL15. On day 5 or 6, T cells were transferred into new wells and subsequently expanded with IL-7 and IL15. Non-transduced (NT) T cells were activated with OKT3/CD28 and expanded in parallel with IL-7 and IL15. 47-CAR expression was determined 3 to 4 days posttransduction.

[0308] Co-Culture Assav

[0309] Recombinant Protein Co-Culture Assay.

[0310] Non-tissue culture 24-well plates were precoated with recombinant human IL13R α 1, IL13R α 2, or IL4R protein, (R&D Systems, Minneapolis, Minn.) at a final concentration of 500 ng/well. Plates were washed once using RPMI, and CAR or NT T cells were plated. After 24 hours, supernatants were harvested and interferon γ (IFN γ) and Interleukin 2 (IL2) release were measured by ELISA according to the manufacturer's instructions (R&D Systems, Minneapolis, Minn.).

[0311] Cell Culture Co-Culture Assay.

[0312] CAR T cells were co-cultured with target cells at a 1:2 effector to target (E:T) ratio in a 24-well plate. NT T cells served as controls. After 24 hours, culture supernatants were harvested, and the presence of IFN γ and IL2 was determined by ELISA according to the manufacturer's instructions (R&D Systems, Minneapolis, Minn.).

[0313] Cytotoxicity Assay

[0314] Standard chromium (⁵¹Cr) release assays were performed as described in Gottschalk et al., Blood 101: 1905-1912 (2003), incorporated herein by reference. Briefly, 1×10^{6} target cells were labeled with 0.1 mCi (3.7MBq)⁵¹Cr and mixed with decreasing numbers of effector cells to give effector to target ratios of 40:1, 20:1, 10:1, and 5:1. Target cells incubated in complete medium alone or in 1% Triton X-100 were used to determine spontaneous and maximum ⁵¹Cr release, respectively. After 4 hours, supernatants were collected and radioactivity was measured in a gamma counter (Cobra Quantum; PerkinElmer; Wellesley; MA). The mean percentage of specific lysis of triplicate wells was calculated according to the following formula: [test releasespontaneous release]/[maximal release-spontaneous release]×100.

[0315] Bioluminescence Imaging

[0316] Isofluorane anesthetized animals were imaged using the IVIS® system (IVIS, Xenogen Corp., Alameda, Calif.) 10-15 minutes after 150 mg/kg D-luciferin (Xeno-

gen) per mouse was injected intraperitoneally. The photons emitted from the luciferase-expressing tumor cells were quantified using Living Image software (Caliper Life Sciences, Hopkinton, Mass.). A pseudo-color image representing light intensity (blue least intense and red most intense) was generated and superimposed over the grayscale reference image. Mice were euthanized when the tumor radiance was greater than 1×10^9 on two occasions or when they met euthanasia criteria (neurological deficits, weight loss, signs of distress) in accordance with the Center for Comparative Medicine at Baylor College of Medicine.

[0317] Statistics

[0318] The differences between groups were evaluated by Student's t test or one-way analysis of variance with post hoc comparison Tukey's test or Dunnett's test. For the in vivo survival data, a Kaplan-Meier survival analysis was used, and statistical analysis was performed using a log rank test. P<0.05 was considered statistically significant.

[0319] For the experiments disclosed in Examples 13-16, the in vitro experiments were performed at least in triplicate, and GraphPad Prism 5 software (GraphPad software, Inc., La Jolla, Calif.) was used for statistical analysis. Measurement data were presented as mean \pm standard deviation (SD). The differences between means were tested by appropriate tests. The significance level used was P<0.05. For the mouse experiments, changes in tumor radiance from baseline at each time point were calculated and compared between groups using t-test. Survival, determined from the time of tumor cell injection, was analyzed by the Kaplan-Meier method and by the log-rank test.

Example 2

[0320] Characterization of Antigen and Screening of Hybridoma Clones Secreting Anti-IL13Rα2 Antibodies [0321] The primary goal of this study was to generate a high affinity monoclonal antibody suitable for targeting of the IL13R α 2 expressed on the surface of tumor cells. We therefore immunized mice and screened the resulting hybridoma clones for reactivity against the antigen, rhIL13Ra2, in its native conformation. A plate-bound ELISA utilizing a hybridoma clone against rhIL13Rα2, YY-23Z, was established for the detection of rhIL13R α 2. The concentration of rhIL13R α 2 absorbed to the plastic at 1 ug/ml was found to be suitable for the detection of antibody binding (FIG. 1A). Next, the rhIL13Ra2hFc was characterized for its "nativity" by utilizing a pair of commercially available antibodies recognizing only the native (found on the cell surface) and denatured (using Western blotting under reducing conditions) forms of IL13R α 2 and for its binding properties to rhIL13R α 2 in ELISA with antibody clones B-D13 and YY-23Z, respectively. Both clones B-D13 and YY-23Z were able to recognize the rhIL13Ra2hFc in a plate-bound ELISA (FIG. 1B). Denaturation of antigen at 95° C. for 5 minutes in the presence of β -mercaptoethanol completely abolished the ability of the antibody clone B-D13 to recognize antigen by ELISA, whereas the YY-23Z clone retained the ability to bind the denatured antigen. Thus, the rhIL13Ra2hFc absorbed to the plastic of ELISA plates containing both native and denatured forms of the protein. Analysis of serum from animals immunized with a fusion of rhIL13R α 2 and hFc revealed the presence of antibodies against both rhIL13Ra2 and human Fc fragment. To select antibodies specific for the IL13Ra2 portion of the fusion, human IgG was included as an additional negative control for the screening of hybridoma populations. Of the 39 screened primary populations, only 15 populations were specific to IL13R α 2, and four were reactive with human IgG. Finally, five clones strongly reacting with native IL13R α 2 were further expanded and recloned. The two clones recognizing only denatured antigen were selected from the separate immunization set with rhIL13Ra2hFc chimera. Supernatants from selected clones were compared for their ability to bind hrIL13R α 2 in a plate-bound ELISA (FIG. 1C) and by Western blotting (FIG. 1D). FIG. 1C shows that clone 47 strongly binds to the antigen in plate-bound ELISA but not by Western blotting, indicating the ability of clone 47 to recognize a native conformation of the antigen. Therefore, clone 47 was selected for further characterization and for further experiments. Clone 47 was found to be of the IgG1 isotype, possessing a k chain.

Example 3

[0322] Specificity of Binding for the IL13R α 2 (Clone 47) mAb to Recombinant Human IL13R α 2 and IL13R α 2 Expressed at the Cell Surface

[0323] We investigated the binding properties of the IL13R α 2 (clone 47) mAb to rhIL13R α 2 versus the commercially available clones 83807 and B-D13 in a platebound ELISA. FIG. 2A shows strong and specific binding of clone 47 to rhIL13Ra2 when compared with clones 83807 and B-D13. Clone 47 reached the plateau of binding at the low concentration of 0.05 µg/ml. None of the antibodies showed binding to human IgG utilized as an additional negative control in these experiments. To further verify the specificity of interaction for clone 47 with human IL13R α 2, a clonal line of CHO cells expressing the full size wild-type human IL13Ra2 (clone 6) was generated. Binding of the antibody to control CHO cells transfected with an empty vector was compared with that of CHO cells expressing IL13Ra2. Again, the IL13Ra2 (clone 47) mAb demonstrated strong and specific binding to $IL13R\alpha 2$ expressed on the cell surface but not to control CHO cells, indicating that this antibody specifically recognizes a native conformation of the IL13Ra2 (FIG. 2B). Clone 47 demonstrated the strongest affinity for IL13Ra2 at the lowest tested concentration of 0.25 µg/ml. Notably, other selected hybridoma clones demonstrated similar specificity of interaction with IL13R α 2 expressed on the cell surface of CHO cells but not with control CHO cells. Data obtained in a plate-bound ELISA also revealed that clone 47 does not interact with the low affinity receptor for IL-13, the IL13R α 1 (FIG. 2C), or mouse recombinant IL13R α 2, further validating the specificity of interaction between clone 47 and IL13R α 2 (FIG. 2D). Clones 83807 and B-D13 did not show binding to mouse rIL13R α 2 in agreement with current understanding of the cross-reactivity of these antibodies with mouse IL13Rα2.

[0324] We next characterized the binding capacity of clone 47 with various glioma cell lines, the patient-derived glioma lines GBM12 and GBM43, and normal human astrocytes. Increased expression of the IL13R α 2 gene relative to normal brain tissue is reported in 44-47% of human GBM resected specimens (3) and in up to 82% (14 of 17) primary cell cultures derived from GBM and normal brain explants (2). FIG. **3**, A and B, show the flow charts of the comparative staining of glioma cells, human astrocytes, and HEK cells expressing recombinant human IL13R α 2 on the

cell surface with the IL13R α 2 (clones 47, 83807, and B-D13) mAb. FIG. 3, A and B, reveal (i) various levels of IL13R α 2 expression on the cell surface and (ii) superior binding of the clone 47 versus clones B-D13 (1.2-4.6-fold difference between the cell lines) and 83807 to the surface of analyzed cell lines. Interestingly, we observed a near complete absence of the binding of clone 83807 to glioma cell lines in contrast to HEK cells expressing IL13R α 2. No binding of clone 47 was detected with normal human astrocytes, confirming the specificity of interaction of clone 47 with human glioma cells expressing IL13R α 2. The expression of IL13R α 2 mRNA in these cells generally correlates with the level of IL13R α 2 expression on the cell surface. Moreover, cells expressing low to no mRNA expression for IL13R α 2, including U118 and primary human astrocytes, demonstrated low to no expression for IL13Ra2 on the cell surface (FIG. 3B). In additional experiments, N10 glioma cells were incubated with either the IL13R α 2 (clone 47) mAb at 1 μg/ml or the IL13Rα2 (clone 47) mAb preincubated with a 10-fold excess of rhIL13R α 2 (FIG. 10) and analyzed by flow cytometry. A significant ablation of interaction between the IL13Ra2 (clone 47) mAb in the presence of a 10-fold excess of rhIL13Ra2 was found when compared with clone 47 alone. Similarly, preincubation of N10 cells with either a 10-fold excess of rhIL-13 or IL13Ra2 (clone 47) mAb almost completely blocked the interaction between the antibody or rhIL-13 and N10 cells (supplemental FIG. 1B), indicating a specificity of recognition between IL13R α 2 expressed on the surface of glioma cells and clone 47 (FIG. 10).

[0325] To verify that the IL13R α 2 (clone 47) mAb possessed the ability to bind IL13R α 2 on the surface of glioma cells in situ, intracranial glioma xenografts of U251 cells expressing green fluorescent protein (GFP) were established in nude mice. Three weeks later, animals were sacrificed, and cells were obtained and placed into in vitro culture conditions. After 48 hours, the cells were collected and stained with control mIgG or IL13R α 2 (clone 47) mAb. Cultured GFP-expressing U251 cells served as a positive control. GFP-positive U251 cells represented about 56% of the total cells (FIG. 3C, panel a), and 96% of the cells were reactive with the IL13Ra2 (clone 47) mAb (FIG. 3C, panel c), whereas GFP-negative cells did not interact with the antibody (FIG. 3C, panel b). These data further confirm that the IL13Ra2 (clone 47) mAb specifically recognizes glioma cells expressing IL13Ra2 in mouse xenografts and is not reactive with other cells from the mouse brain.

Example 4

[0326] Affinity Studies

[0327] Surface plasmon resonance was used to determine the affinity and rate of interaction between the IL13R α 2 (clone 47) mAb and rhIL13R α 2. All measurements were done in comparison with two commercial antibodies against IL13R α 2, clones 83807 and B-D13. FIG. 4 shows the sensorgrams for each antibody. The measurements are summarized in Table 1.

TABLE 1

		nonoclonal antibodies I nan recombinant IL13	0	
mAbs to IL13Rα2	$\substack{\mathbf{k}_{a}\\1/\mathrm{MS}}$	k _d 1/S	$\mathrm{K}_D\mathrm{M}$	R _{max} RU
Clone 47 Clone 83807	9.06e4 ± 322 2.23e4 ± 620	$1.26e-4 \pm 1.07e-6$ $2.31e-3 \pm 1.03e-5$	1.39×10^{-9} 104 × 10 ⁻⁹	390 250
Clone B-D13	1.08e5 ± 5.71e3	4.99e-3 ± 1.45e-4	46.1×10^{-9}	8-16

[0328] The estimation of kinetic parameters was performed as described in Example 1. The dissociation constant (KD) of the complexes was determined as the ratio k_a/k_a . For quantitative analysis, three independent replicates were performed for each sample. Data are expressed as mean±S. E. These data demonstrate that the affinity of IL13R α 2 (clone 47) mAb to recombinant IL13R α 2 exceeds the affinity of commercially available mAb clones 83807 and B-D13 by 75-fold and 33-fold, respectively.

[0329] FIG. **4**A shows that clone 47 demonstrates a prolonged and stable association with rhIL13R α 2 measured over a 30-minute time frame, whereas clones 83807 (FIG. **4**B) and B-D13 (FIG. **4**C) dissociate relatively quickly. The affinity of binding for the IL13R α 2 (clone 47) mAb to rhIL13R α 2 was calculated at 1.39×10^{-9} M. This value exceeded the affinity of the commercially available antibody clones 83807 and B-D13 to rhIL13R α 2 by 75-fold and 33-fold, respectively. Clone 47 demonstrated the highest binding affinity (R_{max}) to rhIL13R α 2 at 390 RU when compared with 250 and 8-16 RU for clones 83807 and B-D13, respectively. These data indicate that the IL13R α 2 (clone 47) mAb possesses properties superior to clones 83807 and B-D13 as well as demonstrates a higher affinity toward rhIL13R α 2.

Example 5

[0330]~ A Monoclonal Antibody Competes with rhIL-13 for Binding to IL13Ra2

[0331] To determine whether the IL13R α 2 (clone 47) mAb possesses inhibitory properties, competitive binding assays utilizing a rhIL13Ra2hFc chimera and HEK cells transiently expressing the human IL13R α 2 were performed. The competitive binding assay was set up in a plate-bound ELISA format. The rhIL13Ra2hFc absorbed to the plate served as the target antigen. To determine whether the IL13R α 2 mAb specifically inhibits the binding of IL-13 to rhIL13R α 2, plates were preincubated with a 100-fold excess of mIgG, the IL13Ra2 (clone 47) mAb, or other IL13Ra2 mAb clones, including 83807, YY-23Z, and B-D13, followed by incubation with rhIL13. FIG. 5A shows that the IL13R α 2 (clone 47) mAb significantly abolished the binding of rhIL-13 to rhIL13Ra2, whereas the IL13Ra2 mAb clones B-D13 and 83807 exhibited significantly less competition for binding of human IL-13.

[0332] To further verify the inhibitory properties of the IL13R α 2 (clone 47) mAb, HEK 293T cells were transfected with an agent encoding wild-type or a 4-amino-acid mutant form of IL13R α 2 cDNA in which Tyr207, Asp271, Tyr315, and Asp318 residues were substituted with Ala. Previously, these residues of the human IL13R α 2 were identified as amino acids required for the interaction with the cognate ligand, IL-13. The presence of all four mutations in one

molecule has been shown to result in near complete loss of the binding of IL-13 to the mutated form of IL13R α 2 (28). After 48 hours, the cells were pretreated with a 20-fold excess of rhIL-13 or the IL13Ra2 (clone 47) mAb, followed by incubation of the IL13R α 2 (clone 47) mAb or rhIL-13, respectively. FIG. 5B shows about 50% binding inhibition of IL13Ra2 (clone 47) mAb by a 20-fold excess of rhIL-13 to wild-type (WT) IL13Ra2 but not to the 4-amino-acid mutant form of IL13Ra2. A 20-fold excess of antibody abolished the binding of rhIL-13 to IL13R α 2 when expressed on the cell surface by 80%, which is similar to the result observed in plate ELISA. The residual binding of IL-13 to the 4-amino-acid mutant form of IL13Ra2 was further decreased by an excess of the IL13R α 2 (clone 47) mAb (FIG. 5C). Collectively, these data indicate that the IL13Ra2 (clone 47) mAb specifically competes with rhIL-13 for the binding site on IL13R α 2. Also, these data indicate that the IL13Ra2 (clone 47) mAb and IL-13 have a significant overlap in their recognition site of the IL13Ra2 molecule.

Example 6

[0333] Role of the Tyr²⁰⁷, Asp²⁷¹, Tyr³¹⁵, and Asp³¹⁸ Residues for IL13R α 2 (Clone 47) mAb Binding

[0334] Taking into consideration that IL-13 and the IL13R α 2 (clone 47) monoclonal antibody can significantly compete with one other for binding of IL13R α 2, we determined whether the residues Tyr207, Asp271, Tyr315, and Asp318 contributing to the interaction of IL-13 with IL13Ra2 (28) were also important for binding of the IL13Rα2 (clone 47) mAb to IL13Rα2. The plasmids encoding cDNA for IL13Ra2 carrying individual mutations of Tyr207, Asp271, Tyr315, or Asp318 residues to Ala or a combination of all four mutations in one molecule were generated and transiently expressed in HEK cells. Binding of the IL13R α 2 (clone 47) mAb to wild-type and mutant forms of IL13Ra2 was analyzed by flow cytometry. The IL13Ra2 mAbs 83807 and B-D13 were used as reference antibodies to exclude a possible influence of variations in the level of expression of wild-type or mutated variants of IL13R α 2 on the surface of HEK cells (FIG. 6A). Data were calculated as a ratio of IL13Ra2 (clone 47) binding to IL13Ra2 when compared with both antibody clones 83807 and B-D13. FIG. 6A demonstrates that the binding of IL13R α 2 (clone 47) mAb was not significantly affected by either the individual mutations or the 4-amino-acid mutant form of IL13R α 2 when compared with wild-type receptor. In contrast, binding of IL-13 to the 4-amino-acid mutant form of IL13R α 2 was nearly abolished (FIG. 6B). These data indicate that the Tyr207, Asp271, Tyr315, and Asp318 residues are not crucial for the interaction of IL13Ra2 (clone 47) mAb with IL13Ra2 but are necessary for binding to IL-13.

Example 7

[0335] N-Linked Glycosylation Affects the Affinity of the IL13R α 2 mAb for IL13R α 2

[0336] N-Linked glycosylation has previously been demonstrated to be important for efficient binding of IL-13 to the cognate receptor, IL13R α 2 (30). Taking into consideration the significant overlap in epitope recognition between the IL13R α 2 (clone 47) mAb and IL-13, we expected N-linked glycosylation of IL13R α 2 to contribute to binding of the IL13R α 2 (clone 47) mAb. To confirm this expectation, rhIL13Ra2hFc was treated with Pngase F to remove N-linked glycosylation from the protein. The binding of the IL13Ra2 (clone 47) mAb to control and deglycosylated target protein was investigated. Treatment of rhIL13Ra2 with Pngase F was performed under native conditions (in the absence of SDS) to avoid denaturation of the rhIL13R α 2 affecting the binding of antibodies. Additional mAbs to IL13Ra2 (clones 83807, B-D13, and YY23Z) and rhIL-13 were included in the assay to demonstrate the specificity of binding. In a plate-bound ELISA, binding of the IL13R α 2 (clone 47) mAb to Pngase F-treated IL13Ra2 was decreased by 35% when compared with untreated protein (n=4; p<0. 001). The binding of the IL13R α 2 (clone 83807) was reduced by 80% when compared with untreated protein and completely absent for the IL13Ra2 mAbs B-D13 and YY-23Z, respectively (n=4; p<0.001) (FIG. 7A). Binding of rhIL-13 with Pngase F-treated rhIL13Ra2 was also significantly diminished. To verify that Pngase F treatment resulted in deglycosylation of the protein, control and Pngase F-treated rhIL13Ra2hFc protein was resolved by Western blot. FIG. 7B shows that Pngase F-treated protein has a lower molecular weight, confirming the removal of N-linked glycans from the IL13R α 2 molecule. Binding of the IL13Rα2 (clone 47) mAb to Pngase F-treated U251 glioma and HEK 293 cells expressing wild-type IL13Ra2 was also decreased by about 30% (n=3; p<0.05) when compared with control untreated cells (FIG. 7C).

Example 8

[0337] Immunohistochemistry

[0338] The ability of the IL13R α 2 (clone 47) mAb to detect IL13Ra2 was evaluated in fresh frozen tissues. Flashfrozen human GBM samples or the U251 glioma flank xenograft was stained with either isotype control mIgG1 or the IL13Ra2 (clone 47) mAb. FIG. 8 shows positive (brown) staining in the two human GBM samples, albeit with different frequency of positive cells in the sample as well as a U251 glioma cell-based glioma xenograft. Positive staining was detected in two of the three GBM samples analyzed, which is consistent with the expectation that fewer than 50% of primary GBM express IL13R α 2 (3). These data are also consistent with the ability of this antibody to recognize the native form of IL13R α 2 expressed on the cell surface and in ELISA applications, as well as the compromised ability of this mAb to detect denatured antigen by Western blotting.

Example 9

[0339] The IL13R α 2 Monoclonal Antibody Prolongs the Survival of Animals with an Intracranial Glioma Xenograft **[0340]** The potential therapeutic properties of the IL13R α 2 (clone 47) mAb were also determined in an orthotopic mouse model of human glioma. U251 glioma cells were intracranially injected into the brain of nude mice alone, in the presence of control mIgG, or with the IL13R α 2 (clone 47) mAb. FIG. **9**A shows that animals in the control PBS (n=15) and mIgG (n=16) groups demonstrated a similar median survival of 27 and 25 days, respectively. In contrast, the survival of animals co-injected with the IL13R α 2 (clone 47) mAb (n=13) was significantly increased to a median of 34 days (p=0.0001; mIgG versus the IL13R α 2 mAb group). Analysis of H&E staining of the glioma xenografts from

brains collected on day 17 revealed a similar pattern of glioma cell distribution in the brain of control groups. In contrast, the tumor mass in the group of animals co-injected with IL13Ra2 mAb was significantly decreased in size (FIG. 9B). Independently, U251 cells were inoculated in the brains of mice and 3 days later injected through the same burr hole with either PBS or the IL13Ra2 (clone 47 or B-D13) mAb as described previously (29). Interestingly, the mice injected with clone 47 demonstrated improvement in median survival when compared with PBS and clone B-D13 groups (35 days versus 27 and 23 days, respectively; n=7; p>0.05) (FIG. 11), similar to what was found in the coinjection experiment (FIG. 9A). Nevertheless, all animals ultimately succumbed to the disease. These data indicate that the IL13R α 2 (clone 47) mAb shows promise in promoting tumor rejection of IL13Ra2-expressing U251 glioma cells in the mouse brain. This finding leads to the expectation that antibody agent incorporating the IL13Ra2-binding domain of the IL13R α 2 (clone 47) mAb will be efficacious in treating a variety of human and non-human cancers characterized by the presentation of IL13R α 2, such as IL13R α 2expressing glioma cells and other malignant cell types.

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Example 10

[0388] A Single-Chain Antibody for Selective Targeting of IL13Rα2-Expressing Brain Tumors

[0389] IL13R α 2 is overexpressed in a majority of highgrade astrocytomas and other malignancies, and has been validated as a target for therapeutic applications in various preclinical models. However, current IL13-based therapeutic agents lack specificity due to interaction with the IL13Ra1 receptor, which is widely expressed by normal or healthy cells. The generation of a targeting agent that strictly binds to IL13Rα2 would significantly expand the therapeutic potential for the treatment of IL13Ra2-expressing cancers. Recently, a monoclonal antibody 47 (mAb47) has been developed and extensively characterized. The mAb47 exclusively binds to a native form of human IL13R α 2. Using mAb47, a single-chain antibody (scFv) fragment was engineered from mAb47 expressed by the parental hybridoma cell line. The single-chain antibody (scFv) fragment was tested for its targeting properties as a soluble agent, and an adenovirus (Ad) with a modified fiber incorporating scFv47 as a targeting motif was agented.

[0390] The phage-display approach was utilized for selection of a functional combination of variable heavy (VH) and light (VL) chains from established hybridoma cells producing mAb47. Purified phages displaying scFv47 were tested for their interaction with IL13R α 2hFc recombinant protein, i.e., a fusion of IL13R α 2 and the Fc region of an antibody. A competitive ELISA was utilized to verify that the parental mAb47 and the scFv47 fragment bind to the same epitope. The soluble form of scFv47 expressed in *E. coli* and CHO cells was analyzed by SDS-PAGE, and tested for stability and targeting properties. To generate IL13R α 2-specific Ad,

the fiber of a replication-deficient Ad5 encoding green fluorescent protein was replaced with a chimeric fiber gene composed of a T4 fibritin trimerization domain linked at its C-terminal to scFV47 (AdFFscFv47-CMV-GFP). To generate viral particles, an agent encoding the adenoviral genome was rescued in HEK293F28 cells, propagated, and purified. IL13R α 2⁺ and IL13R α 2⁻ U251 cell lines were established via stable transfection with either control or IL13R α 2specific shRNAs (U251-IL13R α 2.KO), respectively. The AdFFscFv47-CMV-GFP virus was tested for targeting properties in these U251 cell lines and in IL13R α 2-expressing U87 cells.

[0391] The biopanning-selected pool of phages, as well several individual clones, demonstrated specific binding to IL13Ra2hFc protein, but not to hIgG in plate ELISA. Binding of scFv47-displayed phages to IL13R α 2 was completely abolished by mAb47, but not by control IgG or other tested IL13R α 2 mAbs, thus confirming the same IL13R α 2 epitope was recognized by scFv47 as was recognized by the parental mAb47. Similarly to phage-displayed scFv47, the soluble scFv47 showed specific binding to IL123R α 2, but not to IL13Ra1. Interaction of Ad5FFscFv47-CMV-GFP was also specific to IL13Ra2-expressing U251 cells, as judged by flow cytometry for GFP expression in U251-IL13Ra2⁺ versus U251-IL13Ra2.KO cells. Furthermore, GFP expression in cells infected with Ad5FFscFv47-CMV-GFP strongly correlated with the level of surface expression of IL13Ra2. The specificity of viral infection was further validated in a U251 glioma model.

[0392] The data validate scFv47 as a highly selective IL13R α 2 targeting agent that provides a soluble, singlechain biologic useful in diagnosing and treating IL13R α 2expressing cancers, such as gliomas, colon cancers (see Example 12) and others.

Example 11

[0393] Generation of an IL13Rα2-CAR

[0394] To generate an IL13R α 2-specific T cell, an IL13Ra2-specific chimeric antigen receptor (CAR) was initially constructed. A codon-optimized minigene was synthesized that contained the immunoglobulin heavy-chain leader peptide and the heavy and light chains of the IL13Ra2-specific single-chain variable fragment (scFv) separated by a linker (the scFv was derived from hybridoma 47, Balyasnikova et al. J Biol. Chem. 2012; 287(36):30215-30277). The minigene was subcloned into an SFG retroviral vector containing the human IgG1-CH2CH3 domain, a CD28 transmembrane domain, and costimulatory domains derived from CD28 and the CD32-chain. CD3/CD28-activated human T cells were transduced with RD114pseudotyped retroviral particles and subsequently expanded using IL2. Functional analysis revealed that T cells expressing IL13Ra2-specific CARs (IL13Ra2-CAR T cells) recognized recombinant IL13Ra2 protein as judged by cytokine production (IFNy and IL2; FIGS. 19 and 20), and killed IL13Ra2-positive cells in a cytotoxicity assay (FIG. 18). Non-transduced (NT) T cells did not produce cytokines and had no cytolytic activity.

Example 12

[0395] Redirecting T Cells to IL13R α 2-Positive Pediatric Glioma

[0396] IL13R α 2 is aberrantly expressed in Glioblastoma Multiforme and is, therefore, a promising target for CAR T-cell immunotherapy. The antigen recognition domain of CARs normally consists of a single-chain variable fragment (scFv), but current IL13R α 2-specific CARs use IL13 muteins as an antigen recognition domain. IL13 muteinbased CARs, however, have been shown to also recognize IL13R α 1, raising significant safety concerns. To overcome this obstacle, a high affinity IL13R α 2-specific scFv has been agented. This scFv is used in developing a scFv-based IL13R α 2-specific CAR (IL13R α 2-CAR), which, when expressed in T cells, will provide IL13R α 2-CAR T cells having cytotoxic effector function.

[0397] Antigen-specific T cells were incorporated into an effective immunotherapy for diffuse intrinsic pontine glioma (DIPG) and glioblastoma (GBM), which are the most aggressive, uniformly fatal, primary human brain tumors in children. IL13R α 2 is expressed at a high frequency in both DIPG and GBM, but not in normal brain, making it a promising target for T-cell immunotherapy, including scFv-based therapy, scFv-CAR T-cell-based therapy, and scFv fusions to other frameworks providing effector function, such as BiTEs and scFv-CAR-NKs. IL13-binding CARs have been generated using mutated forms of IL13 as CAR binding domains, but these CARs also recognize IL13R α 1, raising significant toxicity concerns.

[0398] To overcome this limitation, a high-affinity IL13R α 2-specific scFv that does not recognize IL13R α 1 was generated. A panel of IL13R α 2-CARs were agented that contain the IL13R α 2-specific scFv as an ectodomain, a short hinge (SH) or a long hinge (LH), a CD28 transmembrane domain, and endodomains that contain signaling domains derived from CD3 ζ and co-stimulatory molecules (e.g., CD28. ζ , CD137. ζ , CD28.CD137. ζ , CD28.CD134. ζ). IL13R α 2-CAR T cells were generated by retroviral transduction, and effector function was determined in vitro, using co-culture and cytotoxicity assays, and in vivo, using the U373 brain xenograft model (FIG. **21**).

[0399] Expression of all CARs in T cells was similar, as judged by Western blot analyses. CAR cell-surface expression varied, however, depending on the hinge and endodomain of the agent. In cytotoxicity assays, the various IL13R α 2-CAR T cells only killed target cells that expressed IL13Ra2 and not IL13Ra1, confirming specificity (FIG. 18). While all IL13R α 2-CAR T cells secreted significant levels of IFNy in co-culture assays with the IL13R α 2⁺ glioma cell line U373 (FIG. 19), only short-hinge CAR T cells secreted significant amounts of IL2 (FIG. 20). T cells expressing IL13R α 2-CARs with a deleted endodomain (IL13R α 2 Δ -CAR) secreted no cytokines, confirming that cytokine production depends on the presence of a functional IL13Rα2-CAR. In vivo, injection of IL13Rα2.SH.CD28.ζ-CAR T cells into U373-bearing mice resulted in regression of glioma xenografts, as judged by bioluminescence imaging (FIG. 21). IL13Rα2.LH.CD28.ζ- or IL13Rα2.Δ-CAR T cells had no antitumor effects. The data establish that a CAR that only recognizes IL13Rα2 and not IL13Rα1 was generated, and that CAR preferentially targets tumor cells expressing IL13Ra2. Comparison of several IL13Ra2-CARs revealed that a CAR with a SH and a CD28.5 endodomain resulted in significant T cell activation, as judged by IL2 production and in vivo anti-glioma activity. The results show that adoptive immunotherapy of primary human brain tumors, e.g., high-grade gliomas, in children is both feasible and promising.

Example 13

[0400] Generation of 47-CAR T Cells

[0401] Two retroviral vectors encoding CARs based on scFv47 (47-CARs; FIG. 31A)^{24,25} were initially generated. Both CARs contained an N-terminal leader sequence, a codon-optimized synthetic gene encoding scFv47, a spacer region, a CD28 transmembrane domain, and signaling domains derived from CD28 and CD3.2 (FIG. 31A). The spacer region was either the IgG1 hinge (16 amino acids; short spacer region (SSR); 47-CAR.SSR.CD28.ζ) or the IgG1-CH2CH3 domain (293 amino acids; long spacer region (LSR); 47-CAR.LSR.CD28.ζ). As controls, LSR and SSR 47-CARs without signaling domains were constructed (47-CAR.SSR.A, 47-CAR.LSR.A; FIG. 31A). CD3/CD28activated T cells from healthy donors were transduced with RD114-pseudotyped retroviral particles, and 4 to 5 days post-transduction, T-cell phenotype and CAR expression was determined by FACS analysis. CARs were expressed on the cell surface, and the transduction efficiency ranged from 69.2%-98.5% with no significant differences between constructs (FIG. 31B, C). Expression of full-length 47-CAR. SSR.CD28. c and 47-CAR.LSR.CD28. was confirmed by Western blot using a CD3. ζ antibody for detection (FIG. 31D). Phenotypic analysis revealed a mixture of CD4- and CD8-positive T cells. While the ratio of CD8- to CD4positive T cells was about 3:1 for 47-CAR.SSR.CD28.ζ, 47-CAR.SSR.A, and 47-CAR.LSR.A T-cell lines, it was about 1.5:1 for 47-CAR.LSR.CD28. (FIG. 2).

Example 14

[0402] 47-CAR T Cells Only Recognize IL13Rα2

[0403] To initially determine the specificity of 47-CARs, T cells expressing 47-CAR.SSR.CD28.ζ, 47-CAR.LSR. CD28.ζ, M47-CAR.SSR.Δ, or M47-CAR.LSR.Δ were cultured on tissue culture plates that were uncoated or coated with recombinant proteins encoding IL13R α 1, IL13R α 2, or IL4R. Non-transduced (NT) T cells and T cells expressing an IL13mutein-CAR.LSR.CD28. ζ 10 that recognizes IL13Ra1 and IL13Ra2, served as controls. T cells expressing 47-CAR.SSR.CD28.ζ or 47-CAR.LSR.CD28.ζ produced significant levels of IFNy (p<0.001) when stimulated with recombinant IL13Ra2 proteins in comparison to IL13Rα1- or IL4R-stimulated T cells (FIG. 33A). In contrast, T cells expressing 47-CAR.SSR.Δ or 47-CAR.LSR.Δ produced no IFNy in response to all three proteins, indicating that IFNy production depends on an intact 47-CAR signaling domain. 47-CAR.LSR.CD28. CT cells also produced low levels of IFNy without activation, indicating baseline T-cell activation, which was confirmed by intracellular staining for phosphorylated CD3. ζ (FIG. 34). IL13mutein-CAR.LSR.CD28. T cells produced significant levels of IFNy in the presence of IL13R α 1 (p<0.001) and IL13R α 2 (p<0.05) in comparison to NT T cells.

[0404] The specificity of 47-CAR T cells was then confirmed using cell lines that were negative for IL13R α 1 and IL13R α 2 (Raji), positive for IL13R α 1 (293T-GFP cells), or positive for IL13R α 1 and IL13R α 2 (U373, 293T-GFP/ IL13R α 2; FIG. **35**). T cells expressing 47-CAR.SSR.CD28.

ζ, 47-CAR.LSR.CD28.ζ, 47-CAR.SSR.Δ, or 47-CAR. LSR.Δ were co-cultured with Raji, 293T-GFP, or 293T-GFP/ IL13Ra2 cells. NT T cells served as controls. After 24 hours, media was collected and the concentrations of IFNy and IL2 were determined by ELISA. 47-CAR.SSR.CD28. cnd 47-CAR.LSR.CD28. CT cells produced significant amounts of IFNy only in the presence of U373 or 293T-GFP/ IL13Rα2 cells (FIG. 33B) with SSR.CAR T cells producing significantly more IFNy than LSR.CAR T cells (p<0.001). 47-CAR.SSR.CD28. CT cells produced also significant amounts of IL2 in the presence of 293T-GFP/IL13R α 2 and U373 cells, while 47-CAR.LSR.CD28. CT cells did not (FIG. 33C). NT-T cells and T cells expressing 47-CAR. SSR. Δ or 47-CAR.LSR. Δ produced no IFN γ or IL2 in response to any target cells. Finally, we confirmed the specificity of 47-CAR T cells in standard cytotoxicity assays using Raji, 293T-GFP, 293T-GFP/IL13Rα2, and U373 as targets (FIG. 33D).

Example 15

[0405] Generation of Short Spacer Region (SSR) 47-CARs with CD28.OX40/41BB

[0406] While the results described above demonstrated that 47-CAR T cells only recognize IL13R α 2, as judged by cytokine production and cytolytic activity, the results also highlighted differences between LSR and SSR 47-CARs. Because only 47-CAR.SSRs produced IL2 in the presence of IL13R α 2-positive target cells, the focus in the next set of experiments was shifted to 47-CARs with SSRs, and additional CARs were generated with CD28.OX40. ζ , CD28. 41BB. ζ or 41BB. ζ endodomains (FIG. **36**A). CAR T cells were generated by retroviral transduction and CAR expression was determined by FACS analysis (FIG. **36**B, C) and Western blot (FIG. **36**D). While all CARs were expressed, as judged by Western blot analysis, 47-CAR.SSR.CD28. 41BB. ζ was not expressed on the cell surface, and was excluded from further analysis.

Example 16

[0407] Comparison of Short Spacer Region 47-CARs [0408] To compare the ability of 47-CAR.SSR T cells to produce IFNy and IL2 in response to antigen exposure, co-culture assays were performed with U373 cells. T cells expressing 47-CAR.SSR. Δ served as controls. All 47-CAR. SSRs with functional endodomains induced IFNy and IL2 production in the presence of U373 cells; however, 47-CAR. SSR.41BB.ζ T cells produced significantly less (p<0.05) IFNy in comparison to 47-CAR.SSR.CD28. c and 47-CAR. SSR.CD28.OX40.5 T cells (FIG. 37A). 47-CAR.SSR. CD28. T cells produced the highest amount of IL2, followed by 47-CAR.SSR.41BB.5 and 47-CAR.SSR.CD28. OX40.ζ T cells. In cytotoxicity assays, no significant difference was observed between all three constructs using Raji, 293T-GFP, 293T-GFP/IL13Rα2, and U373 cells as targets (FIG. 37B).

[0409] Because all three 47-CAR.SSRs T cells with functional endodomains produced IL2, all three constructs were tested in an orthotopic U373 glioma xenograft mouse model in which T cells are directly injected into tumors.⁶ The model allows for serial bioluminescence imaging because U373 cells are genetically modified to express an eGFP. ffLuc fusion protein (U373.eGFP.ffLuc). On day 0, U373. eGFP.ffLuc cells were injected stereotactically into brains of SCID mice and, on day 7, T cells expressing 47-CAR.SSR. 47-CAR.SSR.41BB.ζ, CD28.ζ, 47-CAR.SSR.CD28. OX40. ζ or 47-CAR.SSR. Δ were injected intratumorally. While mice treated with 47-CAR.SSR.A T cells showed continuous tumor growth within 4 days of T-cell injection, mice treated with 47-CAR.SSR T cells that had functional endodomains did not (FIG. 38A, B). Comparison of bioluminescence imaging results revealed no significant difference between 47-CAR.SSR.∆ T cells and the 47-CAR.SSR T cells groups on the day of T-cell injection. Mice treated with 47-CAR.SSR.CD28.ζ or 47-CAR.SSR.CD28.OX40.ζ T cells, however, had significantly lower tumor signals as early as one day post-treatment in comparison to mice treated with 47-CAR.SSR. Δ T cells (p=0.012; Table 2). This resulted in a significant survival advantage for 47-CAR. SSR.CD28. c or 47-CAR.SSR.CD28.OX40. c T-cell-treated mice (p=0.0002 and p=0.0092; FIG. 40C). While 47-CAR. SSR.41BB. CT-cell-treated mice responded slower, resulting in a significant difference between 47-CAR.SSR.A T-cell treated on day 14 (p=0.005; Table 2), treatment with this CAR T cell also resulted in a significant survival advantage (p=0.0039; FIG. 5C FIG. 40C). 47-CAR.SSR.CD28.ζ T-cell-treated mice had the longest median survival (84 days). There was no statistical difference, however, in comparison to the median survival of 47-CAR.SSR.41BB.2 (63 days) or 47-CAR.SSR.CD28.OX40.5 (56 days) T-celltreated mice.

TABLE 2

Tumor signal comp	arison
	\mathbf{P}^{\dagger}
Day 7	
SSR.Δ vs. SSR.41BB.ζ SSR.Δ vs. SSR.CD28.ζ SSR.Δ vs. SSR.CD28.OX40.ζ Day 8	0.917 0.111 0.917
SSR.Δ vs. SSR.41BB.ζ SSR.Δ vs. SSR.CD28.ζ SSR.Δ vs. SSR.CD28.OX40.ζ Day 14	0.835 0.012 0.023
SSR.A vs. SSR.41BB.ζ SSR.A vs. SSR.CD28.ζ SSR.A vs. SSR.CD28.OX40.ζ Day 21	0.005 0.015 0.015
SSR.A vs. SSR.41BB.ζ SSR.A vs. SSR.CD28.ζ SSR.A vs. SSR.CD28.OX40.ζ Day 28	0.010 0.010 0.012
SSR.Δ vs. SSR.41BB.ζ SSR.Δ vs. SSR.CD28.ζ SSR.Δ vs. SSR.CD28.OX40.ζ	0.051 0.008 0.034

[†]Wilcoxon rank-sum test

[0410] While 47-CAR T cells had potent anti-glioma activity, mice developed recurrent gliomas. To investingate the etiology of tumor recurrence, U373 cells were isolated from two tumor-bearing mice that had been treated either with 47-CAR.SSR.CD28. ζ or 47-CAR.SSR.CD28.OX40. ζ T cells. FACS analysis after short-term culture revealed cell surface expression of IL13R α 2, and these cells were readily killed by 47-CAR T cells in cytotoxicity assays (FIG. **39**). Next, the persistence of T-cells was determined by geneti-

cally modifying T cells with 47-CAR.SSR.CD28. ξ and eGFP.ffLuc (Luc/47-CAR T cells), and injecting them into U373 tumor-bearing mice. T cells persisted for less than 7 days. Without wishing to be bound by theory, limited persistence appears to be the most likely explanation for tumor recurrence (FIG. **40**).

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Example 17

[0452] Transgenic Expression of IL15 Increases 47-CAR T-Cell Persistence Resulting in Enhanced Anti-Tumor Activity

[0453] IL13Ra2-CAR.CD28. CT cells expressing IL15 (IL13R α 2-CAR.IL15 T cells) were generated by double transducing T cells with retroviruses containing expression cassettes encoding i) IL13R α 2-CAR.CD28. ζ or ii) IL15, Δ Nerve Growth Factor Receptor (Δ NGFR), and inducible Caspase 9 (iC9) separated by 2A sequences. Suitable 2A sequences include any 2A sequence known in the art, as exemplified by the 2A amino acid sequence from porcine teschovirus-1 (SEQ ID NO:109) encoded by the polynucleotide sequence set forth as SEQ ID NO:110, the 2A amino acid sequence from Thoseaasigna virus (SEQ ID NO:111) encoded by the polynucleotide sequence set forth as SEQ ID NO:112, the 2A amino acid sequence from Equine rhinitis A virus (ERAV) (SEQ ID NO:113) encoded by the polynucleotide sequence set forth as SEQ ID NO:114, or the 2A amino acid sequence from Foot and Mouth Disease Virus (FMDV) (SEQ ID NO:115) encoded by the polynucleotide sequence set forth as SEQ ID NO:116. Kim et al., PLoS One 6(4):1-8 (2011). The effector function of IL13R α 2-CAR.IL15 T cells was determined in vitro using standard assays, and in the U373 GBM xenograft model.

[0454] Double transduction of CD3/CD28-activated T cells resulted in T-cell lines that expressed both transgenes in 45-50% of T cells. At base line, IL13R α 2-CAR.IL15 T cells produced on average 69.5 pg/ml of IL15. Production was significantly increased after CD3 or antigen-specific T-cell stimulation (176.7 pg/ml; n=6; p<0.001). IL13R α 2-CAR.IL15 T cells were as efficient as IL13R α 2-CAR T cells in killing IL13R α 2-positive GBM cells in vitro. After intratumoral injection into U373 glioma-bearing mice, IL13R α 2-CAR.IL15 T cells (p<0.05). This resulted in a significant increase in progression-free (98 versus 49 days; p=0.004) and overall survival (p=0.006) of treated mice.

[0455] The data disclosed in this Example demonstrate that transgenic expression of IL15 enhances the in vivo persistence of IL13R α 2-CAR T cells, resulting in improved anti-glioma activity.

[0456] Each of the references cited herein is hereby incorporated by reference in its entirety or in relevant part, as would be apparent from the context of the citation.

[0457] From the disclosure herein it will be appreciated that, although specific embodiments of the disclosure have been described herein for purposes of illustration, various modifications may be made without deviating from the spirit and scope of the disclosure.

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53

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Asn Glu Lys Ala Leu Gly Arg Lys Ile Asn Ser Trp Glu Ser Ser Arg 50 55 60	
Ser Gly His Ser Phe Leu Ser Asn Leu His Leu Arg Asn Gly Glu Leu 65 70 75 80	
Val Ile His Glu Lys Gly Phe Tyr Tyr Ile Tyr Ser Gln Thr Tyr Phe 85 90 95	
Arg Phe Gln Glu Glu Ile Lys Glu Asn Thr Lys Asn Asp Lys Gln Met 100 105 110	
Val Gln Tyr Ile Tyr Lys Tyr Thr Ser Tyr Pro Asp Pro Ile Leu Leu 115 120 125	

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Met Lys Ser Ala Arg Asn Ser Cys Trp Ser Lys Asp Ala Glu Tyr Gly 135 130 140 Leu Tyr Ser Ile Tyr Gln Gly Gly Ile Phe Glu Leu Lys Glu Asn Asp 155 145 150 160 Arg Ile Phe Val Ser Val Thr Asn Glu His Leu Ile Asp Met Asp His 170 165 175 Glu Ala Ser Phe Phe Gly Ala Phe Leu Val Gly 180 185 <210> SEQ ID NO 28 <211> LENGTH: 107 <212> TYPE: PRT <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: Synthetic Polypeptide <400> SEQUENCE: 28 Arg Ala Asp Ala Ala Pro Thr Val Ser Ile Phe Pro Pro Ser Ser Glu 10 5 15 1 Gln Leu Thr Ser Gly Gly Ala Ser Val Val Cys Phe Leu Asn Asn Phe 20 25 30 Tyr Pro Lys Asp Ile Asn Val Lys Trp Lys Ile Asp Gly Ser Glu Arg 35 40 Gln Asn Gly Val Leu Asn Ser Trp Thr Asp Gln Asp Ser Lys Asp Ser 50 55 60 Thr Tyr Ser Met Ser Ser Thr Leu Thr Leu Thr Lys Asp Glu Tyr Glu 65 70 75 80 Arg His Asn Ser Tyr Thr Cys Glu Ala Thr His Lys Thr Ser Thr Ser 85 90 95 Pro Ile Val Lys Ser Phe Asn Arg Asn Glu Cys 100 105 <210> SEQ ID NO 29 <211> LENGTH: 324 <212> TYPE: DNA <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: Synthetic Polynucleotide <400> SEQUENCE: 29 cgggctgatg ctgcaccaac tgtatccatc ttcccaccat ccagtgagca gttaacatct 60 ggaggtgcct cagtcgtgtg cttcttgaac aacttctacc ccaaagacat caatgtcaag 120 tggaagattg atggcagtga acgacaaaat ggcgtcctga acagttggac tgatcaggac 180 agcaaagaca gcacctacag catgagcagc accctcacgt tgaccaagga cgagtatgaa 240 cgacataaca gctatacctg tgaggccact cacaagacat caacttcacc cattgtcaag 300 agcttcaaca ggaatgagtg ttag 324

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Phe	Pro	Glu 35	Pro	Val	Thr	Val	Thr 40	Trp	Asn	Ser	Gly	Ser 45	Leu	Ser	Ser
Gly	Val 50	His	Thr	Phe	Pro	Ala 55	Val	Leu	Gln	Ser	Asp 60	Leu	Tyr	Thr	Leu
Ser 65	Ser	Ser	Val	Thr	Val 70	Pro	Ser	Ser	Thr	Trp 75	Pro	Ser	Glu	Thr	Val 80
Thr	Суз	Asn	Val	Ala 85	His	Pro	Ala	Ser	Ser 90	Thr	Lys	Val	Asp	Lys 95	Гля
Ile	Val	Pro	Arg 100		Суз	Gly	Суз	Lys 105	Pro	Суз	Ile	Суз	Thr 110	Val	Pro
Glu	Val	Ser 115	Ser	Val	Phe	Ile	Phe 120	Pro	Pro	Lys	Pro	Lys 125	Asp	Val	Leu
Thr	Ile 130	Thr	Leu	Thr	Pro	Lys 135	Val	Thr	Суз	Val	Val 140	Val	Asp	Ile	Ser
Lys 145	Asp	Asp	Pro	Glu	Val 150	Gln	Phe	Ser	Trp	Phe 155	Val	Asp	Asp	Val	Glu 160
Val	His	Thr	Ala	Gln 165	Thr	Gln	Pro	Arg	Glu 170	Glu	Gln	Phe	Asn	Ser 175	Thr
Phe	Arg	Ser	Val 180	Ser	Glu	Leu	Pro	Ile 185	Met	His	Gln	Asp	Trp 190	Leu	Asn
Gly	Lys	Glu 195	Phe	Lys	Суз	Arg	Val 200	Asn	Ser	Ala	Ala	Phe 205	Pro	Ala	Pro
Ile	Glu 210	Lys	Thr	Ile	Ser	Lys 215	Thr	Гла	Gly	Arg	Pro 220	Lys	Ala	Pro	Gln
Val 225	Tyr	Thr	Ile	Pro	Pro 230	Pro	Lys	Glu	Gln	Met 235	Ala	Lys	Asp	Lys	Val 240
Ser	Leu	Thr	Сүз	Met 245	Ile	Thr	Asp	Phe	Phe 250	Pro	Glu	Asp	Ile	Thr 255	Val
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Pro	Ile	Met 275	Asp	Thr	Asp	Gly	Ser 280	Tyr	Phe	Val	Tyr	Ser 285	Lys	Leu	Asn
Val	Gln 290	Lys	Ser	Asn	Trp	Glu 295	Ala	Gly	Asn	Thr	Phe 300	Thr	Суз	Ser	Val
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Ser	Pro	Gly	Lys												
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Pro Gly Ala Ser Val Lys Leu Ser Cys Lys Ala Ser Gly Tyr Thr Phe 35 40 45								
Ser Asn Tyr Leu Met Asn Trp Val Lys Gln Arg Pro Glu Gln Asp Leu 50								
Asp Trp Ile Gly Arg Ile Asp Pro Tyr Asp Gly Asp Ile Asp Tyr Asn65707580								
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Ile Ser Cys Arg Ala Ser Glu Ser Val Asp Asn Tyr Gly Ile Ser Phe 180 185 190	
Met Asn Trp Phe Gln Gln Lys Pro Gly Gln Pro Pro Lys Leu Leu Ile 195 200 205	
Tyr Ala Ala Ser Arg Gln Gly Ser Gly Val Pro Ala Arg Phe Ser Gly 210 215 220	
Ser Gly Ser Gly Thr Asp Phe Ser Leu Asn Ile His Pro Met Glu Glu 225 230 235 240	
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Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn 50 55 60											
Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg 65 70 75 80											
Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val 85 90 95											
Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser 100 105 110											
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Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly 195 200 205											
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35 40 45	
Pro Thr Arg Lys His Tyr Gln Pro Tyr Ala Pro Pro Arg Asp Phe 50 55 60	Ala
Ala Tyr Arg Ser Arg Val Lys Phe Ser Arg Ser Ala Asp Ala Pro 65 70 75	Ala 80
Tyr Gln Gln Gly Gln Asn Gln Leu Tyr Asn Glu Leu Asn Leu Gly 85 90 95	Arg
Arg Glu Glu Tyr Asp Val Leu Asp Lys Arg Arg Gly Arg Asp Pro 100 105 110	Glu
Met Gly Gly Lys Pro Arg Arg Lys Asn Pro Gln Glu Gly Leu Tyr 115 120 125	Asn
Glu Leu Gln Lys Asp Lys Met Ala Glu Ala Tyr Ser Glu Ile Gly 130 135 140	Met
Lys Gly Glu Arg Arg Gly Lys Gly His Asp Gly Leu Tyr Glr 145 150 155	Gly 160
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1		01 u	5	272	201		1.05	10				012	15	110				
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Pro Lys	Pro 35	Гла	Asp	Thr	Leu	Met 40	Ile	Ser	Arg	Thr	Pro 45	Glu	Val	Thr				
Cys Val 50	Val	Val	Asp	Val	Ser 55	His	Glu	Asp	Pro	Glu 60	Val	Гла	Phe	Asn				
Irp Tyr 65	Val	Asp	Gly	Val 70	Glu	Val	His	Asn	Ala 75	Lys	Thr	Lys	Pro	Arg 80				
Glu Glu	Gln	Tyr	Asn 85	Ser	Thr	Tyr	Arg	Val 90	Val	Ser	Val	Leu	Thr 95	Val				
Leu His	Gln	Asp 100	Trp	Leu	Asn	Gly	Lys 105	Glu	Tyr	Lys	СЛа	Lys 110	Val	Ser				
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Glu Leu 145	Thr	Гла	Asn	Gln 150	Val	Ser	Leu	Thr	Cys 155	Leu	Val	Lys	Gly	Phe 160				
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Val Thr Val Ala Phe Ile Ile Phe Trp Val Arg Ser Lys Arg Ser Arg 260 265 270
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Thr Arg Lys His Tyr Gln Pro Tyr Ala Pro Pro Arg Asp Phe Ala Ala 290 295 300
Tyr Arg Ser Arg Val Lys Phe Ser Arg Ser Ala Asp Ala Pro Ala Tyr 305 310 315 320
Gln Gln Gly Gln Asn Gln Leu Tyr Asn Glu Leu Asn Leu Gly Arg Arg 325 330 335
Glu Glu Tyr Asp Val Leu Asp Lys Arg Arg Gly Arg Asp Pro Glu Met
340 345 350 Gly Gly Lys Pro Arg Arg Lys Asn Pro Gln Glu Gly Leu Tyr Asn Glu
355 360 365 Leu Gln Lys Asp Lys Met Ala Glu Ala Tyr Ser Glu Ile Gly Met Lys
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Pro Pro Arg
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gtcaagttca actggtacgt ggacggcgtg gaggtgcata atgccaagac aaagccgcgg 240
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 Gly Cys Cys Cys Thr Ala Thr Gly Cys Cys Cys Ala Cys Cys Ala

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Gly	Gly	Cys 435	Суз	Thr	Gly	Thr	Ala 440	Суз	Ala	Ala	Thr	Gly 445	Ala	Ala	Суз
Thr	Gly 450	Суз	Ala	Gly	Ala	Ala 455	Ala	Gly	Ala	Thr	Ala 460	Ala	Gly	Ala	Thr
Gly 465	Gly	Суз	Gly	Gly	Ala 470	Gly	Gly	Суз	Суз	Thr 475	Ala	Суз	Ala	Gly	Thr 480
Gly	Ala	Gly	Ala	Thr 485	Thr	Gly	Gly	Gly	Ala 490	Thr	Gly	Ala	Ala	Ala 495	Gly
Gly	Суз	Gly	Ala 500	Gly	Суз	Gly	Суз	Суз 505	Gly	Gly	Ala	Gly	Gly 510	Gly	Gly
Суз	Ala	Ala 515	Gly	Gly	Gly	Gly	Cys 520	Ala	Суз	Gly	Ala	Thr 525	Gly	Gly	Сув
Сүз	Thr 530	Thr	Thr	Ala	Сүз	Суя 535	Ala	Gly	Gly	Gly	Thr 540	Сүз	Thr	Сүз	Ala
Gly 545	Thr	Ala	Суз	Ala	Gly 550	Сув	Суз	Ala	Суз	Сув 555	Ala	Ala	Gly	Gly	Ala 560
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Суз	Ala	Суз	Ala 580	Thr	Gly	Суз	Ala	Gly 585	Gly	Суз	Суз	Суз	Thr 590	Gly	Сув
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Pro	Gly	Ala 35	Ser	Val	ГЛа	Leu	Ser 40	Сүз	Lys	Ala	Ser	Gly 45	Tyr	Thr	Phe
Ser	Asn 50	Tyr	Leu	Met	Asn	Trp 55	Val	Lys	Gln	Arg	Pro 60	Glu	Gln	Asp	Leu
Asp 65	Trp	Ile	Gly	Arg	Ile 70	Asp	Pro	Tyr	Asp	Gly 75	Asp	Ile	Asp	Tyr	Asn 80
Gln	Asn	Phe	Lys	Asp 85	ГÀа	Ala	Ile	Leu	Thr 90	Val	Asp	ГÀа	Ser	Ser 95	Ser
Thr	Ala	Tyr	Met 100	Gln	Leu	Ser	Ser	Leu 105	Thr	Ser	Glu	Asp	Ser 110	Ala	Val
Tyr	Tyr	Суз	Ala	Arg	Gly	Tyr	Gly	Thr	Ala	Tyr	Gly	Val	Asp	Tyr	Trp

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Thr	Gln	Ser	Pro	Ala 165	Ser	Leu	Ala	Val	Ser 170	Leu	Gly	Gln	Arg	Ala 175	Thr
Ile	Ser	Суз	Arg 180	Ala	Ser	Glu	Ser	Val 185	Asp	Asn	Tyr	Gly	Ile 190	Ser	Phe
Met	Asn	Trp 195	Phe	Gln	Gln	ГЛа	Pro 200	Gly	Gln	Pro	Pro	Lys 205	Leu	Leu	Ile
Tyr	Ala 210	Ala	Ser	Arg	Gln	Gly 215	Ser	Gly	Val	Pro	Ala 220	Arg	Phe	Ser	Gly
Ser 225	Gly	Ser	Gly	Thr	Asp 230	Phe	Ser	Leu	Asn	Ile 235	His	Pro	Met	Glu	Glu 240
Asp	Asp	Thr	Ala	Met 245	Tyr	Phe	Суз	Gln	Gln 250	Ser	ГЛа	Glu	Val	Pro 255	Trp
Thr	Phe	Gly	Gly 260	Gly	Thr	Гла	Leu	Glu 265	Ile	Lys	Ala	Glu	Asp 270	Pro	Ala
Glu	Pro	Lys 275	Ser	Pro	Asp	Lys	Thr 280	His	Thr	Сүз	Pro	Pro 285	Cys	Pro	Ala
Pro	Glu 290	Leu	Leu	Gly	Gly	Pro 295	Ser	Val	Phe	Leu	Phe 300	Pro	Pro	Lys	Pro
Lys 305	Asp	Thr	Leu	Met	Ile 310	Ser	Arg	Thr	Pro	Glu 315	Val	Thr	Сув	Val	Val 320
Val	Asp	Val	Ser	His 325	Glu	Asp	Pro	Glu	Val 330	Lys	Phe	Asn	Trp	Tyr 335	Val
Asp	Gly	Val	Glu 340	Val	His	Asn	Ala	Lys 345	Thr	Lys	Pro	Arg	Glu 350	Glu	Gln
Tyr	Asn	Ser 355	Thr	Tyr	Arg	Val	Val 360	Ser	Val	Leu	Thr	Val 365	Leu	His	Gln
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385					390	Lys				395		-	-		400
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-			420			Thr	-	425		-	_		430		
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Lys	Thr 450	Thr	Pro	Pro	Val	Leu 455	Asp	Ser	Asp	Gly	Ser 460	Phe	Phe	Leu	Tyr
Ser 465	Lys	Leu	Thr	Val	Asp 470	Γλa	Ser	Arg	Trp	Gln 475	Gln	Gly	Asn	Val	Phe 480
Ser	Суз	Ser	Val	Met 485	His	Glu	Ala	Leu	His 490	Asn	His	Tyr	Thr	Gln 495	Lys
Ser	Leu	Ser	Leu 500	Ser	Pro	Gly	Lys	Lys 505	Asp	Pro	Lys	Phe	Trp 510	Val	Leu
Val	Val	Val 515	Gly	Gly	Val	Leu	Ala 520	Сүз	Tyr	Ser	Leu	Leu 525	Val	Thr	Val

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Ala Phe Ile Ile Phe Trp Val Arg Ser Lys Arg Ser Arg Leu Leu His Ser Asp Tyr Met Asn Met Thr Pro Arg Arg Pro Gly Pro Thr Arg Lys His Tyr Gln Pro Tyr Ala Pro Pro Arg Asp Phe Ala Ala Tyr Arg Ser Arg Val Lys Phe Ser Arg Ser Ala Asp Ala Pro Ala Tyr Gln Gln Gly Gln Asn Gln Leu Tyr Asn Glu Leu Asn Leu Gly Arg Arg Glu Glu Tyr Asp Val Leu Asp Lys Arg Arg Gly Arg Asp Pro Glu Met Gly Gly Lys Pro Arg Arg Lys Asn Pro Gln Glu Gly Leu Tyr Asn Glu Leu Gln Lys 625 630 635 Asp Lys Met Ala Glu Ala Tyr Ser Glu Ile Gly Met Lys Gly Glu Arg Arg Arg Gly Lys Gly His Asp Gly Leu Tyr Gln Gly Leu Ser Thr Ala Thr Lys Asp Thr Tyr Asp Ala Leu His Met Gln Ala Leu Pro Pro Arg <210> SEQ ID NO 54 <211> LENGTH: 2064 <212> TYPE: PRT <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: Synthetic Polynucleotide <400> SEQUENCE: 54 Ala Thr Gly Gly Ala Cys Thr Gly Gly Ala Thr Cys Thr Gly Gly Cys Gly Cys Ala Thr Cys Cys Thr Gly Thr Thr Thr Cys Thr Cys Gly Thr Gly Cys Cys Cys Ala Thr Thr Cys Thr Cys Ala Gly Gly Thr Gly Cys $% \left({{\left({{{\left({{{}_{{\rm{S}}}} \right)}} \right)}} \right)$ Ala Gly Cys Thr Gly Cys Ala Gly Cys Ala Gly Cys Cys Thr Gly Gly 65 70 75 80 Cys Gly Cys Thr Gly Ala Ala Cys Thr Cys Gly Thr Gly Cys Gly Gly Cys Cys Ala Gly Gly Cys Gly Cys Thr Thr Cys Thr Gly Thr Gly Ala Ala Gly Cys Thr Gly Ala Gly Cys Thr Gly Thr Ala Ala Ala Gly Cys Cys Ala Gly Cys Gly Gly Cys Thr Ala Cys Ala Cys Cys Thr Thr Cys Ala Gly Cys Ala Ala Cys Thr Ala Cys Cys Thr Gly Ala Thr Gly Ala Ala Cys Thr Gly Gly Gly Thr Cys Ala Ala Gly Cys Ala Gly Cys Gly Gly Cys Cys Cys Gly Ala Gly Cys Ala Gly Gly Ala Cys Cys Thr Gly

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Gly Ala Thr Thr Gly Gly Ala Thr Cys Gly Gly Cys Ala Gly Ala Ala Thr Cys Gly Ala Cys Cys Cys Thr Ala Cys Gly Ala Cys Gly Gly Cys Gly Ala Cys Ala Thr Cys Gly Ala Cys Thr Ala Cys Ala Ala Cys Cys Ala Gly Ala Ala Cys Thr Thr Cys Ala Ala Gly Gly Ala Cys Ala Ala Gly Gly Cys Cys Ala Thr Cys Cys Thr Gly Ala Cys Cys Gly Thr 260 265 270 Gly Gly Ala Cys Ala Ala Gly Ala Gly Cys Ala Gly Cys 275 280 285 Ala Cys Cys Gly Cys Cys Thr Ala Cys Ala Thr Gly Cys Ala Gly Cys 290 295 Thr Gly Thr Cys Cys Ala Gly Cys Cys Thr Gly Ala Cys Cys Ala Gly 305 310 315 320 Cys Gly Ala Gly Gly Ala Cys Ala Gly Cys Gly Cys Cys Gly Thr Gly 325 330 335 Thr Ala Cys Thr Ala Cys Thr Gly Cys Gly Cys Cys Ala Gly Ala Gly Gly Cys Thr Ala Cys Gly Gly Cys Ala Cys Ala Gly Cys Cys Thr Ala Cys Gly Gly Cys Gly Thr Gly Gly Ala Cys Thr Ala Thr Thr Gly Gly Gly Gly Cys Cys Ala Gly Gly Gly Cys Ala Cys Ala Ala Gly Cys Gly Thr Gly Ala Cys Cys Gly Thr Gly Thr Cys Cys Ala Gly Cys Gly Cys 405 410 415 Cys Ala Ala Gly Ala Cys Cys Ala Cys Cys Cys Cys Cys Cys Thr 420 425 430 Ala Ala Gly Cys Thr Gly Gly Ala Ala Gly Ala Gly Gly Gly Cys Gly 435 440 445 Ala Gly Thr Thr Cys Thr Cys Cys Gly Ala Gly Gly Cys Cys Gly Gly Gly Thr Gly Gly Ala Cys Ala Thr Thr Gly Thr Gly Cys Thr Gly Ala Cys Ala Cys Ala Gly Thr Cys Thr Cys Cys Ala Gly Cys Cys Ala 485 490 495 Gly Cys Cys Thr Gly Gly Cys Cys Gly Thr Gly Thr Cys Cys Thr 500 505 510 Gly Gly Gly Ala Cys Ala Gly Ala Gly Ala Gly Cys Cys Ala Cys Cys Ala Thr Cys Ala Gly Cys Thr Gly Thr Ala Gly Gly Gly Cys Cys Ala Gly Cys Gly Ala Gly Ala Gly Cys Gly Thr Gly Gly Ala Cys Ala Ala Cys Thr Ala Cys Gly Gly Cys Ala Thr Cys Ala Gly Cys Thr Thr Cys Ala Thr Gly Ala Ala Thr Thr Gly Gly Thr Thr Cys Cys Ala Gly Cys

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Ala	Cys 690	Thr	Thr	Сув	Ala	Gly 695	Cya	Суз	Thr	Gly	Ala 700	Ala	Cys	Ala	Thr			
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СЛа	Ala	Ala 755	Ala	Gly	Ala	Gly	Gly 760	Thr	Gly	Сув	СЛа	Суз 765	Thr	Gly	Gly			
Ala	Cys 770	Сүв	Thr	Thr	Thr	Gly 775	Gly	Сув	Gly	Gly	Ala 780	Gly	Gly	Сув	Ala			
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Gly	Ala	Суз	Суз	Gly 885	Thr	СЛа	Ala	Gly	Thr 890	Суз	Thr	Thr	Суз	Сув 895	Thr			
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Суз	Cys 1355	Thr	Суз	Cys	Cys	Gly 1360		Gly	Суз	Thr	Gly 1365	Gly	Ala	Сүз
Thr	Cys 1370	Суз	Gly	Ala	Суз	Gly 1375	Gly	Сув	Thr	Суз	Cys 1380	Thr	Thr	Сүз

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Ala	Gly 1415	СЛа	Ala	Gly	Gly	Thr 1420		Gly	Суз	Ala	Gly 1425	Суз	Ala	Gly
Gly	Gly 1430	Gly	Ala	Ala	Cys	Gly 1435		Cys	Thr	Thr	Cys 1440	Thr	Cys	Ala
Thr	Gly 1445	Суз	Thr	Суа	Суз	Gly 1450		Gly	Ala	Thr	Gly 1455	Суз	Ala	Thr
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Ala	Thr 1595	Thr	Thr	Thr	Суз	Thr 1600	Gly	Gly	Gly	Thr	Gly 1605	Ala	Gly	Gly
Ala	Gly 1610	Thr	Ala	Ala	Gly	Ala 1615	Gly	Gly	Ala	Gly	Cys 1620	Ala	Gly	Gly
САа	Thr 1625	Сүз	Сүз	Thr	Gly	Cys 1630	Ala	Cys	Ala	Gly	Thr 1635	Gly	Ala	Сүз
Thr	Ala 1640	Сүз	Ala	Thr	Gly	Ala 1645	Ala	Cys	Ala	Thr	Gly 1650	Ala	Сүз	Thr
САа	Cys 1655	Сүз	Сүз	Gly	Cys	Cys 1660	Gly	Cys	Сүз	САа	Cys 1665	Gly	Gly	Gly
Сүз	Cys 1670	Суз	Ala	Cys	Cys	Cys 1675	Gly	Cys	Ala	Ala	Gly 1680	Cys	Ala	Thr
Thr	Ala 1685	Сүз	СЛа	Ala	Gly	Cys 1690	Суз	Сув	Thr	Ala	Thr 1695	Gly	Сүз	Сүа
Сүз	Cys 1700	Ala	CÀa	Cys	Ala	Cys 1705	Gly	Cys	Gly	Ala	Cys 1710	Thr	Thr	Сүз
Gly	Cys 1715	Ala	Gly	Суа	Суз	Thr 1720		Thr	Суз	Gly	Cys 1725	Thr	Суа	СЛа
Ala	Gly 1730	Ala	Gly	Thr	Gly	Ala 1735		Gly	Thr	Thr	Cys 1740	Ala	Gly	Сүз
Ala	Gly 1745	Gly	Ala	Gly	Сув	Gly 1750	Сув	Ala	Gly	Ala	Cys 1755	Gly	Сув	Сүз

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CAa	Cys 1760	Сүз	Gly	Суз	Gly	Thr 1765	Ala	Cys	Сүз	Ala	Gly 1770	Суз	Ala	Gly					
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Thr	Ala 1790	Thr	Ala	Ala	Cys	Gly 1795	Ala	Gly	Суз	Thr	Cys 1800	Ala	Ala	Thr					
Cys	Thr 1805	Ala	Gly	Gly	Ala	Cys 1810	Gly	Ala	Ala	Gly	Ala 1815	Gly	Ala	Gly					
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Gly	Ala 1835	Суз	Ala	Ala	Gly	Ala 1840	Gly	Ala	Суз	Gly	Thr 1845	Gly	Gly	СЛа					
Cya	Gly 1850		Gly	Ala	Cys	Cys 1855	Сув	Thr	Gly	Ala	Gly 1860	Ala	Thr	Gly					
Gly	Gly 1865		-	Gly	Ala	Ala 1870	Ala	Gly	Cys	Суз	Gly 1875	Ala	Gly	Ala					
Ala	Gly 1880		Ala	Ala	Gly	Ala 1885	Ala	Cys	Суз	Суа	Thr 1890	Суз	Ala	Gly					
Gly	Ala 1895	Ala	Gly	Gly	Cys	Cys 1900	Thr	Gly	Thr	Ala	Cys 1905	Ala	Ala	Thr					
Gly	Ala 1910	Ala	Cys	Thr	Gly	Cys 1915	Ala	Gly	Ala	Ala	Ala 1920	Gly	Ala	Thr					
Ala	Ala 1925		Ala	Thr	Gly	Gly 1930	Cys	Gly	Gly	Ala	Gly 1935	Gly	Cys	Сүв					
Thr	Ala 1940		Ala	Gly	Thr	Gly 1945	Ala	Gly	Ala	Thr	Thr 1950	Gly	Gly	Gly					
Ala	Thr 1955	Gly	Ala	Ala	Ala	Gly 1960	Gly	Cys	Gly	Ala	Gly 1965	Суз	Gly	Сув					
Cya		Gly	Ala	Gly	Gly	Gly 1975	Gly	Суз	Ala	Ala		Gly	Gly	Gly					
Cya					Thr	Gly 1990	Gly	Cys	Суз	Thr		Thr	Ala	Суа					
Cya					Thr	Cys 2005	Thr	Cys	Ala	Gly		Ala	Суз	Ala					
Gly		Суз	Ala	Cys	Суз	Ala 2020	Ala	Gly	Gly	Ala		Ala	Суз	Cys					
Thr		Суз	Gly	Ala	Cys	Gly 2035	Суз	Суз	Суз	Thr		Суз	Ala	Суз					
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	2060																		
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Pro	Gly	Ala 35	Ser	Val	Lys	Leu	Ser 40	Суз	Lys	Ala	Ser	Gly 45	Tyr	Thr	Phe
Ser	Asn 50	Tyr	Leu	Met	Asn	Trp 55	Val	Lys	Gln	Arg	Pro 60	Glu	Gln	Asp	Leu
Asp 65	Trp	Ile	Gly	Arg	Ile 70	Asp	Pro	Tyr	Asp	Gly 75	Asp	Ile	Asp	Tyr	Asn 80
Gln	Asn	Phe	ГЛа	Asp 85	ГЛа	Ala	Ile	Leu	Thr 90	Val	Asp	ГÀЗ	Ser	Ser 95	Ser
Thr	Ala	Tyr	Met 100	Gln	Leu	Ser	Ser	Leu 105	Thr	Ser	Glu	Asp	Ser 110	Ala	Val
Tyr	Tyr	Cys 115	Ala	Arg	Gly	Tyr	Gly 120	Thr	Ala	Tyr	Gly	Val 125	Asp	Tyr	Trp
Gly	Gln 130	Gly	Thr	Ser	Val	Thr 135	Val	Ser	Ser	Ala	Lys 140	Thr	Thr	Pro	Pro
Lys 145	Leu	Glu	Glu	Gly	Glu 150	Phe	Ser	Glu	Ala	Arg 155	Val	Asp	Ile	Val	Leu 160
Thr	Gln	Ser	Pro	Ala 165	Ser	Leu	Ala	Val	Ser 170	Leu	Gly	Gln	Arg	Ala 175	Thr
Ile	Ser	Суз	Arg 180	Ala	Ser	Glu	Ser	Val 185	Asp	Asn	Tyr	Gly	Ile 190	Ser	Phe
Met	Asn	Trp 195	Phe	Gln	Gln	Lys	Pro 200	Gly	Gln	Pro	Pro	Lys 205	Leu	Leu	Ile
Tyr	Ala 210	Ala	Ser	Arg	Gln	Gly 215	Ser	Gly	Val	Pro	Ala 220	Arg	Phe	Ser	Gly
Ser 225	Gly	Ser	Gly	Thr	Asp 230	Phe	Ser	Leu	Asn	Ile 235	His	Pro	Met	Glu	Glu 240
Asp	Asp	Thr	Ala	Met 245	Tyr	Phe	Суз	Gln	Gln 250	Ser	Lys	Glu	Val	Pro 255	Trp
Thr	Phe	Gly	Gly 260	Gly	Thr	Lys	Leu	Glu 265	Ile	Lys	Asp	Leu	Glu 270	Pro	Lys
Ser	Суз	Asp 275	ГЛа	Thr	His	Thr	Cys 280	Pro	Pro	Суз	Pro	Asp 285	Pro	Lys	Phe
Trp	Val 290	Leu	Val	Val	Val	Gly 295	Gly	Val	Leu	Ala	Суз 300	Tyr	Ser	Leu	Leu
Val 305	Thr	Val	Ala	Phe	Ile 310	Ile	Phe	Trp	Val	Arg 315	Ser	ГЛа	Arg	Ser	Arg 320
Leu	Leu	His	Ser	Asp 325	Tyr	Met	Asn	Met	Thr 330	Pro	Arg	Arg	Pro	Gly 335	Pro
Thr	Arg	Гла	His 340	Tyr	Gln	Pro	Tyr	Ala 345	Pro	Pro	Arg	Asp	Phe 350	Ala	Ala
Tyr	Arg	Ser 355	Arg	Val	Lys	Phe	Ser 360	Arg	Ser	Ala	Asp	Ala 365	Pro	Ala	Tyr
Gln	Gln 370	Gly	Gln	Asn	Gln	Leu 375	Tyr	Asn	Glu	Leu	Asn 380	Leu	Gly	Arg	Arg
Glu 385		Tyr	Asp	Val	Leu 390		Lys	Arg	Arg	Gly 395		Asp	Pro	Glu	Met 400
	Gly	Lys	Pro	_		ГЛа	Asn	Pro			Gly	Leu	Tyr		
Leu	Gln	Lys	Asp	405 Lys	Met	Ala	Glu	Ala	410 Tyr	Ser	Glu	Ile	Gly	415 Met	Lys

420 425 430 Gly Glu Arg Arg Arg Gly Lys Gly His Asp Gly Leu Tyr Gln Gly Leu 440 435 445 Ser Thr Ala Thr Lys Asp Thr Tyr Asp Ala Leu His Met Gln Ala Leu 450 455 460 Pro Pro Arq 465 <210> SEQ ID NO 56 <211> LENGTH: 1401 <212> TYPE: DNA <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: Synthetic Polynucleotide <400> SEQUENCE: 56 atggaetgga tetggegeat cetgtttete gtgggageeg ceacaggege ceatteteag 60 120 gtgcagctgc agcagcctgg cgctgaactc gtgcggccag gcgcttctgt gaagctgagc tgtaaagcca gcggctacac cttcagcaac tacctgatga actgggtcaa gcagcggccc 180 gagcaggacc tggattggat cggcagaatc gacccctacg acggcgacat cgactacaac 240 cagaacttca aggacaaggc catcctgacc gtggacaaga gcagcagcac cgcctacatg 300 cagetgteca geetgaceag egaggaeage geegtgtaet aetgegeeag aggetaegge 360 420 acageetaeg gegtggaeta ttggggeeag ggeaeaageg tgaeegtgte eagegeeaag accacccccc ctaagctgga agagggcgag ttctccgagg cccgggtgga cattgtgctg 480 acacagtete cagecageet ggeegtgtee etgggaeaga gageeaceat cagetgtagg 540 gccagcgaga gcgtggacaa ctacggcatc agcttcatga attggttcca gcagaagccc 600 ggccagcccc ccaagctgct gatctatgcc gccagcagac agggcagcgg agtgcctgcc 660 agattttctg gcagcggctc cggcaccgac ttcagcctga acatccaccc tatggaagag 720 gacgacaccg ccatgtactt ttgccagcag agcaaagagg tgccctggac ctttggcgga 780 ggcaccaagc tggaaatcaa ggatctcgag cccaaatctt gtgacaaaac tcacacatgc 840 900 ccaccgtgcc cggatcccaa attttgggtg ctggtggtgg ttggtggagt cctggcttgc tatagettge tagtaacagt ggeetttatt attttetggg tgaggagtaa gaggageagg 960 ctcctgcaca gtgactacat gaacatgact ccccgccgcc ccgggcccac ccgcaagcat 1020 taccageeet atgeeeeace acgegaette geageetate geteeagagt gaagtteage 1080 aggagegeag aegeceeege gtaceageag ggeeagaaee agetetataa egageteaat 1140 ctaggacgaa gagaggagta cgatgttttg gacaagagac gtggccggga ccctgagatg 1200 1260 qqqqqaaaqc cqaqaaqqaa qaaccctcaq qaaqqcctqt acaatqaact qcaqaaaqat aagatggcgg aggcctacag tgagattggg atgaaaggcg agcgccggag gggcaagggg 1320 cacgatggcc tttaccaggg tctcagtaca gccaccaagg acacctacga cgcccttcac 1380 atgcaggeee tgeeeeteg e 1401 <210> SEQ ID NO 57 <211> LENGTH: 257

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Суз	Pro	Ala	Pro 20	Glu	Leu	Leu	Gly	Gly 25	Pro	Ser	Val	Phe	Leu 30	Phe	Pro
Pro	Lys	Pro 35	Lys	Asp	Thr	Leu	Met 40	Ile	Ser	Arg	Thr	Pro 45	Glu	Val	Thr
Суз	Val 50	Val	Val	Asp	Val	Ser 55	His	Glu	Asp	Pro	Glu 60	Val	Lys	Phe	Asn
Trp 65	Tyr	Val	Asp	Gly	Val 70	Glu	Val	His	Asn	Ala 75	Lys	Thr	Lys	Pro	Arg 80
Glu	Glu	Gln	Tyr	Asn 85	Ser	Thr	Tyr	Arg	Val 90	Val	Ser	Val	Leu	Thr 95	Val
Leu	His	Gln	Asp 100	Trp	Leu	Asn	Gly	Lys 105	Glu	Tyr	Lys	Суз	Lys 110	Val	Ser
Asn	Lys	Ala 115	Leu	Pro	Ala	Pro	Ile 120	Glu	Lys	Thr	Ile	Ser 125	Lys	Ala	Lys
Gly	Gln 130	Pro	Arg	Glu	Pro	Gln 135	Val	Tyr	Thr	Leu	Pro 140	Pro	Ser	Arg	Asp
Glu 145	Leu	Thr	Lys	Asn	Gln 150	Val	Ser	Leu	Thr	Cys 155	Leu	Val	Lys	Gly	Phe 160
Tyr	Pro	Ser	Asp	Ile 165	Ala	Val	Glu	Trp	Glu 170	Ser	Asn	Gly	Gln	Pro 175	Glu
Asn	Asn	Tyr	Lys 180	Thr	Thr	Pro	Pro	Val 185	Leu	Asp	Ser	Asp	Gly 190	Ser	Phe
Phe	Leu	Tyr 195	Ser	Lys	Leu	Thr	Val 200	Asp	Lys	Ser	Arg	Trp 205	Gln	Gln	Gly
Asn	Val 210	Phe	Ser	Суз	Ser	Val 215	Met	His	Glu	Ala	Leu 220	His	Asn	His	Tyr
Thr 225	Gln	Lys	Ser	Leu	Ser 230	Leu	Ser	Pro	Gly	Lys 235	Lys	Asp	Pro	Lys	Phe 240
Trp	Val	Leu	Val	Val 245	Val	Gly	Gly	Val	Leu 250	Ala	Суз	Tyr	Ser	Leu 255	Leu
His															
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gatecegeeg ageceaate teetgacaaa acteacat geecaecgtg eecageacet gaacteetgg ggggaeegte agtetteete teeeceaa aaceeaagga eaceeteg ateteeegga eecetgaggt eacatgegtg gtggtggaeg tgageeagga agaeeetgag gteaagttee aetggtaeg ggaeggegt gaggtgeata atgeeaagae aaageeggg gaggageagt acaacageae gtaeegtgtg gteagegtee teaeegteet geaeeaggae tggetgaatg geaaggagta eaagtgeaag gteteeaaea aageeeteee ageeeeate

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60

120

180

240

300

360

420

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Tyr	Tyr	Cys 115	Ala	Arg	Gly	Tyr	Gly 120	Thr	Ala	Tyr	Gly	Val 125	Asp	Tyr	Trp
Gly	Gln 130	Gly	Thr	Ser	Val	Thr 135	Val	Ser	Ser	Ala	Lys 140	Thr	Thr	Pro	Pro
Lys 145	Leu	Glu	Glu	Gly	Glu 150	Phe	Ser	Glu	Ala	Arg 155	Val	Asp	Ile	Val	Leu 160
Thr	Gln	Ser	Pro	Ala 165	Ser	Leu	Ala	Val	Ser 170	Leu	Gly	Gln	Arg	Ala 175	Thr
Ile	Ser	Суз	Arg 180	Ala	Ser	Glu	Ser	Val 185	Asp	Asn	Tyr	Gly	Ile 190	Ser	Phe
Met	Asn	Trp 195	Phe	Gln	Gln	ГЛа	Pro 200	Gly	Gln	Pro	Pro	Lys 205	Leu	Leu	Ile
Tyr	Ala 210	Ala	Ser	Arg	Gln	Gly 215	Ser	Gly	Val	Pro	Ala 220	Arg	Phe	Ser	Gly
Ser 225	Gly	Ser	Gly	Thr	Asp 230	Phe	Ser	Leu	Asn	Ile 235	His	Pro	Met	Glu	Glu 240
Asp	Asp	Thr	Ala	Met 245	Tyr	Phe	Суз	Gln	Gln 250	Ser	Lys	Glu	Val	Pro 255	Trp
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Pro	Glu 290	Leu	Leu	Gly	Gly	Pro 295	Ser	Val	Phe	Leu	Phe 300	Pro	Pro	Lys	Pro
Lys 305	Asp	Thr	Leu	Met	Ile 310	Ser	Arg	Thr	Pro	Glu 315	Val	Thr	Суз	Val	Val 320
Val	Asp	Val	Ser	His 325	Glu	Asp	Pro	Glu	Val 330	Гла	Phe	Asn	Trp	Tyr 335	Val
Asp	Gly	Val	Glu 340	Val	His	Asn	Ala	Lys 345	Thr	Lys	Pro	Arg	Glu 350	Glu	Gln
Tyr	Asn	Ser 355	Thr	Tyr	Arg	Val	Val 360	Ser	Val	Leu	Thr	Val 365	Leu	His	Gln
Aap	Trp 370	Leu	Asn	Gly	Гла	Glu 375	Tyr	Гла	Суз	Lys	Val 380	Ser	Asn	Lys	Ala
Leu 385	Pro	Ala	Pro	Ile	Glu 390	Гла	Thr	Ile	Ser	Lys 395	Ala	Lys	Gly	Gln	Pro 400
Arg	Glu	Pro	Gln	Val 405	Tyr	Thr	Leu	Pro	Pro 410	Ser	Arg	Asp	Glu	Leu 415	Thr
Lys	Asn	Gln	Val 420	Ser	Leu	Thr	Сув	Leu 425	Val	Lys	Gly	Phe	Tyr 430	Pro	Ser
Aap	Ile	Ala 435	Val	Glu	Trp	Glu	Ser 440	Asn	Gly	Gln	Pro	Glu 445	Asn	Asn	Tyr
Lys	Thr 450	Thr	Pro	Pro	Val	Leu 455	Asp	Ser	Asp	Gly	Ser 460	Phe	Phe	Leu	Tyr
Ser 465	Lys	Leu	Thr	Val	Asp 470	Lys	Ser	Arg	Trp	Gln 475	Gln	Gly	Asn	Val	Phe 480
Ser	Суз	Ser	Val	Met 485	His	Glu	Ala	Leu	His 490	Asn	His	Tyr	Thr	Gln 495	Lys

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85

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Cys 225	Ala	Gly	Суз	Ala	Gly 230	Сүз	Ala	Сув	Суз	Gly 235	Суз	Суз	Thr	Ala	Cys 240
Ala	Thr	Gly	Сүз	Ala 245	Gly	Сүз	Thr	Gly	Thr 250	Сүз	Сүз	Ala	Gly	Cys 255	Сүз
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Сүз	Gly	Cys 275	Cys	Gly	Thr	Gly	Thr 280	Ala	Cys	Thr	Ala	Cys 285	Thr	Gly	Суз
Gly	Cys 290	Суз	Ala	Gly	Ala	Gly 295	Gly	Суз	Thr	Ala	Сув 300	Gly	Gly	Суз	Ala
Суя 305	Ala	Gly	Суз	Суз	Thr 310	Ala	Суз	Gly	Gly	Cys 315	Gly	Thr	Gly	Gly	Ala 320
Сүз	Thr	Ala	Thr	Thr 325	Gly	Gly	Gly	Gly	Суз 330	Суз	Ala	Gly	Gly	Gly 335	Суз
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-	370	-	-	-	-	Thr 375			-	-	380	-	-		
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	450		-	-	-	Thr 455	-	-	-		460		-		-
465	-	•	•		470	Суз			•	475	•	-		-	480
	-	-	-	485	-	Ala	-	-	490		-		-	495	-
			500			Ala		505					510		
-		515	-			СЛа	520		-			525		-	-
Thr	Thr 530	Сув	Сув	Ala	Gly	Сув 535	Ala	Gly	Ala	Ala	G1y 540	СЛа	Сув	Сув	Gly
Gly 545	Сув	Сув	Ala	Gly	Сув 550	СЛа	Суз	Сув	Суз	Сув 555	Ala	Ala	Gly	Сув	Thr 560
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Ala	Gly	Суз	Ala 580	Gly	Ala	Суз	Ala	Gly 585	Gly	Gly	Сүз	Ala	Gly 590	Суз	Gly
Gly	Ala	Gly 595	Thr	Gly	Сүз	Суз	Thr 600	Gly	Суз	Суз	Ala	Gly 605	Ala	Thr	Thr

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 710
 715
 720
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Thr	Gly 1070		Gly	Gly	Суз	Cys 1075		Gly	Gly	Gly	Cys 1080	Ala	Суз	Ala
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Cya	Cys 1115		Суа	Сув	Thr	Ala 1120		Gly	Cys	Thr	Gly 1125	Gly	Ala	Ala
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Thr		Thr	Суз	Суз	Ala		Суз	Сув	Ala	Gly	1170 Cys	Суз	Thr	Gly
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Сув		Gly	Ala	Gly	Ala		Суз	Сув	Ala	Сув	1200 Сув	Ala	Thr	Cys
-	1205	-		-		1210	-	-		-	1215 Cys			-
	1220					1225					1230 Cys			
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	1265					1270					Cys 1275			
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СЛа	Cys 1295		Суз	Суз	Сүз	Ala 1300		Gly	Суз	Thr	Gly 1305	Сүз	Thr	Gly
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Ala	Gly 1325	Ala	Суз	Ala	Gly	Gly 1330		Cys	Ala	Gly	Cys 1335	Gly	Gly	Ala
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Thr	Cys 1355		Gly	Gly	Сүз	Ala 1360	-	Сув	Gly	Gly	Сув 1365		Сув	Суз
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Glu Pro Gln Glu Ile Asn Phe Pro Asp Asp Leu Pro Gly Ser Asn Thr 20 25 30
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115 120 125 Leu Gln Val Ala Ala Val Asp Ser Ser Val Pro Arg Thr Ala Glu Leu 130 135 140 Ala Gly Ile Thr Thr Leu Asp Asp Pro Leu Gly His Met Pro Glu Arg 145 150 155 160 Phe Asp Ala Phe Ile Cys Tyr Cys Pro Ser Asp Ile 165 170 <210> SEQ ID NO 79 <211> LENGTH: 516 <212> TYPE: DNA <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: Synthetic Polynucleotide <400> SEQUENCE: 79 atggetgetg geggaeetgg egeeggatet getgeteetg tgtetageae aageageetg 60 cctctggccg ccctgaacat gagagtgcgg agaaggctga gcctgttcct gaacgtgcgg 120 acacaggtgg ccgccgattg gacagccctg gccgaggaaa tggacttcga gtacctggaa 180 atccggcagc tggaaaccca ggccgaccct acaggcagac tgctggatgc ttggcagggc 240 agaccaggcg cttctgtggg aaggctgctg gaactgctga ccaagctggg cagggacgac 300 gtgctgctgg aactgggccc tagcatcgaa gaggactgcc agaagtacat cctgaagcag 360 cagcaggaag aggccgagaa gcctctgcag gtggcagccg tggatagcag cgtgccaaga 420 acageegage tggeeggeat caceaceetg gatgateete tgggeeacat geeegagaga 480 ttcgacgcct tcatctgcta ctgccccagc gacatc 516 <210> SEQ ID NO 80 <211> LENGTH: 18 <212> TYPE: PRT <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: Synthetic Polypeptide <400> SEQUENCE: 80 Ala Lys Thr Thr Pro Pro Lys Leu Glu Glu Gly Glu Phe Ser Glu Ala 5 1 10 15 Arg Val <210> SEQ ID NO 81 <211> LENGTH: 42 <212> TYPE: PRT <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: Synthetic Polypeptide <400> SEOUENCE: 81 Lys Arg Gly Arg Lys Lys Leu Leu Tyr Ile Phe Lys Gln Pro Phe Met 1 5 10 15 Arg Pro Val Gln Thr Thr Gln Glu Glu Asp Gly Cys Ser Cys Arg Phe 20 25 30 Pro Glu Glu Glu Glu Gly Gly Cys Glu Leu 35 40 <210> SEQ ID NO 82

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Val Gln		Thr	Gln	Glu	Glu		Gly	Суз	Ser	Суз		Phe	Pro	Glu					
50 Glu Glu	Glu	Glv	Glv	Cvs	55 Glu	Leu	Ara	Val	Lvs	60 Phe	Ser	Ara	Ser	Ala					
65				70					75					80					
Asp Ala	Pro	Ala	Tyr 85	Gln	Gln	Gly	Gln	Asn 90	Gln	Leu	Tyr	Asn	Glu 95	Leu					
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Arg Asp	Pro 115	Glu	Met	Gly	Gly	Lys 120	Pro	Arg	Arg	ГЛЗ	Asn 125	Pro	Gln	Glu					
Gly Leu 130	Tyr	Asn	Glu	Leu	Gln 135	Lys	Asp	Lys	Met	Ala 140	Glu	Ala	Tyr	Ser					
Glu Ile	Gly	Met	Lys	-		Arg	Arg	Arg	_		Gly	His	Asp	-					
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Pro Thr Arg Lys His Tyr Gln Pro Tyr Ala Pr 50 55	Pro Pro Arg Asp Phe Ala 60
Ala Tyr Arg Ser Arg Asp Gln Arg Leu Pro Pr	Pro Asp Ala His Lys Pro
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85 90 Ala His Ser Thr Leu Ala Lys Ile Arg Val Ly	95 Lvs Phe Ser Arg Ser Ala
100 105	110
Asp Ala Pro Ala Tyr Gln Gln Gly Gln Asn Gl 115 120	Gin Leu Tyr Asn Giu Leu 125
Asn Leu Gly Arg Arg Glu Glu Tyr Asp Val Le 130 135	Leu Asp Lys Arg Arg Gly 140
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Gly Leu Tyr Asn Glu Leu Gln Lys Asp Lys Me 165 170	Met Ala Glu Ala Tyr Ser 175
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Asp Pro	Glu 435	Met	Gly	Gly	Гла	Pro 440	Arg	Arg	Lys	Asn	Pro 445	Gln	Glu	Gly		
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	Pro Gly Ala Ser 35	: Val Ly		Ser Cy: 40	a Lya Ala	a Ser	Gly 45	Tyr	Thr	Phe			
	Ser Asn Tyr Leu 50	ı Met As	n Trp ' 55	Val Ly:	3 Gln Arq	g Pro 60	Glu	Gln	Asp	Leu			
	Asp Trp Ile Gly 65	/ Arg Il 70		Pro Tyj	r Asp Gly 75	y Asp	Ile	Asp	Tyr	Asn 80			
	Gln Asn Phe Lys	8 Asp Ly 85	vs Ala	Ile Leu	ı Thr Va 90	l Asp	ГЛа	Ser	Ser 95	Ser			
	Thr Ala Tyr Met 100		eu Ser (Ser Leu 105		r Glu	Asp	Ser 110	Ala	Val			
	Tyr Tyr Cys Ala 115	ι Arg Gl		Gly Thi 120	r Ala Ty:	r Gly	Val 125	Asp	Tyr	Trp			
	Gly Gln Gly Thr 130	: Ser Va	al Thr ' 135	Val Sei	r Ser Ala	a Lys 140	Thr	Thr	Pro	Pro			
	Lys Leu Glu Glu 145	ı Gly Gl 15		Ser Glu	ı Ala Arg 15!		Asp	Ile	Val	Leu 160			
	Thr Gln Ser Pro	Ala Se 165	er Leu .	Ala Val	l Ser Leu 170	ı Gly	Gln	Arg	Ala 175	Thr			
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1.-31. (canceled)

32. An IL13R α 2-specific chimeric antigen receptor (CAR) comprising:

(A) an ectodomain comprising each of the amino acid sequences of:

(i)	(SEO	ID	NO:	1)
NYLMN;				-,
(ii)	(SEQ	тр	NO·	2)
RIDPYDGDIDYNQNFKD;	(522	10	110.	2,
(III)	(SEO	тп	NO	3)
GYGTAYGVDY ;	(559		110.	5,
(iv)	(SEQ	TD	NO ·	4)
RASESVDNYGISFMN;	(-,
(v)	(SEO	ID	NO :	5)
AASRQGSG ; and	(220	10		0,
(vi)	(SEQ	TD	NO ·	6)
QQSKEVPWT;				- /

(B) a spacer region;

(C) a transmembrane domain; and

(D) an endodomain selected from the group consisting of CD3.ζ, CD28.ζ, CD28.OX40.ζ, CD28.41 BB.ζ and 41BB.ζ.

33.-35. (canceled)

36. The IL13R α 2-specific chimeric antigen receptor of claim **32** wherein the spacer region comprises SEQ ID NO:103, SEQ ID NO:35, or SEQ ID NO:37.

37. The IL13R α 2-specific chimeric antigen receptor of claim **32** wherein the transmembrane domain is the transmembrane domain of CD28 or CD8 α .

38. The IL13R α 2-specific CAR of claim **37**, wherein the transmembrane domain of CD28 comprises the amino acid sequence of SEQ ID NO: 39.

39. The IL13R α 2-specific CAR of claim **32**, wherein the endodomain comprises a signaling domain of one or more of: CD137, CD134, CD27, CD40, ICOS, or Myd88.

40. (canceled)

41. The IL13R α 2-specific CAR of claim **32**, wherein the CD3 zeta chain signaling domain comprises the amino acid sequence of SEQ ID NO: 41.

42. The IL13R α 2-specific CAR of claim 32, comprising the amino acid sequence of SEQ ID NO: 47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, or at least one of SEQ ID NO:7 and SEQ ID NO:8.

43. (canceled)

44. (canceled)

45. The IL13R α 2-specific CAR of claim **42**, wherein the amino acid sequence of SEQ ID NO: 7 is fused to the amino acid sequence of SEQ ID NO: 8 through a linker.

46. The IL13R α 2-specific CAR of claim **45**, wherein the linker comprises the amino acid sequence of EEGEFSEAR (SEQ ID NO 10).

47. The IL13R α 2-specific CAR of claim 46, comprising the amino acid sequence of SEQ ID NO: 13.

48. (canceled)

49. A nucleic acid encoding the IL13R α 2-specific CAR of claim **32**.

50. The nucleic acid of claim **49**, comprising the sequence of SEQ ID NO: 34, 36, 38, 40, 42, 44, 46, 48, 50, 54, 56, or 65.

51. The nucleic acid of claim **49**, wherein the nucleic acid is localized in a vector.

52. (canceled)

53. The vector of claim **51**, wherein the vector is localized in a host cell.

54. The host cell of claim **53**, which is a human host cell. **55**. The host cell of claim **53**, which is a T-lymphocyte or a natural killer cell.

56.-59. (canceled)

60. The IL13R α 2-specific CAR of claim **32** wherein the CAR is combined with a pharmaceutically acceptable carrier in a pharmaceutical composition.

61. A method of treating a cancer in a subject, comprising administering to the subject a population of cells comprising the IL13R α 2-specific CAR of claim **32**, in an amount effective to treat the cancer in the subject.

62. The method of claim **61**, wherein the cancer is gliobastoma multiforme or colon cancer.

63. The method of claim **61**, wherein the population of cells is obtained from the subject.

64. The method of claim 63, wherein the cells are T-lymphocytes or natural killer cells obtained from the subject.

65. (canceled)

66. The nucleic acid of claim **47**, wherein the nucleic acid is combined with a pharmaceutically acceptable carrier in a pharmaceutical composition.

* * * * *