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## (54) COMPOSITIONS AND METHODS RELATED TO PROTEIN A (SPA) VARIANTS

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#### (57) ABSTRACT

The present invention concerns methods and compositions for treating or preventing a bacterial infection, particularly infection by a *Staphylococcus* bacterium. The invention provides methods and compositions for stimulating an immune response against the bacteria. In certain embodiments, the methods and compositions involve a non-toxigenic Protein A (SpA) variant.

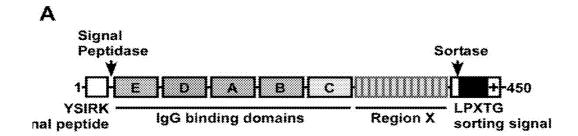


FIG. 1A

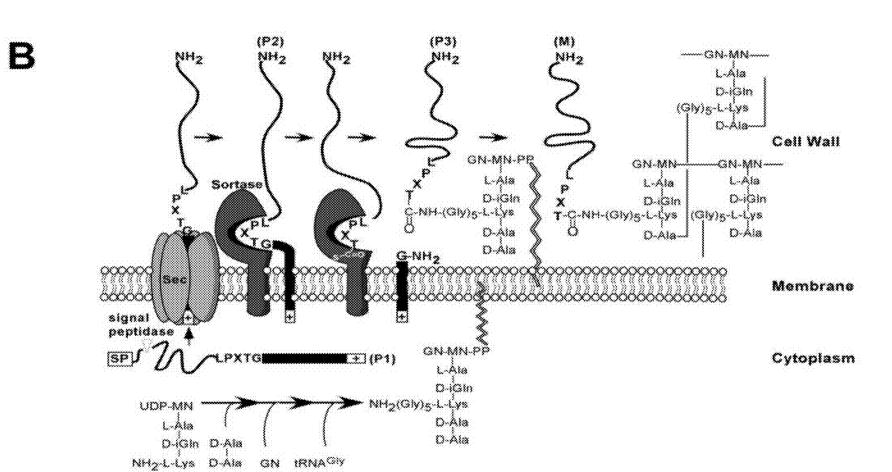


FIG. 1B

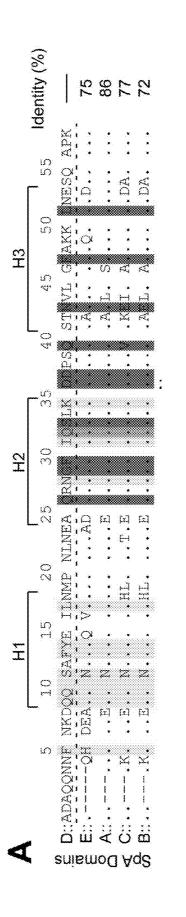


FIG. 2A

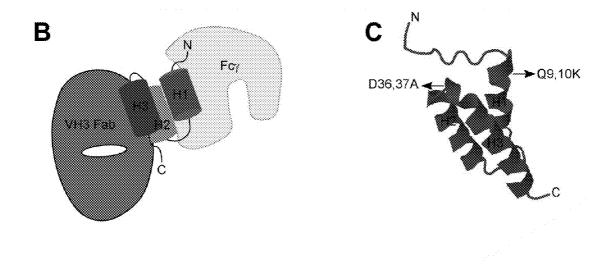


FIG. 2B-2C

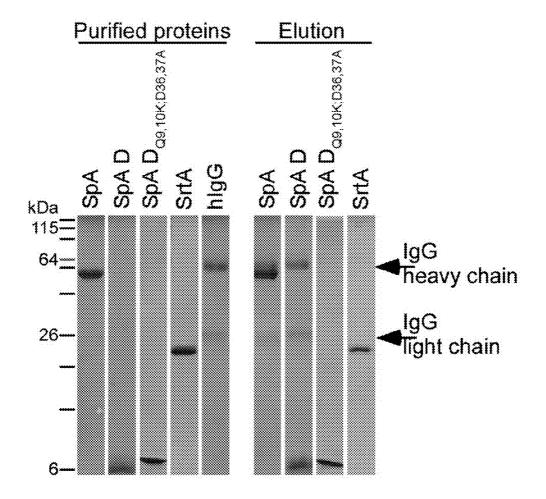
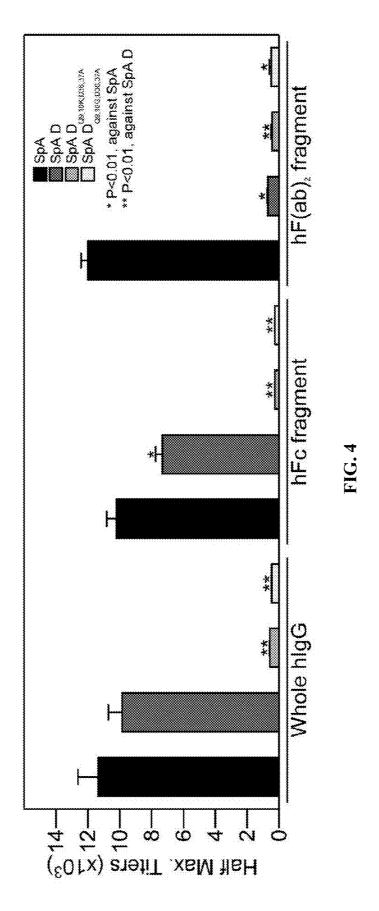


FIG. 3



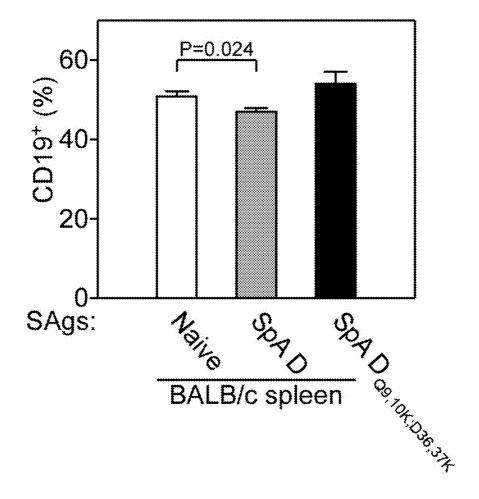


FIG. 5

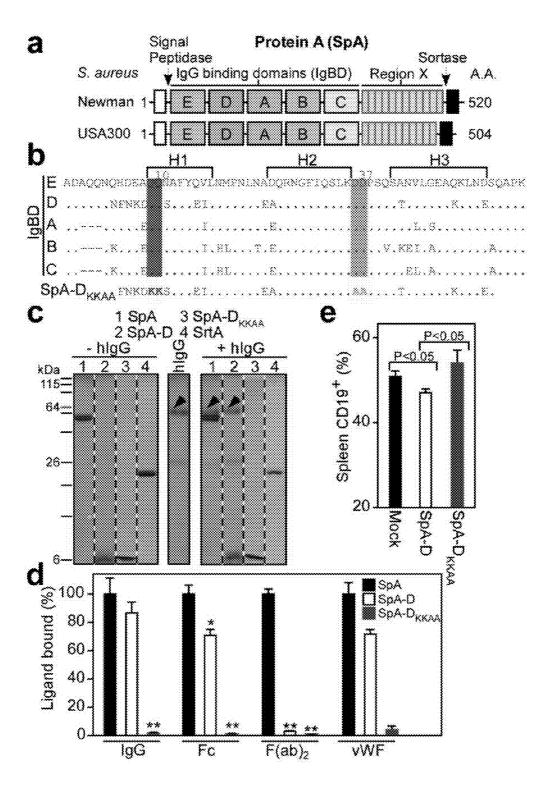
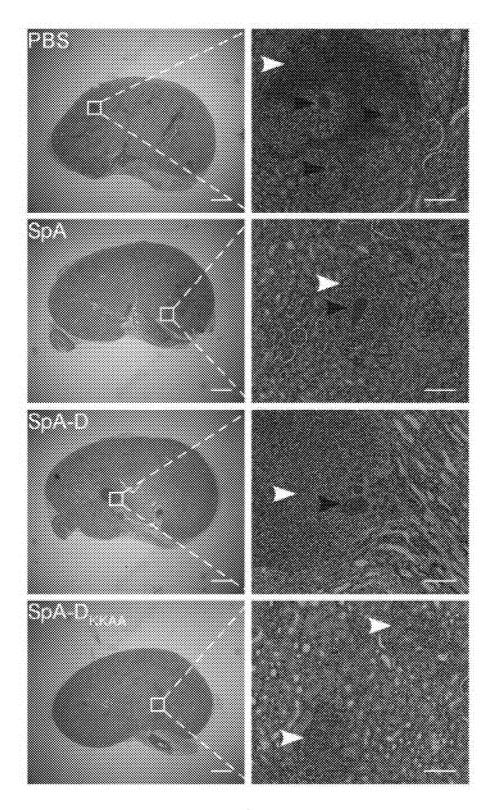


FIG. 6



**FIG.** 7

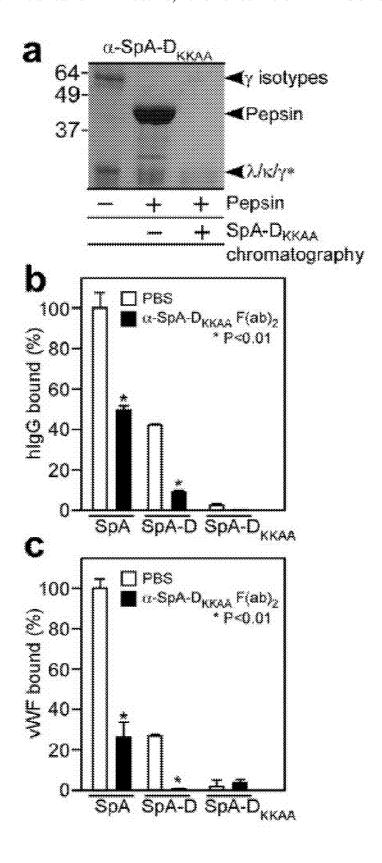


FIG. 8

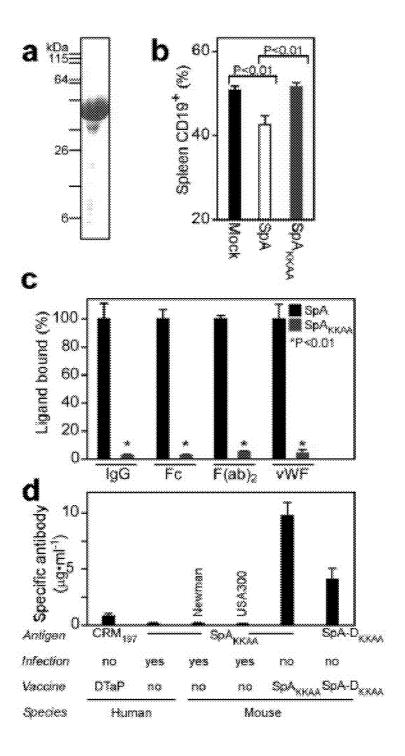


FIG. 9

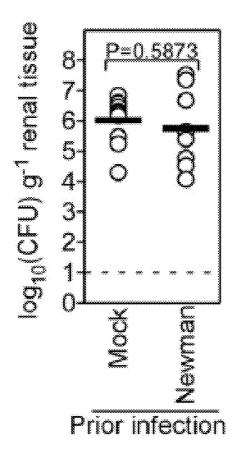


FIG. 10

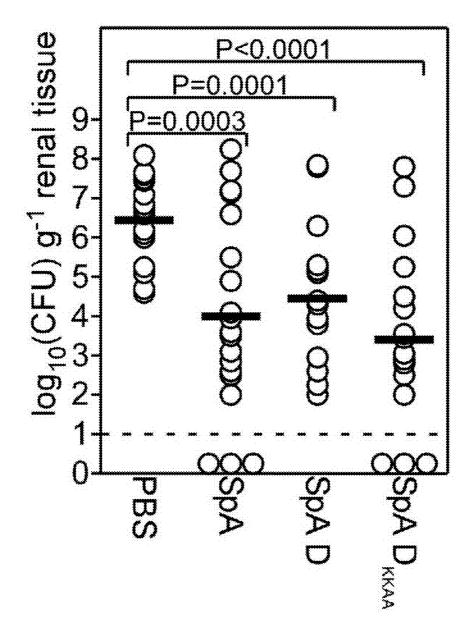


FIG. 11

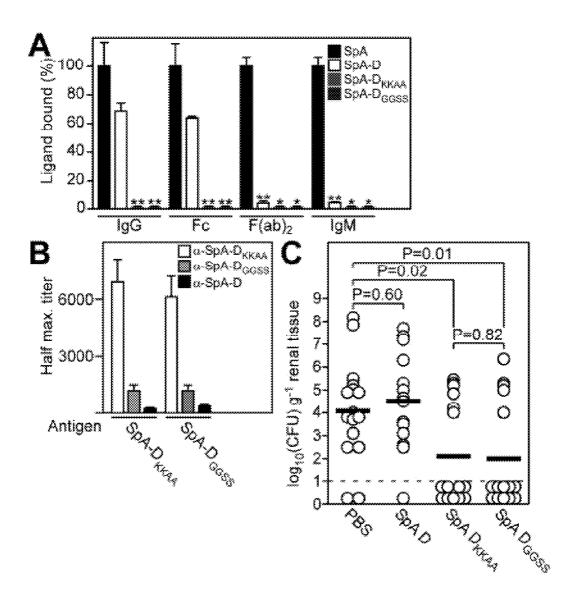


FIG. 12A-12C

### COMPOSITIONS AND METHODS RELATED TO PROTEIN A (SPA) VARIANTS

[0001] This application claims the benefit of priority to U.S. Provisional Patent Application Ser. No. 61/361,218 filed Jul. 2, 2010, and 61/370,725 filed Aug. 4, 2010, hereby incorporated by reference in their entirety.

[0002] This invention was made with government support under AI057153, AI052474, and GM007281 awarded by the National Institutes of Health. The government has certain rights in the invention.

#### BACKGROUND OF THE INVENTION

[0003] I. Field of the Invention

[0004] The present invention relates generally to the fields of immunology, microbiology, and pathology. More particularly, it concerns methods and compositions involving bacterial Protein A variants, which can be used to invoke an immune response against the bacteria.

[0005] II. Background

[0006] The number of both community acquired and hospital acquired infections have increased over recent years with the increased use of intravascular devices. Hospital acquired (nosocomial) infections are a major cause of morbidity and mortality, more particularly in the United States, where it affects more than 2 million patients annually. The most frequent infections are urinary tract infections (33% of the infections), followed by pneumonia (15.5%), surgical site infections (14.8%) and primary bloodstream infections (13%) (Emorl and Gaynes, 1993).

[0007] The major nosocomial pathogens include Staphylococcus aureus, coagulase-negative Staphylococci (mostly Staphylococcus epidermidis), enterococcus spp., Escherichia coli and Pseudomonas aeruginosa. Although these pathogens cause approximately the same number of infections, the severity of the disorders they can produce combined with the frequency of antibiotic resistant isolates balance this ranking towards S. aureus and S. epidermidis as being the most significant nosocomial pathogens.

[0008] Staphylococci can cause a wide variety of diseases in humans and other animals through either toxin production or invasion. Staphylococcal toxins are also a common cause of food poisoning, as the bacteria can grow in improperly-stored food.

[0009] Staphylococcus epidermidis is a normal skin commensal which is also an important opportunistic pathogen responsible for infections of impaired medical devices and infections at sites of surgery. Medical devices infected by S. epidermidis include cardiac pacemakers, cerebrospinal fluid shunts, continuous ambulatory peritoneal dialysis catheters, orthopedic devices and prosthetic heart valves.

[0010] Staphylococcus aureus is the most common cause of nosocomial infections with a significant morbidity and mortality. It is the cause of some cases of osteomyelitis, endocarditis, septic arthritis, pneumonia, abscesses, and toxic shock syndrome. S. aureus can survive on dry surfaces, increasing the chance of transmission. Any S. aureus infection can cause the staphylococcal scalded skin syndrome, a cutaneous reaction to exotoxin absorbed into the blood-stream. It can also cause a type of septicemia called pyaemia that can be life-threatening. Problematically, Methicillin-resistant Staphylococcus aureus (MRSA) has become a major cause of hospital-acquired infections.

[0011] S. aureus and S. epidermidis infections are typically treated with antibiotics, with penicillin being the drug of choice, whereas vancomycin is used for methicillin resistant isolates. The percentage of staphylococcal strains exhibiting wide-spectrum resistance to antibiotics has become increasingly prevalent, posing a threat for effective antimicrobial therapy. In addition, the recent emergence of vancomycin resistant S. aureus strain has aroused fear that MRSA strains are emerging and spreading for which no effective therapy is available.

[0012] An alternative to antibiotic treatment for staphylococcal infections is under investigation that uses antibodies directed against staphylococcal antigens. This therapy involves administration of polyclonal antisera (WO00/15238, WO00/12132) or treatment with monoclonal antibodies against lipoteichoic acid (WO98/57994).

[0013] An alternative approach would be the use of active vaccination to generate an immune response against staphylococci. The *S. aureus* genome has been sequenced and many of the coding sequences have been identified (WO02/094868, EP0786519), which can lead to the identification of potential antigens. The same is true for *S. epidermidis* (WO01/34809). As a refinement of this approach, others have identified proteins that are recognized by hyperimmune sera from patients who have suffered staphylococcal infection (WO01/98499, WO02/059148).

[0014] S. aureus secretes a plethora of virulence factors into the extracellular milieu (Archer, 1998; Dinges et al., 2000; Foster, 2005; Shaw et al., 2004; Sibbald et al., 2006). Like most secreted proteins, these virulence factors are translocated by the Sec machinery across the plasma membrane. Proteins secreted by the Sec machinery bear an N-terminal leader peptide that is removed by leader peptidase once the pre-protein is engaged in the Sec translocon (Dalbey and Wickner, 1985; van Wely et al., 2001). Recent genome analysis suggests that Actinobacteria and members of the Firmicutes encode an additional secretion system that recognizes a subset of proteins in a Sec-independent manner (Pallen, 2002). ESAT-6 (early secreted antigen target 6 kDa) and CFP-10 (culture filtrate antigen 10 kDa) of Mycobacterium tuberculosis represent the first substrates of this novel secretion system termed ESX-1 or Snm in M. tuberculosis (Andersen et al., 1995; Hsu et al., 2003; Pym et al., 2003; Stanley et al., 2003). In S. aureus, two ESAT-6 like factors designated EsxA and EsxB are secreted by the Ess pathway (ESAT-6 secretion system) (Burts et al., 2005).

[0015] The first generation of vaccines targeted against *S. aureus* or against the exoproteins it produces have met with limited success (Lee, 1996). There remains a need to develop effective vaccines against staphylococcal infections. Additional compositions for treating staphylococcal infections are also needed.

#### SUMMARY OF THE INVENTION

[0016] Protein A (SpA)(SEQ ID NO:33), a cell wall anchored surface protein of *Staphylococcus aureus*, provides for bacterial evasion from innate and adaptive immune responses. Protein A binds immunoglobulins at their Fc portion, interacts with the VH3 domain of B cell receptors inappropriately stimulating B cell proliferation and apotosis, binds to von Willebrand factor A1 domains to activate intracellular clotting, and also binds to the TNF Receptor-1 to contribute to the pathogenesis of staphylococcal pneumonia. Due to the fact that Protein A captures immunoglobulin and

displays toxic attributes, the possibility that this surface molecule may function as a vaccine in humans has not been rigorously pursued. Here the inventors demonstrate that Protein A variants no longer able to bind to immunoglobulins, which are thereby removed of their toxigenic potential, i.e., are non-toxigenic, stimulate humoral immune responses that protect against staphylococcal disease.

[0017] In certain embodiments the SpA variant is a full length SpA variant comprising a variant A, B, C, D, and/or E domain. In certain aspects, the SpA variant comprises or consists of the amino acid sequence that is 80, 90, 95, 98, 99, or 100% identical to the amino acid sequence of SEQ ID NO:34 In other embodiments the SpA variant comprises a segment of SpA. The SpA segment can comprise at least or at most 1, 2, 3, 4, 5 or more IgG binding domains. The IgG domains can be at least or at most 1, 2, 3, 4, 5 or more variant A, B, C, D, or E domains. In certain aspects the SpA variant comprises at least or at most 1, 2, 3, 4, 5, or more variant A domains. In a further aspect the SpA variant comprises at least or at most 1, 2, 3, 4, 5, or more variant B domains. In still a further aspect the SpA variant comprises at least or at most 1, 2, 3, 4, 5, or more variant C domains. In yet a further aspect the SpA variant comprises at least or at most 1, 2, 3, 4, 5, or more variant D domains. In certain aspects the SpA variant comprises at least or at most 1, 2, 3, 4, 5, or more variant E domains. In a further aspect the SpA variant comprises a combination of A, B, C, D, and E domains in various combinations and permutations. The combinations can include all or part of a SpA signal peptide segment, a SpA region X segment, and/or a SpA sorting signal segment. In other aspects the SpA variant does not include a SpA signal peptide segment, a SpA region X segment, and/or a SpA sorting signal segment. In certain aspects a variant A domain comprises a substitution at position(s) 7, 8, 34, and/or 35 of SEQ ID NO:4. In another aspect a variant B domain comprises a substitution at position(s) 7, 8, 34, and/or 35 of SEQ ID NO:6. In still anther aspect a variant C domain comprises a substitution at position(s) 7, 8, 34, and/or 35 of SEQ ID NO:5. In certain aspects a variant D domain comprises a substitution at position(s) 9, 10, 36, and/or 37 of SEQ ID NO:2. In a further aspect a variant E domain comprises a substitution at position (s) 6, 7, 33, and/or 34 of SEQ ID NO:3.

[0018] In certain aspects, an SpA domain D variant or its equivalent can comprise a mutation at position 9 and 36; 9 and 37; 9 and 10; 36 and 37; 10 and 36; 10 and 37; 9, 36, and 37; 10, 36, and 37, 9, 10 and 36; or 9, 10 and 37 of SEQ ID NO:2. In a further aspect, analogous mutations can be included in one or more of domains A, B, C, or E.

[0019] In further aspects, the amino acid glutamine (Q) at position 9 of SEQ ID NO:2 (or its analogous amino acid in other SpA domains) can be replaced with an alanine (A), an asparagine (N), an aspartic acid (D), a cysteine (C), a glutamic acid (E), a phenylalanine (F), a glycine (G), a histidine (H), an isoleucine (I), a lysine (K), a leucine (L), a methionine (M), a proline (P), a serine (S), a threonine (T), a valine (V), a tryptophane (W), or a tyrosine (Y). In some aspects the glutamine at position 9 can be substituted with an arginine (R). In a further aspect, the glutamine at position 9 of SEQ ID NO:2, or its equivalent, can be substituted with a lysine or a glycine. Any 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more of the substitutions can be explicitly excluded.

[0020] In another aspect, the amino acid glutamine (Q) at position 10 of SEQ ID NO:2 (or its analogous amino acid in other SpA domains) can be replaced with an alanine (A), an

asparagine (N), an aspartic acid (D), a cysteine (C), a glutamic acid (E), a phenylalanine (F), a glycine (G), a histidine (H), an isoleucine (I), a lysine (K), a leucine (L), a methionine (M), a proline (P), a serine (S), a threonine (T), a valine (V), a tryptophane (W), or a tyrosine (Y). In some aspects the glutamine at position 10 can be substituted with an arginine (R). In a further aspect, the glutamine at position 10 of SEQ ID NO:2, or its equivalent, can be substituted with a lysine or a glycine. Any 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more of the substitutions can be explicitly excluded.

[0021] In certain aspects, the aspartic acid (D) at position 36 of SEQ ID NO:2 (or its analogous amino acid in other SpA domains) can be replaced with an alanine (A), an asparagine (N), an arginine (R), a cysteine (C), a phenylalanine (F), a glycine (G), a histidine (H), an isoleucine (I), a lysine (K), a leucine (L), a methionine (M), a proline (P), a glutamine (Q), a serine (S), a threonine (T), a valine (V), a tryptophane (W), or a tyrosine (Y). In some aspects the aspartic acid at position 36 can be substituted with a glutamic acid (E). In certain aspects, an aspartic acid at position 36 of SEQ ID NO:2, or its equivalent, can be substituted with an alanine or a serine. Any 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more of the substitutions can be explicitly excluded.

[0022] In another aspect, the aspartic acid (D) at position 37 of SEQ ID NO:2 (or its analogous amino acid in other SpA domains) can be replaced with an alanine (A), a an asparagine (N), an arginine (R), a cysteine (C), a phenylalanine (F), a glycine (G), a histidine (H), an isoleucine (I), a lysine (K), a leucine (L), a methionine (M), a proline (P), a glutamine (Q), a serine (S), a threonine (T), a valine (V), a tryptophane (W), or a tyrosine (Y). In some aspects the aspartic acid at position 37 can be substituted with a glutamic acid (E). In certain aspects, an aspartic acid at position 37 of SEQ ID NO:2, or its equivalent, can be substituted with an alanine or a serine. Any 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more of the substitutions can be explicitly excluded.

[0023] In a particular embodiment the amino at position 9 of SEQ ID NO:2 (or an analogous amino acid in another SpA domain) is replaced by an alanine (A), a glycine (G), an isoleucine (I), a leucine (L), a proline (P), a serine (S), or a valine (V), In certain aspects the amino acid at position 9 of SEQ ID NO:2 is replaced by a glycine. In a further aspect the amino acid at position 9 of SEQ ID NO:2 is replaced by a lysine.

[0024] In a particular embodiment the amino at position 10 of SEQ ID NO:2 (or an analogous amino acid in another SpA domain) is replaced by an alanine (A), a glycine (G), an isoleucine (I), a leucine (L), a proline (P), a serine (S), or a valine (V), In certain aspects the amino acid at position 10 of SEQ ID NO:2 is replaced by a glycine. In a further aspect the amino acid at position 10 of SEQ ID NO:2 is replaced by a lysine.

[0025] In a particular embodiment the amino at position 36 of SEQ ID NO:2 (or an analogous amino acid in another SpA domain) is replaced by an alanine (A), a glycine (G), an isoleucine (I), a leucine (L), a proline (P), a serine (S), or a valine (V), In certain aspects the amino acid at position 36 of SEQ ID NO:2 is replaced by a serine. In a further aspect the amino acid at position 36 of SEQ ID NO:2 is replaced by an alanine

[0026] In a particular embodiment the amino at position 37 of SEQ ID NO:2 (or an analogous amino acid in another SpA domain) is replaced by an alanine (A), a glycine (G), an isoleucine (I), a leucine (L), a proline (P), a serine (S), or a

valine (V), In certain aspects the amino acid at position 37 of SEQ ID NO:2 is replaced by a serine. In a further aspect the amino acid at position 37 of SEQ ID NO:2 is replaced by an alanine

[0027] In certain aspects the SpA variant includes a substitution of (a) one or more amino acid substitution in an IgG Fc binding sub-domain of SpA domain A, B, C, D, and/or E that disrupts or decreases binding to IgG Fc, and (b) one or more amino acid substitution in a  $V_{H}3$  binding sub-domain of SpA domain A, B, C, D, and/or E that disrupts or decreases binding to  $V_{H}3$ . In still further aspects the amino acid sequence of a SpA variant comprises an amino acid sequence that is at least 50%, 60%, 70%, 80%, 90%, 95%, or 100% identical, including all values and ranges there between, to the amino acid sequence of SEQ ID NOs:2-6.

[0028] In a further aspect the SpA variant includes (a) one or more amino acid substitution in an IgG Fc binding subdomain of SpA domain D, or at a corresponding amino acid position in other IgG domains, that disrupts or decreases binding to IgG Fc, and (b) one or more amino acid substitution in a  $V_H$ 3 binding sub-domain of SpA domain D, or at a corresponding amino acid position in other IgG domains, that disrupts or decreases binding to  $V_H$ 3. In certain aspects amino acid residue F5, Q9, Q10, S11, F13, Y14, L17, N28, 131, and/or K35 (SEQ ID NO:2, QQNNFNKDQQSAFYEILN-MPNLNEAQRNGFIQSLKDDPSQSTNVLGEAKKLNES) of the IgG Fc binding sub-domain of domain D are modified or substituted. In certain aspects amino acid residue Q26, G29, F30, S33, D36, D37, Q40, N43, and/or E47 (SEQ ID NO:2) of the  $V_H$ 3 binding sub-domain of domain D are modified or substituted such that binding to Fc or  $V_H3$  is attenuated. In further aspects corresponding modifications or substitutions can be engineered in corresponding positions of the domain A, B, C, and/or E. Corresponding positions are defined by alignment of the domain D amino acid sequence with one or more of the amino acid sequences from other IgG binding domains of SpA, for example see FIG. 2A. In certain aspects the amino acid substitution can be any of the other 20 amino acids. In a further aspect conservative amino acid substitutions can be specifically excluded from possible amino acid substitutions. In other aspects only non-conservative substitutions are included. In any event, any substitution or combination of substitutions that reduces the binding of the domain such that SpA toxicity is significantly reduced is contemplated. The significance of the reduction in binding refers to a variant that produces minimal to no toxicity when introduced into a subject and can be assessed using in vitro methods described herein.

[0029] In certain embodiments, a variant SpA comprises at least or at most 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more variant SpA domain D peptides. In certain aspects 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, or 19 or more amino acid residues of the variant SpA are substituted or modifiedincluding but not limited to amino acids F5, Q9, Q10, S11, F13, Y14, L17, N28, 131, and/or K35 (SEQ ID NO:2) of the IgG Fc binding sub-domain of domain D and amino acid  $residue\,Q26,G29,F30,S33,D36,D37,Q40,N43,and/or\,E47$ (SEQ ID NO:2) of the  $V_H$ 3 binding sub-domain of domain D. In one aspect of the invention glutamine residues at position 9 and/or 10 of SEQ ID NO:2 (or corresponding positions in other domains) are mutated. In another aspect, aspartic acid residues 36 and/or 37 of SEQ ID NO:2 (or corresponding positions in other domains) are mutated. In a further aspect, glutamine 9 and 10, and aspartic acid residues 36 and 37 are mutated. Purified non-toxigenic SpA or SpA-D mutants/variants described herein are no longer able to significantly bind (i.e., demonstrate attenuated or disrupted binding affinity) Fey or  $F(ab)_2 V_H 3$  and also do not stimulate B cell apoptosis. These non-toxigenic Protein A variants can be used as subunit vaccines and raise humoral immune responses and confer protective immunity against S. aureus challenge. Compared to wild-type full-length Protein A or the wild-type SpA-domain D, immunization with SpA-D variants resulted in an increase in Protein A specific antibody. Using a mouse model of staphylococcal challenge and abscess formation, it was observed that immunization with the non-toxigenic Protein A variants generated significant protection from staphylococcal infection and abscess formation. As virtually all S. aureus strains express Protein A, immunization of humans with the non-toxigenic Protein A variants can neutralize this virulence factor and thereby establish protective immunity. In certain aspects the protective immunity protects or ameliorates infection by drug resistant strains of Staphylococcus, such as USA300 and other MRSA strains.

[0030] Embodiments include the use of Protein A variants in methods and compositions for the treatment of bacterial and/or staphylococcal infection. This application also provides an immunogenic composition comprising a Protein A variant or immunogenic fragment thereof. In certain aspects, the immunogenic fragment is a Protein A domain D segment. Furthermore, the present invention provides methods and compositions that can be used to treat (e.g., limiting staphylococcal abscess formation and/or persistence in a subject) or prevent bacterial infection. In some cases, methods for stimulating an immune response involve administering to the subject an effective amount of a composition including or encoding all or part of a Protein A variant polypeptide or antigen, and in certain aspects other bacterial proteins. Other bacterial proteins include, but are not limited to (i) a secreted virulence factor, and/or a cell surface protein or peptide, or (ii) a recombinant nucleic acid molecule encoding a secreted virulence factor, and/or a cell surface protein or peptide.

[0031] In other aspects, the subject can be administered all or part of a Protein A variant, such as a variant Protein A domain D segment. The polypeptide of the invention can be formulated in a pharmaceutically acceptable composition. The composition can further comprise one or more of at least or at most 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, or 19 additional staphylococcal antigen or immunogenic fragment thereof (e.g., Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla (e.g., H35 mutants), IsdC, SasF, vWbp, or vWh). Additional staphylococcal antigens that can be used in combination with a Protein A variant include, but are not limited to 52 kDa vitronectin binding protein (WO 01/60852), Aaa (GenBank CAC80837), Aap (GenBank accession AJ249487), Ant (Gen-Bank accession NP\_372518), autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibringen binding protein (U.S. Pat. No. 6,008, 341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg2+ transporter, MHC II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/ saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein (see PCT publications WO2007/113222, WO2007/113223, WO2006/ 032472, WO2006/032475, WO2006/032500, each of which is incorporated herein by reference in their entirety). The staphylococcal antigen or immunogenic fragment can be administered concurrently with the Protein A variant. The staphylococcal antigen or immunogenic fragment and the Protein A variant can be administered in the same composition. The Protein A variant can also be a recombinant nucleic acid molecule encoding a Protein A variant. A recombinant nucleic acid molecule can encode the Protein A variant and at least one staphylococcal antigen or immunogenic fragment thereof. As used herein, the term "modulate" or "modulation" encompasses the meanings of the words "enhance," or "inhibit." "Modulation" of activity may be either an increase or a decrease in activity. As used herein, the term "modulator" refers to compounds that effect the function of a moiety, including up-regulation, induction, stimulation, potentiation, inhibition, down-regulation, or suppression of a protein, nucleic acid, gene, organism or the like.

[0032] In certain embodiments the methods and compositions use or include or encode all or part of the Protein A variant or antigen. In other aspects, the Protein A variant may be used in combination with secreted factors or surface antigens including, but not limited to one or more of an isolated Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, or vWh polypeptide or immunogenic segment thereof. Additional staphylococcal antigens that can be used in combination with a Protein A variant include, but are not limited to 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase, autolysin amidase, Cna, collagen

binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/ 0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg2+ transporter, MHC II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/ Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein. In certain embodiments, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 or more of Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, vWh, 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibringen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg2+ transporter, MHC II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/ saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein. can be specifically excluded from a formulation of the invention.

[0033] The following table lists the various combinations of SpA variants and various other Staphyloccal antigens

TABLE 1

							SpA	and	stap	hylo	coc	cal a	ıntig	en c	omb.	inati	ons.							
Eap	+	+	+		+	+	+	+	+	+	-	+	+	+	+	+	+	- 4		+	+	+	+	+
Ebh		+	+		+	+	+	+	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+
Emp			+		+	+	+	+	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+
EsaB					+	+	+	+	+	+	+	+	+	+	+	+	+	- 4	-	+	+	+	+	+
EsaC						+	+	+	+	+	+	+	+	+	+	+	+	- 4	-	+	+	+	+	+
EsxA							+	+	+	+	+	+	+	+	+	+	+	- 4	-	+	+	+	+	+
EsxB								+	+	+	+	+	+	+	+	+	+	- 4	-	+	+	+	+	+
SdrC									+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+
SdrD										+	+	+	+	+	+	+	+	+	-	+	+	+	+	+
SdrE											+	+	+	+	+	+	+	+	-	+	+	+	+	+
IsdA													+	+	+	+	+	- 4	-	+	+	+	+	+
IsdB														+	+	+	+	+	-	+	+	+	+	+
ClfA.															+	+	+	4	-	+	+	+	+	+
ClfB																+	+	+	-	+	+	+	+	+
Coa																	+	- 4	-	+	+	+	+	+
Hla																		+	-	+	+	+	+	+
$\mathrm{Hla}_{H35A}$																				+	+	+	+	+
IsdC																					+	+	+	+
SasF																						+	+	+
vWbp																							+	+
vWh																								+
Ebh		+	+	+	+	+	-		+	+	+	+	+		+	+	+	+	+	+	+	-		+
Emp			+	+	+	+			+	+	+	+	+			+	+	+	+	+	+	. +		+
EsaB				+	+	+			+	+	+	+	+			+	+	+	+	+	+			+
EsaC					+	4			+	+	+	+	+			+	+	+		+				+
EsaC EsxA					+														+		+			
						+			+	+	+	+	+			+	+	+	+	+	+			+
EsxB							+		+	+	+	+	+			+	+	+	+	+	+			+
SdrC									+	+	+	+	+		+	+	+	+	+	+	+	+		+
SdrD										+	+	+	+		+	+	+	+	+	+	+	+		+
SdrE											+	+	+		+	+	+	+	+	+	+	+		+

TABLE 1-continued

	SpA and staphylococcal antigen combinations.
IsdA IsdB ClfA ClfB Coa Hla Hla <sub>H35.4</sub> IsdC SasF vWbp	+ + + + + + + + + + + + + + + + + + +
Emp EsaB EsaC EsxA EsxA EsxB SdrC SdrD SdrE IsdA IsdB Coa Hla Hla <sub>H35,4</sub> IsdC SasF vWbp	
EsaB EsaC EsxA EsxB SdrC SdrD SdrE IsdA IsdB ClfA ClfB Coa Hla Hla <sub>H354</sub> IsdC SasF vWbp	+         +
EsaC EsxA EsxB SdrC SdrD SdrE IsdA IsdB ClfA ClfB Coa Hla Hla <sub>H35A</sub> IsdC SasF vWbp	
EsxA EsxB SdrC SdrD SdrE IsdA	+ + + + + + + + + + + + + + + + + + +

TABLE 1-continued

	SnA	and eta	nhvlo	oncoal	antigen	comb	inetiz	ne						
	SpA :	and sta	ьпАго	coccal										
IsdB ClfA ClfB					+	+	+ + +	++++++	++++++	++++++	+ + +	+ + +	+ + +	+ + +
Coa Hla								+	++	++	+	++	+	++
$Hla_{H35A}$										+	+	+	+	+
IsdC SasF											+	+	+	+
vWbp vWh													+	+
EsxB SdrC	+ +	+		+		+	+	++	++	++	++	++	++	+
SdrD		+	+	+	+	+	+	+	+	+	+	+	+	+
SdrE IsdA			+	+		+	+	+	+	+	+	+	+	+
IsdB OlfA					+	+	+	+	+	+	+	+	+	+
OlfB						+	+	+	+	+	+	+	+	+
Coa Hla								+	+	+	+	+	+	+
Hla <sub>H35A</sub>									+	+	+	+	+	+
sdC SasF											+	++	++	+
vWbp												+	+	+
vWh														+
SdrC SdrD	+	+	++	+	+ +		+	+	+	++	+	+	+	+
SdrE sdA			+	+	+		+	+	+	+	+	+	+	+
sdA sdB				+	+		+ +	+	+	+	+	+	+	+
OlfA OlfB							+	+	+	+	+	+	+	+
Coa							+	+	+	+	+	+	+	+
Ha									+	+	+	+	+	+
Hla <sub>H35A</sub> sdC										+	+	+	+	+
SasF vWbp												+	+	++
vWh														+
SdrD		+ +		+ +					+	++	++	++	++	+
SAWE.														+
sdA		+		+ +					+	+	+	+	+	+
$\operatorname{sdA}$ $\operatorname{sdB}$		+			+ +	+		+ -	+ +	++	+	+	+	+
sdA sdB ClfA ClfB		+		+ +	+	+		+ -	+	+	+	+	+	
sdA sdB ElfA ElfB Coa		+		+ +	+ +	+ + +		+	+ + + + +	+ + + +	+ + + +	+ + + +	+ + + +	+ + +
sdA sdB ElfA ElfB Coa Hla Hla <sub>#354</sub>		+		+ +	+ +	+ + +		+	+ + + +	+ + +	+ + + + + +	+ + + + + +	+ + + + + +	+ + + + +
sdA sdB ElfA ElfB Coa Hla Hla <sub>H354</sub> sdC SasF		+		+ +	+ +	+ + +		+	+ + + + +	+ + + + + +	+ + + + + +	+ + + + +	+ + + + + +	+ + + +
sdA sdB ClfA ClfB Coa Hla Hla <sub>H35.4</sub> sdC sasF		+		+ +	+ +	+ + +		+	+ + + + +	+ + + + + +	+ + + + + +	+ + + + + +	+ + + + + +	+ + + + +
sdA sdB ClifA ClifB Coa Hla Hla <sub>H35.4</sub> sdC Sas F Wbp Wh		+	+	+ +	+	+ + +	+	+	+ + + + + + + + +	+ + + + + +	+ + + + + + + +	+ + + + + + + + +	+ + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +
sdA sdB ElfA ElfB Coa Hla Hla <sub>H354</sub> sdC SasF /Wbp /Wh				+ + +	+ +	+ + +	+ +	+	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +
sdA sdB ClfA ClfB Coa Hla Hla Hla <sub>H35,4</sub> sdC SasF SWbp Wh SdrE sdA sdB			+	+ +	+	+ + + + + + + + + + + + + + + + + + + +	+ + + + +	+	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +
sdA sdB ClfA ClfA ClfB Coa Hla Hla Hla H35.4 sdC SasF WWbp Wh SdrE sdA sdB			+	+ + +	+ + +	+ + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+++++++++++++++++++++++++++++++++++++++	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +
sdA sdB ClfA ClfB Coa Hla Hla <sub>H35A</sub> sdC SasF rWbp rWh SdrE sdA sdB ClfA ClfB ClfB			+	+ + +	+ + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + +	+	+++++++++++++++++++++++++++++++++++++++	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +
isdA isdB ClfA ClfB Coa Hla Hla <sub>H35.4</sub> isdC SasF rWbp Wh SdrE isdA isdB ClfA ClfB Coa Hla Hla <sub>H35.4</sub>			+	+ + +	+ + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+++++++++++++++++++++++++++++++++++++++	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +
SdrE  IsdA  IsdA  IsdB  ClifA  ClifB  Coa  Hlla  Hla <sub>H35.4</sub> IsdC  SasF  vWbp  vWh  SdrE  IsdA  IsdB  ClifA  ClifB  Coa  Hlla  Hla <sub>H35.4</sub> IsdB  ClifA  ClifB  Coa  Hlla  Hla <sub>H35.4</sub> IsdC  SasF			+	+ + +	+ + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+++++++++++++++++++++++++++++++++++++++	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +
sdA sdB ElfA ElfB Coa Hla Hla <sub>H354</sub> sdC SasF Wbp Wh  SdrE sdA sdB ElfA ElfB ElfB Ecoa Hla Hla <sub>H354</sub> sdC SasF			+	+ + +	+ + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+++++++++++++++++++++++++++++++++++++++	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +
sdA sdB ClfA ClfB Coa Hla Hla <sub>H354</sub> sdC SasF Wbp Wh SdrE sdA sdB ClfB Cloa Hla Hla <sub>H354</sub> sdC SasF VWbp VWh SdrE SdA			+	+ + + + + + + + + + + + + + + + + + + +	+ + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+++++++++++++++++++++++++++++++++++++++	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +
sdA sdB ClffA ClffB Cloa Hla Hla <sub>H35.4</sub> sdC sasF Wbp Wh SdrE sdA sdB ClffA ClffB Cloa Hla Hla <sub>H35.4</sub> sdC SasF Wbp Wh SdC SasF			+ +	+ + +	+ + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+++++++++++++++++++++++++++++++++++++++	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +
sdA sdB ClfA ClfB Coa Hla Hla <sub>H35A</sub> sdC SasF Wbp Wh SdrE sdA sdB ClfA ClfB Coa Hla Hla <sub>H35A</sub> sdC SasF VWbp VWh			+ +	+ + + + + + + + + + + + + + + + + + + +	+ + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+++++++++++++++++++++++++++++++++++++++	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +	+ + + + + + + + + + + + + + + + + + + +

TABLE 1-continued

SIB		C!- 4		1				41					
## # # # # # # # # # # # # # # # # # #		SpA an	ıd staphy	1000008	u anti	gen co	mbina	tions.					
asaF Wbp Wbp Wbg Wh	Hla <sub>H35A</sub>								+				
Wbp  Wbh  Wh										+			
### ### ### ### ### ### ### ### ### ##											+		
### ### ### ### ### ### ### ### ### ##	Wh											т	
### # # # # # # # # # # # # # # # # #													•
## # # # # # # # # # # # # # # # # # #				+									
1					+								
Hallans						+							
Harrass							+						
sidC sasF Wbp Wbp  LEEA  + + + + + + + + + + + + + + + + + + +								+					
assF Wbp Wbp Wh Wh Wh Wh Wh Wh Wh  CIGA	lla <sub>H35A</sub>								+				
Wbp Wh Wh Wh										+			
### CIEA											+		
SEEA												+	
### ### ### ### ### ### ### ### ### ##	. МП												
Coa					+								
Ila						+	+	+	+	+	+	+	+
Haggssa side C s							+	+	+	+	+	+	+
sdC wbp wbp wbp wh h  ClfB								+	+	+	+	+	+
sdC wbp wbp wbp wh h  ClfB									+	+	+	+	+
Wbp Wh Wh Wh	sdC									+	+	+	+
## CHB    Coa	SasF										+	+	+
Harrian	Wbp											+	+
Coa	Wh												
Coa	OlfB					+	+	+	+	+	+	+	+
Hala													
Illayassa       +        +       +       +       +       +       +       +       +       +       +       +       +       +       +       +        +       +       +       +       +       +       +       +       +       +       +       +       +       +       +        + <t< td=""><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></t<>													
sdC													
## ## ## ## ## ## ## ## ## ## ## ## ##	sdC								•				
Wbp Wh  Coa										•			
Coa											•		
Ha	Wh											•	
Ha	`03												
Allay354       + + + + + + + + + + + + + + + + + + +							-						
sdC													
asF       + + + + + + + + + + + + + + + + + + +	edC								-				
Wbp       + + + + + + + + + + + + + + + + + + +										-			
## ## ## ## ## ## ## ## ## ## ## ## ##											т		
All and safe and the safe	Wh												
All and safe and the safe	TI.											-	
sdC								+					
sasF       + + + + +         Wbp       + + + + +         Wh       + + + + + +         sdC       + + + + +         asF       + + + +         Wbp       + + + +         sdC       + + + +         sasF       + + + +         Wbp       + + + + + <td>11aH35A</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td>+</td> <td></td> <td></td> <td></td> <td></td>	11aH35A								+				
Wbp       + + + + + + + + + + + + + + + + + + +										+			
### ### ### ### ### ### ### ### ### ##											+		
Ila <sub>H35.4</sub>												+	
sdC       +	AA TI												+
sdC       +	$\mathrm{Ila}_{H35A}$								+	+	+	+	+
Wbp       + + +         wh       + + +         sdC       + + +         asF       + + +         Wbp	sdC									+	+	+	+
Wh         +	SasF										+	+	+
Wh         +	Wbp												
dasF     + + + +       Wbp     + + +       Wh     + + +       dasF     + + +       Wbp     + + +       Wh     + + +       Wbp     + + +       Wbp     + + +       Wbp     + + +       Wh     + + +	Wh												
dasF     + + + +       Wbp     + + +       Wh     + + +       dasF     + + +       Wbp     + + +       Wh     + + +       Wbp     + + +       Wbp     + + +       Wbp     + + +       Wh     + + +	sdC									+	+	+	+
Wbp       +										•			
Wh         +           fasF         +         +         +           Wbp         +         +         +           Wbp         +         +         +           Wh         +         +         +													
SasF       + + + +         Wbp       + + +         Wh       + + +         Wbp       + + +         Wh       - + +												т	
Wbp       +       +         Wbp       +       +         Wh       +       +													•
Wh + Wbp + + Wh +											+	+	+
Wh + Wbp + + Wh +													
Wh +													
Wh +	SasF Wbp Wh											·	+
XVI.	Wbp Wh												
	Wbp Wh Wbp												+

[0034] In still further aspects, the isolated Protein A variant is multimerized, e.g., dimerized or a linear fusion of two or more polypeptides or peptide segments. In certain aspects of the invention, a composition comprises multimers or concatamers of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 or more isolated cell surface proteins or segments thereof. Concatamers are linear polypeptides having one or more repeating peptide units. SpA polypeptides or fragments can be consecutive or separated by a spacer or other peptide sequences, e.g., one or more additional bacterial peptide. In a further aspect, the other polypeptides or peptides contained in the multimer or concatamer can include, but are not limited to 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 of Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, vWh or immunogenic fragments thereof. Additional staphylococcal antigens that can be used in combination with a Protein A variant include, but are not limited to 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibringen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg2+ transporter, MHC II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein.

[0035] The term "Protein A variant" or "SpA variant" refers to polypeptides that include a SpA IgG domain having two or more amino acid substitutions that disrupt binding to Fc and  $V_{H}3$ . In certain aspect, a SpA variant includes a variant domain D peptide, as well as variants of SpA polypeptides and segments thereof that are non-toxigenic and stimulate an immune response against staphylococcus bacteria Protein A and/or bacteria expressing such.

[0036] Embodiments of the present invention include methods for eliciting an immune response against a staphylococcus bacterium or staphylococci in a subject comprising providing to the subject an effective amount of a Protein A variant or a segment thereof. In certain aspects, the methods for eliciting an immune response against a staphylococcus bacterium or staphylococci in a subject comprising providing to the subject an effective amount of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or more secreted proteins and/or cell surface proteins or segments/fragments thereof. A secreted protein or cell surface protein includes, but is not limited to Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, and/or vWh proteins and immunogenic fragments thereof. Additional staphylococcal antigens that can be used in combination with a Protein A variant include, but are not limited to 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg2+ transporter, MHC II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein.

[0037] Embodiments of the invention include compositions that include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to Protein A, or a second protein or peptide that is a secreted bacterial protein or a bacterial cell surface protein. In a further embodiment of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a Protein A domain D polypeptide (SEQ ID NO:2), domain E (SEQ ID NO:3), domain A (SEQ ID NO:4), domain C (SEQ ID NO:5), domain B (SEQ ID NO:6), or a nucleic acid sequence encoding a Protein A domain D, domain E, domain A, domain C, or domain B polypeptide. In certain aspects a Protein A polypeptide segment will have an amino acid sequence of SEQ ID NO:8. Similarity or identity, with identity being preferred, is known in the art and a number of different programs can be used to identify whether a protein (or nucleic acid) has sequence identity or similarity to a known sequence. Sequence identity and/or similarity is determined using standard techniques known in the art, including, but not limited to, the local sequence identity algorithm of Smith & Waterman (1981), by the sequence identity alignment algorithm of Needleman & Wunsch (1970), by the search for similarity method of Pearson & Lipman (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Drive, Madison, Wis.), the Best Fit sequence program described by Devereux et al. (1984), preferably using the default settings, or by inspection. Preferably, percent identity is calculated by using alignment tools known to and readily ascertainable to those of skill in the art. Percent identity is essentially the number of identical amino acids divided by the total number of amino acids compared times one hundred.

[0038] Still further embodiments include methods for stimulating in a subject a protective or therapeutic immune response against a *staphylococcus* bacterium comprising administering to the subject an effective amount of a composition including (i) a SpA variant, e.g., a variant SpA domain D polypeptide or peptide thereof; or, (ii) a nucleic acid molecule encoding such a SpA variant polypeptide or peptide thereof, or (iii) administering a SpA variant domain D polypeptide with any combination or permutation of bacterial proteins described herein. In a preferred embodiment the composition is not a *staphylococcus* bacterium. In certain aspects the subject is a human or a cow. In a further aspect the composition is formulated in a pharmaceutically acceptable formulation. The staphylococci may be *Staphylococcus aureus*.

[0039] Yet still further embodiments include vaccines comprising a pharmaceutically acceptable composition having an isolated SpA variant polypeptide, or any other combination or permutation of protein(s) or peptide(s) described herein, wherein the composition is capable of stimulating an immune

response against a *staphylococcus* bacterium. The vaccine may comprise an isolated SpA variant polypeptide, or any other combination or permutation of protein(s) or peptide(s) described. In certain aspects of the invention the isolated SpA variant polypeptide, or any other combination or permutation of protein(s) or peptide(s) described are multimerized, e.g., dimerized or concatamerized. In a further aspect, the vaccine composition is contaminated by less than about 10, 9, 8, 7, 6, 5, 4, 3, 2, 1, 0.5, 0.25, 0.05% (or any range derivable therein) of other Staphylococcal proteins. A composition may further comprise an isolated non-SpA polypeptide. Typically the vaccine comprises an adjuvant. In certain aspects a protein or peptide of the invention is linked (covalently or non-covalently) to the adjuvant, preferably the adjuvant is chemically conjugated to the protein.

[0040] In still yet further embodiments, a vaccine composition is a pharmaceutically acceptable composition having a recombinant nucleic acid encoding all or part of a SpA variant polypeptide, or any other combination or permutation of protein(s) or peptide(s) described herein, wherein the composition is capable of stimulating an immune response against a staphylococcus bacteria. The vaccine composition may comprise a recombinant nucleic acid encoding all or part of a SpA variant polypeptide, or any other combination or permutation of protein(s) or peptide(s) described herein. In certain embodiments the recombinant nucleic acid contains a heterologous promoter. Preferably the recombinant nucleic acid is a vector. More preferably the vector is a plasmid or a viral vector. In some aspects the vaccine includes a recombinant, non-staphylococcus bacterium containing the nucleic acid. The recombinant non-staphylococci may be Salmonella or another gram-positive bacteria. The vaccine may comprise a pharmaceutically acceptable excipient, more preferably an adjuvant.

[0041] Still further embodiments include methods for stimulating in a subject a protective or therapeutic immune response against a staphylococcus bacterium comprising administering to the subject an effective amount of a composition of a SpA variant polypeptide or segment/fragment thereof and further comprising one or more of a Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, or vWh protein or peptide thereof. In a preferred embodiment the composition comprises a non-staphylococcus bacterium. In a further aspect the composition is formulated in a pharmaceutically acceptable formulation. The staphylococci for which a subject is being treated may be Staphylococcus aureus. Methods of the invention also include SpA variant compositions that contain 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or more secreted virulence factors and/or cell surface proteins, such as Eap, Ebh, Emp, EsaC,  ${\sf EsxA}, {\sf EsxB}, {\sf SdrC}, \bar{\sf SdrD}, {\sf SdrE}, {\sf IsdA}, {\sf IsdB}, {\sf ClfA}, {\sf ClfB}, {\sf Coa},$ Hla, IsdC, SasF, vWbp, or vWh in various combinations. In certain aspects a vaccine formulation includes Eap, Ebh, Emp, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, and vWh. In certain aspects an antigen combination can include (1) a SpA variant and IsdA; (2) SpA variant and ClfB; (3) SpA variant and SdrD; (4) SpA variant and Hla or Hla variant; (5) SpA variant and ClfB, SdrD, and Hla or Hla variant; (6) SpA variant, IsdA, SdrD, and Hla or Hla variant; (7) SpA variant, IsdA, ClfB, and Hla or Hla variant; (8) SpA variant, IsdA, ClfB, and SdrD; (9) SpA variant, IsdA, ClfB, SdrD and Hla or Hla variant; (10) SpA variant, IsdA, ClfB, and SdrD; (11) SpA variant, IsdA,

SdrD, and Hla or Hla variant; (12) SpA variant, IsdA, and Hla or Hla variant; (13) SpA variant, IsdA, ClfB, and Hla or Hla variant; (14) SpA variant, ClfB, and SdrD; (15) SpA variant, ClfB, and Hla or Hla variant; or (16) SpA variant, SdrD, and Hla or Hla variant.

[0042] In certain aspects, a bacterium delivering a composition of the invention will be limited or attenuated with respect to prolonged or persistent growth or abscess formation. In yet a further aspect, SpA variant(s) can be overexpressed in an attenuated bacterium to further enhance or supplement an immune response or vaccine formulation.

[0043] The term "EsxA protein" refers to a protein that includes isolated wild-type EsxA polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria EsxA proteins.

[0044] The term "EsxB protein" refers to a protein that includes isolated wild-type EsxB polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria EsxB proteins.

[0045] The term "SdrD protein" refers to a protein that includes isolated wild-type SdrD polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria SdrD proteins.

[0046] The term "SdrE protein" refers to a protein that includes isolated wild-type SdrE polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria SdrE proteins.

[0047] The term "IsdA protein" refers to a protein that includes isolated wild-type IsdA polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria IsdA proteins.

[0048] The term "IsdB protein" refers to a protein that includes isolated wild-type IsdB polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria IsdB proteins.

[0049] The term "Eap protein" refers to a protein that includes isolated wild-type Eap polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria Eap proteins.

[0050] The term "Ebh protein" refers to a protein that includes isolated wild-type Ebh polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria Ebh proteins.

[0051] The term "Emp protein" refers to a protein that includes isolated wild-type Emp polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria Emp proteins.

[0052] The term "EsaB protein" refers to a protein that includes isolated wild-type EsaB polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria EsaB proteins.

[0053] The term "EsaC protein" refers to a protein that includes isolated wild-type EsaC polypeptides from *staphy*-

*lococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria EsaC proteins.

[0054] The term "SdrC protein" refers to a protein that includes isolated wild-type SdrC polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria SdrC proteins.

[0055] The term "ClfA protein" refers to a protein that includes isolated wild-type ClfA polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria ClfA proteins.

[0056] The term "ClfB protein" refers to a protein that includes isolated wild-type ClfB polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria ClfB proteins.

[0057] The term "Coa protein" refers to a protein that includes isolated wild-type Coa polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria Coa proteins.

[0058] The term "Hla protein" refers to a protein that includes isolated wild-type Hla polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria Hla proteins.

[0059] The term "IsdC protein" refers to a protein that includes isolated wild-type IsdC polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria IsdC proteins.

[0060] The term "SasF protein" refers to a protein that includes isolated wild-type SasF polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria SasF proteins.

[0061] The term "vWbp protein" refers to a protein that includes isolated wild-type vWbp (von Willebrand factor binding protein) polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria vWbp proteins.

[0062] The term "vWh protein" refers to a protein that includes isolated wild-type vWh (von Willebrand factor binding protein homolog) polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria vWh proteins.

[0063] An immune response refers to a humoral response, a cellular response, or both a humoral and cellular response in an organism. An immune response can be measured by assays that include, but are not limited to, assays measuring the presence or amount of antibodies that specifically recognize a protein or cell surface protein, assays measuring T-cell activation or proliferation, and/or assays that measure modulation in terms of activity or expression of one or more cytoking.

[0064] In still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an EsxA protein. In

certain aspects the EsxA protein will have all or part of the amino acid sequence of SEQ ID NO:11.

[0065] In still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an EsxB protein. In certain aspects the EsxB protein will have all or part of the amino acid sequence of SEQ ID NO:12.

[0066] In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an SdrD protein. In certain aspects the SdrD protein will have all or part of the amino acid sequence of SEQ ID NO:13.

[0067] In further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an SdrE protein. In certain aspects the SdrE protein will have all or part of the amino acid sequence of SEQ ID NO:14.

[0068] In still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an IsdA protein. In certain aspects the IsdA protein will have all or part of the amino acid sequence of SEQ ID NO:15.

[0069] In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an IsdB protein. In certain aspects the IsdB protein will have all or part of the amino acid sequence of SEQ ID NO:16.

[0070] Embodiments of the invention include compositions that include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a EsaB protein. In certain aspects the EsaB protein will have all or part of the amino acid sequence of SEQ ID NO:17.

[0071] In a further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a ClfB protein. In certain aspects the ClfB protein will have all or part of the amino acid sequence of SEQ ID NO:18.

[0072] In still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an IsdC protein. In certain aspects the IsdC protein will have all or part of the amino acid sequence of SEQ ID NO:19.

[0073] In yet further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a SasF protein. In certain aspects the SasF protein will have all or part of the amino acid sequence of SEQ ID NO:20.

[0074] In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a SdrC protein. In certain aspects the SdrC protein will have all or part of the amino acid sequence of SEQ ID NO:21.

[0075] In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein

that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a ClfA protein. In certain aspects the ClfA protein will have all or part of the amino acid sequence of SEQ ID NO:22.

[0076] In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an Eap protein. In certain aspects the Eap protein will have all or part of the amino acid sequence of SEQ ID NO:23.

[0077] In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an Ebh protein. In certain aspects the Ebh protein will have all or part of the amino acid sequence of SEQ ID NO:24.

[0078] In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an Emp protein. In certain aspects the Emp protein will have all or part of the amino acid sequence of SEQ ID NO:25.

[0079] In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an EsaC protein. In certain aspects the EsaC protein will have all or part of the amino acid sequence of SEQ ID NO:26. Sequence of EsaC polypeptides can be found in the protein databases and include, but are not limited to accession numbers ZP\_02760162 (GI:168727885), NP\_645081.1 (GI: 21281993), and NP\_370813.1 (GI:15923279), each of which is incorporated herein by reference as of the priority date of this application.

[0080] In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a Coa protein. In certain aspects the Coa protein will have all or part of the amino acid sequence of SEQ ID NO:27.

[0081] In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a Hla protein. In certain aspects the Hla protein will have all or part of the amino acid sequence of SEQ ID NO:28.

[0082] In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a vWa protein. In certain aspects the vWa protein will have all or part of the amino acid sequence of SEQ ID NO:29.

[0083] In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a vWbp protein. In certain aspects the vWbp protein will have all or part of the amino acid sequence of SEQ ID NO:32.

[0084] In certain aspects, a polypeptide or segment/fragment can have a sequence that is at least 85%, at least 90%, at least 95%, at least 98%, or at least 99% or more identical to the amino acid sequence of the reference polypeptide. The term "similarity" refers to a polypeptide that has a sequence that has a certain percentage of amino acids that are either

identical with the reference polypeptide or constitute conservative substitutions with the reference polypeptides.

[0085] The polypeptides described herein may include 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, or more variant amino acids within at least, or at most 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 101, 102, 103, 104, 105, 106, 107, 108, 109, 110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 125, 126, 127, 128, 129, 130, 131, 132, 133, 134, 135, 136, 137, 138, 139, 140, 141, 142, 143, 144, 145, 146, 147, 148, 149, 150, 151, 152, 153, 154, 155, 156, 157, 158, 159, 160, 161, 162, 163, 164, 165, 166, 167, 168, 169, 170, 171, 172, 173, 174, 175, 176, 177, 178, 179, 180, 181, 182, 183, 184, 185, 186, 187, 188, 189, 190, 191, 192, 193, 194, 195, 196, 197, 198, 199, 200, 201, 202, 203, 204, 205, 206, 207, 208, 209, 210, 211, 212, 213, 214, 215, 216, 217, 218,  $219,\,220,\,221,\,222,\,223,\,224,\,225,\,226,\,227,\,228,\,229,\,230,$ 231, 232, 233, 234, 235, 236, 237, 238, 239, 240, 241, 242, 243, 244, 245, 246, 247, 248, 249, 250, 300, 400, 500, 550, 1000 or more contiguous amino acids, or any range derivable therein, of SEQ ID NO:2-30, or SEQ ID NO:32-34.

[0086] A polypeptide segment as described herein may include 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 101, 102, 103, 104, 105, 106, 107, 108, 109, 110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 125, 126, 127, 128, 129, 130, 131, 132, 133, 134, 135, 136, 137, 138, 139, 140, 141, 142, 143, 144, 145, 146, 147, 148, 149, 150, 151, 152, 153, 154, 155, 156, 157, 158, 159, 160, 161, 162, 163, 164, 165, 166, 167, 168, 169, 170, 171, 172, 173, 174, 175, 176, 177, 178, 179, 180, 181, 182, 183, 184, 185, 186, 187, 188, 189, 190, 191, 192, 193, 194, 195, 196, 197, 198, 199, 200, 201, 202, 203, 204, 205, 206, 207, 208, 209, 210, 211, 212, 213, 214, 215, 216, 217, 218, 219, 220, 221, 222, 223, 224, 225, 226, 227, 228, 229, 230, 231, 232, 233, 234, 235, 236, 237, 238, 239, 240, 241, 242, 243, 244, 245, 246, 247, 248, 249, 250, 300, 400, 500, 550, 1000 or more contiguous amino acids, or any range derivable therein, of SEQ ID NO:2-30, or SEQ ID NO:33-34.

[0087] The compositions may be formulated in a pharmaceutically acceptable composition. In certain aspects of the invention the *staphylococcus* bacterium is an *S. aureus* bacterium.

[0088] In further aspects, a composition may be administered more than one time to the subject, and may be administered 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20 or more times. The administration of the compositions include, but is not limited to oral, parenteral, subcutaneous, intramuscular, intravenous, or various combinations thereof, including inhalation or aspiration.

[0089] In still further embodiments, a composition comprises a recombinant nucleic acid molecule encoding a polypeptide described herein or segments/fragments thereof. Typically a recombinant nucleic acid molecule encoding a polypeptide described herein contains a heterologous promoter. In certain aspects, a recombinant nucleic acid molecule

ecule of the invention is a vector, in still other aspects the vector is a plasmid. In certain embodiments the vector is a viral vector. In certain aspects a composition includes a recombinant, non-staphylococcus bacterium containing or expressing a polypeptide described herein. In particular aspects the recombinant non-staphylococcus bacteria is Salmonella or another gram-positive bacteria. A composition is typically administered to mammals, such as human subjects, but administration to other animals that are capable of eliciting an immune response is contemplated. In further aspects the staphylococcus bacterium containing or expressing the polypeptide is Staphylococcus aureus. In further embodiments the immune response is a protective immune response.

ments the immune response is a protective immune response. [0090] In further embodiments a composition comprises a recombinant nucleic acid molecule encoding all or part of one or more of a Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, SpA, vWbp, or vWh protein or peptide or variant thereof. Additional staphylococcal antigens that can be used in combination with the polypeptides described herein include, but are not limited to 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg2+ transporter, MHC II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/ saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein. In particular aspects, a bacteria is a recombinant non-staphylococcus bacteria, such as a Salmonella or other gram-positive bacteria.

[0091] Compositions of the invention are typically administered to human subjects, but administration to other animals that are capable of eliciting an immune response to a *staphylococcus* bacterium is contemplated, particularly cattle, horses, goats, sheep and other domestic animals, i.e., mammals

[0092] In certain aspects the *staphylococcus* bacterium is a *Staphylococcus aureus*. In further embodiments the immune response is a protective immune response. In still further aspects, the methods and compositions of the invention can be used to prevent, ameliorate, reduce, or treat infection of tissues or glands, e.g., mammary glands, particularly mastitis and other infections. Other methods include, but are not limited to prophylactically reducing bacterial burden in a subject not exhibiting signs of infection, particularly those subjects suspected of or at risk of being colonized by a target bacteria, e.g., patients that are or will be at risk or susceptible to infection during a hospital stay, treatment, and/or recovery.

[0093] Any embodiment discussed with respect to one aspect of the invention applies to other aspects of the invention as well. In particular, any embodiment discussed in the context of a SpA variant polypeptide or peptide or nucleic acid may be implemented with respect to other antigens, such as Eap, Ebh, Emp, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, vWh, 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap,

Ant, autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg2+ transporter, MHC II analogue (U.S. Pat. No. 5,648, 240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein (or nucleic acids), and vice versa. It is also understood that any one or more of Eap, Ebh, Emp, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, vWh, 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibringen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg2+ transporter, MHC II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein can be specifically excluded from a claimed composition.

[0094] Embodiments of the invention include compositions that contain or do not contain a bacterium. A composition may or may not include an attenuated or viable or intact staphylococcal bacterium. In certain aspects, the composition comprises a bacterium that is not a staphylococcal bacterium or does not contain staphylococcal bacteria. In certain embodiments a bacterial composition comprises an isolated or recombinantly expressed staphylococcal Protein A variant or a nucleotide encoding the same. The composition may be or include a recombinantly engineered staphylococcus bacterium that has been altered in a way that comprises specifically altering the bacterium with respect to a secreted virulence factor or cell surface protein. For example, the bacteria may be recombinantly modified to express more of the virulence factor or cell surface protein than it would express if unmodified.

[0095] The term "isolated" can refer to a nucleic acid or polypeptide that is substantially free of cellular material, bacterial material, viral material, or culture medium (when produced by recombinant DNA techniques) of their source of origin, or chemical precursors or other chemicals (when chemically synthesized). Moreover, an isolated compound refers to one that can be administered to a subject as an isolated compound; in other words, the compound may not simply be considered "isolated" if it is adhered to a column or embedded in an agarose gel. Moreover, an "isolated nucleic acid fragment" or "isolated peptide" is a nucleic acid or protein fragment that is not naturally occurring as a fragment and/or is not typically in the functional state.

[0096] Moieties of the invention, such as polypeptides, peptides, antigens, or immunogens, may be conjugated or linked covalently or noncovalently to other moieties such as adjuvants, proteins, peptides, supports, fluorescence moieties, or labels. The term "conjugate" or "immunoconjugate" is broadly used to define the operative association of one moiety with another agent and is not intended to refer solely to any type of operative association, and is particularly not limited to chemical "conjugation." Recombinant fusion proteins are particularly contemplated. Compositions of the invention may further comprise an adjuvant or a pharmaceutically acceptable excipient. An adjuvant may be covalently or non-covalently coupled to a polypeptide or peptide of the invention. In certain aspects, the adjuvant is chemically conjugated to a protein, polypeptide, or peptide.

[0097] The term "providing" is used according to its ordinary meaning to indicate "to supply or furnish for use." In some embodiments, the protein is provided directly by administering the protein, while in other embodiments, the protein is effectively provided by administering a nucleic acid that encodes the protein. In certain aspects the invention contemplates compositions comprising various combinations of nucleic acid, antigens, peptides, and/or epitopes.

[0098] The subject will have (e.g., are diagnosed with a staphylococcal infection), will be suspected of having, or will be at risk of developing a staphylococcal infection. Compositions of the present invention include immunogenic compositions wherein the antigen(s) or epitope(s) are contained in an amount effective to achieve the intended purpose. More specifically, an effective amount means an amount of active ingredients necessary to stimulate or elicit an immune response, or provide resistance to, amelioration of, or mitigation of infection. In more specific aspects, an effective amount prevents, alleviates or ameliorates symptoms of disease or infection, or prolongs the survival of the subject being treated. Determination of the effective amount is well within the capability of those skilled in the art, especially in light of the detailed disclosure provided herein. For any preparation used in the methods of the invention, an effective amount or dose can be estimated initially from in vitro studies, cell culture, and/or animal model assays. For example, a dose can be formulated in animal models to achieve a desired immune response or circulating antibody concentration or titer. Such information can be used to more accurately determine useful doses in humans.

[0099] The embodiments in the Example section are understood to be embodiments of the invention that are applicable to all aspects of the invention.

[0100] The use of the term "or" in the claims is used to mean "and/or" unless explicitly indicated to refer to alternatives only or the alternatives are mutually exclusive, although the disclosure supports a definition that refers to only alternatives and "and/or." It is also contemplated that anything listed using the term "or" may also be specifically excluded.

[0101] Throughout this application, the term "about" is used to indicate that a value includes the standard deviation of error for the device or method being employed to determine the value.

[0102] Following long-standing patent law, the words "a" and "an," when used in conjunction with the word "comprising" in the claims or specification, denotes one or more, unless specifically noted.

[0103] Other objects, features and advantages of the present invention will become apparent from the following

detailed description. It should be understood, however, that the detailed description and the specific examples, while indicating specific embodiments of the invention, are given by way of illustration only, since various changes and modifications within the spirit and scope of the invention will become apparent to those skilled in the art from this detailed description.

#### DESCRIPTION OF THE DRAWINGS

[0104] So that the matter in which the above-recited features, advantages and objects of the invention as well as others which will become clear are attained and can be understood in detail, more particular descriptions and certain embodiments of the invention briefly summarized above are illustrated in the appended drawings. These drawings form a part of the specification. It is to be noted, however, that the appended drawings illustrate certain embodiments of the invention and therefore are not to be considered limiting in their scope.

[0105] FIGS. 1A-1B. (FIG. 1A) Primary structure of the Protein A precursor with an N-terminal YSIRK motif signal peptide, five immunoglobulin binding domains as tandem repeats designated E, D, A, B, C, region X, and the LPXTG sorting signal. (FIG. 1B) Following synthesis of the Protein A precursor, staphylococci secrete this product via the Sec pathway, and sortase A cleaves the LPXTG sorting signal between the T and G residues. Nucleophilic attack of the amino group within lipid II at the sortase-Protein A thioester-linked intermediate forms the amide bond that links Protein A to the cell wall envelope and enables its display on the bacterial surface. [0106] FIG. 2. Three dimensional model of the molecular interactions between the SpA-domain D of Protein A, the VH3 Fab domain of the B cell receptor, and of the Fcy domain of immunoglobulin. The model is derived from two crystal structures (Graille et al., 2000 and Gouda et al., 1992) that revealed side chain residues involved in the formation of ionic bonds that enable these complexes. Gln-9 and Gln-10 of SpA-D promote binding to Fcy, whereas Asp-36 and Asp-37 enable complex formation with VH3 Fab.

[0107] FIG. 3. Left panel—Coomassie Blue stained SDS-PAGE reveals the migrational position of purified His-tagged SpA, SpA-D, SpA-D $_{Q9,10K;D36,374}$ , human IgG, and sortase A (SrtA), a control protein. Right panel—Coomassie Blue stained SDS-PAGE to reveal the elution of Protein A immunoglobulin complexes eluted following affinity chromatography of human IgG on Ni-NTA columns pre-charged with His-tagged SpA, SpA-D, SpA-D $_{Q9,10K;D36,374}$  or SrtA.

[0108] FIG. 4. ELISA assays to quantify human immunoglobulin (hIgG), human F(ab)<sub>2</sub> IgG fragments and human Fc fragments of immunoglobulin (hFc). Plates were coated with equal amounts of His-tagged SpA, SpA-D, SpA-D<sub>Q9,10K;D36,37.4</sub> or SrtA. hIgG-HRP, F(ab)<sub>2</sub>-HRP and hFc-HRP were added onto the plates and incubated for an hour. Absorbance at 450 nm was recorded and plotted to determine the half maximal titers.

[0109] FIG. 5. Purified SpA-D, SpA-D $_{Q9,10K;D36,37A}$  or a PBS mock control were injected into the peritoneum of mice and analyzed for their ability to reduce the B cell population in the spleen of experimental BALB/c mice. Animals were killed 4 hours following injection, their spleen removed, tissue homogenized and stained with CD19 antibodies directed against B cells. The number of B cells was quantified by FACS sorting.

[0110] FIG. 6 Generation of a non-toxigenic protein A vaccine. a, Translational protein A (SpA) product of *S. aureus* 

Newman and USA300 LAC with an N-terminal signal peptide (white box), five immunoglobulin binding domains (Ig-BDs designated E, D, A, B and C), variable region X and C-terminal sorting signal (black box). b, Amino acid sequence of the five IgBDs as well as nontoxigenic SpA- $D_{KK44}$ , with the positions of triple  $\alpha$ -helical bundles (H1, H2 and H3) as well as glutamine (Q) 9, 10 and aspartate (D) 36, 37 indicated. c, Coomassie Blue-stained SDS-PAGE of SpA, SpA-D, SpA-D<sub>KKAA</sub> or SrtA purified on Ni-NTA sepharose in the presence or absence of human immunoglobulin (hIgG). d, ELISA examining the association of immobilized SpA, SpA-D or SpA-D $_{KKAA}$  with human IgG as well as its Fc or F(ab)<sub>2</sub> fragments and von Willebrand factor (vWF). e, CD19+B lymphocytes in splenic tissue of BALB/c mice that had been mock immunized or treated with SpA-D or SpA- $D_{KKAA}$  were quantified by FACS.

[0111] FIG. 7 Non-toxigenic protein A vaccine prevents abscess formation. Histopathology of renal tissue isolated during necropsy of BALB/c mice that had been mock immunized (PBS) or vaccinated with SpA, SpA-D as well as SpA-D<sub>KKAA</sub> and challenged with *S. aureus* Newman. Thin sectioned tissues were stained with hematoxylin-eosin. White arrows identify polymorphonuclear leukocyte (PMN) infiltrates. Dark arrows identify staphylococcal abscess communities

[0112] FIG. 8 Antibodies raised by the non-toxigenic protein A vaccine block the B cell superantigen function of SpA. a, Rabbit antibodies raised against SpA- $D_{KKAA}$  were purified on a matrix with immobilized antigen and analyzed by Coomassie Blue-stained SDS-PAGE. Antibodies were cleaved with pepsin and F(ab)2 fragments were purified by a second round of affinity chromatography on SpA- $D_{KKAA}$  matrix. b, SpA- $D_{KKAA}$  specific F(ab)2 interfere with the binding of SpA or SpA-D to human immunoglobulin (hIgG) or, c, to von Willebrand Factor (vWF).

[0113] FIG. 9 Full-length non-toxigenic protein A generates improved immune responses. a, Full-length SpAKKAA was purified on Ni-NTA sepharose and analyzed by Coomassie-Blue stained SDS-PAGE. b, CD19+B lymphocytes in splenic tissue of BALB/c mice that had been mock immunized or treated with SpA or  $SpA_{KKAA}$  were quantified by FACS. c, ELISA examining the association of immobilized SpA or SpA $_{KKAA}$  with human IgG as well as its Fc or F(ab)2 fragments or von Willebrand factor (vWF). d, Human or mouse serum antibody titers to diphtheria toxoid (CRM197) and non-toxigenic  $\mathrm{SpA}_{\mathit{KKAA}}$  or  $\mathrm{SpA-D}_{\mathit{KKAA}}.$  Human volunteers with a history of DTaP immunization and staphylococcal infection (n=16) as well as mice (n=20) that had been infected with S. aureus Newman or USA 300 LAC or immunized with  $SpA_{KKAA}$  or  $SpA-D_{KKAA}$  were examined by quantitative dot blot.

[0114] FIG. 10 Staphylococcal infection does not generate protective immunity. BALB/c mice (n=20) were infected with *S. aureus* Newman or mock challenged (PBS) for thirty days and infection cleared with chloramphenicol treatment. Both cohorts of animals were then challenged with *S. aureus* Newman and bacterial load (CFU) in kidney tissue homogenate analyzed following necropsy on day 4.

[0115] FIG. 11 Comparison of abscess formation in mice treated with PBS, SpA, SpA-D, and SpA-D<sub>KKAA</sub>.

[0116] FIGS. 12A-12C (A) ELISA examining the association of immobilized SpA, SpA-D, SpA-DKKAA or SpA-DGGSS with human IgG as well as its Fc or F(ab)2 fragments and IgM. Statistical significance of SpA-DKKAA and SpA-

DGGSS binding to each ligand was compared against SpA-D; SpA-D binding was compared against SpA (n=3); \* signifies P<0.05; \*\* signifies P<0.01. (B) ELISA examining the level of cross-reactive antibodies of hyper-immune sera samples collected from actively immunized mice (n=5) with SpA-D, SpA-DKKAA and SpA-DGGSS. (C) Abscess formation in mice treated with PBS, SpA-D, SpA-D $_{KKAA}$  and SpA-D $_{GGsS}$ .

#### DETAILED DESCRIPTION

[0117] Staphylococcus aureus is a commensal of the human skin and nares, and the leading cause of bloodstream, skin and soft tissue infections (Klevens et al., 2007). Recent dramatic increases in the mortality of staphylococcal diseases are attributed to the spread of methicillin-resistant *S. aureus* (MRSA) strains often not susceptible to antibiotics (Kennedy et al., 2008). In a large retrospective study, the incidence of MRSA infections was 4.6% of all hospital admissions in the United States (Klevens et al., 2007). The annual health care costs for 94,300 MRSA infected individuals in the United States exceed \$2.4 billion (Klevens et al., 2007). The current MRSA epidemic has precipitated a public health crisis that needs to be addressed by development of a preventive vaccine (Boucher and Corey, 2008). To date, an FDA licensed vaccine that prevents *S. aureus* diseases is not available.

[0118] The inventors describe here the use of Protein A, a cell wall anchored surface protein of staphylococci, for the generation of variants that can serve as subunit vaccines. The pathogenesis of staphylococcal infections is initiated as bacteria invade the skin or blood stream via trauma, surgical wounds, or medical devices (Lowy, 1998). Although the invading pathogen may be phagocytosed and killed, staphylococci can also escape innate immune defenses and seed infections in organ tissues, inducing inflammatory responses that attract macrophages, neutrophils, and other phagocytes (Lowy, 1998). The responsive invasion of immune cells to the site of infection is accompanied by liquefaction necrosis as the host seeks to prevent staphylococcal spread and allow for removal of necrotic tissue debris (Lam et al., 1963). Such lesions can be observed by microscopy as hypercellular areas containing necrotic tissue, leukocytes, and a central nidus of bacteria (Lam et al., 1963). Unless staphylococcal abscesses are surgically drained and treated with antibiotics, disseminated infection and septicemia produce a lethal outcome (Sheagren, 1984).

#### II. Staphylococcal Antigens

[0119] A. Staphylcoccal Protein A (SpA)

[0120] All Staphylococcus aureus strains express the structural gene for Protein A (spa) (Jensen, 1958; Said-Salim et al., 2003), a well characterized virulence factor whose cell wall anchored surface protein product (SpA) encompasses five highly homologous immunoglobulin binding domains designated E, D, A, B, and C (Sjodahl, 1977). These domains display ~80% identity at the amino acid level, are 56 to 61 residues in length, and are organized as tandem repeats (Uhlen et al., 1984). SpA is synthesized as a precursor protein with an N-terminal YSIRK/GS signal peptide and a C-terminal LPXTG motif sorting signal (DeDent et al., 2008; Schneewind et al., 1992). Cell wall anchored Protein A is displayed in great abundance on the staphylococcal surface (DeDent et al., 2007; Sjoquist et al., 1972). Each of its immunoglobulin binding domains is composed of anti-parallel

 $\alpha$ -helices that assemble into a three helix bundle and bind the Fc domain of immunoglobulin G (IgG) (Deisenhofer, 1981; Deisenhofer et al., 1978), the VH3 heavy chain (Fab) of IgM (i.e., the B cell receptor) (Graille et al., 2000), the von Willebrand factor at its A1 domain [vWF AI is a ligand for platelets] (O'Seaghdha et al., 2006) and the tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ) receptor I (TNFRI) (Gomez et al., 2006), which is displayed on surfaces of airway epithelia (Gomez et al., 2004; Gomez et al., 2007).

[0121] SpA impedes neutrophil phagocytosis of staphylococci through its attribute of binding the Fc component of IgG (Jensen, 1958; Uhlen et al., 1984). Moreover, SpA is able to activate intravascular clotting via its binding to von Willebrand factor AI domains (Hartleib et al., 2000). Plasma proteins such as fibrinogen and fibronectin act as bridges between staphylococci (CIfA and CIfB) and the platelet integrin GPIIb/IIIa (O'Brien et al., 2002), an activity that is supplemented through Protein A association with vWF AI, which allows staphylococci to capture platelets via the GPIb-α platelet receptor (Foster, 2005; O'Seaghdha et al., 2006). SpA also binds TNFRI and this interaction contributes to the pathogenesis of staphylococcal pneumonia (Gomez et al., 2004). SpA activates proinflammatory signaling through TNFR1 mediated activation of TRAF2, the p38/c-Jun kinase, mitogen activate protein kinase (MAPK) and the Rel-transcription factor NF-KB. SpA binding further induces TNFR1 shedding, an activity that appears to require the TNF-converting enzyme (TACE)(Gomez et al., 2007). All of the aforementioned SpA activities are mediated through its five IgG binding domains and can be perturbed by the same amino acid substitutions, initially defined by their requirement for the interaction between Protein A and human IgG1 (Cedergren et al., 1993.

[0122] SpA also functions as a B cell superantigen by capturing the Fab region of VH3 bearing IgM, the B cell receptor (Gomez et al., 2007; Goodyear et al., 2003; Goodyear and Silverman, 2004; Roben et al., 1995). Following intravenous challenge, staphylococcal Protein A (SpA) mutations show a reduction in staphylococcal load in organ tissues and dramatically diminished ability to form abscesses (described herein). During infection with wildtype S. aureus, abscesses are formed within forty-eight hours and are detectable by light microscopy of hematoxylin-eosin stained, thin-sectioned kidney tissue, initially marked by an influx of polymorphonuclear leukocytes (PMNs). On day 5 of infection, abscesses increase in size and enclosed a central population of staphylococci, surrounded by a layer of eosinophilic, amorphous material and a large cuff of PMNs. Histopathology revealed massive necrosis of PMNs in proximity to the staphylococcal nidus at the center of abscess lesions as well as a mantle of healthy phagocytes. The inventors also observed a rim of necrotic PMNs at the periphery of abscess lesions, bordering the eosinophilic pseudocapsule that separated healthy renal tissue from the infectious lesion. Staphylococcal variants lacking Protein A are unable to establish the histopathology features of abscesses and are cleared during infection.

[0123] In previous studies, Cedergren et al. (1993) engineered five individual substitutions in the Fc frgament binding sub-domain of the B domain of SpA, L17D, N28A, I31A and K35A. These authors created these proteins to test data gathered from a three dimensional structure of a complex between one domain of SpA and Fc<sub>1</sub>. Cedergren et al. determined the effects of these mutations on stability and binding,

but did not contemplate use of such substitutions for the production of a vaccine antigen.

[0124] Brown et al. (1998) describe studies designed to engineer new proteins based on SpA that allow the use of more favorable elution conditions when used as affinity ligands. The mutations studied included single mutations of Q13A, Q14H, N15A, N15H, F17H, Y18F, L21H, N32H, or K39H. Brown et al. report that Q13A, N15A, N15H, and N32H substitutions made little difference to the dissociation constant values and that the Y18F substitution resulted in a 2 fold decrease in binding affinity as compared to wild type SpA. Brown et al. also report that L21H and F17H substitutions decrease the binding affinity by five-fold and a hundredfold respectively. The authors also studied analogous substitutions in two tandem domains. Thus, the Brown et al. studies were directed to generating a SpA with a more favorable elution profile, hence the use of His substitutions to provide a pH sensitive alteration in the binding affinity. Brown et al. is silent on the use of SpA as a vaccine antigen.

[0125] Graille et al. (2000) describe a crystal structure of domain D of SpA and the Fab fragment of a human IgM antibody. Graille et al. define by analysis of a crystal structure the D domain amino acid residues that interact with the Fab fragment as residues Q26, G29, F30, Q32, S33, D36, D37, Q40, N43, E47, or L51, as well as the amino acid residues that form the interface between the domain D sub-domains. Graille et al. define the molecular interactions of these two proteins, but is silent in regard to any use of substitutions in the interacting residues in producing a vaccine antigen.

[0126] O'Seaghdha et al. (2006) describe studies directed at elucidating which sub-domain of domain D binds vWF. The authors generated single mutations in either the Fc or VH3 binding sub-domains, i.e., amino acid residues F5A, Q9A, Q10A, F13A, Y14A, L17A, N28A, I31A, K35A, G29A, F30A, S33A, D36A, D37A, Q40A, E47A, or Q32A. The authors discovered that vWF binds the same sub-domain that binds Fc. O'Seaghda et al. define the sub-domain of domain D responsible for binding vWF, but is silent in regard to any use of substitutions in the interacting residues in producing a vaccine antigen.

[0127] Gomez et al. (2006) describe the identification of residues responsible for activation of the TNFR1 by using single mutations of F5A, F13A, Y14A, L17A, N21A, I31A, Q32A, and K35A. Gomez et al. is silent in regard to any use of substitutions in the interacting residues in producing a vaccine antigen.

[0128] Recombinant affinity tagged Protein A, a polypeptide encompassing the five IgG domains (EDCAB) (Sjodahl, 1977) but lacking the C-terminal Region X (Guss et al., 1984), was purified from recombinant E. coli and used as a vaccine antigen (Stranger-Jones et al., 2006). Because of the attributes of SpA in binding the Fc portion of IgG, a specific humoral immune response to Protein A could not be measured (Stranger-Jones et al., 2006). The inventors have overcome this obstacle through the generation of SpA-DQ9,10K; D36,37A. BALB/c mice immunized with recombinant Protein A (SpA) displayed significant protection against intravenous challenge with S. aureus strains: a 2.951 log reduction in staphylococcal load as compared to the wildtype (P>0.005; Student's t-test) (Stranger-Jones et al., 2006). SpA specific antibodies may cause phagocytic clearance prior to abscess formation and/or impact the formation of the aforementioned eosinophilic barrier in abscesses that separate staphylococcal communities from immune cells since these do not form during infection with Protein A mutant strains. Each of the five SpA domains (i.e., domains formed from three helix bundles designated E, D, A, B, and C) exerts similar binding properties (Jansson et al., 1998). The solution and crystal structure of the domain D has been solved both with and without the Fc and VH3 (Fab) ligands, which bind Protein A in a non-competitive manner at distinct sites (Graille et al., 2000). Mutations in residues known to be involved in IgG binding (FS, Q9, Q10, S11, F13, Y14, L17, N28, 131 and K35) are also required for vWF AI and TNFR1 binding (Cedergren et al., 1993; Gomez et al., 2006; O'Seaghdha et al., 2006), whereas residues important for the VH3 interaction (Q26, G29, F30, S33, D36, D37, Q40, N43, E47) appear to have no impact on the other binding activities (Graille et al., 2000; Jansson et al., 1998). SpA specifically targets a subset of B cells that express VH3 family related IgM on their surface, i.e., VH3 type B cell receptors (Roben et al., 1995). Upon interaction with SpA, these B cells proliferate and commit to apoptosis, leading to preferential and prolonged deletion of innate-like B lymphocytes (i.e., marginal zone B cells and follicular B2 cells)(Goodyear et al., 2003; Goodyear et al., 2004).

[0129] Molecular Basis of Protein A Surface Display and Function.

[0130] Protein A is synthesized as a precursor in the bacterial cytoplasm and secreted via its YSIRK signal peptide at the cross wall, i.e. the cell division septum of staphylococci (FIG. 1) (DeDent et al., 2007; DeDent et al., 2008). Following cleavage of the C-terminal LPXTG sorting signal, Protein A is anchored to bacterial peptidoglycan crossbridges by sortase A (Mazmanian et al., 1999; Schneewind et al., 1995; Mazmanian et al., 2000). Protein A is the most abundant surface protein of staphylococci; the molecule is expressed by virtually all S. aureus strains (Cespedes et al., 2005; Kennedy et al., 2008; Said-Salim et al., 2003). Staphylococci turn over 15-20% of their cell wall per division cycle (Navarre and Schneewind, 1999). Murine hydrolases cleave the glycan strands and wall peptides of peptidoglycan, thereby releasing Protein A with its attached C-terminal cell wall disaccharide tetrapeptide into the extracellular medium (Ton-That et al., 1999). Thus, by physiological design, Protein A is both anchored to the cell wall and displayed on the bacterial surface but also released into surrounding tissues during host infection (Marraffini et al., 2006).

[0131] Protein A captures immunoglobulins on the bacterial surface and this biochemical activity enables staphylococcal escape from host innate and acquired immune responses (Jensen, 1958; Goodyear et al., 2004). Interestingly, region X of Protein A (Guss et al., 1984), a repeat domain that tethers the IgG binding domains to the LPXTG sorting signal/cell wall anchor, is perhaps the most variable portion of the staphylococcal genome (Said-Salim, 2003; Schneewind et al., 1992). Each of the five immunoglobulin binding domains of Protein A (SpA), formed from three helix bundles and designated E, D, A, B, and C, exerts similar structural and functional properties (Sjodahl, 1977; Jansson et al., 1998). The solution and crystal structure of the domain D has been solved both with and without the Fc and  $V_H3$  (Fab) ligands, which bind Protein A in a non-competitive manner at distinct sites (Graille 2000).

[0132] In the crystal structure complex, the Fab interacts with helix II and helix III of domain D via a surface composed of four VH region  $\beta$ -strands (Graille 2000). The major axis of helix II of domain D is approximately 50° to the orientation of

the strands, and the interhelical portion of domain D is most proximal to the C0 strand. The site of interaction on Fab is remote from the Ig light chain and the heavy chain constant region. The interaction involves the following domain D residues: Asp-36 of helix II, Asp-37 and Gln-40 in the loop between helix II and helix III and several other residues (Graille 2000). Both interacting surfaces are composed predominantly of polar side chains, with three negatively charged residues on domain D and two positively charged residues on the 2A2 Fab buried by the interaction, providing an overall electrostatic attraction between the two molecules. Of the five polar interactions identified between Fab and domain D, three are between side chains. A salt bridge is formed between Arg-H19 and Asp-36 and two hydrogen bonds are made between Tyr-H59 and Asp-37 and between Asn-H82a and Ser-33. Because of the conservation of Asp-36 and Asp-37 in all five IgG binding domains of Protein A, the inventors mutated these residues.

[0133] The SpA-D sites responsible for Fab binding are structurally separate from the domain surface that mediates Fcy binding. The interaction of Fcy with domain D primarily involves residues in helix I with lesser involvement of helix II (Gouda et al., 1992; Deisenhofer, 1981). With the exception of the Gln-32, a minor contact in both complexes, none of the residues that mediate the Fcy interaction are involved in Fab binding. To examine the spatial relationship between these different Ig-binding sites, the SpA domains in these complexes have been superimposed to construct a model of a complex between Fab, the SpA-domain D, and the Fcy molecule. In this ternary model, Fab and Fcy form a sandwich about opposite faces of the helix II without evidence of steric hindrance of either interaction. These findings illustrate how, despite its small size (i.e., 56-61 aa), an SpA domain can simultaneously display both activities, explaining experimental evidence that the interactions of Fab with an individual domain are noncompetitive. Residues for the interaction between SpA-D and Fcy are Gln-9 and Gln-10.

[0134] In contrast, occupancy of the Fc portion of IgG on the domain D blocks its interaction with vWF A1 and probably also TNFR1 (O'Seaghdha et al., 2006). Mutations in residues essential for IgGFc binding (F5, Q9, Q10, S11, F13, Y14, L17, N28, 131 and K35) are also required for vWF A1 and TNFR1 binding (O'Seaghdha et al., 2006; Cedergren et al., 1993; Gomez et al., 2006), whereas residues critical for the VH3 interaction (Q26, G29, F30, S33, D36, D37, Q40, N43, E47) have no impact on the binding activities of IgG Fc, vWF A1 or TNFR1 (Jansson et al., 1998; Graille et al., 2000). The Protein A immunoglobulin Fab binding activity targets a subset of B cells that express V<sub>H</sub>3 family related IgM on their surface, i.e., these molecules function as VH3type B cell receptors (Roben et al., 1995). Upon interaction with SpA, these B cells rapidly proliferate and then commit to apoptosis, leading to preferential and prolonged deletion of innate-like B lymphocytes (i.e., marginal zone B cells and follicular B2 cells) (Goodyear and Silverman, 2004; Goodyear and Silverman, 2003). More than 40% of circulating B cells are targeted by the Protein A interaction and the  $V_H$ 3 family represents the largest family of human B cell receptors to impart protective humoral responses against pathogens (Goodyear and Silverman, 2004; Goodyear and Silverman, 2003). Thus, Protein A functions analogously to staphylococcal superantigens (Roben et al., 1995), albeit that the latter class of molecules, for example SEB, TSST-1, TSST-2, form complexes with the T cell receptor to inappropriately stimulate host immune

responses and thereby precipitating characteristic disease features of staphylococcal infections (Roben et al., 1995; Tiedemann et al., 1995). Together these findings document the contributions of Protein A in establishing staphylococcal infections and in modulating host immune responses.

[0135] In sum, Protein A domains can viewed as displaying two different interfaces for binding with host molecules and any development of Protein A based vaccines must consider the generation of variants that do not perturb host cell signaling, platelet aggregation, sequestration of immunoglobulins or the induction of B cell proliferation and apoptosis. Such Protein A variants should also be useful in analyzing vaccines for the ability of raising antibodies that block the aforementioned SpA activities and occupy the five repeat domains at their dual binding interfaces. This goal is articulated and pursued here for the first time and methods are described in detail for the generation of Protein A variants that can be used as a safe vaccine for humans. To perturb IgG Fcy, vWF AI and TNFR1 binding, glutamine (Q) 9 and 10 [numbering derived from the SpA domain D as described in Uhlen et al., 1984] were mutated, and generated lysine substitutions for both glutamines with the expectation that these abolish the ligand attributes at the first binding interface. To perturb IgM Fab VH3 binding, aspartate (D) 36 and 37 were mutated, each of which is required for the association with the B cell receptor. D36 and D37 were both substituted with alanine Q9,10K and D36.37A mutations are here combined in the recombinant molecule SpA-DQ9,10K;D36,37A and tested for the binding attributes of Protein A. Further, SpA-D and SpA-DQ9,10K; D36,37A are subjected to immunization studies in mice and rabbits and analyzed for [1] the production of specific antibodies (SpA-D Ab); [2] the ability of SpA-D Ab to block the association between Protein A and its four different ligands; and, [3] the attributes of SpA-D Ab to generate protective immunity against staphylococcal infections. (See Examples section below).

#### [0136] B. Staphylococcal Coagulases

[0137] Coagulases are enzymes produced by Staphylococcus bacteria that convert fibrinogen to fibrin. Coa and vW<sub>h</sub> activate prothrombin without proteolysis (Friedrich et al., 2003). The coagulase.prothrombin complex recognizes fibringen as a specific substrate, converting it directly into fibrin. The crystal structure of the active complex revealed binding of the D1 and D2 domains to prothrombin and insertion of its Ile1-Val<sup>2</sup> N-terminus into the Ile<sup>16</sup> pocket, inducing a functional active site in the zymogen through conformational change (Friedrich et al., 2003). Exosite I of α-thrombin, the fibringen recognition site, and proexosite I on prothrombin are blocked by the D2 of Coa (Friedrich et al., 2003). Nevertheless, association of the tetrameric (Coa.prothrombin), complex binds fibringen at a new site with high affinity (Panizzi et al., 2006). This model explains the coagulant properties and efficient fibringen conversion by coagulase (Panizzi et al., 2006).

[0138] Fibrinogen is a large glycoprotein (Mr ~340,000), formed by three pairs of  $A\alpha$ -,  $B\beta$ -, and  $\gamma$ -chains covalently linked to form a "dimer of trimers," where A and B designate the fibrinopeptides released by thrombin cleavage (Panizzi et al., 2006). The elongated molecule folds into three separate domains, a central fragment E that contains the N-termini of all six chains and two flanking fragments D formed mainly by the C-termini of the  $B\beta$ - and  $\gamma$ -chains. These globular domains are connected by long triple-helical structures. Coagulase-prothrombin complexes, which convert human

fibrinogen to the self-polymerizing fibrin, are not targeted by circulating thrombin inhibitors (Panizzi et al., 2006). Thus, staphylococcal coagulases bypass the physiological blood coagulation pathway.

[0139] All S. aureus strains secrete coagulase and vWbp (Bjerketorp et al., 2004; Field and Smith, 1945). Although early work reported important contributions of coagulase to the pathogenesis of staphylococcal infections (Ekstedt and Yotis, 1960; Smith et al., 1947), more recent investigations with molecular genetics tools challenged this view by observing no virulence phenotypes with endocarditis, skin abscess and mastitis models in mice (Moreillon et al., 1995; Phonimdaeng et al., 1990). Generating isogenic variants of S. aureus Newman, a fully virulent clinical isolate (Duthie et al., 1952), it is described herein that coa mutants indeed display virulence defects in a lethal bacteremia and renal abscess model in mice. In the inventors experience, S. aureus 8325-4 is not fully virulent and it is presumed that mutational lesions in this strain may not be able to reveal virulence defects in vivo. Moreover, antibodies raised against Coa or vWbp perturb the pathogenesis of S. aureus Newman infections to a degree mirroring the impact of gene deletions. Coa and vWbp contribute to staphylococcal abscess formation and lethal bacteremia and may also function as protective antigens in subunit vaccines.

[0140] Biochemical studies document the biological value of antibodies against Coa and vWbp. By binding to antigen and blocking its association with clotting factors, the antibodies prevent the formation of Coa.prothrombin and vWbp. prothrombin complexes. Passive transfer studies revealed protection of experimental animals against staphylococcal abscess formation and lethal challenge by Coa and vWbp antibodies. Thus, Coa and vWbp neutralizing antibodies generate immune protection against staphylococcal disease.

[0141] Earlier studies revealed a requirement of coagulase for resisting phagocytosis in blood (Smith et al., 1947) and the inventors observed a similar phenotype for Δcoa mutants in lepirudin-treated mouse blood (see Example 3 below). As vWbp displays higher affinity for human prothrombin than the mouse counterpart, it is suspected the same may be true for  $\Delta vWbp$  variants in human blood. Further, expression of Coa and vWbp in abscess lesions as well as their striking distribution in the eosinophilic pseudocapsule surrounding (staphylococcal abscess communities (SACs) or the peripheral fibrin wall, suggest that secreted coagulases contribute to the establishment of these lesions. This hypothesis was tested and, indeed, Δcoa mutants were defective in the establishment of abscesses. A corresponding test, blocking Coa function with specific antibodies, produced the same effect. Consequently, it is proposed that the clotting of fibrin is a critical event in the establishment of staphylococcal abscesses that can be targeted for the development of protective vaccines. Due to their overlapping function on human prothrombin, both Coa and vWbp are considered excellent candidates for vaccine development.

[0142] C. Other Staphylococcal Antigens

[0143] Research over the past several decades identified *S. aureus* exotoxins, surface proteins and regulatory molecules as important virulence factors (Foster, 2005; Mazmanian et al., 2001; Novick, 2003). Much progress has been achieved regarding the regulation of these genes. For example, staphylococci perform a bacterial census via the secretion of autoinducing peptides that bind to a cognate receptor at threshold concentration, thereby activating phospho-relay reactions

and transcriptional activation of many of the exotoxin genes (Novick, 2003). The pathogenesis of staphylococcal infections relies on these virulence factors (secreted exotoxins, exopolysaccharides, and surface adhesins). The development of staphylococcal vaccines is hindered by the multifaceted nature of staphylococcal invasion mechanisms. It is well established that live attenuated micro-organisms are highly effective vaccines; immune responses elicited by such vaccines are often of greater magnitude and of longer duration than those produced by non-replicating immunogens. One explanation for this may be that live attenuated strains establish limited infections in the host and mimic the early stages of natural infection. Embodiments of the invention are directed to compositions and methods including variant SpA polypeptides and peptides, as well as other immunogenic extracellular proteins, polypeptides, and peptides (including both secreted and cell surface proteins or peptides) of gram positive bacteria for the use in mitigating or immunizing against infection. In particular embodiments the bacteria is a staphylococcus bacteria. Extracellular proteins, polypeptides, or peptides include, but are not limited to secreted and cell surface proteins of the targeted bacteria.

[0144] The human pathogen S. aureus secretes EsxA and EsxB, two ESAT-6 like proteins, across the bacterial envelope (Burts et al., 2005, which is incorporated herein by reference). Staphylococcal esxA and esxB are clustered with six other genes in the order of transcription: esxA esaA essA esaB essB essC esaC esxB. The acronyms esa, ess, and esx stand for ESAT-6 secretion accessory, system, and extracellular, respectively, depending whether the encoded proteins play an accessory (esa) or direct (ess) role for secretion, or are secreted (esx) in the extracellular milieu. The entire cluster of eight genes is herein referred to as the Ess cluster. EsxA, esxB, essA, essB, and essC are all required for synthesis or secretion of EsxA and EsxB. Mutants that fail to produce EsxA, EsxB, and EssC display defects in the pathogenesis of S. aureus murine abscesses, suggesting that this specialized secretion system may be a general strategy of human bacterial pathogenesis. Secretion of non-WXG100 substrates by the ESX-1 pathway has been reported for several antigens including EspA, EspB, Rv3483c, and Rv3615c (Fortune et al., 2005; MacGurn et al., 2005; McLaughlin et al., 2007; Xu et al., 2007). The alternate ESX-5 pathway has also been shown to secrete both WXG100 and non-WXG100 proteins in pathogenic mycobacteria (Abdallah et al., 2007; Abdallah et al., 2006).

[0145] The Staphylococcus aureus Ess pathway can be viewed as a secretion module equipped with specialized transport components (Ess), accessory factors (Esa) and cognate secretion substrates (Esx). EssA, EssB and EssC are required for EsxA and EsxB secretion. Because EssA, EssB and EssC are predicted to be transmembrane proteins, it is contemplated that these proteins form a secretion apparatus. Some of the proteins in the ess gene cluster may actively transport secreted substrates (acting as motor) while others may regulate transport (regulator). Regulation may be achieved, but need not be limited to, transcriptional or posttranslational mechanisms for secreted polypeptides, sorting of specific substrates to defined locations (e.g., extracellular medium or host cells), or timing of secretion events during infection. At this point, it is unclear whether all secreted Esx proteins function as toxins or contribute indirectly to pathogenesis.

[0146] Staphylococci rely on surface protein mediated-adhesion to host cells or invasion of tissues as a strategy for escape from immune defenses. Furthermore, S. aureus utilize surface proteins to sequester iron from the host during infection. The majority of surface proteins involved in staphylococcal pathogenesis carry C-terminal sorting signals, i.e., they are covalently linked to the cell wall envelope by sortase. Further, staphylococcal strains lacking the genes required for surface protein anchoring, i.e., sortase A and B, display a dramatic defect in the virulence in several different mouse models of disease. Thus, surface protein antigens represent a validated vaccine target as the corresponding genes are essential for the development of staphylococcal disease and can be exploited in various embodiments of the invention. The sortase enzyme superfamily are Gram-positive transpeptidases responsible for anchoring surface protein virulence factors to the peptidoglycan cell wall layer. Two sortase isoforms have been identified in Staphylococcus aureus, SrtA and SrtB. These enzymes have been shown to recognize a LPXTG motif in substrate proteins. The SrtB isoform appears to be important in heme iron acquisition and iron homeostasis, whereas the SrtA isoform plays a critical role in the pathogenesis of Gram-positive bacteria by modulating the ability of the bacterium to adhere to host tissue via the covalent anchoring of adhesins and other proteins to the cell wall peptidoglycan. In certain embodiments the SpA variants described herein can be used in combination with other staphylococcal proteins such as Coa, Eap, Ebh, Emp, EsaC, EsaB, EsxA, EsxB, Hla, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, IsdC, SasF, vWbp, and/or vWh proteins.

[0147] Certain aspects of the invention include methods and compositions concerning proteinaceous compositions including polypeptides, peptides, or nucleic acid encoding SpA variant(s) and other staphylococcal antigens such as other proteins transported by the Ess pathway, or sortase substrates. These proteins may be modified by deletion, insertion, and/or substitution.

[0148] The Esx polypeptides include the amino acid sequence of Esx proteins from bacteria in the Staphylococcus genus. The Esx sequence may be from a particular staphylococcus species, such as Staphylococcus aureus, and may be from a particular strain, such as Newman. In certain embodiments, the EsxA sequence is SAV0282 from strain Mu50 (which is the same amino acid sequence for Newman) and can be accessed using Genbank Accession Number Q99WU4 (gil68565539), which is hereby incorporated by reference. In other embodiments, the EsxB sequence is SAV0290 from strain Mu50 (which is the same amino acid sequence for Newman) and can be accessed using Genbank Accession Number Q99WT7 (gil68565532), which is hereby incorporated by reference. In further embodiments, other polypeptides transported by the Ess pathway may be used, the sequences of which may be identified by one of skill in the art using databases and internet accessible resources.

[0149] The sortase substrate polypeptides include, but are not limited to the amino acid sequence of SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, IsdC or SasF proteins from bacteria in the *Staphylococcus* genus. The sortase substrate polypeptide sequence may be from a particular *staphylococcus* species, such as *Staphylococcus aureus*, and may be from a particular strain, such as Newman. In certain embodiments, the SdrD sequence is from strain N315 and can be accessed using Genbank Accession Number NP\_373773.1 (gil15926240), which is incorporated by reference. In other

embodiments, the SdrE sequence is from strain N315 and can be accessed using Genbank Accession Number NP\_373774.1 (gil15926241), which is incorporated by reference. In other embodiments, the IsdA sequence is SAV1130 from strain Mu50 (which is the same amino acid sequence for Newman) and can be accessed using Genbank Accession Number NP\_371654.1 (gi|15924120), which is incorporated by reference. In other embodiments, the IsdB sequence is SAV1129 from strain Mu50 (which is the same amino acid sequence for Newman) and can be accessed using Genbank Accession Number NP\_371653.1 (gi|15924119), which is incorporated by reference. In further embodiments, other polypeptides transported by the Ess pathway or processed by sortase may be used, the sequences of which may be identified by one of skill in the art using databases and internet accessible resources.

[0150] Examples of various proteins that can be used in the context of the present invention can be identified by analysis of database submissions of bacterial genomes, including but not limited to accession numbers NC\_002951 (GI:57650036 and GenBank CP000046), NC\_002758 (GI:57634611 and GenBank BA000017), NC\_002745 (GI:29165615 and GenBank BA000018), NC\_003923 (GI:21281729 and GenBank BA000033), NC\_002952 (GI:49482253 and GenBank BX571856), NC\_002953 (GI:49484912 and GenBank BX571857), NC\_007793 (GI:87125858 and GenBank CP000255), NC\_007795 (GI:87201381 and GenBank CP000253) each of which are incorporated by reference.

[0151] As used herein, a "protein" or "polypeptide" refers to a molecule comprising at least ten amino acid residues. In some embodiments, a wild-type version of a protein or polypeptide are employed, however, in many embodiments of the invention, a modified protein or polypeptide is employed to generate an immune response. The terms described above may be used interchangeably. A "modified protein" or "modified polypeptide" or a "variant" refers to a protein or polypeptide whose chemical structure, particularly its amino acid sequence, is altered with respect to the wild-type protein or polypeptide. In some embodiments, a modified/variant protein or polypeptide has at least one modified activity or function (recognizing that proteins or polypeptides may have multiple activities or functions). It is specifically contemplated that a modified/variant protein or polypeptide may be altered with respect to one activity or function yet retain a wild-type activity or function in other respects, such as immunogenic-

[0152] In certain embodiments the size of a protein or polypeptide (wild-type or modified) may comprise, but is not limited to, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, 200, 210, 220, 230, 240, 250, 275, 300, 325, 350, 375, 400, 425, 450, 475, 500, 525, 550, 575, 600, 625, 650, 675, 700, 725, 750, 775, 800, 825, 850, 875, 900, 925, 950, 975, 1000, 1100, 1200, 1300, 1400, 1500, 1750, 2000, 2250, 2500 amino molecules or greater, and any range derivable therein, or derivative of a corresponding amino sequence described or referenced herein. It is contemplated that polypeptides may be mutated by truncation, rendering them shorter than their corresponding wild-type form, but also they might be altered by fusing or conjugating a heterologous protein sequence with a particular function (e.g., for targeting or localization, for enhanced immunogenicity, for purification purposes, etc.).

[0153] As used herein, an "amino molecule" refers to any amino acid, amino acid derivative, or amino acid mimic known in the art. In certain embodiments, the residues of the proteinaceous molecule are sequential, without any non-amino molecule interrupting the sequence of amino molecule residues. In other embodiments, the sequence may comprise one or more non-amino molecule moieties. In particular embodiments, the sequence of residues of the proteinaceous molecule may be interrupted by one or more non-amino molecule moieties.

[0154] Accordingly, the term "proteinaceous composition" encompasses amino molecule sequences comprising at least one of the 20 common amino acids in naturally synthesized proteins, or at least one modified or unusual amino acid.

[0155] Proteinaceous compositions may be made by any technique known to those of skill in the art, including (i) the expression of proteins, polypeptides, or peptides through standard molecular biological techniques, (ii) the isolation of proteinaceous compounds from natural sources, or (iii) the chemical synthesis of proteinaceous materials. The nucleotide as well as the protein, polypeptide, and peptide sequences for various genes have been previously disclosed, and may be found in the recognized computerized databases. One such database is the National Center for Biotechnology Information's Genbank and GenPept databases (on the World Wide Web at ncbi.nlm.nih.gov/). The coding regions for these genes may be amplified and/or expressed using the techniques disclosed herein or as would be known to those of ordinary skill in the art.

[0156] Amino acid sequence variants of SpA, coagulases and other polypeptides of the invention can be substitutional, insertional, or deletion variants. A variation in a polypeptide of the invention may affect 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29,30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, or more non-contiguous or contiguous amino acids of the polypeptide, as compared to wild-type. A variant can comprise an amino acid sequence that is at least 50%, 60%, 70%, 80%, or 90%, including all values and ranges there between, identical to any sequence provided or referenced herein, e.g., SEQ ID NO:2-8 or SEQ ID NO:11-30, A variant can include 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, or more substitute amino acids. A polypeptide processed or secreted by the Ess pathway or other surface proteins (see Table 1) or sortase substrates from any staphylococcus species and strain are contemplated for use in compositions and methods described herein.

[0157] Deletion variants typically lack one or more residues of the native or wild-type protein. Individual residues can be deleted or a number of contiguous amino acids can be deleted. A stop codon may be introduced (by substitution or insertion) into an encoding nucleic acid sequence to generate a truncated protein. Insertional mutants typically involve the addition of material at a non-terminal point in the polypeptide. This may include the insertion of one or more residues. Terminal additions, called fusion proteins, may also be generated. These fusion proteins include multimers or concatamers of one or more peptide or polypeptide described or referenced herein.

[0158] Substitutional variants typically contain the exchange of one amino acid for another at one or more sites

within the protein, and may be designed to modulate one or more properties of the polypeptide, with or without the loss of other functions or properties. Substitutions may be conservative, that is, one amino acid is replaced with one of similar shape and charge. Conservative substitutions are well known in the art and include, for example, the changes of: alanine to serine; arginine to lysine; asparagine to glutamine or histidine; aspartate to glutamate; cysteine to serine; glutamine to asparagine; glutamate to aspartate; glycine to proline; histidine to asparagine or glutamine; isoleucine to leucine or valine; leucine to valine or isoleucine; lysine to arginine; methionine to leucine or isoleucine; phenylalanine to tyrosine, leucine or methionine; serine to threonine; threonine to serine; tryptophan to tyrosine; tyrosine to tryptophan or phenylalanine; and valine to isoleucine or leucine. Alternatively, substitutions may be non-conservative such that a function or activity of the polypeptide is affected. Non-conservative changes typically involve substituting a residue with one that is chemically dissimilar, such as a polar or charged amino acid for a nonpolar or uncharged amino acid, and vice versa.

TABLE 3

		Co	odon	Tab]	le
Amino Acids		Code	ons		
Alanine	AlaA	GCA	GCC	GCG	GCU
Cysteine	Cys C	UGC	UGU		
Aspartic acid	Asp D	GAC	GAU		
Glutamic acid	Glu E	GAA	GAG		
Phenylalanine	Phe F	UUC	טטט		
Glycine	GlyG	GGA	GGC	GGG	GGU
Histidine	HisH	CAC	CAU		
Isoleucine	IleI	AUA	AUC	AUU	

TABLE 2

		Lacinpi	ny sum	ice prot	ems of a	S. aureus str	11115.	
SAV#	SA#	Surface	MW2	Mu50	N315	Newman	MRSA252*	MSSA476*
SAV0111	SA0107	Spa	492	450	450	520	516	492
SAV2503	SA2291	FnBPA	1015	1038	1038	741	_	1015
SAV2502	SA2290	FnBPB	943	961	961	677	965	957
SAV0811	SA0742	ClfA	946	935	989	933	1029	928
SAV2630	SA2423	ClfB	907	877	877	913	873	905
Np	Np	Can	1183	_	_	_	1183	1183
SAV0561	SA0519	SdrC	955	953	953	947	906	957
SAV0562	SA0520	SdrD	1347	1385	1385	1315	_	1365
SAV0563	SA0521	SdrE	1141	1141	1141	1166	1137	1141
Np	Np	Pls	_	_	_	_	_	_
SAV2654	SA2447	SasA	2275	2271	2271	2271	1351	2275
SAV2160	SA1964	SasB	686	2481	2481	2481	2222	685
	SA1577	SasC	2186	213	2186	2186	2189	2186
SAV0134	SA0129	SasD	241	241	241	241	221	241
SAV1130	SA0977	SasE/IsdA	350	350	350	350	354	350
SAV2646	SA2439	SasF	635	635	635	635	627	635
SAV2496		SasG	1371	525	927	_	_	1371
SAV0023	SA0022	SasH	772	_	772	772	786	786
SAV1731	SA1552	SasI	895	891	891	891	534	895
SAV1129	SA0976	SasJ/IsdB	645	645	645	645	652	645
	SA2381	SasK	198	211	211	_	_	197
	Np	SasL	_	232	_	_	_	_
SAV1131	•	IsdC	227	227	227	227	227	227

[0159] Proteins of the invention may be recombinant, or synthesized in vitro. Alternatively, a non-recombinant or recombinant protein may be isolated from bacteria. It is also contemplated that a bacteria containing such a variant may be implemented in compositions and methods of the invention. Consequently, a protein need not be isolated.

[0160] The term "functionally equivalent codon" is used herein to refer to codons that encode the same amino acid, such as the six codons for arginine or serine, and also refers to codons that encode biologically equivalent amino acids (see Table 2, below).

TABLE 3-continued

	Codon Table
Amino Acids	Codons
Lysine	Lys K AAA AAG
Leucine	Leu L UUA UUG CUA CUC CUG CUU
Methionine	Met M AUG

TABLE 3-continued

	Codon Table
Amino Acids	Codons
Asparagine	Asn N AAC AAU
Proline	Pro P CCA CCC CCG CCU
Glutamine	GlnQ CAA CAG
Arginine	Arg R AGA AGG CGA CGC CGG CGU
Serine	Sers AGC AGU UCA UCC UCG UCU
Threonine	Thr T ACA ACC ACG ACU
Valine	ValV GUA GUC GUG GUU
Tryptophan	Trp W UGG
Tyrosine	Tyr Y UAC UAU

[0161] It also will be understood that amino acid and nucleic acid sequences may include additional residues, such as additional N- or C-terminal amino acids, or 5' or 3' sequences, respectively, and yet still be essentially as set forth in one of the sequences disclosed herein, so long as the sequence meets the criteria set forth above, including the maintenance of biological protein activity (e.g., immunogenicity) where protein expression is concerned. The addition of terminal sequences particularly applies to nucleic acid sequences that may, for example, include various non-coding sequences flanking either of the 5' or 3' portions of the coding region.

[0162] The following is a discussion based upon changing of the amino acids of a protein to create a variant polypeptide or peptide. For example, certain amino acids may be substituted for other amino acids in a protein structure with or without appreciable loss of interactive binding capacity with structures such as, for example, antigen-binding regions of antibodies or binding sites on substrate molecules. Since it is the interactive capacity and nature of a protein that defines that protein's functional activity, certain amino acid substitutions can be made in a protein sequence, and in its underlying DNA coding sequence, and nevertheless produce a protein with a desirable property. It is thus contemplated by the inventors that various changes may be made in the DNA sequences of genes.

[0163] It is contemplated that in compositions of the invention, there is between about 0.001 mg and about 10 mg of total polypeptide, peptide, and/or protein per ml. The concentration of protein in a composition can be about, at least about or at most about 0.001, 0.010, 0.050, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1.0, 1.5, 2.0, 2.5, 3.0, 3.5, 4.0, 4.5, 5.0, 5.5, 6.0,6.5, 7.0, 7.5, 8.0, 8.5, 9.0, 9.5, 10.0 mg/ml or more (or any range derivable therein). Of this, about, at least about, or at most about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85,86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100% may be an SpA variant or a coagulase, and may be used in combination with other peptides or polypeptides, such as other bacterial peptides and/or antigens.

[0164] The present invention contemplates the administration of variant SpA polypeptides or peptides to effect a preventative therapy or therapeutic effect against the development of a disease or condition associated with infection by a *staphylococcus* pathogen.

[0165] In certain aspects, combinations of staphylococcal antigens are used in the production of an immunogenic composition that is effective at treating or preventing staphylococcal infection. Staphylococcal infections progress through several different stages. For example, the staphylococcal life cycle involves commensal colonization, initiation of infection by accessing adjoining tissues or the bloodstream, and/or anaerobic multiplication in the blood. The interplay between S. aureus virulence determinants and the host defense mechanisms can induce complications such as endocarditis, metastatic abscess formation, and sepsis syndrome. Different molecules on the surface of the bacterium are involved in different steps of the infection cycle. Combinations of certain antigens can elicit an immune response which protects against multiple stages of staphylococcal infection. The effectiveness of the immune response can be measured either in animal model assays and/or using an opsonophagocytic assay.

[0166] D. Polypeptides and Polypeptide Production

[0167] The present invention describes polypeptides, peptides, and proteins and immunogenic fragments thereof for use in various embodiments of the present invention. For example, specific polypeptides are assayed for or used to elicit an immune response. In specific embodiments, all or part of the proteins of the invention can also be synthesized in solution or on a solid support in accordance with conventional techniques. Various automatic synthesizers are commercially available and can be used in accordance with known protocols. See, for example, Stewart and Young, (1984); Tam et al., (1983); Merrifield, (1986); and Barany and Merrifield (1979), each incorporated herein by reference.

[0168] Alternatively, recombinant DNA technology may be employed wherein a nucleotide sequence which encodes a peptide of the invention is inserted into an expression vector, transformed or transfected into an appropriate host cell and cultivated under conditions suitable for expression.

[0169] One embodiment of the invention includes the use of gene transfer to cells, including microorganisms, for the production and/or presentation of polypeptides or peptides. The gene for the polypeptide or peptide of interest may be transferred into appropriate host cells followed by culture of cells under the appropriate conditions. The generation of recombinant expression vectors, and the elements included therein, are well known in the art and briefly discussed herein. Alternatively, the protein to be produced may be an endogenous protein normally synthesized by the cell that is isolated and purified.

[0170] Another embodiment of the present invention uses autologous B lymphocyte cell lines, which are transfected with a viral vector that expresses an immunogen product, and more specifically, a protein having immunogenic activity. Other examples of mammalian host cell lines include, but are not limited to Vero and HeLa cells, other B- and T-cell lines, such as CEM, 721.221, H9, Jurkat, Raji, as well as cell lines of Chinese hamster ovary, W138, BHK, COS-7, 293, HepG2, 3T3, RIN and MDCK cells. In addition, a host cell strain may be chosen that modulates the expression of the inserted sequences, or that modifies and processes the gene product in the manner desired. Such modifications (e.g., glycosylation) and processing (e.g., cleavage) of protein products may be

important for the function of the protein. Different host cells have characteristic and specific mechanisms for the post-translational processing and modification of proteins. Appropriate cell lines or host systems can be chosen to ensure the correct modification and processing of the foreign protein expressed.

[0171] A number of selection systems may be used including, but not limited to HSV thymidine kinase, hypoxanthine-guanine phosphoribosyltransferase, and adenine phosphoribosyltransferase genes, in tk-, hgprt- or aprt-cells, respectively. Also, anti-metabolite resistance can be used as the basis of selection: for dhfr, which confers resistance to trimethoprim and methotrexate; gpt, which confers resistance to mycophenolic acid; neo, which confers resistance to the aminoglycoside G418; and hygro, which confers resistance to hygromycin.

[0172] Animal cells can be propagated in vitro in two modes: as non-anchorage-dependent cells growing in suspension throughout the bulk of the culture or as anchorage-dependent cells requiring attachment to a solid substrate for their propagation (i.e., a monolayer type of cell growth).

[0173] Non-anchorage dependent or suspension cultures from continuous established cell lines are the most widely used means of large scale production of cells and cell products. However, suspension cultured cells have limitations, such as tumorigenic potential and lower protein production than adherent cells.

[0174] Where a protein is specifically mentioned herein, it is preferably a reference to a native or recombinant protein or optionally a protein in which any signal sequence has been removed. The protein may be isolated directly from the staphylococcal strain or produced by recombinant DNA techniques. Immunogenic fragments of the protein may be incorporated into the immunogenic composition of the invention. These are fragments comprising at least 10 amino acids, 20 amino acids, 30 amino acids, 40 amino acids, 50 amino acids, or 100 amino acids, including all values and ranges there between, taken contiguously from the amino acid sequence of the protein. In addition, such immunogenic fragments are immunologically reactive with antibodies generated against the Staphylococcal proteins or with antibodies generated by infection of a mammalian host with Staphylococci. Immunogenic fragments also include fragments that when administered at an effective dose, (either alone or as a hapten bound to a carrier), elicit a protective or therapeutic immune response against Staphylococcal infection, in certain aspects it is protective against S. aureus and/or S. epidermidis infection. Such an immunogenic fragment may include, for example, the protein lacking an N-terminal leader sequence, and/or a transmembrane domain and/or a C-terminal anchor domain. In a preferred aspect the immunogenic fragment according to the invention comprises substantially all of the extracellular domain of a protein which has at least 80% identity, at least 85% identity, at least 90% identity, at least 95% identity, or at least 97-99% identity, including all values and ranges there between, to a sequence selected segment of a polypeptide described or referenced herein.

[0175] Also included in immunogenic compositions of the invention are fusion proteins composed of one or more Staphylococcal proteins, or immunogenic fragments of staphylococcal proteins. Such fusion proteins may be made recombinantly and may comprise one portion of at least 1, 2, 3, 4, 5, or 6 staphylococcal proteins or segments. Alternatively, a fusion protein may comprise multiple portions of at least 1, 2,

3, 4 or 5 staphylococcal proteins. These may combine different Staphylococcal proteins and/or multiples of the same protein or protein fragment, or immunogenic fragments in the same protein (forming a multimer or a concatamer). Alternatively, the invention also includes individual fusion proteins of Staphylococcal proteins or immunogenic fragments thereof, as a fusion protein with heterologous sequences such as a provider of T-cell epitopes or purification tags, for example:  $\beta$ -galactosidase, glutathione-S-transferase, green fluorescent proteins (GFP), epitope tags such as FLAG, myc tag, poly histidine, or viral surface proteins such as influenza virus haemagglutinin, or bacterial proteins such as tetanus toxoid, diphtheria toxoid, or CRM197.

### II. Nucleic Acids

[0176] In certain embodiments, the present invention concerns recombinant polynucleotides encoding the proteins, polypeptides, peptides of the invention. The nucleic acid sequences for SpA, coagulases and other bacterial proteins are included, all of which are incorporated by reference, and can be used to prepare peptides or polypeptides.

[0177] As used in this application, the term "polynucle-otide" refers to a nucleic acid molecule that either is recombinant or has been isolated free of total genomic nucleic acid. Included within the term "polynucleotide" are oligonucle-otides (nucleic acids of 100 residues or less in length), recombinant vectors, including, for example, plasmids, cosmids, phage, viruses, and the like. Polynucleotides include, in certain aspects, regulatory sequences, isolated substantially away from their naturally occurring genes or protein encoding sequences. Polynucleotides may be single-stranded (coding or antisense) or double-stranded, and may be RNA, DNA (genomic, cDNA or synthetic), analogs thereof, or a combination thereof. Additional coding or non-coding sequences may, but need not, be present within a polynucleotide.

[0178] In this respect, the term "gene," "polynucleotide," or "nucleic acid" is used to refer to a nucleic acid that encodes a protein, polypeptide, or peptide (including any sequences required for proper transcription, post-translational modification, or localization). As will be understood by those in the art, this term encompasses genomic sequences, expression cassettes, cDNA sequences, and smaller engineered nucleic acid segments that express, or may be adapted to express, proteins, polypeptides, domains, peptides, fusion proteins, and mutants. A nucleic acid encoding all or part of a polypeptide may contain a contiguous nucleic acid sequence of: 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, 200, 210, 220, 230, 240, 250, 260, 270, 280, 290, 300, 310, 320, 330, 340, 350, 360, 370, 380, 390, 400, 410, 420, 430, 440, 441, 450, 460, 470, 480, 490, 500, 510, 520, 530, 540, 550, 560, 570, 580, 590, 600, 610, 620, 630, 640, 650, 660, 670, 680, 690, 700, 710, 720, 730, 740, 750, 760, 770, 780, 790, 800, 810, 820, 830, 840, 850, 860, 870, 880, 890, 900, 910, 920, 930, 940, 950, 960, 970, 980, 990, 1000, 1010, 1020, 1030, 1040, 1050, 1060, 1070, 1080, 1090, 1095, 1100, 1500, 2000, 2500, 3000, 3500, 4000, 4500, 5000, 5500, 6000, 6500, 7000, 7500, 8000, 9000, 10000, or more nucleotides, nucleosides, or base pairs, including all values and ranges therebetween, of a polynucleotide encoding one or more amino acid sequence described or referenced herein. It also is contemplated that a particular polypeptide may be encoded by nucleic acids containing variations having

slightly different nucleic acid sequences but, nonetheless, encode the same or substantially similar protein (see Table 3 above).

[0179] In particular embodiments, the invention concerns isolated nucleic acid segments and recombinant vectors incorporating nucleic acid sequences that encode a variant SpA or coagulase. The term "recombinant" may be used in conjunction with a polynucleotide or polypeptide and generally refers to a polypeptide or polynucleotide produced and/or manipulated in vitro or that is a replication product of such a molecule.

[0180] In other embodiments, the invention concerns isolated nucleic acid segments and recombinant vectors incorporating nucleic acid sequences that encode a variant SpA or coagulase polypeptide or peptide to generate an immune response in a subject. In various embodiments the nucleic acids of the invention may be used in genetic vaccines.

[0181] The nucleic acid segments used in the present invention can be combined with other nucleic acid sequences, such as promoters, polyadenylation signals, additional restriction enzyme sites, multiple cloning sites, other coding segments, and the like, such that their overall length may vary considerably. It is therefore contemplated that a nucleic acid fragment of almost any length may be employed, with the total length preferably being limited by the ease of preparation and use in the intended recombinant nucleic acid protocol. In some cases, a nucleic acid sequence may encode a polypeptide sequence with additional heterologous coding sequences, for example to allow for purification of the polypeptide, transport, secretion, post-translational modification, or for therapeutic benefits such as targeting or efficacy. As discussed above, a tag or other heterologous polypeptide may be added to the modified polypeptide-encoding sequence, wherein "heterologous" refers to a polypeptide that is not the same as the modified polypeptide.

[0182] In certain other embodiments, the invention concerns isolated nucleic acid segments and recombinant vectors that include within their sequence a contiguous nucleic acid sequence from SEQ ID NO:1 (SpA domain D) or SEQ ID NO:3 (SpA) or any other nucleic acid sequences encoding coagulases or other secreted virulence factors and/or surface proteins including proteins transported by the Ess pathway, processed by sortase, or proteins incorporated herein by reference

[0183] In certain embodiments, the present invention provides polynucleotide variants having substantial identity to the sequences disclosed herein; those comprising at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% or higher sequence identity, including all values and ranges there between, compared to a polynucleotide sequence of this invention using the methods described herein (e.g., BLAST analysis using standard parameters).

[0184] The invention also contemplates the use of polynucleotides which are complementary to all the above described polynucleotides.

[0185] A. Vectors

[0186] Polypeptides of the invention may be encoded by a nucleic acid molecule comprised in a vector. The term "vector" is used to refer to a carrier nucleic acid molecule into which a heterologous nucleic acid sequence can be inserted for introduction into a cell where it can be replicated and expressed. A nucleic acid sequence can be "heterologous," which means that it is in a context foreign to the cell in which the vector is being introduced or to the nucleic acid in which

is incorporated, which includes a sequence homologous to a sequence in the cell or nucleic acid but in a position within the host cell or nucleic acid where it is ordinarily not found. Vectors include DNAs, RNAs, plasmids, cosmids, viruses (bacteriophage, animal viruses, and plant viruses), and artificial chromosomes (e.g., YACs). One of skill in the art would be well equipped to construct a vector through standard recombinant techniques (for example Sambrook et al., 2001; Ausubel et al., 1996, both incorporated herein by reference). In addition to encoding a variant SpA polypeptide the vector can encode other polypeptide sequences such as a one or more other bacterial peptide, a tag, or an immunogenicity enhancing peptide. Useful vectors encoding such fusion proteins include pIN vectors (Inouye et al., 1985), vectors encoding a stretch of histidines, and pGEX vectors, for use in generating glutathione S-transferase (GST) soluble fusion proteins for later purification and separation or cleavage.

[0187] The term "expression vector" refers to a vector containing a nucleic acid sequence coding for at least part of a gene product capable of being transcribed. In some cases, RNA molecules are then translated into a protein, polypeptide, or peptide. Expression vectors can contain a variety of "control sequences," which refer to nucleic acid sequences necessary for the transcription and possibly translation of an operably linked coding sequence in a particular host organism. In addition to control sequences that govern transcription and translation, vectors and expression vectors may contain nucleic acid sequences that serve other functions as well and are described herein.

[0188] 1. Promoters and Enhancers

[0189] A "promoter" is a control sequence. The promoter is typically a region of a nucleic acid sequence at which initiation and rate of transcription are controlled. It may contain genetic elements at which regulatory proteins and molecules may bind such as RNA polymerase and other transcription factors. The phrases "operatively positioned," "operatively linked," "under control," and "under transcriptional control" mean that a promoter is in a correct functional location and/or orientation in relation to a nucleic acid sequence to control transcriptional initiation and expression of that sequence. A promoter may or may not be used in conjunction with an "enhancer," which refers to a cis-acting regulatory sequence involved in the transcriptional activation of a nucleic acid sequence.

[0190] Naturally, it may be important to employ a promoter and/or enhancer that effectively directs the expression of the DNA segment in the cell type or organism chosen for expression. Those of skill in the art of molecular biology generally know the use of promoters, enhancers, and cell type combinations for protein expression (see Sambrook et al., 2001, incorporated herein by reference). The promoters employed may be constitutive, tissue-specific, or inducible and in certain embodiments may direct high level expression of the introduced DNA segment under specified conditions, such as large-scale production of recombinant proteins or peptides.

[0191] Various elements/promoters may be employed in the context of the present invention to regulate the expression of a gene. Examples of such inducible elements, which are regions of a nucleic acid sequence that can be activated in response to a specific stimulus, include but are not limited to Immunoglobulin Heavy Chain (Banerji et al., 1983; Gilles et al., 1983; Grosschedl et al., 1985; Atchinson et al., 1986, 1987; Imler et al., 1987; Weinberger et al., 1984; Kiledjian et al., 1988; Porton et al.; 1990), Immunoglobulin Light Chain

(Queen et al., 1983; Picard et al., 1984), T Cell Receptor (Luria et al., 1987; Winoto et al., 1989; Redondo et al.; 1990), HLA DQ a and/or DQ 13 (Sullivan et al., 1987), (3 Interferon (Goodbourn et al., 1986; Fujita et al., 1987; Goodbourn et al., 1988), Interleukin-2 (Greene et al., 1989), Interleukin-2 Receptor (Greene et al., 1989; Lin et al., 1990), MHC Class II 5 (Koch et al., 1989), MHC Class II HLA-DRa (Sherman et al., 1989), β-Actin (Kawamoto et al., 1988; Ng et al.; 1989), Muscle Creatine Kinase (MCK) (Jaynes et al., 1988; Horlick et al., 1989; Johnson et al., 1989), Prealbumin (Transthyretin) (Costa et al., 1988), Elastase I (Ornitz et al., 1987), Metallothionein (MTII) (Karin et al., 1987; Culotta et al., 1989), Collagenase (Pinkert et al., 1987; Angel et al., 1987), Albumin (Pinkert et al., 1987; Tronche et al., 1989, 1990),  $\alpha$ -Fetoprotein (Godbout et al., 1988; Campere et al., 1989), γ-Globin (Bodine et al., 1987; Perez-Stable et al., 1990), β-Globin (Trudel et al., 1987), c-fos (Cohen et al., 1987), c-Ha-Ras (Triesman, 1986; Deschamps et al., 1985), Insulin (Edlund et al., 1985), Neural Cell Adhesion Molecule (NCAM) (Hirsh et al., 1990), α1-Antitrypain (Latimer et al., 1990), H2B (TH2B) Histone (Hwang et al., 1990), Mouse and/or Type I Collagen (Ripe et al., 1989), Glucose-Regulated Proteins (GRP94 and GRP78) (Chang et al., 1989), Rat Growth Hormone (Larsen et al., 1986), Human Serum Amyloid A (SAA) (Edbrooke et al., 1989), Troponin I (TN I) (Yutzey et al., 1989), Platelet-Derived Growth Factor (PDGF) (Pech et al., 1989), Duchenne Muscular Dystrophy (Klamut et al., 1990), SV40 (Banerji et al., 1981; Moreau et al., 1981; Sleigh et al., 1985; Firak et al., 1986; Herr et al., 1986; Imbra et al., 1986; Kadesch et al., 1986; Wang et al., 1986; Ondek et al., 1987; Kuhl et al., 1987; Schaffner et al., 1988), Polyoma (Swartzendruber et al., 1975; Vasseur et al., 1980; Katinka et al., 1980, 1981; Tyndell et al., 1981; Dandolo et al., 1983; de Villiers et al., 1984; Hen et al., 1986; Satake et al., 1988; Campbell et al., 1988), Retroviruses (Kriegler et al., 1982, 1983; Levinson et al., 1982; Kriegler et al., 1983, 1984a, b, 1988; Bosze et al., 1986; Miksicek et al., 1986; Celander et al., 1987; Thiesen et al., 1988; Celander et al., 1988; Choi et al., 1988; Reisman et al., 1989), Papilloma Virus (Campo et al., 1983; Lusky et al., 1983; Spandidos and Wilkie, 1983; Spalholz et al., 1985; Lusky et al., 1986; Cripe et al., 1987; Gloss et al., 1987; Hirochika et al., 1987; Stephens et al., 1987), Hepatitis B Virus (Bulla et al., 1986; Jameel et al., 1986; Shaul et al., 1987; Spandau et al., 1988; Vannice et al., 1988), Human Immunodeficiency Virus (Muesing et al., 1987; Hauber et al., 1988; Jakobovits et al., 1988; Feng et al., 1988; Takebe et al., 1988; Rosen et al., 1988; Berkhout et al., 1989; Laspia et al., 1989; Sharp et al., 1989; Braddock et al., 1989), Cytomegalovirus (CMV) IE (Weber et al., 1984; Boshart et al., 1985; Foecking et al., 1986), Gibbon Ape Leukemia Virus (Holbrook et al., 1987; Quinn et al., 1989).

[0192] Inducible elements include, but are not limited to MT II—Phorbol Ester (TFA)/Heavy metals (Palmiter et al., 1982; Haslinger et al., 1985; Searle et al., 1985; Stuart et al., 1985; Imagawa et al., 1987, Karin et al., 1987; Angel et al., 1987b; McNeall et al., 1989); MMTV (mouse mammary tumor virus)—Glucocorticoids (Huang et al., 1981; Lee et al., 1981; Majors et al., 1983; Chandler et al., 1983; Lee et al., 1984; Ponta et al., 1985; Sakai et al., 1988);  $\beta$ -Interferon—poly(rI)x/poly(rc) (Tavernier et al., 1983); Adenovirus 5 E2—ElA (Imperiale et al., 1984); Collagenase—Phorbol Ester (TPA) (Angel et al., 1987a); Stromelysin—Phorbol Ester (TPA) (Angel et al., 1987b); SV40—Phorbol Ester (TPA) (Angel et al., 1987b); Murine MX Gene—Interferon,

Newcastle Disease Virus (Hug et al., 1988); GRP78 Gene—A23187 (Resendez et al., 1988);  $\alpha$ -2-Macroglobulin—IL-6 (Kunz et al., 1989); Vimentin—Serum (Rittling et al., 1989); MHC Class I Gene H-2 $\kappa$ b—Interferon (Blanar et al., 1989); HSP70—ElA/SV40 Large T Antigen (Taylor et al., 1989, 1990a, 1990b); Proliferin—Phorbol Ester/TPA (Mordacq et al., 1989); Tumor Necrosis Factor—PMA (Hensel et al., 1989); and Thyroid Stimulating Hormone a Gene—Thyroid Hormone (Chatterjee et al., 1989).

[0193] The particular promoter that is employed to control the expression of peptide or protein encoding polynucleotide of the invention is not believed to be critical, so long as it is capable of expressing the polynucleotide in a targeted cell, preferably a bacterial cell. Where a human cell is targeted, it is preferable to position the polynucleotide coding region adjacent to and under the control of a promoter that is capable of being expressed in a human cell. Generally speaking, such a promoter might include either a bacterial, human or viral promoter.

[0194] In embodiments in which a vector is administered to a subject for expression of the protein, it is contemplated that a desirable promoter for use with the vector is one that is not down-regulated by cytokines or one that is strong enough that even if down-regulated, it produces an effective amount of a variant SpA for eliciting an immune response. Non-limiting examples of these are CMV IE and RSV LTR. Tissue specific promoters can be used, particularly if expression is in cells in which expression of an antigen is desirable, such as dendritic cells or macrophages. The mammalian MHC I and MHC II promoters are examples of such tissue-specific promoters.

[0195] 2. Initiation Signals and Internal Ribosome Binding Sites (IRES)

[0196] A specific initiation signal also may be required for efficient translation of coding sequences. These signals include the ATG initiation codon or adjacent sequences. Exogenous translational control signals, including the ATG initiation codon, may need to be provided. One of ordinary skill in the art would readily be capable of determining this and providing the necessary signals.

[0197] In certain embodiments of the invention, the use of internal ribosome entry sites (IRES) elements are used to create multigene, or polycistronic, messages. IRES elements are able to bypass the ribosome scanning model of 5' methylated Cap dependent translation and begin translation at internal sites (Pelletier and Sonenberg, 1988; Macejak and Sarnow, 1991). IRES elements can be linked to heterologous open reading frames. Multiple open reading frames can be transcribed together, each separated by an IRES, creating polycistronic messages. Multiple genes can be efficiently expressed using a single promoter/enhancer to transcribe a single message (see U.S. Pat. Nos. 5,925,565 and 5,935,819, herein incorporated by reference).

[0198] 3. Selectable and Screenable Markers

[0199] In certain embodiments of the invention, cells containing a nucleic acid construct of the present invention may be identified in vitro or in vivo by encoding a screenable or selectable marker in the expression vector. When transcribed and translated, a marker confers an identifiable change to the cell permitting easy identification of cells containing the expression vector. Generally, a selectable marker is one that confers a property that allows for selection. A positive selectable marker is one in which the presence of the marker allows for its selection, while a negative selectable marker is one in

which its presence prevents its selection. An example of a positive selectable marker is a drug resistance marker.

[0200] B. Host Cells [0201] As used herein, the terms "cell," "cell line," and "cell culture" may be used interchangeably. All of these terms also include their progeny, which is any and all subsequent generations. It is understood that all progeny may not be identical due to deliberate or inadvertent mutations. In the context of expressing a heterologous nucleic acid sequence, "host cell" refers to a prokaryotic or eukaryotic cell, and it includes any transformable organism that is capable of replicating a vector or expressing a heterologous gene encoded by a vector. A host cell can, and has been, used as a recipient for vectors or viruses. A host cell may be "transfected" or "transformed," which refers to a process by which exogenous nucleic acid, such as a recombinant protein-encoding sequence, is transferred or introduced into the host cell. A transformed cell includes the primary subject cell and its

[0202] Host cells may be derived from prokaryotes or eukaryotes, including bacteria, yeast cells, insect cells, and mammalian cells for replication of the vector or expression of part or all of the nucleic acid sequence(s). Numerous cell lines and cultures are available for use as a host cell, and they can be obtained through the American Type Culture Collection (ATCC), which is an organization that serves as an archive for living cultures and genetic materials (www.atcc.org).

[0203] C. Expression Systems

[0204] Numerous expression systems exist that comprise at least a part or all of the compositions discussed above. Prokaryote- and/or eukaryote-based systems can be employed for use with the present invention to produce nucleic acid sequences, or their cognate polypeptides, proteins and peptides. Many such systems are commercially and widely available.

[0205] The insect cell/baculovirus system can produce a high level of protein expression of a heterologous nucleic acid segment, such as described in U.S. Pat. Nos. 5,871,986, 4,879,236, both herein incorporated by reference, and which can be bought, for example, under the name MAXBAC® 2.0 from INVITROGEN® and BACPACK™ BACULOVIRUS EXPRESSION SYSTEM FROM CLONTECH®.

[0206] In addition to the disclosed expression systems of the invention, other examples of expression systems include STRATAGENE®'s COMPLETE CONTROL™ Inducible Mammalian Expression System, which involves a synthetic ecdysone-inducible receptor, or its pET Expression System, an E. coli expression system. Another example of an inducible expression system is available from INVITROGEN®, which carries the T-REX<sup>TM</sup> (tetracycline-regulated expression) System, an inducible mammalian expression system that uses the full-length CMV promoter. INVITROGEN® also provides a yeast expression system called the Pichia methanolica Expression System, which is designed for highlevel production of recombinant proteins in the methylotrophic yeast Pichia methanolica. One of skill in the art would know how to express a vector, such as an expression construct, to produce a nucleic acid sequence or its cognate polypeptide, protein, or peptide.

## III. Polysaccharides

[0207] The immunogenic compositions of the invention may further comprise capsular polysaccharides including one or more of PIA (also known as PNAG) and/or S. aureus Type V and/or type VIII capsular polysaccharide and/or S. epidermidis Type I, and/or Type II and/or Type III capsular polysaccharide.

[0208] A. PIA (PNAG)

[0209] It is now clear that the various forms of staphylococcal surface polysaccharides identified as PS/A, PIA and SAA are the same chemical entity—PNAG (Maira-Litran et al., 2004). Therefore the term PIA or PNAG encompasses all these polysaccharides or oligosaccharides derived from them.

[0210] PIA is a polysaccharide intercellular adhesin and is composed of a polymer of  $\beta$ -(1 $\rightarrow$ 6)-linked glucosamine substituted with N-acetyl and O-succinyl constituents. This polysaccharide is present in both S. aureus and S. epidermidis and can be isolated from either source (Joyce et al., 2003; Maira-Litran et al., 2002). For example, PNAG may be isolated from S. aureus strain MN8m (WO04/43407). PIA isolated from S. epidermidis is a integral constituent of biofilm. It is responsible for mediating cell-cell adhesion and probably also functions to shield the growing colony from the host's immune response. The polysaccharide previously known as poly-N-succinyl-β-(1→6)-glucosamine (PNSG) recently shown not to have the expected structure since the identification of N-succinylation was incorrect (Maira-Litran et al., 2002). Therefore the polysaccharide formally known as PNSG and now found to be PNAG is also encompassed by the term PIA.

[0211] PIA (or PNAG) may be of different sizes varying from over 400 kDa to between 75 and 400 kDa to between 10 and 75 kDa to oligosaccharides composed of up to 30 repeat units (of  $\beta$ -(1 $\rightarrow$ 6)-linked glucosamine substituted with N-acetyl and O-succinyl constituents). Any size of PIA polysaccharide or oligosaccharide may be use in an immunogenic composition of the invention, in one aspect the polysaccharide is over 40 kDa. Sizing may be achieved by any method known in the art, for instance by microfluidization, ultrasonic irradiation or by chemical cleavage (WO 03/53462, EP497524, EP497525). In certain aspects PIA (PNAG) is at least or at most 40-400 kDa, 40-300 kDa, 50-350 kDa, 60-300 kDa, 50-250 kDa and 60-200 kDa.

[0212] PIA (PNAG) can have different degree of acetylation due to substitution on the amino groups by acetate. PIA produced in vitro is almost fully substituted on amino groups (95-100%). Alternatively, a deacetylated PIA (PNAG) can be used having less than 60%, 50%, 40%, 30%, 20%, 10% acetylation. Use of a deacetylated PIA (PNAG) is preferred since non-acetylated epitopes of PNAG are efficient at mediating opsonic killing of Gram positive bacteria, preferably S. aureus and/or S. epidermidis. In certain aspects, the PIA (PNAG) has a size between 40 kDa and 300 kDa and is deacetylated so that less than 60%, 50%, 40%, 30% or 20% of amino groups are acetylated.

[0213] The term deacetylated PNAG (dPNAG) refers to a PNAG polysaccharide or oligosaccharide in which less than 60%, 50%, 40%, 30%, 20% or 10% of the amino agroups are acetylated. In certain aspects, PNAG is deaceylated to form dPNAG by chemically treating the native polysaccharide. For example, the native PNAG is treated with a basic solution such that the pH rises to above 10. For instance the PNAG is treated with 0.1-5 M, 0.2-4 M, 0.3-3 M, 0.5-2 M, 0.75-1.5 M or 1 M NaOH, KOH or NH<sub>4</sub>OH. Treatment is for at least 10 to 30 minutes, or 1, 2, 3, 4, 5, 10, 15 or 20 hours at a temperature of 20-100, 25-80, 30-60 or 30-50 or 35-45° C. dPNAG may be prepared as described in WO 04/43405.

[0214] The polysaccharide(s) can be conjugated or unconjugated to a carrier protein.

[0215] B. Type 5 and Type 8 Polysaccharides from S. aureus

**[0216]** Most strains of *S. aureus* that cause infection in man contain either Type 5 or Type 8 polysaccharides. Approximately 60% of human strains are Type 8 and approximately 30% are Type 5. The structures of Type 5 and Type 8 capsular polysaccharide antigens are described in Moreau et al., (1990) and Fournier et al., (1984). Both have FucNAcp in their repeat unit as well as ManNAcA which can be used to introduce a sulfhydryl group. The structures are:

[**0217**] Type 5

[0218]  $\rightarrow$ 4)- $\beta$ -D-ManNAcA(3OAc)-(1 $\rightarrow$ 4)- $\alpha$ -L-FucNAc (1 $\rightarrow$ 3)- $\beta$ -D-FucNAc-(1 $\rightarrow$ 

[0219] Type 8

[0220]  $\rightarrow$ 3)- $\beta$ -D-ManNAcA(4OAc)-(1 $\rightarrow$ 3)- $\alpha$ -L-FucNAc (1 $\rightarrow$ 3)- $\beta$ -D-FucNAc-(1 $\rightarrow$ 

[0221] Recently (Jones, 2005) NMR spectroscopy revised the structures to:

[0222] Type 5

[0223]  $\rightarrow$ 4)- $\beta$ -D-ManNAcA-(1 $\rightarrow$ 4)- $\alpha$ -L-FucNAc

(3OAc)- $(1\rightarrow 3)$ - $\beta$ -D-FucNAc- $(1\rightarrow$ 

[0224] Type 8

[0225]  $\rightarrow$ 3)- $\beta$ -D-ManNAcA(4OAc)-(1 $\rightarrow$ 3)- $\alpha$ -L-FucNAc (1 $\rightarrow$ 3)- $\alpha$ -D-FucNAc(1 $\rightarrow$ 

**[0226]** Polysaccharides may be extracted from the appropriate strain of *S. aureus* using method well known to of skill in the art, See U.S. Pat. No. 6,294,177. For example, ATCC 12902 is a Type 5 *S. aureus* strain and ATCC 12605 is a Type 8 *S. aureus* strain.

**[0227]** Polysaccharides are of native size or alternatively may be sized, for instance by microfluidisation, ultrasonic irradiation, or by chemical treatment. The invention also covers oligosaccharides derived from the type 5 and 8 polysaccharides from *S. aureus*. The type 5 and 8 polysaccharides included in the immunogenic composition of the invention are preferably conjugated to a carrier protein as described below or are alternatively unconjugated. The immunogenic compositions of the invention alternatively contains either type 5 or type 8 polysaccharide.

[0228] C. S. aureus 336 Antigen

[0229] In an embodiment, the immunogenic composition of the invention comprises the

[0230] S. aureus 336 antigen described in U.S. Pat. No. 6,294,177. The 336 antigen comprises f3-linked hexosamine, contains no O-acetyl groups, and specifically binds to antibodies to S. aureus Type 336 deposited under ATCC 55804. In an embodiment, the 336 antigen is a polysaccharide which is of native size or alternatively may be sized, for instance by microfluidisation, ultrasonic irradiation, or by chemical treatment. The invention also covers oligosaccharides derived from the 336 antigen. The 336 antigen can be unconjugated or conjugated to a carrier protein.

[0231] D. Type I, II and III Polysaccharides from S. epider-midis

[0232] Amongst the problems associated with the use of polysaccharides in vaccination, is the fact that polysaccharides per se are poor immunogens. It is preferred that the polysaccharides utilized in the invention are linked to a protein carrier which provide bystander T-cell help to improve immunogenicity. Examples of such carriers which may be conjugated to polysaccharide immunogens include the Diphtheria and Tetanus toxoids (DT, DT CRM197 and TT respec-

tively), Keyhole Limpet Haemocyanin (KLH), and the purified protein derivative of Tuberculin (PPD), *Pseudomonas aeruginosa* exoprotein A (rEPA), protein D from *Haemophilus influenzae*, pneumolysin or fragments of any of the above. Fragments suitable for use include fragments encompassing T-helper epitopes. In particular the protein D fragment from H. influenza will preferably contain the N-terminal ½ of the protein. Protein D is an IgD-binding protein from *Haemophilus influenzae* (EP 0 594 610 B1) and is a potential immunogen. In addition, staphylococcal proteins may be used as a carrier protein in the polysaccharide conjugates of the invention.

[0233] A carrier protein that would be particularly advantageous to use in the context of a staphylococcal vaccine is staphylococcal alpha toxoid. The native form may be conjugated to a polysaccharide since the process of conjugation reduces toxicity. Preferably genetically detoxified alpha toxins such as the His35Leu or His35Arg variants are used as carriers since residual toxicity is lower. Alternatively the alpha toxin is chemically detoxified by treatment with a cross-linking reagent, formaldehyde or glutaraldehyde. A genetically detoxified alpha toxin is optionally chemically detoxified, preferably by treatment with a cross-linking reagent, formaldehyde or glutaraldehyde to further reduce toxicity.

[0234] The polysaccharides may be linked to the carrier protein(s) by any known method (for example those methods described in U.S. Pat. Nos. 4,372,945, 4,474,757, and 4,356, 170). Preferably, CDAP conjugation chemistry is carried out (see WO95/08348). In CDAP, the cyanylating reagent 1-cyano-dimethylaminopyridinium tetrafluoroborate (CDAP) is preferably used for the synthesis of polysaccharide-protein conjugates. The cyanilation reaction can be performed under relatively mild conditions, which avoids hydrolysis of the alkaline sensitive polysaccharides. This synthesis allows direct coupling to a carrier protein.

[0235] Conjugation preferably involves producing a direct linkage between the carrier protein and polysaccharide. Optionally a spacer (such as adipic dihydride (ADH)) may be introduced between the carrier protein and the polysaccharide.

### IV. Immune Response and Assays

[0236] As discussed above, the invention concerns evoking or inducing an immune response in a subject against a variant SpA or coagulase peptide. In one embodiment, the immune response can protect against or treat a subject having, suspected of having, or at risk of developing an infection or related disease, particularly those related to staphylococci. One use of the immunogenic compositions of the invention is to prevent nosocomial infections by inoculating a subject prior to undergoing procedures in a hospital or other environment having an increased risk of infection.

### [0237] A. Immunoassays

[0238] The present invention includes the implementation of serological assays to evaluate whether and to what extent an immune response is induced or evoked by compositions of the invention. There are many types of immunoassays that can be implemented. Immunoassays encompassed by the present invention include, but are not limited to, those described in U.S. Pat. No. 4,367,110 (double monoclonal antibody sandwich assay) and U.S. Pat. No. 4,452,901 (west-

ern blot). Other assays include immunoprecipitation of labeled ligands and immunocytochemistry, both in vitro and in vivo.

[0239] Immunoassays generally are binding assays. Certain preferred immunoassays are the various types of enzyme linked immunosorbent assays (ELISAs) and radioimmunoassays (RIA) known in the art. Immunohistochemical detection using tissue sections is also particularly useful. In one example, antibodies or antigens are immobilized on a selected surface, such as a well in a polystyrene microtiter plate, dipstick, or column support. Then, a test composition suspected of containing the desired antigen or antibody, such as a clinical sample, is added to the wells. After binding and washing to remove non specifically bound immune complexes, the bound antigen or antibody may be detected. Detection is generally achieved by the addition of another antibody, specific for the desired antigen or antibody, that is linked to a detectable label. This type of ELISA is known as a "sandwich ELISA." Detection also may be achieved by the addition of a second antibody specific for the desired antigen, followed by the addition of a third antibody that has binding affinity for the second antibody, with the third antibody being linked to a detectable label.

[0240] Competition ELISAs are also possible implementations in which test samples compete for binding with known amounts of labeled antigens or antibodies. The amount of reactive species in the unknown sample is determined by mixing the sample with the known labeled species before or during incubation with coated wells. The presence of reactive species in the sample acts to reduce the amount of labeled species available for binding to the well and thus reduces the ultimate signal. Irrespective of the format employed, ELISAs have certain features in common, such as coating, incubating or binding, washing to remove non specifically bound species, and detecting the bound immune complexes.

[0241] Antigen or antibodies may also be linked to a solid support, such as in the form of plate, beads, dipstick, membrane, or column matrix, and the sample to be analyzed is applied to the immobilized antigen or antibody. In coating a plate with either antigen or antibody, one will generally incubate the wells of the plate with a solution of the antigen or antibody, either overnight or for a specified period. The wells of the plate will then be washed to remove incompletelyadsorbed material. Any remaining available surfaces of the wells are then "coated" with a nonspecific protein that is antigenically neutral with regard to the test antisera. These include bovine serum albumin (BSA), casein, and solutions of milk powder. The coating allows for blocking of nonspecific adsorption sites on the immobilizing surface and thus reduces the background caused by nonspecific binding of antisera onto the surface.

[0242] B. Diagnosis of Bacterial Infection

[0243] In addition to the use of proteins, polypeptides, and/ or peptides, as well as antibodies binding these polypeptides, proteins, and/or peptides, to treat or prevent infection as described above, the present invention contemplates the use of these polypeptides, proteins, peptides, and/or antibodies in a variety of ways, including the detection of the presence of Staphylococci to diagnose an infection, whether in a patient or on medical equipment which may also become infected. In accordance with the invention, a preferred method of detecting the presence of infections involves the steps of obtaining a sample suspected of being infected by one or more staphylococcal bacteria species or strains, such as a sample taken

from an individual, for example, from one's blood, saliva, tissues, bone, muscle, cartilage, or skin. Following isolation of the sample, diagnostic assays utilizing the polypeptides, proteins, peptides, and/or antibodies of the present invention may be carried out to detect the presence of staphylococci, and such assay techniques for determining such presence in a sample are well known to those skilled in the art and include methods such as radioimmunoassay, western blot analysis and ELISA assays. In general, in accordance with the invention, a method of diagnosing an infection is contemplated wherein a sample suspected of being infected with staphylococci has added to it the polypeptide, protein, peptide, antibody, or monoclonal antibody in accordance with the present invention, and staphylococci are indicated by antibody binding to the polypeptides, proteins, and/or peptides, or polypeptides, proteins, and/or peptides binding to the antibodies in the sample.

[0244] Accordingly, antibodies in accordance with the invention may be used for the prevention of infection from staphylococcal bacteria (i.e., passive immunization), for the treatment of an ongoing infection, or for use as research tools. The term "antibodies" as used herein includes monoclonal, polyclonal, chimeric, single chain, bispecific, simianized, and humanized or primatized antibodies as well as Fab fragments, such as those fragments which maintain the binding specificity of the antibodies, including the products of an Fab immunoglobulin expression library. Accordingly, the invention contemplates the use of single chains such as the variable heavy and light chains of the antibodies. Generation of any of these types of antibodies or antibody fragments is well known to those skilled in the art. Specific examples of the generation of an antibody to a bacterial protein can be found in U.S. Patent Application Pub. No. 20030153022, which is incorporated herein by reference in its entirety.

[0245] Any of the above described polypeptides, proteins, peptides, and/or antibodies may be labeled directly with a detectable label for identification and quantification of staphylococcal bacteria. Labels for use in immunoassays are generally known to those skilled in the art and include enzymes, radioisotopes, and fluorescent, luminescent and chromogenic substances, including colored particles such as colloidal gold or latex beads. Suitable immunoassays include enzyme-linked immunosorbent assays (ELISA).

[0246] C. Protective Immunity

[0247] In some embodiments of the invention, proteinaceous compositions confer protective immunity to a subject. Protective immunity refers to a body's ability to mount a specific immune response that protects the subject from developing a particular disease or condition that involves the agent against which there is an immune response. An immunogenically effective amount is capable of conferring protective immunity to the subject.

[0248] As used herein in the specification and in the claims section that follows, the term polypeptide or peptide refer to a stretch of amino acids covalently linked there amongst via peptide bonds. Different polypeptides have different functionalities according to the present invention. While according to one aspect, a polypeptide is derived from an immunogen designed to induce an active immune response in a recipient, according to another aspect of the invention, a polypeptide is derived from an antibody which results following the elicitation of an active immune response in, for example, an animal, and which can serve to induce a passive

immune response in the recipient. In both cases, however, the polypeptide is encoded by a polynucleotide according to any possible codon usage.

[0249] As used herein the phrase "immune response" or its equivalent "immunological response" refers to the development of a humoral (antibody mediated), cellular (mediated by antigen-specific T cells or their secretion products) or both humoral and cellular response directed against a protein, peptide, carbohydrate, or polypeptide of the invention in a recipient patient. Such a response can be an active response induced by administration of immunogen or a passive response induced by administration of antibody, antibody containing material, or primed T-cells. A cellular immune response is elicited by the presentation of polypeptide epitopes in association with Class I or Class II MHC molecules, to activate antigen-specific CD4 (+) T helper cells and/or CD8 (+) cytotoxic T cells. The response may also involve activation of monocytes, macrophages, NK cells, basophils, dendritic cells, astrocytes, microglia cells, eosinophils or other components of innate immunity. As used herein "active immunity" refers to any immunity conferred upon a subject by administration of an antigen.

[0250] As used herein "passive immunity" refers to any immunity conferred upon a subject without administration of an antigen to the subject. "Passive immunity" therefore includes, but is not limited to, administration of activated immune effectors including cellular mediators or protein mediators (e.g., monoclonal and/or polyclonal antibodies) of an immune response. A monoclonal or polyclonal antibody composition may be used in passive immunization for the prevention or treatment of infection by organisms that carry the antigen recognized by the antibody. An antibody composition may include antibodies that bind to a variety of antigens that may in turn be associated with various organisms. The antibody component can be a polyclonal antiserum. In certain aspects the antibody or antibodies are affinity purified from an animal or second subject that has been challenged with an antigen(s). Alternatively, an antibody mixture may be used, which is a mixture of monoclonal and/or polyclonal antibodies to antigens present in the same, related, or different microbes or organisms, such as gram-positive bacteria, gramnegative bacteria, including but not limited to staphylococcus bacteria.

[0251] Passive immunity may be imparted to a patient or subject by administering to the patient immunoglobulins (Ig) and/or other immune factors obtained from a donor or other non-patient source having a known immunoreactivity. In other aspects, an antigenic composition of the present invention can be administered to a subject who then acts as a source or donor for globulin, produced in response to challenge with the antigenic composition ("hyperimmune globulin"), that contains antibodies directed against Staphylococcus or other organism. A subject thus treated would donate plasma from which hyperimmune globulin would then be obtained, via conventional plasma-fractionation methodology, and administered to another subject in order to impart resistance against or to treat staphylococcus infection. Hyperimmune globulins according to the invention are particularly useful for immunecompromised individuals, for individuals undergoing invasive procedures or where time does not permit the individual to produce their own antibodies in response to vaccination. See U.S. Pat. Nos. 6,936,258, 6,770,278, 6,756,361, 5,548, 066, 5,512,282, 4,338,298, and 4,748,018, each of which is incorporated herein by reference in its entirety, for exemplary methods and compositions related to passive immunity.

[0252] For purposes of this specification and the accompanying claims the terms "epitope" and "antigenic determinant" are used interchangeably to refer to a site on an antigen to which B and/or T cells respond or recognize. B-cell epitopes can be formed both from contiguous amino acids or noncontiguous amino acids juxtaposed by tertiary folding of a protein. Epitopes formed from contiguous amino acids are typically retained on exposure to denaturing solvents whereas epitopes formed by tertiary folding are typically lost on treatment with denaturing solvents. An epitope typically includes at least 3, and more usually, at least 5 or 8-10 amino acids in a unique spatial conformation. Methods of determining spatial conformation of epitopes include, for example, x-ray crystallography and 2-dimensional nuclear magnetic resonance. See, e.g., Epitope Mapping Protocols (1996). Antibodies that recognize the same epitope can be identified in a simple immunoassay showing the ability of one antibody to block the binding of another antibody to a target antigen. T-cells recognize continuous epitopes of about nine amino acids for CD8 cells or about 13-15 amino acids for CD4 cells. T cells that recognize the epitope can be identified by in vitro assays that measure antigen-dependent proliferation, as determined by <sup>3</sup>H-thymidine incorporation by primed T cells in response to an epitope (Burke et al., 1994), by antigendependent killing (cytotoxic T lymphocyte assay, Tigges et al., 1996) or by cytokine secretion.

[0253] The presence of a cell-mediated immunological response can be determined by proliferation assays (CD4 (+) T cells) or CTL (cytotoxic T lymphocyte) assays. The relative contributions of humoral and cellular responses to the protective or therapeutic effect of an immunogen can be distinguished by separately isolating IgG and T-cells from an immunized syngeneic animal and measuring protective or therapeutic effect in a second subject.

[0254] As used herein and in the claims, the terms "antibody" or "immunoglobulin" are used interchangeably and refer to any of several classes of structurally related proteins that function as part of the immune response of an animal or recipient, which proteins include IgG, IgD, IgE, IgA, IgM and related proteins.

[0255] Under normal physiological conditions antibodies are found in plasma and other body fluids and in the membrane of certain cells and are produced by lymphocytes of the type denoted B cells or their functional equivalent. Antibodies of the IgG class are made up of four polypeptide chains linked together by disulfide bonds. The four chains of intact IgG molecules are two identical heavy chains referred to as H-chains and two identical light chains referred to as L-chains.

[0256] In order to produce polyclonal antibodies, a host, such as a rabbit or goat, is immunized with the antigen or antigen fragment, generally with an adjuvant and, if necessary, coupled to a carrier. Antibodies to the antigen are subsequently collected from the sera of the host. The polyclonal antibody can be affinity purified against the antigen rendering it monospecific.

[0257] Monoclonal antibodies can be produced by hyperimmunization of an appropriate donor with the antigen or ex-vivo by use of primary cultures of splenic cells or cell lines derived from spleen (Anavi, 1998; Huston et al., 1991; Johnson et al., 1991; Mernaugh et al., 1995). [0258] As used herein and in the claims, the phrase "an immunological portion of an antibody" includes a Fab fragment of an antibody, a Fv fragment of an antibody, a heavy chain of an antibody, a light chain of an antibody, a heterodimer consisting of a heavy chain and a light chain of an antibody, a variable fragment of a light chain of an antibody, a variable fragment of a light chain of an antibody, a variable fragment of an antibody, which is also known as scFv. In addition, the term includes chimeric immunoglobulins which are the expression products of fused genes derived from different species, one of the species can be a human, in which case a chimeric immunoglobulin is said to be humanized. Typically, an immunological portion of an antibody competes with the intact antibody from which it was derived for specific binding to an antigen.

[0259] Optionally, an antibody or preferably an immunological portion of an antibody, can be chemically conjugated to, or expressed as, a fusion protein with other proteins. For purposes of this specification and the accompanying claims, all such fused proteins are included in the definition of antibodies or an immunological portion of an antibody.

[0260] As used herein the terms "immunogenic agent" or "immunogen" or "antigen" are used interchangeably to describe a molecule capable of inducing an immunological response against itself on administration to a recipient, either alone, in conjunction with an adjuvant, or presented on a display vehicle.

[0261] D. Treatment Methods

[0262] A method of the present invention includes treatment for a disease or condition caused by a *staphylococcus* pathogen. An immunogenic polypeptide of the invention can be given to induce an immune response in a person infected with *staphylococcus* or suspected of having been exposed to *staphylococcus*. Methods may be employed with respect to individuals who have tested positive for exposure to *staphylococcus* or who are deemed to be at risk for infection based on possible exposure.

[0263] In particular, the invention encompasses a method of treatment for staphylococcal infection, particularly hospital acquired nosocomial infections. The immunogenic compositions and vaccines of the invention are particularly advantageous to use in cases of elective surgery. Such patients will know the date of surgery in advance and could be inoculated in advance. The immunogenic compositions and vaccines of the invention are also advantageous to use to inoculate health care workers.

[0264] In some embodiments, the treatment is administered in the presence of adjuvants or carriers or other staphylococcal antigens. Furthermore, in some examples, treatment comprises administration of other agents commonly used against bacterial infection, such as one or more antibiotics.

[0265] The use of peptides for vaccination can require, but not necessarily, conjugation of the peptide to an immunogenic carrier protein, such as hepatitis B surface antigen, keyhole limpet hemocyanin, or bovine serum albumin. Methods for performing this conjugation are well known in the art.

V. Vaccine and Other Pharmaceutical Compositions and Administration

[0266] A. Vaccines

[0267] The present invention includes methods for preventing or ameliorating staphylococcal infections, particularly hospital acquired nosocomial infections. As such, the invention contemplates vaccines for use in both active and passive

immunization embodiments. Immunogenic compositions, proposed to be suitable for use as a vaccine, may be prepared from immunogenic SpA polypeptide(s), such as a SpA domain D variant, or immunogenic coagulases. In other embodiments SpA or coagulases can be used in combination with other secreted virulence proteins, surface proteins or immunogenic fragments thereof. In certain aspects, antigenic material is extensively dialyzed to remove undesired small molecular weight molecules and/or lyophilized for more ready formulation into a desired vehicle.

[0268] Other options for a protein/peptide-based vaccine involve introducing nucleic acids encoding the antigen(s) as DNA vaccines. In this regard, recent reports described construction of recombinant vaccinia viruses expressing either 10 contiguous minimal CTL epitopes (Thomson, 1996) or a combination of B cell, cytotoxic T-lymphocyte (CTL), and T-helper (Th) epitopes from several microbes (An, 1997), and successful use of such constructs to immunize mice for priming protective immune responses. Thus, there is ample evidence in the literature for successful utilization of peptides, peptide-pulsed antigen presenting cells (APCs), and peptide-encoding constructs for efficient in vivo priming of protective immune responses. The use of nucleic acid sequences as vaccines is exemplified in U.S. Pat. Nos. 5,958,895 and 5,620,

[0269] The preparation of vaccines that contain polypeptide or peptide sequence(s) as active ingredients is generally well understood in the art, as exemplified by U.S. Pat. Nos. 4,608,251; 4,601,903; 4,599,231; 4,599,230; 4,596,792; and 4,578,770, all of which are incorporated herein by reference. Typically, such vaccines are prepared as injectables either as liquid solutions or suspensions: solid forms suitable for solution in or suspension in liquid prior to injection may also be prepared. The preparation may also be emulsified. The active immunogenic ingredient is often mixed with excipients that are pharmaceutically acceptable and compatible with the active ingredient. Suitable excipients are, for example, water, saline, dextrose, glycerol, ethanol, or the like and combinations thereof. In addition, if desired, the vaccine may contain amounts of auxiliary substances such as wetting or emulsifying agents, pH buffering agents, or adjuvants that enhance the effectiveness of the vaccines. In specific embodiments, vaccines are formulated with a combination of substances, as described in U.S. Pat. Nos. 6,793,923 and 6,733,754, which are incorporated herein by reference.

[0270] Vaccines may be conventionally administered parenterally, by injection, for example, either subcutaneously or intramuscularly. Additional formulations which are suitable for other modes of administration include suppositories and, in some cases, oral formulations. For suppositories, traditional binders and carriers may include, for example, polyalkalene glycols or triglycerides: such suppositories may be formed from mixtures containing the active ingredient in the range of about 0.5% to about 10%, preferably about 1% to about 2%. Oral formulations include such normally employed excipients as, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate and the like. These compositions take the form of solutions, suspensions, tablets, pills, capsules, sustained release formulations or powders and contain about 10% to about 95% of active ingredient, preferably about 25% to about 70%.

[0271] The polypeptides and polypeptide-encoding DNA constructs may be formulated into a vaccine as neutral or salt

forms. Pharmaceutically-acceptable salts include the acid addition salts (formed with the free amino groups of the peptide) and those that are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. [0272] Typically, vaccines are administered in a manner compatible with the dosage formulation, and in such amount as will be therapeutically effective and immunogenic. The quantity to be administered depends on the subject to be treated, including the capacity of the individual's immune system to synthesize antibodies and the degree of protection desired. Precise amounts of active ingredient required to be administered depend on the judgment of the practitioner. However, suitable dosage ranges are of the order of several hundred micrograms of active ingredient per vaccination. Suitable regimes for initial administration and booster shots are also variable, but are typified by an initial administration followed by subsequent inoculations or other administrations.

**[0273]** The manner of application may be varied widely. Any of the conventional methods for administration of a vaccine are applicable. These are believed to include oral application within a solid physiologically acceptable base or in a physiologically acceptable dispersion, parenterally, by injection and the like. The dosage of the vaccine will depend on the route of administration and will vary according to the size and health of the subject.

[0274] In certain instances, it will be desirable to have multiple administrations of the vaccine, e.g., 2, 3, 4, 5, 6 or more administrations. The vaccinations can be at 1, 2, 3, 4, 5, 6, 7, 8, to 5, 6, 7, 8, 9, 10, 11, 12 twelve week intervals, including all ranges there between. Periodic boosters at intervals of 1-5 years will be desirable to maintain protective levels of the antibodies. The course of the immunization may be followed by assays for antibodies against the antigens, as described in U.S. Pat. Nos. 3,791,932; 4,174,384 and 3,949, 064

[0275] 1. Carriers

[0276] A given composition may vary in its immunogenicity. It is often necessary therefore to boost the host immune system, as may be achieved by coupling a peptide or polypeptide to a carrier. Exemplary and preferred carriers are keyhole limpet hemocyanin (KLH) and bovine serum albumin (BSA). Other albumins such as ovalbumin, mouse serum albumin, or rabbit serum albumin can also be used as carriers. Means for conjugating a polypeptide to a carrier protein are well known in the art and include glutaraldehyde, m-maleimidobencoyl-N-hydroxysuccinimide ester, carbodiimyde, and bis-biazotized benzidine.

[0277] 2. Adjuvants

[0278] The immunogenicity of polypeptide or peptide compositions can be enhanced by the use of non-specific stimulators of the immune response, known as adjuvants. Suitable adjuvants include all acceptable immunostimulatory compounds, such as cytokines, toxins, or synthetic compositions. A number of adjuvants can be used to enhance an antibody response against a variant SpA polypeptide or coagulase, or any other bacterial protein or combination contemplated herein. Adjuvants can (1) trap the antigen in the body to cause a slow release; (2) attract cells involved in the immune response to the site of administration; (3) induce proliferation or activation of immune system cells; or (4) improve the spread of the antigen throughout the subject's body.

[0279] Adjuvants include, but are not limited to, oil-in-water emulsions, water-in-oil emulsions, mineral salts, polynucleotides, and natural substances. Specific adjuvants that may be used include IL-1, IL-2, IL-4, IL-7, IL-12, γ-interferon, GMCSP, BCG, aluminum salts, such as aluminum hydroxide or other aluminum compound, MDP compounds, such as thur-MDP and nor-MDP, CGP (MTP-PE), lipid A, and monophosphoryl lipid A (MPL). RIBI, which contains three components extracted from bacteria, MPL, trehalose dimycolate (TDM), and cell wall skeleton (CWS) in a 2% squalene/Tween 80 emulsion. MHC antigens may even be used. Others adjuvants or methods are exemplified in U.S. Pat. Nos. 6,814,971, 5,084,269, 6,656,462, each of which is incorporated herein by reference).

[0280] Various methods of achieving adjuvant affect for the vaccine includes use of agents such as aluminum hydroxide or phosphate (alum), commonly used as about 0.05 to about 0.1% solution in phosphate buffered saline, admixture with synthetic polymers of sugars (Carbopol®) used as an about 0.25% solution, aggregation of the protein in the vaccine by heat treatment with temperatures ranging between about 70° to about 101° C. for a 30-second to 2-minute period, respectively. Aggregation by reactivating with pepsin-treated (Fab) antibodies to albumin; mixture with bacterial cells (e.g., C. parvum), endotoxins or lipopolysaccharide components of Gram-negative bacteria; emulsion in physiologically acceptable oil vehicles (e.g., mannide mono-oleate (Aracel A)); or emulsion with a 20% solution of a perfluorocarbon (Fluosol-DA®) used as a block substitute may also be employed to produce an adjuvant effect.

[0281] Examples of and often preferred adjuvants include complete Freund's adjuvant (a non-specific stimulator of the immune response containing killed *Mycobacterium tuberculosis*), incomplete Freund's adjuvants, and aluminum hydroxide.

[0282] In some aspects, it is preferred that the adjuvant be selected to be a preferential inducer of either a Th1 or a Th2 type of response. High levels of Th1-type cytokines tend to favor the induction of cell mediated immune responses to a given antigen, while high levels of Th2-type cytokines tend to favor the induction of humoral immune responses to the antigen

[0283] The distinction of Th1 and Th2-type immune response is not absolute. In reality an individual will support an immune response which is described as being predominantly Th1 or predominantly Th2. However, it is often convenient to consider the families of cytokines in terms of that described in murine CD4+ T cell clones by Mosmann and Coffman (Mosmann, and Coffman, 1989). Traditionally, Th1-type responses are associated with the production of the INF- $\gamma$  and IL-2 cytokines by T-lymphocytes. Other cytokines often directly associated with the induction of Th1-type immune responses are not produced by T-cells, such as IL-12. In contrast, Th2-type responses are associated with the secretion of IL-4, IL-5, IL-6, IL-10.

[0284] In addition to adjuvants, it may be desirable to co-administer biologic response modifiers (BRM) to enhance immune responses. BRMs have been shown to upregulate T cell immunity or downregulate suppresser cell activity. Such BRMs include, but are not limited to, Cimetidine (CIM; 1200 mg/d) (Smith/Kline, PA); or low-dose Cyclophosphamide (CYP; 300 mg/m²) (Johnson/Mead, NJ) and cytokines such as γ-interferon, IL-2, or IL-12 or genes encoding proteins involved in immune helper functions, such as B-7.

[0285] B. Lipid Components and Moieties

[0286] In certain embodiments, the present invention concerns compositions comprising one or more lipids associated with a nucleic acid or a polypeptide/peptide. A lipid is a substance that is insoluble in water and extractable with an organic solvent. Compounds other than those specifically described herein are understood by one of skill in the art as lipids, and are encompassed by the compositions and methods of the present invention. A lipid component and a non-lipid may be attached to one another, either covalently or non-covalently.

[0287] A lipid may be a naturally occurring lipid or a synthetic lipid. However, a lipid is usually a biological substance. Biological lipids are well known in the art, and include for example, neutral fats, phospholipids, phosphoglycerides, steroids, terpenes, lysolipids, glycosphingolipids, glucolipids, sulphatides, lipids with ether and ester-linked fatty acids and polymerizable lipids, and combinations thereof.

[0288] A nucleic acid molecule or a polypeptide/peptide, associated with a lipid may be dispersed in a solution containing a lipid, dissolved with a lipid, emulsified with a lipid, mixed with a lipid, combined with a lipid, covalently bonded to a lipid, contained as a suspension in a lipid or otherwise associated with a lipid. A lipid or lipid-poxvirus-associated composition of the present invention is not limited to any particular structure. For example, they may also simply be interspersed in a solution, possibly forming aggregates which are not uniform in either size or shape. In another example, they may be present in a bilayer structure, as micelles, or with a "collapsed" structure. In another non-limiting example, a lipofectamine (Gibco BRL)-poxvirus or Superfect (Qiagen)-poxvirus complex is also contemplated.

[0289] In certain embodiments, a composition may comprise about 1%, about 2%, about 3%, about 4% about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, about 20%, about 21%, about 22%, about 23%, about 24%, about 25%, about 26%, about 27%, about 28%, about 29%, about 30%, about 31%, about 32%, about 33%, about 34%, about 35%, about 36%, about 37%, about 38%, about 39%, about 40%, about 41%, about 42%, about 43%, about 44%, about 45%, about 46%, about 47%, about 48%, about 49%, about 50%, about 51%, about 52%, about 53%, about 54%, about 55%, about 56%, about 57%, about 58%, about 59%, about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or any range therebetween, of a particular lipid, lipid type, or non-lipid component such as an adjuvant, antigen, peptide, polypeptide, sugar, nucleic acid or other material disclosed herein or as would be known to one of skill in the art. In a non-limiting example, a composition may comprise about 10% to about 20% neutral lipids, and about 33% to about 34% of a cerebroside, and about 1% cholesterol. In another non-limiting example, a liposome may comprise about 4% to about 12% terpenes, wherein about 1% of the micelle is specifically lycopene, leaving about 3% to about 11% of the liposome as comprising other terpenes; and about 10% to about 35% phosphatidyl choline, and about 1% of a non-lipid component. Thus, it is contemplated that compositions of the present invention may comprise any of the lipids, lipid types or other components in any combination or percentage range.

[0290] C. Combination Therapy

[0291] The compositions and related methods of the present invention, particularly administration of a secreted virulence factor or surface protein, including a variant SpA polypeptide or peptide, and/or other bacterial peptides or proteins to a patient/subject, may also be used in combination with the administration of traditional therapies. These include, but are not limited to, the administration of antibiotics such as streptomycin, ciprofloxacin, doxycycline, gentamycin, chloramphenicol, trimethoprim, sulfamethoxazole, ampicillin, tetracycline or various combinations of antibiotics.

[0292] In one aspect, it is contemplated that a polypeptide vaccine and/or therapy is used in conjunction with antibacterial treatment. Alternatively, the therapy may precede or follow the other agent treatment by intervals ranging from minutes to weeks. In embodiments where the other agents and/or a proteins or polynucleotides are administered separately, one would generally ensure that a significant period of time did not expire between the time of each delivery, such that the agent and antigenic composition would still be able to exert an advantageously combined effect on the subject. In such instances, it is contemplated that one may administer both modalities within about 12-24 h of each other or within about 6-12 h of each other. In some situations, it may be desirable to extend the time period for administration significantly, where several days (2, 3, 4, 5, 6 or 7) to several weeks (1, 2, 3, 4, 5, 6, 7 or 8) lapse between the respective administrations.

[0293] Various combinations may be employed, for example antibiotic therapy is "A" and the immunogenic molecule given as part of an immune therapy regime, such as an antigen, is "B":

 ${\tt A/B/A~B/A/B~B/B/A~A/A/B~A/B/B~B/A/A~A/B/B/B}$ 

B/A/B/B B/B/B/A B/B/A/B A/A/B/B A/B/A/B

A/B/B/A B/B/A/A B/A/B/A B/A/A/B A/A/A/B

B/A/A/A A/B/A/A A/A/B/A

[0294] Administration of the immunogenic compositions of the present invention to a patient/subject will follow general protocols for the administration of such compounds, taking into account the toxicity, if any, of the SpA composition, or other compositions described herein. It is expected that the treatment cycles would be repeated as necessary. It also is contemplated that various standard therapies, such as hydration, may be applied in combination with the described therapy.

[0295] D. General Pharmaceutical Compositions

[0296] In some embodiments, pharmaceutical compositions are administered to a subject. Different aspects of the present invention involve administering an effective amount of a composition to a subject. In some embodiments of the present invention, staphylococcal antigens, members of the Ess pathway, including polypeptides or peptides of the Esa or Esx class, and/or members of sortase substrates may be administered to the patient to protect against infection by one or more *staphylococcus* pathogens. Alternatively, an expression vector encoding one or more such polypeptides or pep-

tides may be given to a patient as a preventative treatment. Additionally, such compounds can be administered in combination with an antibiotic or an antibacterial. Such compositions will generally be dissolved or dispersed in a pharmaceutically acceptable carrier or aqueous medium.

[0297] In addition to the compounds formulated for parenteral administration, such as those for intravenous or intramuscular injection, other pharmaceutically acceptable forms include, e.g., tablets or other solids for oral administration; time release capsules; and any other form currently used, including creams, lotions, mouthwashes, inhalants and the like.

[0298] The active compounds of the present invention can be formulated for parenteral administration, e.g., formulated for injection via the intravenous, intramuscular, sub-cutaneous, or even intraperitoneal routes. The preparation of an aqueous composition that contains a compound or compounds that increase the expression of an MHC class I molecule will be known to those of skill in the art in light of the present disclosure. Typically, such compositions can be prepared as injectables, either as liquid solutions or suspensions; solid forms suitable for use to prepare solutions or suspensions upon the addition of a liquid prior to injection can also be prepared; and, the preparations can also be emulsified.

[0299] Solutions of the active compounds as free base or pharmacologically acceptable salts can be prepared in water suitably mixed with a surfactant, such as hydroxypropylcellulose. Dispersions can also be prepared in glycerol, liquid polyethylene glycols, and mixtures thereof and in oils. Under ordinary conditions of storage and use, these preparations contain a preservative to prevent the growth of microorganisms

[0300] The pharmaceutical forms suitable for injectable use include sterile aqueous solutions or dispersions; formulations including sesame oil, peanut oil, or aqueous propylene glycol; and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions. In all cases the form must be sterile and must be fluid to the extent that it may be easily injected. It also should be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms, such as bacteria and fungi.

[0301] The proteinaceous compositions may be formulated into a neutral or salt form. Pharmaceutically acceptable salts, include the acid addition salts (formed with the free amino groups of the protein) and which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups can also be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, histidine, procaine and the like.

[0302] The carrier also can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and vegetable oils. The proper fluidity can be maintained, for example, by the use of a coating, such as lecithin, by the maintenance of the required particle size in the case of dispersion, and by the use of surfactants. The prevention of the action of microorganisms can be brought about by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, and the like. In many cases, it will

be preferable to include isotonic agents, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

[0303] Sterile injectable solutions are prepared by incorporating the active compounds in the required amount in the appropriate solvent with various of the other ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the various sterilized active ingredients into a sterile vehicle which contains the basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum-drying and freeze-drying techniques, which yield a powder of the active ingredient, plus any additional desired ingredient from a previously sterile-filtered solution thereof

[0304] Administration of the compositions according to the present invention will typically be via any common route. This includes, but is not limited to oral, nasal, or buccal administration. Alternatively, administration may be by orthotopic, intradermal, subcutaneous, intramuscular, intraperitoneal, intranasal, or intravenous injection. In certain embodiments, a vaccine composition may be inhaled (e.g., U.S. Pat. No. 6,651,655, which is specifically incorporated by reference). Such compositions would normally be administered as pharmaceutically acceptable compositions that include physiologically acceptable carriers, buffers or other excipients. As used herein, the term "pharmaceutically acceptable" refers to those compounds, materials, compositions, and/or dosage forms which are, within the scope of sound medical judgment, suitable for contact with the tissues of human beings and animals without excessive toxicity, irritation, allergic response, or other problem complications commensurate with a reasonable benefit/risk ratio. The term "pharmaceutically acceptable carrier," means a pharmaceutically acceptable material, composition or vehicle, such as a liquid or solid filler, diluent, excipient, solvent or encapsulating material, involved in carrying or transporting a chemical agent.

[0305] For parenteral administration in an aqueous solution, for example, the solution should be suitably buffered, if necessary, and the liquid diluent first rendered isotonic with sufficient saline or glucose. These particular aqueous solutions are especially suitable for intravenous, intramuscular, subcutaneous, and intraperitoneal administration. In this connection, sterile aqueous media which can be employed will be known to those of skill in the art in light of the present disclosure. For example, one dosage could be dissolved in isotonic NaCl solution and either added to hypodermoclysis fluid or injected at the proposed site of infusion, (see for example, Remington's Pharmaceutical Sciences, 1990). Some variation in dosage will necessarily occur depending on the condition of the subject. The person responsible for administration will, in any event, determine the appropriate dose for the individual subject.

[0306] An effective amount of therapeutic or prophylactic composition is determined based on the intended goal. The term "unit dose" or "dosage" refers to physically discrete units suitable for use in a subject, each unit containing a predetermined quantity of the composition calculated to produce the desired responses discussed above in association with its administration, i.e., the appropriate route and regi-

men. The quantity to be administered, both according to number of treatments and unit dose, depends on the protection desired.

[0307] Precise amounts of the composition also depend on the judgment of the practitioner and are peculiar to each individual. Factors affecting dose include physical and clinical state of the subject, route of administration, intended goal of treatment (alleviation of symptoms versus cure), and potency, stability, and toxicity of the particular composition.

[0308] Upon formulation, solutions will be administered in a manner compatible with the dosage formulation and in such amount as is therapeutically or prophylactically effective. The formulations are easily administered in a variety of dosage forms, such as the type of injectable solutions described above.

[0309] E. In Vitro, Ex Vivo, or In Vivo Administration

[0310] As used herein, the term in vitro administration refers to manipulations performed on cells removed from or outside of a subject, including, but not limited to cells in culture. The term ex vivo administration refers to cells which have been manipulated in vitro, and are subsequently administered to a subject. The term in vivo administration includes all manipulations performed within a subject.

[0311] In certain aspects of the present invention, the compositions may be administered either in vitro, ex vivo, or in vivo. In certain in vitro embodiments, autologous B-lymphocyte cell lines are incubated with a virus vector of the instant invention for 24 to 48 hours or with a variant SpA and/or cogaulase and/or any other composition described herein for two hours. The transduced cells can then be used for in vitro analysis, or alternatively for ex vivo administration. U.S. Pat. Nos. 4,690,915 and 5,199,942, both incorporated herein by reference, disclose methods for ex vivo manipulation of blood mononuclear cells and bone marrow cells for use in therapeutic applications.

[0312] F. Antibodies And Passive Immunization

[0313] Another aspect of the invention is a method of preparing an immunoglobulin for use in prevention or treatment of staphylococcal infection comprising the steps of immunizing a recipient or donor with the vaccine of the invention and isolating immunoglobulin from the recipient or donor. An immunoglobulin prepared by this method is a further aspect of the invention. A pharmaceutical composition comprising the immunoglobulin of the invention and a pharmaceutically acceptable carrier is a further aspect of the invention which could be used in the manufacture of a medicament for the treatment or prevention of staphylococcal disease. A method for treatment or prevention of staphylococcal infection comprising a step of administering to a patient an effective amount of the pharmaceutical preparation of the invention is a further aspect of the invention.

[0314] Inocula for polyclonal antibody production are typically prepared by dispersing the antigenic composition in a physiologically tolerable diluent such as saline or other adjuvants suitable for human use to form an aqueous composition. An immunostimulatory amount of inoculum is administered to a mammal and the inoculated mammal is then maintained for a time sufficient for the antigenic composition to induce protective antibodies.

[0315] The antibodies can be isolated to the extent desired by well known techniques such as affinity chromatography (Harlow and Lane, 1988). Antibodies can include antiserum preparations from a variety of commonly used animals, e.g. goats, primates, donkeys, swine, horses, guinea pigs, rats or man.

[0316] An immunoglobulin produced in accordance with the present invention can include whole antibodies, antibody fragments or subfragments. Antibodies can be whole immunoglobulins of any class (e.g., IgG, IgM, IgA, IgD or IgE), chimeric antibodies or hybrid antibodies with dual specificity to two or more antigens of the invention. They may also be fragments (e.g., F(ab')2, Fab', Fab, Fv and the like) including hybrid fragments. An immunoglobulin also includes natural, synthetic, or genetically engineered proteins that act like an antibody by binding to specific antigens to form a complex.

[0317] A vaccine of the present invention can be administered to a recipient who then acts as a source of immunoglobulin, produced in response to challenge from the specific vaccine. A subject thus treated would donate plasma from which hyperimmune globulin would be obtained via conventional plasma fractionation methodology. The hyperimmune globulin would be administered to another subject in order to impart resistance against or treat staphylococcal infection. Hyperimmune globulins of the invention are particularly useful for treatment or prevention of staphylococcal disease in infants, immune compromised individuals, or where treatment is required and there is no time for the individual to produce antibodies in response to vaccination.

[0318] An additional aspect of the invention is a pharmaceutical composition comprising two of more monoclonal antibodies (or fragments thereof; preferably human or humanised) reactive against at least two constituents of the immunogenic composition of the invention, which could be used to treat or prevent infection by Gram positive bacteria, preferably staphylococci, more preferably *S. aureus* or *S. epidermidis*. Such pharmaceutical compositions comprise monoclonal antibodies that can be whole immunoglobulins of any class, chimeric antibodies, or hybrid antibodies with specificity to two or more antigens of the invention. They may also be fragments (e.g., F(ab')2, Fab', Fab, Fv and the like) including hybrid fragments.

[0319] Methods of making monoclonal antibodies are well known in the art and can include the fusion of splenocytes with myeloma cells (Kohler and Milstein, 1975; Harlow and Lane, 1988). Alternatively, monoclonal Fv fragments can be obtained by screening a suitable phage display library (Vaughan et al., 1998). Monoclonal antibodies may be humanized or part humanized by known methods.

#### VI. Examples

[0320] The following examples are given for the purpose of illustrating various embodiments of the invention and are not meant to limit the present invention in any fashion. One skilled in the art will appreciate readily that the present invention is well adapted to carry out the objects and obtain the ends and advantages mentioned, as well as those objects, ends and advantages inherent herein. The present examples, along with the methods described herein are presently representative of preferred embodiments, are exemplary, and are not intended as limitations on the scope of the invention. Changes therein and other uses which are encompassed within the spirit of the invention as defined by the scope of the claims will occur to those skilled in the art.

### Example 1

Non-Toxigenic Protein a Variants as Subunit Vaccines to Prevent Staphylococcus Aureus Infections

[0321] A. Results

[0322]An Animal Model for S. aureus Infection

[0323] BALB/c mice were infected by intravenous injection with  $1\times10^7$  CFU of the human clinical isolate S. aureus Newman (Baba et al., 2007). Within 6 hours following infection, 99.999% of staphylococci disappeared from the blood stream and were distributed via the vasculature. Staphylococcal dissemination to peripheral tissues occurred rapidly, as the bacterial load in kidney and other peripheral organ tissues reached 1×10<sup>5</sup> CFU g<sup>-1</sup> within the first three hours. The staphylococcal load in kidney tissues increased by 1.5 log CFU within twenty-four hours. Forty-eight hours following infection, mice developed disseminated abscesses in multiple organs, detectable by light microscopy of hematoxylin-eosin stained, thin-sectioned kidney tissue. The initial abscess diameter was 524 µM (±65 µM); lesions were initially marked by an influx of polymorphonuclear leukocytes (PMNs) and harbored no discernable organization of staphylococci, most of which appeared to reside within PMNs. On day 5 of infection, abscesses increased in size and enclosed a central population of staphylococci, surrounded by a layer of eosinophilic, amorphous material and a large cuff of PMNs. Histopathology revealed massive necrosis of PMNs in proximity to the staphylococcal nidus at the center of abscess lesions as well as a mantle of healthy phagocytes. A rim of necrotic PMNs were observed at the periphery of abscess lesions, bordering eosigenate spread on agar media for colony formation. On day 5 of infection, a mean of  $1\times10^6$  CFU g<sup>-1</sup> renal tissue for S. aureus Newman was observed. To quantify abscess formation, kidneys were visually inspected, and each individual organ was given a score of one or zero. The final sum was divided by the total number of kidneys to calculate percent surface abscesses (Table 4). In addition, randomly chosen kidneys were fixed in formalin, embedded, thin sectioned, and stained with hematoxylin-eosin. For each kidney, four sagittal sections at 200 µM intervals were viewed by microscopy. The numbers of lesions were counted for each section and averaged to quantify the number of abscesses within the kidneys. S. aureus Newman caused 4.364±0.889 abscesses per kidney, and surface abscesses were observed on 14 out of 20 kidneys (70%) (Table 4).

[0325] When examined by scanning electron microscopy, S. aureus Newman was located in tightly associated lawns at the center of abscesses. Staphylococci were contained by an amorphous pseudocapsule that separated bacteria from the cuff of abscesses leukocytes. No immune cells were observed in these central nests of staphylococci, however occasional red blood cells were located among the bacteria. Bacterial populations at the abscess center, designated staphylococcal abscess communities (SAC), appeared homogenous and coated by an electron-dense, granular material. The kinetics of the appearance of infectious lesions and the morphological attributes of abscesses formed by S. aureus Newman were similar to those observed following mouse infection with S. aureus USA300 (LAC), the current epidemic communityacquired methicillin-resistant S. aureus (CA-MRSA) clone in the United States (Diep et al., 2006).

TABLE 4

	Genet	c requirements	for S. aureus Newm	an abscess forma	tion in mice	
				Abscess	formation in kidn	ey tissue
	Staphylo	coccal load in k	idney tissue		<sup>e</sup> Number of	
Genotype	<sup>a</sup> log <sub>10</sub> CFU g <sup>-1</sup> tissue	<sup>b</sup> Significance (P-value)	<sup>c</sup> Reduction (log <sub>10</sub> CFU g <sup>-1</sup> )	<sup>d</sup> Surface abscesses (%)	abscesses per kidney	<sup>f</sup> Significance (P-value)
wild-type ΔsrtA spa	6.141 ± 0.192 4.095 ± 0.347 5.137 ± 0.374	— 6.7 × 10 <sup>-6</sup> 0.0144	2.046 1.004	70 0 13	4.364 ± 0.889 0.000 ± 0.000 0.375 ± 0.374	0.0216 0.0356

<sup>&</sup>lt;sup>a</sup>Means of staphylococcal load calculated as  $\log_{10}$  CFU g<sup>-1</sup> in homogenized renal tissues 5 days following infection in cohorts of fifteen BALB/c mice per challenge strain. Standard error of the means (±SEM) is indicated.
<sup>b</sup>Statistical significance was calculated with the Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

nophilic, amorphous material that separates healthy renal tissue from lesions. Abscesses eventually reached a diameter of 1,524 µM on day 15 or 36. At later time intervals, the staphylococcal load was increased to 10<sup>4</sup>-10<sup>6</sup> CFU g<sup>-1</sup> and growing abscess lesions migrated towards the organ capsule. Peripheral lesions were prone to rupture, thereby releasing necrotic material and staphylococci into the peritoneal cavity or the retroperitoneal space. These events resulted in bacteremia as well as a secondary wave of abscesses, eventually precipitating a lethal outcome.

[0324] To enumerate staphylococcal load in renal tissue, animals were killed, their kidneys excised and tissue homo[0326] S. aureus Protein A (spa) Mutants are Avirulent and Cannot Form Abscesses

[0327] Sortase A is a transpeptidase that immobilizes nineteen surface proteins in the envelope of S. aureus strain Newman (Mazmanian et al., 1999; Mazmanian et al., 2000). Earlier work identified sortase A as a virulence factor in multiple animal model systems, however the contributions of this enzyme and its anchored surface proteins to abscess formation or persistence have not yet been revealed (Jonsson et al., 2002; Weiss et al., 2004). Compared to the wild-type parent (Baba et al., 2007), an isogenic srtA variant (AsrtA) failed to form abscess lesions on either macroscopic or histopathology

<sup>&</sup>lt;sup>c</sup>Reduction in bacterial load calculated as log<sub>10</sub> CFU g<sup>-1</sup>.

<sup>&</sup>lt;sup>d</sup>Abscess formation in kidney tissues five days following infection was measured by macroscopic inspection (% positive)

<sup>&</sup>quot;Histopathology of hematoxylin-eosin stained, thin sectioned kidneys from eight to ten animals; the average number of abscesses per kidney was recorded and averaged again for the final mean (±SEM).

Statistical significance was calculated with the Students t-test and P-values recorded; P-values < 0.05 were deemed

examination on days 2, 5, or 15. In mice infected with the strA mutant, only 1×10<sup>4</sup> CFU g<sup>-1</sup> was recovered from kidney tissue on day 5 of infection, which is a 2.046 log<sub>10</sub> CFU g<sup>-</sup> reduction compared to the wild-type parent strain (P=6.73× 10<sup>-6</sup>). A similar defect was observed for the srtA mutant of MRSA strain USA300 (data not shown). Scanning electron microscopy showed that srtA mutants were highly dispersed and often associated with leukocytes in otherwise healthy renal tissue. On day fifteen following infection, srtA mutants were cleared from renal tissues, a  $\geq$ 3.5  $\log_{10}$  CFU g<sup>-1</sup> reduction compared to the wild-type (Table 3). Thus, sortase A anchored surface proteins enable the formation of abscess lesions and the persistence of bacteria in host tissues, wherein staphylococci replicate as communities embedded in an extracellular matrix and shielded from surrounding leukocytes by an amorphous pseudocapsule.

[0328] Sortase A anchors a large spectrum of proteins with LPXTG motif sorting signals to the cell wall envelope, thereby providing for the surface display of many virulence factors (Mazmanian et al., 2002). To identify surface proteins required for staphylococcal abscess formation, bursa aurealis insertions were introduced in 5' coding sequences of genes that encode polypeptides with LPXTG motif proteins (Bae et al., 2004) and these mutations were transduced into S. aureus Newman. Mutations in the structural gene for Protein A (spa) reduced the staphylococcal load in infected mouse kidney tissues by 1.004 log<sub>10</sub> (P=0.0144). When analyzed for their ability to form abscesses in kidney tissues by histopathology, we observed that the spa mutants were unable to form abscesses as compared with the wild-type parent strain S. aureus Newman (wild-type S. aureus Newman 4.364±0.889 abscesses per kidney vs. the isogenic spa mutant with 0.375±0.374 lesions; P=0.0356).

[0329] Protein A Blocks Innate and Adaptive Immune Responses.

[0330] Studies identified Protein A as a critical virulence factor during the pathogenesis of S. aureus infections. Earlier work demonstrated that Protein A impedes phagocytosis of staphylococci by binding the Fc component of immunoglobulin (Jensen 1958; Uhlén et al., 1984), activates platelet aggregation via the von Willebrand factor (Hartleib et al., 2000), functions as a B cell superantigen by capturing the F(ab)<sub>2</sub> region of VH3 bearing IgM (Roben et al., 1995), and, through its activation of TNFR1, can initiate staphylococcal pneumonia (Gomez et al., 2004). Due to the fact that Protein A captures immunoglobulin and displays toxic attributes, the possibility that this surface molecule may function as a vaccine in humans has not been rigorously pursued. The inventors demonstrate for the first time that Protein A variants no longer able to bind to immunoglobulins, vWF and TNFR-1 are removed of their toxigenic potential and are able to stimulate humoral immune responses that protect against staphylococcal disease.

[0331] Molecular Basis of Protein A Surface Display and Function.

[0332] Protein A is synthesized as a precursor in the bacterial cytoplasm and secreted via its YSIRK signal peptide at the cross wall, i.e., the cell division septum of staphylococci (FIG. 1). (DeDent et al., 2007; DeDent et al., 2008). Following cleavage of the C-terminal LPXTG sorting signal, Protein A is anchored to bacterial peptidoglycan crossbridges by sortase A (Schneewind et al., 1995; Mazmanian et al., 1999; Mazmanian et al., 2000). Protein A is the most abundant surface protein of staphylococci; the molecule is expressed by

virtually all *S. aureus* strains (SaId-Salim et al., 2003; Cespedes et al., 2005; Kennedy et al., 2008). Staphylococci turn over 15-20% of their cell wall per division cycle (Navarre and Schneewind 1999). Murine hydrolases cleave the glycan strands and wall peptides of peptidoglycan, thereby releasing Protein A with its attached C-terminal cell wall disaccharide tetrapeptide into the extracellular medium (Ton-That et al., 1999). Thus, by physiological design, Protein A is both anchored to the cell wall and displayed on the bacterial surface but also released into surrounding tissues during host infection (Marraffini et al., 2006).

[0333] Protein A captures immunoglobulins on the bacterial surface and this biochemical activity enables staphylococcal escape from host innate and acquired immune responses (Jensen 1958; Goodyear and Silverman 2004). Interestingly, region X of Protein A (Guss et al., 1984), a repeat domain that tethers the IgG binding domains to the LPXTG sorting signal/cell wall anchor, is perhaps the most variable portion of the staphylococcal genome (Schneewind et al., 1992; SaId-Salim et al., 2003). Each of the five immunoglobulin binding domains of Protein A (SpA), formed from three helix bundles and designated E, D, A, B, and C, exerts similar structural and functional properties (Sjödahl 1977; Jansson et al., 1998). The solution and crystal structure of domain D has been solved both with and without the Fc and  $V_H$ 3 (Fab) ligands, which bind Protein A in a non-competitive manner at distinct sites (Graille et al., 2000).

[0334] In the crystal structure complex, the Fab interacts with helix II and helix III of domain D via a surface composed of four VH region β-strands (Graille et al., 2000). The major axis of helix II of domain D is approximately 50° to the orientation of the strands, and the interhelical portion of domain D is most proximal to the C0 strand. The site of interaction on Fab is remote from the Ig light chain and the heavy chain constant region. The interaction involves the following domain D residues: Asp-36 of helix II as well as Asp-37 and Gln-40 in the loop between helix II and helix III, in addition to several other residues with SpA-D (Graille et al., 2000). Both interacting surfaces are composed predominantly of polar side chains, with three negatively charged residues on domain D and two positively charged residues on the 2A2 Fab buried by the interaction, providing an overall electrostatic attraction between the two molecules. Of the five polar interactions identified between Fab and domain D, three are between side chains. A salt bridge is formed between Arg-H19 and Asp-36 and two hydrogen bonds are made between Tyr-H59 and Asp-37 and between Asn-H82a and Ser-33. Because of the conservation of Asp-36 and Asp-37 in all five IgG binding domains of Protein A, these residues were selected for mutagenesis.

[0335] The SpA-D sites responsible for Fab binding are structurally separate from the domain surface that mediates Fey binding. The interaction of Fey with domain B primarily involves residues in helix I with lesser involvement of helix II (Deisenhofer 1981; Gouda et al., 1992). With the exception of the Gln-32, a minor contact in both complexes, none of the residues that mediate the Fey interaction are involved in Fab binding. To examine the spatial relationship between these different Ig-binding sites, the SpA domains in these complexes have been superimposed to construct a model of a complex between Fab, the SpA-domain D, and the Fey molecule. In this ternary model, Fab and Fey form a sandwich about opposite faces of the helix II without evidence of steric hindrance of either interaction. These findings illustrate how,

despite its small size (i.e., 56-61 aa), a SpA domain can simultaneously display both activities, explaining experimental evidence that the interactions of Fab with an individual domain are noncompetitive. Residues for the interaction between SpA-D and Fey are Gln-9 and Gln-10.

[0336] In contrast, occupancy of the Fc portion of IgG on the domain D blocks its interaction with vWF A1 and probably also TNFR1 (O'Seaghdha et al., 2006). Mutations in residues essential for IgG Fc binding (F5, Q9, Q10, S11, F13, Y14, L17, N28, 131 and K35) are also required for vWF A1 and TNFR1 binding (Cedergren et al., 1993; Gomez et al., 2006; O'Seaghdha et al. 2006), whereas residues critical for the  $V_H3$  interaction (Q26, G29, F30, S33, D36, D37, Q40, N43, E47) have no impact on the binding activities of IgG Fc, vWF A1 or TNFR1 (Jansson et al., 1998; Graille et al., 2000). The Protein A immunoglobulin Fab binding activity targets a subset of B cells that express VH3 family related IgM on their surface, i.e. these molecules function as VH3 type B cell receptors (Roben et al., 1995). Upon interaction with SpA, these B cells rapidly proliferate and then commit to apoptosis, leading to preferential and prolonged deletion of innate-like B lymphocytes (i.e. marginal zone B cells and follicular B2 cells) (Goodyear and Silverman 2003; Goodyear and Silverman 2004). It is important to note that more than 40% of circulating B cells are targeted by the Protein A interaction and the VH3 family represents the largest family of human B cell receptors to impart protective humoral responses against pathogens (Goodyear and Silverman 2003; Goodyear and Silverman 2004). Thus, Protein A functions analogously to staphylococcal superantigens (Roben et al., 1995), albeit that the latter class of molecules, for example SEB, TSST-1, TSST-2, form complexes with the T cell receptor to inappropriately stimulate host immune responses and thereby precipitating characteristic disease features of staphylococcal infections (Roben et al., 1995; Tiedemann et al., 1995). Together these findings document the contributions of Protein A in establishing staphylococcal infections and in modulating host immune responses.

[0337] Non-Toxigenic Variant of Protein A.

[0338] The inventors have developed a non-toxigenic variant of staphylococcal Protein A and, with this reagent in hand, aimed for the first time to measure the immune response of animals to Protein A immunization. Further, the inventors address whether immunization of animals with a non-toxigenic variant of Protein A could generate immune responses that raise protective immunity against staphylococcal infection.

[0339] To perturb the IgG Fc, vWFA1 and TNFR1 binding activities of Protein A, glutamine (Q) residues 9 and 10 [the numbering here is derived from that established for the SpA domain D] were modified generating lysine or glycine substitutions for both glutamines with the expectation that these substitutions abolish the ion bonds formed between wild-type Protein A and its ligands. The added effect of the dual lysine substitutions may be that these positively charged residues institute a repellent charge for immunoglobulins. To perturb IgM Fab VH3 binding, the inventors selected the aspartate (D) residues 36 and 37 of SpA-D, each of which is required for the association of Protein A with the B cell receptor. D36 and D37 were both substituted with alanine. The Q9,10K and D36,37A mutations were combined in the recombinant molecule  $SpA-D_{Q9,10K;D36,37A}$  and examined for the binding attributes of Protein A.

[0340] In brief, the Protein A (spa) genomic sequence of Staphylococcus aureus N315 was PCR amplified with the (GCTGCACATATGGCGCAACACGATprimers GAAGCTCAAC [5' primer](SEQ ID NO:35) and AGTG-GATCCTTATGCTTTGTTAGCATCTGC [3' primer] (SEQ ID NO:36)), cloned into the pET15b vector (pYSJ1, codons 48-486) (Stranger-Jones, et al., 2006) and recombinant plasmid transformed into E. coli BL21(DE3) (Studier et al., 1990). The Protein A product derived from pYSJ1 harbors SpA residues 36-265 fused to the N-terminal His tag (MGSSHHHHHHSSGLVPRGS (SEQ ID NO:37)). Following IPTG inducible expression, recombinant N-terminal His<sub>6</sub>-tagged SpA was purified by affinity chromatography on Ni-NTA resin (Stranger-Jones et al., 2006). The domain D of SpA (SpA-D) was PCR amplified with a pair of specific primers (AACATATGTTCAACAAAGATCAACAAAGC [5' primer] (SEQ ID NO:38) and AAGGATCCAGATTCGTT-TAATTTTTAGC [3' primer] (SEQ ID NO:39)), sub-cloned into the pET15b vector (pHAN1, spa codons 212-261) and recombinant plasmid transformed into E. coli BL21(DE3) to express and purify recombinant N-terminal His6-tagged protein. To generate mutations in the SpA-D coding sequence, sets of two pairs of primers were synthesized (for D to A CTTCATTCAAAGTCTTAAAGCCGCsubstitutions: CCCAAGCCAAAGCACTAAC [5' primer] (SEQ ID NO:40) and GTTAGTGCTTTGGCTTGGGGCGCCTT-TAAGACTTTGAATGAAG [3' primer] (SEQ ID NO:41); for Q to K substitutions CATATGTTCAACAAA-GATAAAAAAAGCGCCTTCTATGAAATC [5' primer] (SEQ ID NO:42) and GATTTCATAGAAGGCGCTTTTTT-TATCTTTGTTGAACATATG [3' primer] (SEQ ID NO:43); for Q to G substitutions CATATGTTCAACAAAGATG-GAGGAAGCGCCTTCTATGAAATC [5' primer] (SEQ ID GATTTCATAGAAGGCGCTTCCTCand CATCTTTGTTGAACATATG' [3' primer] (SEQ ID NO:45). Primers were used for quick-change mutagenesis protocols. Following mutagenesis, DNA sequences were confirmed for each of the recombinant proteins: SpA, SpA-D and SpA-D<sub>Q9</sub>,  $_{10G;D36,37A}$  and SpA-D $_{Q9,10K;D36,37A}$ . All proteins were purified from lysates of recombinant  $E.\ coli$  using Ni-NTA chromatography and subsequently dialyzed against PBS and stored at 4° C.

[0341] To measure binding of immunoglobulin to Protein A and its variants, 200 µg of purified protein was diluted into a 1 ml volume using column buffer (50 mM Tris-HCl, 150 mM NaCl, pH7.5) and then loaded onto a pre-equilibrated Ni-NTA column (1 ml bed volume). Columns were washed with 10 ml of column buffer. 200 μg of purified human IgG was diluted in a total volume of 1 ml column buffer and then applied to each of the columns charged with Protein A and its variants. The columns were subsequently washed with 5 ml wash buffer (10 mM imidazole in column buffer) and 5 ml column buffer. Protein samples were eluted with 2 ml elution buffer (500 mM imidazole in column buffer), fractions collected and aliquots subjected to SDS-PAGE gel electrophoresis, followed by Coomassie-Blue staining. As shown in FIG. 3, wild-type Protein A (SpA) and its SpA-domain D both retained immunogobulin during chromatography. In contrast, the SpA-D<sub>Q9,10K;D36,37A</sub> variant did not bind to immunoglobulin.

[0342] To quantify the binding of Protein A and its variants to the Fc portion of immunoglobulin and the VH3 domain of Fab, HRP conjugated human immunoglobulin G [hIgG], the Fc portion of human IgG [hFc] and the F(ab)<sub>2</sub> portion of

human IgG [hF(ab)<sub>2</sub>] as well as ELISA assays were used to quantify the relative amount binding to Protein A and its variants. The data in FIG. 4 demonstrate the binding of SpA and SpA-D to hIgG and hFc, whereas SpA-D  $_{Q9,10G;D36,37A}$ and SpA-D<sub>Q9,10K;D36,37A</sub> displayed only background binding activities. SpA bound similar amounts of hFc and hF(ab)<sub>2</sub>, however the binding of SpA-D to hF(ab), was reduced compared to full length SpA. This result suggests that the presence of multiple IgG binding domains may cooperatively increase the ability of Protein A to bind to the B cell receptor. When compared with the reduced binding power of SpA-D for hF(ab)2, of the two variants only SPA-DQ9,10K;D36,37A displayed a significant reduction in the ability to bind the VH3 domain of immunoglobulin. To examine the toxigenic attributes of SpA-D and its variants, purified proteins were injected into mice, which were sacrificed after 4 hours to remove their spleens. Organ tissue was homogenized, capsular material removed and B cells stained with fluorescent CD19 antibodies. Following FACS analysis to quantify the abundance of B cells in splenic tissues, it was observed that SpA-D caused a 5% drop in the B cell count compared to a mock (PBS) control (FIG. 5). In contrast, SpA-D<sub>Q9,10K;D36</sub>, 374 did not cause a reduction in B-cell counts, indicating that the mutant molecule had lost its toxigenic attributes of stimulating B cell proliferation and death (FIG. 5). In summary, amino acid substitutions in the SpA-D residues Q9, Q10, D36, and D37 abolished the ability of Protein A domains to bind immunoglobulins or exert toxigenic functions in human and animal tissues.

[0343] Non-Toxigenic Protein A Variants Elicit Vaccine Protection.

[0344] To test whether or not Protein A and its variants can function as vaccine antigens, SpA, SpA-D, SpA-D $_{Q9,10K;D36,374}$ , and SpA-D $_{Q9,10K;D36,374}$  were emulsified with complete or incomplete Freund's adjuvant and immunized 4 week old BALB/c mice on day 1 and day 11 with 50  $\mu$ g of purified protein. Cohort of animals (n=5) were analyzed for humoral immune responses to immunization by bleeding the animals before (day 0) and after the immunization schedule (day 21). Table 5 indicates that immunized mice generated only a modest humoral immune response directed at wild-type Protein A or its SpA-D module, whereas the amount of antibody raised following immunization with SpA-D $_{Q9,10K;D36,374}$  or SpA-

 $D_{O9,10K;D36,37A}$  was increased four to five fold. Following intravenous challenge with 1×10<sup>7</sup> CFU S. aureus Newman, animals were killed on day 4, their kidneys removed and either analyzed for staphylococcal load (by plating tissue homogenate on agar plates and enumerating colony forming units, CFU) or histopathology. As expected, mock (PBS) immunized mice (n=19) harbored 6.46  $\log_{10}$  (±0.25) CFU in kidney tissue and infectious lesions were organized into 3.7 (±1.2) abscesses per organ (n=10)(Table 5). Immunization of animals with SpA led to a 2.51 log<sub>10</sub> CFU reduction on day 5 (P=0.0003) with 2.1 (±1.2) abscesses per organ. The latter data indicate that there was no significant reduction in abscess formation (P=0.35). Immunization with SpA-D generated similar results: a 2.03 log<sub>10</sub> CFU reduction on day 5 (P=0. 0001) with 1.5 (±0.8) abscesses per organ (P=0.15). In contrast, immunization with SpA-D  $_{Q9,10K;D36,37A}$  or SpA-D  $_{Q9,}$ 10G;D36,37A created increased protection, with 3.07 log<sub>10</sub> and 3.03 log<sub>10</sub> CFU reduction on day 4, respectively (statistical significance P<0.0001 for both observations). Further, immunization with both SpA-D<sub>Q9,10K;D36,37A</sub> and SpA-D<sub>Q9,10G</sub>; D36,37A generated significant protection from staphylococcal abscess formation, as only  $0.5 (\pm 0.4)$  and  $0.8 (\pm 0.5)$  infectious lesions per organ (P=0.02 and P=0.04) were identified. Thus, immunization with non-toxigenic Protein A variants generates increased humoral immune responses for Protein A and provides protective immunity against staphylococcal challenge. These data indicate that Protein A is an ideal candidate for a human vaccine that prevents S. aureus disease.

[0345] These exciting results have several implications for the design of a human vaccine. First, the generation of substitution mutations that affect the ability of the immunoglobulin binding domains of Protein A, either alone or in combination of two or more domains, can generate non-toxigenic variants suitable for vaccine development. It seems likely that a combination of mutant IgG binding domains closely resembling the structure of Protein A can generate even better humoral immune responses as is reported here for the SpAdomain D alone. Further, a likely attribute of Protein A specific antibodies may be that the interaction of antigen binding sites with the microbial surface can neutralize the ability of staphylococci to capture immunoglobulins via their Fc portion or to stimulate the B cell receptor via the VH3 binding activities.

TABLE 5

Non-toxigenic Protein A variants as vaccine antigens that prevent S. aureus disease

		ial load in kidn umber of mice	•		Abs	scess formatic	on in mice (n = num	ber of mice)	
Antigen	<sup>a</sup> log <sub>10</sub> CFU g <sup>−1</sup>	<sup>b</sup> Reduction	<sup>c</sup> P value	IgG titer	<sup>d</sup> Surface abscess	Reduction	<sup>e</sup> Histopathology	Reduction	√P value
Mock	$6.46 \pm 0.25$ (n = 19)	_	_	<100	14/19 (70%)	_	$3.7 \pm 1.2$ (n = 10)	_	_
SpA	$3.95 \pm 0.56$ (n = 20)	2.51	0.0003	1706 ± 370	10/20 (50%)	32%	$2.1 \pm 1.2$ (n = 10)	2.2	0.35
SpA-D	$4.43 \pm 0.41$ (n = 18)	2.03	0.0001	381 ± 27	10/18 (55%)	25%	$1.5 \pm 0.8$ (n = 10)	2.2	0.15
SpA-D1	$3.39 \pm 0.50$ (n = 19)	3.07	<0.0001	5600 ± 801	6/20 (30%)	59%	$0.5 \pm 0.4$ (n = 10)	3.2	0.02

TABLE 5-continued

	N	Ion-toxigenic l	Protein A v	ariants as vacci	ine antigens	that prevent S	s. aureus disease		
		ial load in kidr umber of mice			Abs	scess formation	on in mice (n = num	ber of mice)	
Antigen	<sup>a</sup> log <sub>10</sub> CFU g <sup>−1</sup>	<sup>b</sup> Reduction	<sup>c</sup> P value	IgG titer	<sup>d</sup> Surface abscess	Reduction	<sup>e</sup> Histopathology	Reduction	f <sub>P</sub> value
SpA-D2	$3.43 \pm 0.46$ (n = 19)	3.03	<0.0001	3980 ± 676	6/19 (32%)	57%	$0.8 \pm 0.5$ $(n = 10)$	2.9	0.04

<sup>&</sup>quot;Means of staphylococcal load calculated as  $\log_{10}$  CFU g<sup>-1</sup> in homogenized renal tissues 4 days following infection in cohorts of 18 to 20 BALB/c mice. Standard error of the means ( $\pm$ SEM) is indicated. "Statistical significance was calculated with the Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

[0346] Vaccine Protection in Murine Abscess, Murine Lethal Infection, and Murine Pneumonia Models.

[0347] Three animal models have been established for the study of S. aureus infectious disease. These models are used here to examine the level of protective immunity provided via the generation of Protein A specific antibodies.

[0348] Murine Abscess-

[0349] BALB/c mice (24-day-old female, 8-10 mice per group, Charles River Laboratories, Wilmington, Mass.) are immunized by intramuscular injection into the hind leg with purified protein (Chang et al., 2003; Schneewind et al., 1992). Purified SpA, SpA-D or SpA-DQ9,10K;D36,37A (50 µg protein) is administered on days 0 (emulsified 1:1 with complete Freund's adjuvant) and 11 (emulsified 1:1 with incomplete Freund's adjuvant). Blood samples are drawn by retroorbital bleeding on days 0, 11, and 20. Sera are examined by ELISA for IgG titers for specific SpA-D and SpA-DQ9,10K;D36, 37A binding activity. Immunized animals are challenged on day 21 by retroorbital injection of 100 µl of S. aureus Newman or S. aureus USA300 suspension ( $1 \times 10^7$  cfu). For this, overnight cultures of S. aureus Newman are diluted 1:100 into fresh tryptic soy broth and grown for 3 h at 37° C. Staphylococci are centrifuged, washed twice, and diluted in PBS to yield an  $A_{600}$  of 0.4 (1×10<sup>8</sup> cfu per ml). Dilutions are verified experimentally by agar plating and colony formation. Mice are anesthetized by intraperitoneal injection of 80-120 mg of ketamine and 3-6 mg of xylazine per kilogram of body weight and infected by retroorbital injection. On day 5 or 15 following challenge, mice are euthanized by compressed CO<sub>2</sub> inhalation. Kidneys are removed and homogenized in 1% Triton X-100. Aliquots are diluted and plated on agar medium for triplicate determination of cfu. For histology, kidney tissue is incubated at room temperature in 10% formalin for 24 h. Tissues are embedded in paraffin, thin-sectioned, stained with hematoxylinleosin, and examined by microscopy.

[0350] Murine Lethal Infection-

[0351] BALB/c mice (24-day-old female, 8-10 mice per group, Charles River Laboratories, Wilmington, Mass.) are immunized by intramuscular injection into the hind leg with purified SpA, SpA-D or SpA-D  $_{Q9,10K;D36,37A}$  (50  $\mu g$  protein). Vaccine is administered on days 0 (emulsified 1:1 with complete Freund's adjuvant) and 11 (emulsified 1:1 with incomplete Freund's adjuvant). Blood samples are drawn by retroorbital bleeding on days 0, 11, and 20. Sera are examined by ELISA for IgG titers with specific SpA-D and SpA-D<sub>O9,10K</sub>; D36,37A binding activity. Immunized animals are challenged on day 21 by retroorbital injection of 100 µl of S. aureus Newman or S. aureus USA300 suspension  $(15 \times 10^7 \text{ cfu})$  (34). For this, overnight cultures of S. aureus Newman are diluted 1:100 into fresh tryptic soy broth and grown for 3 h at 37° C. Staphylococci are centrifuged, washed twice, diluted in PBS to yield an  $A_{600}$  of 0.4 (1×10<sup>8</sup> cfu per ml) and concentrated. Dilutions are verified experimentally by agar plating and colony formation. Mice are anesthetized by intraperitoneal injection of 80-120 mg of ketamine and 3-6 mg of xylazine per kilogram of body weight. Immunized animals are challenged on day 21 by intraperitoneal inject with  $2\times10^{10}$  cfu of S. aureus Newman or 3-10×10° cfu of clinical S. aureus isolates. Animals are monitored for 14 days, and lethal disease is recorded.

[0352] Murine Pneumonia Model—

[0353] S. aureus strains Newman or USA300 (LAC) are grown at 37° C. in tryptic soy broth/agar to OD<sub>660</sub> 0.5. 50-ml culture aliquots are centrifuged, washed in PBS, and suspended in 750  $\mu$ l PBS for mortality studies (3-4×10<sup>8</sup> CFU per 30-μl volume), or 1,250 μl PBS (2×10<sup>8</sup> CFU per 30-μl volume) for bacterial load and histopathology experiments (2, 3). For lung infection, 7-wk-old C57BL/6J mice (The Jackson Laboratory) are anesthetized before inoculation of 30 μl of S. aureus suspension into the left nare. Animals are placed into the cage in a supine position for recovery and observed for 14 days. For active immunization, 4-wk-old mice receive  $20 \,\mu g \, SpA$ -D or SpA-D $_{Q9,10K;D36,374}$  in CFA on day 0 via the i.m. route, followed by a boost with 20 µg SpA-D or SpA- $D_{Q9,10K;D36,37A}$  in incomplete Freund's adjuvant (IFA) on day 10. Animals are challenged with S. aureus on day 21. Sera are collected before immunization and on day 20 to assess specific antibody production. For passive immunization studies, 7-wk-old mice receive 100 µl of either NRS (normal rabbit serum) or SpA-D-specific rabbit antisera via i.p. injection 24 h before challenge. To assess the pathological correlates of pneumonia, infected animals are killed via forced CO<sub>2</sub> inhalation before removal of both lungs. The right lung is homogenized for enumeration of lung bacterial load. The left lung is placed in 1% formalin and paraffin embedded, thin sectioned, stained with hematoxylin-eosin, and analyzed by microscopy.

[0354] Rabbit Antibodies—

[0355] Purified 200 μg SpA-D or SpA-D<sub>O9,10K;D36,374</sub> is used as an immunogen for the production of rabbit antisera. 200 µg protein is emulsified with CFA for injection at day 0, followed by booster injections with 200 µg protein emulsified

 $<sup>^</sup>b\mathrm{Reduction}$  in bacterial load calculated as  $\log_{10}\mathrm{CFU}\ \mathrm{g}^{-1}$ 

<sup>&</sup>lt;sup>d</sup>Abscess formation in kidney tissues four days following infection was measured by macroscopic inspection (% positive)

<sup>&</sup>quot;Histopathology of hematoxylin-eosin stained, thin sectioned kidneys from ten animals; the number of abscesses per kidney was recorded and averaged for Instigation by the hallow in reason statics, this scattered reasons in the final mean (±SEM).

Statistical significance was calculated with the Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

SpA-D1 and SpA-D2 represent SpA-D<sub>Q9, 10K; D36, 37A</sub> and SpA-D<sub>Q9, 10G; D36, 37A</sub>, respectively.

with IFA on days 21 and 42. Rabbit antibody titers are determined by ELISA. Purified antibodies are obtained by affinity chromatography of rabbit serum on SpA-D or SpA-D  $_{\mathcal{Q}9,10K;}$   $_{\mathcal{D}36,37.4}$  sepharose. The concentration of eluted antibodies is measured by absorbance at  $A_{280}$  and specific antibody titers are determined by ELISA.

[0356] Active Immunization with SpA-Domain D Variants.—

[0357] To determine vaccine efficacy, animals are actively immunized with purified SpA-D or SpAD $_{Q9,10K;D36,37A}$ . As a control, animals are immunized with adjuvant alone. Antibody titers against Protein A preparations are determined using SpA-D or SpA-D $_{Q9,10K;D36,37A}$  as antigens; note that the SpA-D $_{Q9,10K;D36,37A}$  variant cannot bind the Fc or Fab portion of IgG. Using infectious disease models described above, any reduction in bacterial load (murine abscess and pneumonia), histopathology evidence of staphylococcal disease (murine abscess and pneumonia) and protection from lethal disease (murine lethal challenge and pneumonia) is measured.

[0358] Passive Immunization with Affinity Purified Rabbit Polyclonal Antibodies Generated Against SpA-Domain D Variants.

[0359] To determine protective immunity of Protein A specific rabbit antibodies, mice are passively immunized with 5 mg/kg of purified SpA-D or SpA-D<sub>Q9,10K;D36,37,4</sub> derived rabbit antibodies. Both of these antibody preparations are purified by affinity chromatography using immobilized SpA-D or SpA-D<sub>Q9,10K;D36,37,4</sub>. As a control, animals are passively immunized with rV10 antibodies (a plague protective antigen that has no impact on the outcome of staphylococcal infections). Antibody titers against all Protein A preparations are determined using SpA-D<sub>Q9,10K;D36,37,4</sub> as an antigen, as this variant cannot bind the Fc or Fab portion of IgG. Using the infectious disease models described above, the reduction in bacterial load (murine abscess and pneumonia), histopathology evidence of staphylococcal disease (murine abscess and pneumonia), and the protection from lethal disease (murine lethal challenge and pneumonia) is measured.

## Example 2

Non-Toxigenic Protein a Vaccine for Methicillin-Resistant *Staphylococcus aureus* Infection

[0360] Clinical isolates of S. aureus express protein A (Shopsin et al., 1999, whose primary translational product is comprised of an N-terminal signal peptide (DeDent et al., 2008), five Ig-BDs (designated E, D, A, B and C)(Sjödahl, 1977), region X with variable repeats of an eight residue peptide (Guss et al., 1984), and C-terminal sorting signal for the cell wall anchoring of SpA (Schneewind et al., 1992; Schneewind et al., 1995) (FIG. 6). Guided by amino acid homology (Uhlen et al., 1984), the triple α-helical bundle structure of IgBDs (Deisenhofer et al., 1978; Deisenhofer et al., 1981) and their atomic interactions with Fab  $V_H$ 3 (Graille et al., 2000) or Fcy (Gouda et al., 1998), glutamine 9 and 10 were selected as well as aspartate 36 and 37 as critical for the association of SpA with antibodies or B cell receptor, respectively. Substitutions Gln9Lys, Gln10Lys, Asp36Ala and Asp37Ala were introduced into the D domain to generate SpA-D<sub>KK44</sub> (FIG. 6). The ability of isolated SpA-D or SpA-D<sub>KK4.4</sub> to bind human IgG was analyzed by affinity chromatography (FIG. 6). Polyhistidine tagged SpA-D as well as full-length SpA retained human IgG on Ni-NTA, whereas SpA-D $_{KKAA}$  and a negative control (SrtA) did not (FIG. 6). A similar result was observed with von Willebrand factor (Hartleib et al., 2000), which, along with tumor necrosis factor receptor 1 (TNFR1)(Gomez et al., 2004), can also bind protein A via glutamine 9 and 10 (FIG. 6). Human immunoglobulin encompasses  $60-70\% V_H 3$ -type IgG. The inventors distinguish between Fc domain and B cell receptor activation of Igs and measured association of human Fcy and F(ab), fragments, both of which bound to full-length SpA or SpA-D, but not to SpA-D $_{KKAA}$  (FIG. 6). Injection of SpA-D into the peritoneal cavity of mice resulted in B cell expansion followed by apoptotic collapse of CD19+ lymphocytes in spleen tissue of BALB/c mice (Goodyear and Silverman, 2003)(FIG. 6). B cell superantigen activity was not observed following injection with SpA-D $_{KKAA}$ , and TUNEL-staining of splenic tissue failed to detect the increase in apoptotic cells that follows injection of SpA or SpA-D (FIG. 6).

[0361] Naive six week old BALB/c mice were injected with 50 μg each of purified SpA, SpA-D or SpA-D<sub>KKAA</sub> emulsified in CFA and boosted with the same antigen emulsified in IFA. In agreement with the hypothesis that SpA-D promotes the apoptotic collapse of activated clonal B cell populations, the inventors observed a ten-fold higher titer of SpA-D $_{KKAA}$  specific antibodies following immunization of mice with the non-toxigenic variant as compared to the B cell superantigen (Spa-D vs. SpA-D<sub>KK4.4</sub> P<0.0001, Table 6). Antibody titers raised by immunization with full-length SpA were higher than those elicited by SpA-D (P=0.0022), which is likely due to the larger size and reiterative domain structure of this antigen (Table 6). Nevertheless, even SpA elicited lower antibody titers than SpA-D<sub>KKAA</sub> (P=0.0003), which encompasses only 50 amino acids of protein A (520 residues, SEQ ID NO:33). Immunized mice were challenged by intravenous inoculation with S. aureus Newman and the ability of staphylococci to seed abscesses in renal tissues was examined by necropsy four days after challenge. In homogenized renal tissue of mock (PBS/adjuvant) immunized mice, an average staphylococcal load of 6.46 log<sub>10</sub> CFU g<sup>-1</sup> was enumerated (Table 6). Immunization of mice with SpA or SpA-D led to a reduction in staphylococcal load, however SpA-D $_{KKAA}$  vaccinated animals displayed an even greater, 3.07 log<sub>10</sub> CFU g<sup>-1</sup> reduction of S. aureus Newman in renal tissues (P<0.0001, Table 6). Abscess formation in kidneys was analyzed by histopathology (FIG. 7). Mock immunized animals harbored an average of 3.7 (±1.2) abscesses per kidney (Table 6). Vaccination with SpA-D<sub>KK4A</sub> reduced the average number of abscesses to 0.5 (±0.4) (P=0.0204), whereas immunization with SpA or SpA-D did not cause a significant reduction in the number of abscess lesions (Table 6). Lesions from SpA- $D_{KKAA}$  vaccinated animals were smaller in size, with fewer infiltrating PMNs and characteristically lacked staphylococcal abscess communities (Cheng et al., 2009)(FIG. 7). Abscesses in animals that had been immunized with SpA or SpA-D displayed the same overall structure of lesions in mock immunized animals (FIG. 7).

[0362] The inventors examined whether SpA-D $_{KKAA}$  immunization can protect mice against MRSA strains and selected the USA300 LAC isolate for animal challenge (Diep et al., 2006). This highly virulent CA-MRSA strain spread rapidly throughout the United States, causing significant human morbidity and mortality (Kennedy et al., 2008). Compared to adjuvant control mice, SpA-D $_{KKAA}$  immunized animals harbored a 1.07  $\log_{10}$  CFU g<sup>-1</sup> reduction in bacterial

load of infected kidney tissues. Histopathology examination of renal tissue following *S. aureus* USA300 challenge revealed that the average number of abscesses was reduced from  $4.04~(\pm0.8)$  to  $1.6~(\pm0.6)$  (P=0.02774). In contrast, SpA or SpA-D immunization did not cause a significant reduction in bacterial load or abscess formation (Table 6).

[0363] Rabbits were immunized with SpA-D $_{KKAA}$  and specific antibodies were purified on SpA-D $_{KKAA}$  affinity column followed by SDS-PAGE (FIG. 8). SpA-D $_{KKAA}$  specific IgG was cleaved with pepsin to generate Fc $\gamma$  and F(ab) $_2$  fragments, the latter of which were purified by chromatography on SpA-D $_{KKAA}$  column (FIG. 8). Binding of human IgG or vWF to SpA or SpA-D was perturbed by SpA-D $_{KKAA}$  specific F(ab) $_2$ , indicating that SpA-D $_{KKAA}$  derived antibodies neutralize the B cell superantigen function of protein A as well as its interactions with Ig (FIG. 8).

[0364] To further improve the vaccine properties for nontoxigenic protein A, the inventors generated  $\mathrm{SpA}_{KKAA}$ , which includes all five IgBDs with four amino acid substitutions—substitutions corresponding to Gln9Lys, Gln10Lys, Asp36Ala and Asp37Ala of domain D—in each of its five domains (E, D, A, B and C). Polyhistidine tagged  $\mathrm{SpA}_{KKAA}$  was purified by affinity chromatography and analyzed by Coomassie Blue-stained SDS-PAGE (FIG. 9). Unlike full-length  $\mathrm{SpA}$ ,  $\mathrm{SpA}_{KKAA}$  did not bind human  $\mathrm{IgG}$ , Fc and F(ab)<sub>2</sub> or vWF (FIG. 9).  $\mathrm{SpA}_{KKAA}$  failed to display B cell superan-

tigen activity, as injection of the variant into BALB/c mice did not cause a depletion of CD19+ B cells in splenic tissue (FIG. 9). SpA $_{KKAA}$  vaccination generated higher specific antibody titers than SpA-D $_{KKAA}$  immunization and provided mice with elevated protection against *S. aureus* USA300 challenge (Table 6). Four days following challenge, SpA $_{KKAA}$  vaccinated animals harbored 3.54  $\log_{10}$  CFU g<sup>-1</sup> fewer staphylococci in renal tissues (P=0.0001) and also caused a greater reduction in the number of abscess lesions (P=0.0109) (Table 6).

[0365] SpA $_{KKAA}$  was used to immunize rabbits. Rabbit antibodies specific for SpA-D $_{KKAA}$  or SpA $_{KKAA}$  were affinity purified on matrices with immobilized cognate antigen and injected at a concentration of 5 mg kg $^{-1}$  body weight into the peritoneal cavity of BALB/c mice (Table 7). Twenty-four hours later, specific antibody titers were determined in serum and animals challenged by intravenous inoculation with S. aureus Newman. Passive transfer reduced the staphylococcal load in kidney tissues for SpA-D $_{KKAA}$  (P=0.0016) or SpA $_{KAA}$  (P=0.0005) specific antibodies. On histopathology examination, both antibodies reduced the abundance of abscess lesions in the kidneys of mice challenged with S. aureus Newman (Table 7). Together these data reveal that vaccine protection following immunization with SpA-D $_{KKAA}$  or SpA $_{KKAA}$  is conferred by antibodies that neutralize protein S.

TABLE 6

	Im	nmunization	of mice with protein	n A vaccines.					
		Staphylococ	cal load and abscess	s formation in ren	al tissue				
Antigen	"log <sub>10</sub> CFU g <sup>-1</sup>	<sup>b</sup> P-value	<sup>c</sup> Reduction (log <sub>10</sub> CFU g <sup>-1</sup> )	<sup>d</sup> IgG Titer	<sup>e</sup> Number of abscesses	€P-value			
		S. au	reus Newman challe	enge					
Mock	6.46 ± 0.25	_	_	<100	3.7 ± 1.2	_			
SpA	$3.95 \pm 0.56$	0.0003	2.51	1706 ± 370	$2.1 \pm 1.2$	0.3531			
SpA-D	$4.43 \pm 0.41$	0.0001	2.03	$381 \pm 27$	$1.5 \pm 0.8$	0.1430			
$\mathrm{SpA-D}_{K\!K\!A\!A}$	$3.39 \pm 0.50$	< 0.0001	3.07	$5600 \pm 801$	$0.5 \pm 0.4$ $0.0204$				
		S. aureus	USA300 (LAC) ch	allenge					
Mock	7.20 ± 0.24	_	_	<100	4.0 ± 0.8	_			
SpA	$6.81 \pm 0.26$	0.2819	0.39	476 ± 60	$3.3 \pm 1.0$	0.5959			
SpA-D	$6.34 \pm 0.52$	0.1249	0.86	$358 \pm 19$	$2.2 \pm 0.6$	0.0912			
$\mathrm{SpA\text{-}D}_{KKAA}$	$6.00 \pm 0.42$	0.0189	1.20	3710 ± 1147	$1.6 \pm 0.6$	0.0277			
$\mathrm{SpA}_{K\!K\!A\!A}$	$3.66 \pm 0.76$	0.0001	3.54	10200 ± 2476	$1.2 \pm 0.5$	0.0109			

 $<sup>^{\</sup>circ}$ Means of staphylococcal load calculated as  $\log_{10}$  CFU  $g^{-1}$  in homogenized renal tissues 4 days following infection in cohorts of fifteen to twenty BALB/c mice per immunization. Representative of two independent and reproducible animal experiments is shown. Standard error of the means ( $\pm$ SEM) is indicated.

 $<sup>^</sup>b$ Statistical significance was calculated with the unpaired two-tailed Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

 $<sup>^</sup>c$ Reduction in bacterial load calculated as  $\log_{10}$  CFU g $^{-1}$ .

<sup>&</sup>lt;sup>d</sup>Means of five randomly chosen serum IgG titers were measured prior to staphylococcal infection by ELISA.

<sup>&</sup>quot;Histopathology of hematoxylene-eosin stained, thin sectioned kidneys from ten animals; the average number of abscesses per kidney was recorded and averaged again for the final mean (±SEM).

TABLE 7

	Passive immu:	nization of	mice with antibodi	es against prote	in A.	
	St	aphylococo	cal load and abscess	s formation in r	enal tissue	
"Antibody	<sup>b</sup> log <sub>10</sub> CFU g <sup>−1</sup>	<sup>c</sup> P-value	<sup>d</sup> Reduction (log <sub>10</sub> CFU g <sup>-1</sup> )	<sup>e</sup> IgG Titer	Number of abscesses	<sup>c</sup> P-value
Mock α-SpA-D <sub>KKAA</sub> α-SpA <sub>KKAA</sub>	$7.10 \pm 0.14$ $5.53 \pm 0.43$ $5.69 \pm 0.34$	 0.0016 0.0005	1.57 1.41	<100 466 ± 114 1575 ± 152	4.5 ± 0.8 1.9 ± 0.7 1.6 ± 0.5	 0.0235 0.0062

[0366] Following infection with virulent S. aureus, mice do not develop protective immunity against subsequent infection with the same strain (Burts et al., 2008)(FIG. 10). The average abundance of SpA-D $_{KKAA}$  specific IgG in these animals was determined by dot blot as  $0.20 \,\mu g \, ml^{-1}$  (±0.04) and  $0.14 \,\mu g$ ml<sup>-1</sup> (±0.01) for strains Newman and USA300 LAC, respectively (FIG. 9). The minimal concentration of protein A-specific IgG required for disease protection in SpA<sub>KKAA</sub> or SpA- $D_{KKAA}$  vaccinated animals (P 0.0.05  $log_{10}$  reduction in staphylococcal CFU g<sup>-1</sup> renal tissue) was calculated as 4.05 μg ml<sup>-1</sup> (±0.88). Average serum concentration of SpA-specific IgG in adult healthy human volunteers (n=16) was 0.21  $\mu$ g ml<sup>-1</sup> (±0.02). Thus, S. aureus infections in mice or humans are not associated with immune responses that raise significant levels of neutralizing antibodies directed against protein A, which is likely due to the B cell superantigen attributes of this molecule. In contrast, the average serum concentration of IgG specific for diphtheria toxin in human volunteers, 0.068  $\mu g \text{ ml}^{-1}$  (±0.20), was within range for protective immunity against diphtheria (Behring, 1890; Lagergard et al., 1992).

[0367] Clinical S. aureus isolates express protein A, an essential virulence factor whose B cell surperantigen activity and evasive attributes towards opsono-phagocytic clearance are absolutely required for staphylococcal abscess formation (Palmqvist et al., 2005; Cheng et al., 2009; Silverman and Goodyear, 2006). Protein A can thus be thought of as a toxin, essential for pathogenesis, whose molecular attributes must be neutralized in order to achieve protective immunity. By generating non-toxigenic variants unable to bind Igs via Fcy or VH<sub>3</sub>-Fab domains, the inventors measure here for the first time protein A neutralizing immune responses as a correlate for protective immunity against S. aureus infection. In contrast to many methicillin-sensitive strains, CA-MRSA isolate USA300 LAC is significantly more virulent (Cheng et al., 2009). For example, immunization of experimental animals with the surface protein IsdB (Kuklin et al., 2006; Stranger-Jones et al., 2006) raises antibodies that confer protection against S. aureus Newman (Stranger-Jones et al., 2009) but not against USA300 challenge.

[0368] The methods utilized include:

[0369] Bacterial Strains and Growth.

[0370] Staphylococcus aureus strains Newman and USA300 were grown in tryptic soy broth (TSB) at 37° ° C.

Escherichia coli strains DH5a and BL21 (DE3) were grown in Luria-Bertani (LB) broth with 100 μg ml<sup>-1</sup> ampicillin at 37° C.

[0371] Rabbit Antibodies.

[0372] The coding sequence for SpA was PCR-amplified with two primers, gctgcacatatggcgcaacacgatgaagctcaac (SEQ ID NO:35) and agtggatccttatgcttgagctttgttagcatctgc (SEQ ID NO:36) using S. aureus Newman template DNA. SpA-D was PCR-amplified with two primers, aacatatgttcaacaaagatcaacaaagc (SEQ ID NO:38) and aaggatccagattcgtttaattttttagc (SEQ ID NO:39). The sequence for SpA-D $_{KKAA}$ was mutagenized with two sets of primers catatgttcaacaaagataaaaaaaagcgccttctatgaaatc (SEQ ID NO:42) and gatttcatagaaggegetttttttatetttgttgaacatatg (SEQ ID NO:43) for for Q9K, Q10K as well as cttcattcaaagtcttaaagccgccccaagccaaagcactaac (SEQ ID NO:40) and gttagtgctttggcttggggcggctttaagactttgaatgaag (SEQ ID NO:41) for D36A, D37A. The sequence of  $SpA_{KKAA}$  was synthesized by Integrated DNA Technologies, Inc. PCR products were cloned into pET-15b generating N-terminal His6 tagged recombinant protein. Plasmids were transformed into BL21 (DE3). Overnight cultures of transformants were diluted 1:100 into fresh media and grown at 37° C. to an  $OD_{600}$  0.5, at which point cultures were induced with 1 mM isopropyl β-D-1-thiogalatopyranoside (IPTG) and grown for an additional three hours. Bacterial cells were sedimented by centrifugation, suspended in column buffer (50 mM Tris-HCl, pH 7.5, 150 mM NaCl) and disrupted with a French pressure cell at 14,000 psi. Lysates were cleared of membrane and insoluble components by ultracentrifugation at 40,000×g. Proteins in the soluble lysate were subjected to nickel-nitrilotriacetic acid (Ni-NTA, Qiagen) affinity chromatography. Proteins were eluted in column buffer containing successively higher concentrations of imidazole (100-500 mM). Protein concentrations were determined by bicinchonic acid (BCA) assay (Thermo Scientific). For antibody generation, rabbits (6 month old New-Zealand white, female, Charles River Laboratories) were immunized with 500 µg protein emulsified in Complete Freund's Adjuvant (Difco) by subscapular injection. For booster immunizations, proteins emulsified in Incomplete Freund's Adjuvant and injected 24 or 48 days following the initial immunization. On day 60, rabbits were bled and serum recovered.

[0373] Purified antigen (5 mg protein) was covalently linked to HiTrap NHS-activated HP columns (GE Health-

<sup>&</sup>lt;sup>a</sup>Affinity purified antibodies were injected into the peritoneal cavity of BALB/c mice at a concentration of 5 mg  $\cdot$  kg<sup>-1</sup> twenty-four hours prior to intravenous challenge with 1 x 10.º CFU *S. aureus* Newman.

<sup>a</sup>Means of staphylococcal load calculated as  $\log_{10}$  CFU g<sup>-1</sup> in homogenized renal tissues 4 days following infection in cohorts of fifteen BALB/c mice per immunization. Representative of two independent and reproducible animal experiments is shown. Standard error of the means (±SEM) is indicated.

<sup>a</sup>Statistical significance was calculated with the unpaired two-tailed Students t-test and P-values recorded; P-values < 0.05 was a description of the means (±SEM).

were deemed significant.

Reduction in bacterial load calculated as log<sub>10</sub> CFU g<sup>-1</sup>.

<sup>&</sup>lt;sup>e</sup>Means of five randomly chosen serum IgG titers were measured prior to staphylococcal infection by ELISA

Histopathology of hematoxylene-eosin stained, thin sectioned kidneys from ten animals; the average number of abscesses per kidney was recorded and averaged again for the final mean (±SEM).

care). Antigen-matrix was used for affinity chromatography of 10-20 ml of rabbit serum at 4° C. Charged matrix was washed with 50 column volumes of PBS, antibodies eluted with elution buffer (1 M glycine, pH 2.5, 0.5 M NaCl) and immediately neutralized with 1M Tris-HCl, pH 8.5. Purified antibodies were dialyzed overnight against PBS at 4° C.

[0374] F(Ab), Fragments.

[0375] Affinity purified antibodies were mixed with 3 mg of pepsin at 37° ° C. for 30 minutes. The reaction was quenched with 1 M Tris-HCl, pH 8.5 and F(ab)<sub>2</sub> fragments were affinity purified with specific antigen-conjugated HiTrap NHS-activated HP columns. Purified antibodies were dialyzed overnight against PBS at 4° C., loaded onto SDS-PAGE gel and visualized with Coomassie Blue staining

[0376] Active and Passive Immunization.

[0377] BALB/c mice (3 week old, female, Charles River Laboratories) were immunized with 50 µg protein emulsified in Complete Freund's Adjuvant (Difco) by intramuscular injection. For booster immunizations, proteins were emulsified in Incomplete Freund's Adjuvant and injected 11 days following the initial immunization. On day 20 following immunization, 5 mice were bled to obtain sera for specific antibody titers by enzyme-linked immunosorbent assay (ELISA).

[0378] Affinity purified antibodies in PBS were injected at a concentration 5 mg kg<sup>-1</sup> of experimental animal weight into the peritoneal cavity of BALB/c mice (6 week old, female, Charles River Laboratories) 24 hours prior to challenge with *S. aureus*. Animal blood was collected via periorbital vein puncture. Blood cells were removed with heparinized microhematocrit capillary tubes (Fisher) and Z-gel serum separation micro tubes (Sarstedt) were used to collect and measure antigen specific antibody titers by ELISA.

[0379] Mouse Renal Abscess.

[0380] Overnight cultures of S. aureus Newman or USA300 (LAC) were diluted 1:100 into fresh TSB and grown for 2 hours at 37° C. Staphylococci were sedimented, washed and suspended PBS at  $OD_{600}$  of 0.4 (~1×10<sup>8</sup> CFU m1<sup>-1</sup>). Inocula were quantified by spreading sample aliquots on TSA and enumerating colonies formed. BALB/c mice (6 week old, female, Charles River Laboratories) were anesthetized via intraperitoneal injection with 100 mg ml<sup>-</sup> ketamine and 20 mg ml<sup>-1</sup> xylazine per kilogram of body weight. Mice were infected by retro-orbital injection with  $1\times10^7$  CFU of S. aureus Newman or 5×10<sup>6</sup> CFU of S. aureus USA300. On day 4 following challenge, mice were killed by CO<sub>2</sub> inhalation. Both kidneys were removed, and the staphylococcal load in one organ was analyzed by homogenizing renal tissue with PBS, 1% Triton X-100. Serial dilutions of homogenate were spread on TSA and incubated for colony formation. The remaining organ was examined by histopathology. Briefly, kidneys were fixed in 10% formalin for 24 hours at room temperature. Tissues were embedded in paraffin, thin-sectioned, stained with hematoxylin-eosin, and inspected by light microscopy to enumerate abscess lesions. All mouse experiments were performed in accordance with the institutional guidelines following experimental protocol review and approval by the Institutional Biosafety Committee (IBC) and the Institutional Animal Care and Use Committee (IACUC) at the University of Chicago.

[0381] Protein A Binding.

[0382] For human IgG binding, Ni-NTA affinity columns were pre-charged with 200  $\mu$ g of purified proteins (SpA, SpA-D, SpA-D, SpA-D $_{KK44}$ , and SrtA) in column buffer. After wash-

ing, 200 µg of human IgG (Sigma) was loaded onto the column. Protein samples were collected from washes and elutions and subjected to SDS-PAGE gel electrophoresis, followed by Coomassie Blue staining Purified proteins (SpA, SpA $_{KKAA}$ , SpA-D and SpA-D $_{KKAA}$ ) were coated onto MaxiSorp ELISA plates (NUNC) in 0.1M carbonate buffer (pH 9.5) at 1 µg ml $^{-1}$  concentration overnight at 4° C. Plates were next blocked with 5% whole milk followed by incubation with serial dilutions of peroxidase-conjugated human IgG, Fc or F(ab) $_2$  fragments for one hour. Plates were washed and developed using OptEIA ELISA reagents (BD). Reactions were quenched with 1 M phosphoric acid and  $A_{450}$  readings were used to calculate half maximal titer and percent binding.

[0383] von Willebrand Factor (vWF) Binding Assays.

[0384] Purified proteins (SpA, SpA $_{KKAA}$ , SpA D and SpA- $D_{KKAA}$ ) were coated and blocked as described above. Plates were incubated with human vWF at 1  $\mu$ g ml $^{-1}$  concentration for two hours, then washed and blocked with human IgG for another hour. After washing, plates were incubated with serial dilution of peroxidase-conjugated antibody directed against human vWF for one hour. Plates were washed and developed using OptEIA ELISA reagents (BD). Reactions were quenched with 1 M phosphoric acid and  $A_{450}$  readings were used to calculate half maximal titer and percent binding. For inhibition assays, plates were incubated with affinity purified  $F(ab)_2$  fragments specific for SpA- $D_{KKAA}$  at 10  $\mu$ g ml $^{-1}$  concentration for one hour prior to ligand binding assays.

[0385] Splenocyte Apoptosis.

[0386] Affinity purified proteins (150 µg of SpA, SpA-D,  $SpA_{KKAA}$ , and  $SpA-D_{KKAA}$ ) were injected into the peritoneal cavity of BALB/c mice (6 week old, female, Charles River Laboratories). Four hours following injection, animals were killed by CO2 inhalation. Their spleens were removed and homogenized. Cell debris were removed using cell strainer and suspended cells were transferred to ACK lysis buffer (0.15 M NH<sub>4</sub>Cl, 10 mM KHCO<sub>3</sub>, 0.1 mM EDTA) to lyse red blood cells. White blood cells were sedimented by centrifugation, suspended in PBS and stained with 1:250 diluted R-PE conjugated anti-CD19 monoclonal antibody (Invitrogen) on ice and in the dark for one hour. Cells were washed with 1% FBS and fixed with 4% formalin overnight at 4° C. The following day, cells were diluted in PBS and analyzed by flow cytometry. The remaining organ was examined for histopathology. Briefly, spleens were fixed in 10% formalin for 24 hours at room temperature. Tissues were embedded in paraffin, thin-sectioned, stained with the Apoptosis detection kit (Millipore), and inspected by light microscopy.

[0387] Antibody Quantification.

[0388] Sera were collected from healthy human volunteers or BALB/c mice that had been either infected with *S. aureus* Newman or USA300 for 30 days or that had been immunized with SpA-D<sub>KKAA</sub>/SpA<sub>KKAA</sub> as described above. Human/mouse IgG (Jackson Immunology Laboratory), SpA<sub>KKAA</sub>, and CRM<sub>197</sub> were blotted onto nitrocellulose membrane. Membranes were blocked with 5% whole milk, followed by incubation with either human or mouse sera. IRDye 700DX conjugated affinity purified anti-human/mouse IgG (Rockland) was used to quantify signal intensities using the Odyssey<sup>TM</sup> infrared imaging system (Li-cor). Experiments with blood from human volunteers involved protocols that were reviewed, approved and performed under regulatory supervision of The University of Chicago's Institutional Review Board (IRB).

[0389] Statistical Analysis.

[0390] Two tailed Student's t tests were performed to analyze the statistical significance of renal abscess, ELISA, and B cell superantigen data.

### REFERENCES

[0391] The following references, to the extent that they provide exemplary procedural or other details supplementary to those set forth herein, are specifically incorporated herein by reference.

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#### SEQUENCE LISTING

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Asp	Ser 1070		/ Lys	г Туг	. Lys	Val		le P	he G	lu Ly		ro 080	Thr	Gly	Leu
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Gly	Gly 1100		ı Val	. Asr	Val	. Thi		le T	hr A	sp Hi		sp . 110	Asp	Phe	Thr
Leu	Asp 1115		ı Gly	ту1	Tyr	Glu 112		lu G	lu Tì	nr Se		sp 125	Ser	Asp	Ser
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Asp	Ser	Asp	Ser	: Asr	Ser	. Yal	o S∈	er A	sp Se	er As	∌p S	er .	Aap	Ser	Asp

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Ser	Asp 1235		Asp	Ser	Asp	Ser 124		p Sei	Asp	Ser	Asp 1245		. Asl	) Ser	
Asp	Ser 1250	_	Ser	Asp	Ser	Asp 125		r Asp	Ser	Asp	Ser 1260	_	Sei	: Asp	,
Ser	Asp 1265		Asp	Ser	Asp	Ser 127		p Sei	: Asp	Ser	Asp 1275		. Asl	) Ser	
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Asn	Thr				Pro	Glu .				lu S	er Th				er
Thr		Gln		Gln	Asn		Val '		Ala T	hr T	hr Gl	lu Tì		s Pr	0
Gln	Asn	115 Ile	Glu	Lys	Glu	Asn	120 Val :	Lys I	ro S		12 hr As		/s Tl	ır Al	a
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Ile	Lys 210	Leu	Val	Ser	Tyr	Asp 215	Thr	Val	Lys	Asp	Tyr 220	Ala	Tyr	Ile	Arg
Phe 225	Ser	Val	Ser	Asn	Gly 230	Thr	Lys	Ala	Val	Lys 235	Ile	Val	Ser	Ser	Thr 240
His	Phe	Asn	Asn	Lys 245	Glu	Glu	Lys	Tyr	Asp 250	Tyr	Thr	Leu	Met	Glu 255	Phe
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Asn	Asp 50	Thr	Thr	Gln	Ser	Ser 55	Lys	Asn	Asn	Ala	Ser 60	Ala	Asp	Ser	Glu
Lys	Asn	Asn	Met	Ile	Glu 70	Thr	Pro	Gln	Leu	Asn 75	Thr	Thr	Ala	Asn	Asp 80
Thr	Ser	Asp	Ile	Ser 85	Ala	Asn	Thr	Asn	Ser 90	Ala	Asn	Val	Asp	Ser 95	Thr
Thr	Lys	Pro	Met 100	Ser	Thr	Gln	Thr	Ser 105	Asn	Thr	Thr	Thr	Thr 110	Glu	Pro
Ala	Ser	Thr 115	Asn	Glu	Thr	Pro	Gln 120	Pro	Thr	Ala	Ile	Lys 125	Asn	Gln	Ala
Thr	Ala 130	Ala	Lys	Met	Gln	Asp 135	Gln	Thr	Val	Pro	Gln 140	Glu	Ala	Asn	Ser
Gln 145	Val	Asp	Asn	ГЛа	Thr 150	Thr	Asn	Asp	Ala	Asn 155	Ser	Ile	Ala	Thr	Asn 160

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Gln	Thr	Ile	Ser 180	Asn	Ala	Gln	Gly	Thr 185	Ser	ГЛЗ	Pro	Ser	Val 190	Arg	Thr
Arg	Ala	Val 195	Arg	Ser	Leu	Ala	Val 200	Ala	Glu	Pro	Val	Val 205	Asn	Ala	Ala
Asp	Ala 210	ГÀз	Gly	Thr	Asn	Val 215	Asn	Asp	Lys	Val	Thr 220	Ala	Ser	Asn	Phe
Lys 225	Leu	Glu	Lys	Thr	Thr 230	Phe	Asp	Pro	Asn	Gln 235	Ser	Gly	Asn	Thr	Phe 240
Met	Ala	Ala	Asn	Phe 245	Thr	Val	Thr	Asp	Lys 250	Val	Lys	Ser	Gly	Asp 255	Tyr
Phe	Thr	Ala	Lys 260	Leu	Pro	Asp	Ser	Leu 265	Thr	Gly	Asn	Gly	Asp 270	Val	Asp
Tyr	Ser	Asn 275	Ser	Asn	Asn	Thr	Met 280	Pro	Ile	Ala	Asp	Ile 285	Lys	Ser	Thr
Asn	Gly 290	Asp	Val	Val	Ala	Lys 295	Ala	Thr	Tyr	Asp	Ile 300	Leu	Thr	Lys	Thr
Tyr 305	Thr	Phe	Val	Phe	Thr 310	Asp	Tyr	Val	Asn	Asn 315	ГÀв	Glu	Asn	Ile	Asn 320
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Ser	Gly	Thr	Tyr 340	Asp	Ala	Asn	Ile	Asn 345	Ile	Ala	Asp	Glu	Met 350	Phe	Asn
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Pro	Asn 370	Gly	Ala	Asn	Ile	Ser 375	Ser	Gln	Ile	Ile	Gly 380	Val	Asp	Thr	Ala
Ser 385	Gly	Gln	Asn	Thr	Tyr 390	ГÀз	Gln	Thr	Val	Phe 395	Val	Asn	Pro	ГЛа	Gln 400
Arg	Val	Leu	Gly	Asn 405	Thr	Trp	Val	Tyr	Ile 410	ГÀЗ	Gly	Tyr	Gln	Asp 415	Lys
Ile	Glu	Glu	Ser 420	Ser	Gly	ГÀз	Val	Ser 425	Ala	Thr	Asp	Thr	Lys 430	Leu	Arg
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Asp	Pro 450	Asn	Asp	Ser	Asn	Leu 455	Lys	Glu	Val	Thr	Asp 460	Gln	Phe	Lys	Asn
Arg 465	Ile	Tyr	Tyr	Glu	His 470	Pro	Asn	Val	Ala	Ser 475	Ile	Lys	Phe	Gly	Asp 480
Ile	Thr	Lys	Thr	Tyr 485	Val	Val	Leu	Val	Glu 490	Gly	His	Tyr	Asp	Asn 495	Thr
Gly	ГÀв	Asn	Leu 500	ГÀа	Thr	Gln	Val	Ile 505	Gln	Glu	Asn	Val	Asp 510	Pro	Val
Thr	Asn	Arg 515	Asp	Tyr	Ser	Ile	Phe 520	Gly	Trp	Asn	Asn	Glu 525	Asn	Val	Val
Arg	Tyr 530	Gly	Gly	Gly	Ser	Ala 535	Asp	Gly	Asp	Ser	Ala 540	Val	Asn	Pro	Lys
Asp 545	Pro	Thr	Pro	Gly	Pro 550	Pro	Val	Asp	Pro	Glu 555	Pro	Ser	Pro	Asp	Pro 560
Glu	Pro	Glu	Pro	Thr 565	Pro	Asp	Pro	Glu	Pro 570	Ser	Pro	Asp	Pro	Glu 575	Pro

Glu Pro Ser Pro Asp Pro Asp Pro Asp Ser Asp Ser Asp Ser Asp Ser 585 Gly Ser Asp Ser Asp Ser Gly Ser Asp Ser Asp Ser Glu Ser Asp Ser Glu Ser 615 Asp Ser Asp Ser Glu Ser Asp Ser Glu Ser Asp Ser Glu Ser Asp Ser Asp Ser Glu Ser Asp Ser Glu Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser - 680 Asp Ser 690 700 Asp Ser Asp Ser Asp Ser Asp Ser Glu Ser Asp Ser Asp Ser Asp Ser 705  $\phantom{\bigg|}$  710  $\phantom{\bigg|}$  715  $\phantom{\bigg|}$  720 Asp Ser 730 Asp Ser Asp S Asp Ser 760 Asp Ser 790 Asp Ser Asp Ser Arg Val Thr Pro Pro Asn Asn Glu Gln Lys Ala Pro 810 Ser Asn Pro Lys Gly Glu Val Asn His Ser Asn Lys Val Ser Lys Gln 825 His Lys Thr Asp Ala Leu Pro Glu Thr Gly Asp Lys Ser Glu Asn Thr Asn Ala Thr Leu Phe Gly Ala Met Met Ala Leu Leu Gly Ser Leu Leu 855 Leu Phe Arg Lys Arg Lys Gln Asp His Lys Glu Lys Ala <210> SEQ ID NO 19 <211> LENGTH: 227 <212> TYPE: PRT <213> ORGANISM: Staphylococcus sp. <400> SEQUENCE: 19 Met Lys Asn Ile Leu Lys Val Phe Asn Thr Thr Ile Leu Ala Leu Ile 1.0 Ile Ile Ile Ala Thr Phe Ser Asn Ser Ala Asn Ala Asp Ser Gly Thr Leu Asn Tyr Glu Val Tyr Lys Tyr Asn Thr Asn Asp Thr Ser Ile Ala Asn Asp Tyr Phe Asn Lys Pro Ala Lys Tyr Ile Lys Lys Asn Gly

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Met	Ser	Ile	Glu	Gly 85	His	Lys	Glu	Asn	Ile 90	Ile	Ser	Lys	Asn	Thr 95	Ala
Lys	Asp	Glu	Arg 100	Thr	Ser	Glu	Phe	Glu 105	Val	Ser	Lys	Leu	Asn 110	Gly	Lys
Ile	Asp	Gly 115	Lys	Ile	Asp	Val	Tyr 120	Ile	Asp	Glu	Lys	Val 125	Asn	Gly	ГЛа
Pro	Phe 130	Lys	Tyr	Asp	His	His 135	Tyr	Asn	Ile	Thr	Tyr 140	Lys	Phe	Asn	Gly
Pro 145	Thr	Asp	Val	Ala	Gly 150	Ala	Asn	Ala	Pro	Gly 155	Lys	Asp	Asp	Lys	Asn 160
Ser	Ala	Ser	Gly	Ser 165	Asp	Lys	Gly	Ser	Asp 170	Gly	Thr	Thr	Thr	Gly 175	Gln
Ser	Glu	Ser	Asn 180	Ser	Ser	Asn	Lys	Asp 185	Lys	Val	Glu	Asn	Pro 190	Gln	Thr
Asn	Ala	Gly 195	Thr	Pro	Ala	Tyr	Ile 200	Tyr	Ala	Ile	Pro	Val 205	Ala	Ser	Leu
Ala	Leu 210	Leu	Ile	Ala	Ile	Thr 215	Leu	Phe	Val	Arg	Lys 220	Lys	Ser	Lys	Gly
Asn 225	Val	Glu													
	)> SE L> LE	EQ II													
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Lys	Ser	Lys 195	Asp	Ala	Leu	Lys	Asp 200	Ala	Ser	Lys	Asp	Pro 205	Ala	Val	Ser
Thr	Thr 210	Asp	Ser	Asn	His	Glu 215	Val	Ala	Lys	Thr	Pro 220	Asn	Asn	Asp	Gly
Ser 225	Gly	His	Val	Val	Leu 230	Asn	Lys	Phe	Leu	Ser 235	Asn	Glu	Glu	Asn	Gln 240
Ser	His	Ser	Asn	Gln 245	Leu	Thr	Asp	Lys	Leu 250	Gln	Gly	Ser	Asp	Lys 255	Ile
Asn	His	Ala	Met 260	Ile	Glu	ГÀа	Leu	Ala 265	Lys	Ser	Asn	Ala	Ser 270	Thr	Gln
His	Tyr	Thr 275	Tyr	His	Lys	Leu	Asn 280	Thr	Leu	Gln	Ser	Leu 285	Asp	Gln	Arg
Ile	Ala 290	Asn	Thr	Gln	Leu	Pro 295	Lys	Asn	Gln	ГÀа	Ser 300	Asp	Leu	Met	Ser
Glu 305	Val	Asn	Lys	Thr	Lys 310	Glu	Arg	Ile	Lys	Ser 315	Gln	Arg	Asn	Ile	Ile 320
Leu	Glu	Glu	Leu	Ala 325	Arg	Thr	Asp	Asp	330 Lys	Lys	Tyr	Ala	Thr	Gln 335	Ser
Ile	Leu	Glu	Ser 340	Ile	Phe	Asn	Lys	Asp 345	Glu	Ala	Asp	LÀs	Ile 350	Leu	ГЛя
Asp	Ile	Arg 355	Val	Asp	Gly	ГÀз	Thr 360	Asp	Gln	Gln	Ile	Ala 365	Asp	Gln	Ile
Thr	Arg 370	His	Ile	Asp	Gln	Leu 375	Ser	Leu	Thr	Thr	Ser 380	Asp	Asp	Leu	Leu
Thr 385	Ser	Leu	Ile	Asp	Gln 390	Ser	Gln	Asp	Lys	Ser 395	Leu	Leu	Ile	Ser	Gln 400
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Asp	Trp	Thr	Asn 420	Lys	Gly	Leu	Ser	Asn 425	Arg	Gln	Ile	Val	Asp 430	Gln	Leu
Lys	Lys	His 435	Phe	Ala	Ser	Thr	Gly 440	Asp	Thr	Ser	Ser	Asp 445	Asp	Ile	Leu
ГÀа	Ala 450	Ile	Leu	Asn	Asn	Ala 455	Lys	Asp	Lys	Lys	Gln 460	Ala	Ile	Glu	Thr
Ile 465	Leu	Ala	Thr	Arg	Ile 470	Glu	Arg	Gln	Lys	Ala 475	Lys	Leu	Leu	Ala	Asp 480
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Lys	Ser	Ala	Leu 500	Asn	Gly	Lys	Ala	Asp 505	Asp	Leu	Leu	Asn	Leu 510	Gln	ГЛа
Arg	Leu	Asn 515	Gln	Thr	Lys	ГÀз	Asp 520	Ile	Asp	Tyr	Ile	Leu 525	Ser	Pro	Ile
Val	Asn 530	Arg	Pro	Ser	Leu	Leu 535	Asp	Arg	Leu	Asn	Lys 540	Asn	Gly	Lys	Thr
Thr 545	Asp	Leu	Asn	Lys	Leu 550	Ala	Asn	Leu	Met	Asn 555	Gln	Gly	Ser	Asn	Leu 560
Leu	Asp	Ser	Ile	Pro 565	Asp	Ile	Pro	Thr	Pro 570	Lys	Pro	Glu	Lys	Thr 575	Leu
Thr	Leu	Gly	Lys	Gly	Asn	Gly	Leu	Leu	Ser	Gly	Leu	Leu	Asn	Ala	Asp

		580					585					590		
Gly Asn	Val 595	Ser	Leu	Pro	Lys	Ala 600	Gly	Glu	Thr	Ile	Lys 605	Glu	His	Trp
Leu Pro 610	Ile	Ser	Val	Ile	Val 615	Gly	Ala	Met	Gly	Val 620	Leu	Met	Ile	Trp
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Ile Leu	Val 35	Gly	Thr	Thr	Leu	Ile 40	Phe	Gly	Leu	Ser	Gly 45	His	Glu	Ala
Lys Ala 50	Ala	Glu	His	Thr	Asn 55	Gly	Glu	Leu	Asn	Gln 60	Ser	Lys	Asn	Glu
Thr Thr 65	Ala	Pro	Ser	Glu 70	Asn	ГЛа	Thr	Thr	Glu 75	ГЛа	Val	Asp	Ser	Arg 80
Gln Leu	Lys	Asp	Asn 85	Thr	Gln	Thr	Ala	Thr 90	Ala	Asp	Gln	Pro	Lys 95	Val
Thr Met	Ser	Asp 100	Ser	Ala	Thr	Val	Lys 105	Glu	Thr	Ser	Ser	Asn 110	Met	Gln
Ser Pro	Gln 115	Asn	Ala	Thr	Ala	Ser 120	Gln	Ser	Thr	Thr	Gln 125	Thr	Ser	Asn
Val Thr 130	Thr	Asn	Asp	Lys	Ser 135	Ser	Thr	Thr	Tyr	Ser 140	Asn	Glu	Thr	Asp
Lys Ser 145	Asn	Leu	Thr	Gln 150	Ala	ГÀа	Asn	Val	Ser 155	Thr	Thr	Pro	ГЛа	Thr 160
Thr Thr	Ile	Lys	Gln 165	Arg	Ala	Leu	Asn	Arg 170	Met	Ala	Val	Asn	Thr 175	Val
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Asn Tyr 225	Thr	Ile	Asp	Asp 230	Ser	Val	Lys	Glu	Gly 235	Asp	Thr	Phe	Thr	Phe 240
Lys Tyr	Gly	Gln	Tyr 245	Phe	Arg	Pro	Gly	Ser 250	Val	Arg	Leu	Pro	Ser 255	Gln
Thr Gln	Asn	Leu 260	Tyr	Asn	Ala	Gln	Gly 265	Asn	Ile	Ile	Ala	Lys 270	Gly	Ile
Tyr Asp	Ser 275	Lys	Thr	Asn	Thr	Thr 280	Thr	Tyr	Thr	Phe	Thr 285	Asn	Tyr	Val
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Asn	Gln	Lys	Gly 340	Gln	Gln	Leu	Ile	Ser 345	Ser	Thr	Asn	Tyr	Ile 350	Asn	Asn
Glu	Asp	Leu 355	Ser	Arg	Asn	Met	Thr 360	Val	Tyr	Val	Asn	Gln 365	Pro	Lys	Lys
Thr	Tyr 370	Thr	Lys	Glu	Thr	Phe 375	Val	Thr	Asn	Leu	Thr 380	Gly	Tyr	Lys	Phe
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Gln	Phe	Val	Asp	Ser 405	Phe	Thr	Pro	Asp	Thr 410	Ser	ГЛа	Leu	Lys	Asp 415	Val
Thr	Gly	Gln	Phe 420	Asp	Val	Ile	Tyr	Ser 425	Asn	Asp	Asn	Lys	Thr 430	Ala	Thr
Val	Asp	Leu 435	Leu	Asn	Gly	Gln	Ser 440	Ser	Ser	Asp	Lys	Gln 445	Tyr	Ile	Ile
Gln	Gln 450	Val	Ala	Tyr	Pro	Asp 455	Asn	Ser	Ser	Thr	Asp 460	Asn	Gly	Lys	Ile
Asp 465	Tyr	Thr	Leu	Glu	Thr 470	Gln	Asn	Gly	Lys	Ser 475	Ser	Trp	Ser	Asn	Ser 480
Tyr	Ser	Asn	Val	Asn 485	Gly	Ser	Ser	Thr	Ala 490	Asn	Gly	Asp	Gln	Lys 495	Lys
Tyr	Asn	Leu	Gly 500	Asp	Tyr	Val	Trp	Glu 505	Asp	Thr	Asn	ГÀв	Asp 510	Gly	Lys
Gln	Asp	Ala 515	Asn	Glu	Lys	Gly	Ile 520	Lys	Gly	Val	Tyr	Val 525	Ile	Leu	ГЛа
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Lys 545	Tyr	Gln	Phe	Thr	Gly 550	Leu	Ser	Asn	Gly	Thr 555	Tyr	Ser	Val	Glu	Phe 560
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Ala	Asp	Asn 595	Met	Thr	Leu	Asp	Ser 600	Gly	Phe	Tyr	Lys	Thr 605	Pro	Lys	Tyr
Ser	Leu 610	Gly	Asp	Tyr	Val	Trp 615	Tyr	Asp	Ser	Asn	Lys 620	Asp	Gly	ГЛа	Gln
Asp 625	Ser	Thr	Glu	ГÀа	Gly 630	Ile	Lys	Gly	Val	Lys 635	Val	Thr	Leu	Gln	Asn 640
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Pro Pro Asn Ser Pro Lys Asn Gly Thr Asn Ala Ser Asn Lys Asn Glu 935 Ala Lys Asp Ser Lys Glu Pro Leu Pro Asp Thr Gly Ser Glu Asp Glu Ala Asn Thr Ser Leu Ile Trp Gly Leu Leu Ala Ser Leu Gly Ser Leu 970 Leu Leu Phe Arg Arg Lys Lys Glu Asn Lys Asp Lys Lys <210> SEQ ID NO 23 <211> LENGTH: 584 <212> TYPE: PRT <213 > ORGANISM: Staphylococcus sp. <400> SEQUENCE: 23 Met Lys Phe Lys Ser Leu Ile Thr Thr Thr Leu Ala Leu Gly Val Leu Ala Ser Thr Gly Ala Asn Phe Asn Asn Asn Glu Ala Ser Ala Ala Ala Lys Pro Leu Asp Lys Ser Ser Ser Ser Leu His His Gly Tyr Ser Lys 40 Val His Val Pro Tyr Ala Ile Thr Val Asn Gly Thr Ser Gln Asn Ile 55 Leu Ser Ser Leu Thr Phe Asn Lys Asn Gln Asn Ile Ser Tyr Lys Asp Leu Glu Asp Arg Val Lys Ser Val Leu Lys Ser Asp Arg Gly Ile Ser Asp Ile Asp Leu Arg Leu Ser Lys Gln Ala Lys Tyr Thr Val Tyr Phe 105 Lys Asn Gly Thr Lys Lys Val Ile Asp Leu Lys Ala Gly Ile Tyr Thr 120 Ala Asp Leu Ile Asn Thr Ser Glu Ile Lys Ala Ile Asn Ile Asn Val Asp Thr Lys Lys Gln Val Glu Asp Lys Lys Lys Asp Lys Ala Asn Tyr Gln Val Pro Tyr Thr Ile Thr Val Asn Gly Thr Ser Gln Asn Ile Leu Ser Asn Leu Thr Phe Asn Lys Asn Gln Asn Ile Ser Tyr Lys Asp Leu Glu Asp Lys Val Lys Ser Val Leu Glu Ser Asn Arg Gly Ile Thr Asp Val Asp Leu Arg Leu Ser Lys Gln Ala Lys Tyr Thr Val Asn Phe Lys Asn Gly Thr Lys Lys Val Ile Asp Leu Lys Ser Gly Ile Tyr Thr Ala 230 Asn Leu Ile Asn Ser Ser Asp Ile Lys Ser Ile Asn Ile Asn Val Asp 250 Thr Lys Lys His Ile Glu Asn Lys Ala Lys Arg Asn Tyr Gln Val Pro Tyr Ser Ile Asn Leu Asn Gly Thr Ser Thr Asn Ile Leu Ser Asn Leu 280 Ser Phe Ser Asn Lys Pro Trp Thr Asn Tyr Lys Asn Leu Thr Ser Gln

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Val	Ser 1160		n Met	: Asn	Ala	Gly 116		nr G	ln :	Leu	Tyr	His 1170	Ile	Thr	Ala
Thr	Asn 1175		а Туг	His	Lys	Asp 118		er G	lu i	Asp	Phe	Tyr 1185	Ile	Ser	Leu
Lys	Ile 1190		e Asp	Val	Lys	Gln 119		ro G	lu (	Gly	Asp	Gln 1200	Arg	Val	Tyr
Arg	Thr 1205		Thr	Tyr	Asp	Leu 121		nr T	hr i	Asp	Glu	Ile 1215	Ser	Lys	Val
Lys	Gln 1220		≀ Ph∈	e Ile	Asn	Ala		≅n A	rg i	Asp	Val	Ile 1230	Thr	Leu	Ala
Glu	Gly 1235	_	) Ile	e Ser	Val	Thr		sn T	hr :	Pro	Asn	Gly 1245	Ala	Asn	Val
Ser	Thr 1250		• Thr	: Val	Asn	Ile 125		sn L	ys (	Gly	Arg	Leu 1260	Thr	Lys	Ser

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Phe	Ala 1265		Asn	Leu	Ala	Asn 1270		Asn	Phe	Leu	Arg 1275		Val	Asn
Phe	Pro 1280	Gln	Asp	Tyr	Thr	Val 1285		Trp	Thr	Asn	Ala 1290		Ile	Ala
Asn	Arg 1295	Pro	Thr	Asp	Gly	Gly 1300		Ser	Trp	Ser	Asp 1305		His	Lys
Ser	Leu 1310	Ile	Tyr	Arg	Tyr	Asp 1315		Thr	Leu	Gly	Thr 1320		Ile	Thr
Thr	Asn 1325	Asp	Ile	Leu	Thr	Met 1330		Lys	Ala	Thr	Thr 1335		Val	Pro
Gly	Leu 1340	Arg	Asn	Asn	Ile	Thr 1345		Asn	Glu	Lys	Ser 1350		Ala	Glu
Ala	Gly 1355	Gly	Arg	Pro	Asn	Phe 1360		Thr	Thr	Gly	Tyr 1365	Ser	Gln	Ser
Asn	Ala 1370	Thr	Thr	Asp	Gly	Gln 1375		Gln	Phe	Thr	Leu 1380	Asn	Gly	Gln
Val	Ile 1385	Gln	Val	Leu	Asp	Ile 1390		Asn	Pro	Ser	Asn 1395	Gly	Tyr	Gly
Gly	Gln 1400	Pro	Val	Thr	Asn	Ser 1405	Asn	Thr	Arg	Ala	Asn 1410		Ser	Asn
Ser	Thr 1415	Val	Val	Asn	Val	Asn 1420		Pro	Ala	Ala	Asn 1425	Gly	Ala	Gly
Ala	Phe 1430	Thr	Ile	Asp	His	Val 1435		Lys	Ser	Asn	Ser 1440		His	Asn
Ala	Ser 1445	Asp	Ala	Val	Tyr	Lys 1450		Gln	Leu	Tyr	Leu 1455		Pro	Tyr
Gly	Pro 1460	ГÀв	Gln	Tyr	Val	Glu 1465		Leu	Asn	Gln	Asn 1470		Gly	Asn
Thr	Thr 1475	Asp	Ala	Ile	Asn	Ile 1480	_	Phe	Val	Pro	Ser 1485	Asp	Leu	Val
Asn	Pro 1490	Thr	Ile	Ser	Val	Gly 1495	Asn	Tyr	Thr	Asn	His 1500		Val	Phe
Ser	Gly 1505	Glu	Thr	Phe	Thr	Asn 1510		Ile	Thr	Ala	Asn 1515		Asn	Phe
Gly	Val 1520	Gln	Ser	Val	Thr	Val 1525	Pro	Asn	Thr	Ser	Gln 1530		Thr	Gly
	Val 1535	_	Asn	Asn	His	Gln 1540		Val	Ser	Ala	Thr 1545		Pro	Asn
Val	Thr 1550	Ser	Ala	Thr	Asn	Lys 1555		Ile	Asn	Leu	Leu 1560	Ala	Thr	Asp
Thr	Ser 1565	Gly	Asn	Thr	Ala	Thr 1570		Ser	Phe	Asn	Val 1575		Val	Lys
Pro	Leu 1580	Arg	Asp	Lys	Tyr	Arg 1585		Gly	Thr	Ser	Ser 1590	Thr	Ala	Ala
Asn	Pro 1595	Val	Arg	Ile	Ala	Asn 1600		Ser	Asn	Asn	Ala 1605	Thr	Val	Ser
Gln	Ala 1610	Asp	Gln	Thr	Thr	Ile 1615	Ile	Asn	Ser	Leu	Thr 1620	Phe	Thr	Glu
Thr	Val 1625	Pro	Asn	Arg	Ser	Tyr 1630		Arg	Ala	Ser	Ala 1635		Glu	Ile
Thr	Ser	Lys	Thr	Val	Ser	Asn	Val	Ser	Arg	Thr	Gly	Asn	Asn	Ala

	1640					1645					1650			
Asn	Val 1655		Val	Thr	Val	Thr 1660		Gln	Asp	Gly	Thr 1665	Thr	Ser	Thr
Val	Thr 1670	Val	Pro	Val	Lys	His 1675			Pro		Ile 1680	Val	Ala	His
Ser	His 1685		Thr	Val	Gln	Gly 1690			Phe		Ala 1695	Gly	Asn	Gly
Ser	Ser 1700	Ala	Ser	Asp	Tyr	Phe 1705		Leu	Ser	Asn	Gly 1710	Ser	Asp	Ile
Ala	Asp 1715	Ala	Thr	Ile	Thr	Trp 1720			Gly		Ala 1725	Pro	Asn	Lys
Asp	Asn 1730	Thr	Arg	Ile	Gly	Glu 1735		Ile	Thr	Val	Thr 1740	Ala	His	Ile
Leu	Ile 1745		Gly			Thr 1750			Thr	_	Thr 1755	Ala	Thr	Tyr
Lys	Val 1760	Val		Thr		Pro 1765		His	Val	Phe	Glu 1770	Thr	Ala	Arg
Gly	Val 1775	Leu	Tyr	Pro	Gly	Val 1780			Met		Asp 1785	Ala	Lys	Gln
Tyr	Val 1790	-	Pro	Val	Asn	Asn 1795		Trp			Asn 1800	Ala	Gln	His
Met	Asn 1805	Phe	Gln	Phe	Val	Gly 1810	Thr	Tyr		Pro	Asn 1815	Lys	Asp	Val
Val	Gly 1820	Ile				Leu 1825		Arg			Tyr 1830	Asp	Asn	Arg
Gln	Thr 1835	Glu		Leu		Ile 1840		Ser			Lys 1845	Pro	Asp	Pro
Pro	Arg 1850		Asp			Ser 1855					Ala 1860		Leu	Thr
Asn		Glu		Lys	Val	Asn 1870	Asn						Ser	Val
Lys		Phe		Ala	Asp	Asn 1885	Thr	Pro	Leu	Asn		Thr	Asn	Ile
Thr		Gly	Ser		Phe	Ser 1900	Ser	Val	Val	Thr		Ser	Asp	Ala
Leu			Gly	Gly	Ile	Lys 1915	Ala						Met	Asn
Asn	Val	Thr				Gln 1930	Asp				Gln		Val	Thr
Val		Arg	Asn	Glu	Ser	Val	Asp	Ser	Asn	Asp		Ala	Thr	Val
Thr		Thr	Pro	Gln	Leu	1945 Gln	Ala	Thr	Thr	Glu	-	Ala	Val	Phe
Ile	1955 Lys	Gly	Gly	Asp	Gly	1960 Phe		Phe	Gly	His	1965 Val	Glu	Arg	Phe
	1970	Ī	Ī	Ī	Ī	1975	Ī		·		1980			
тте	1985	ASI	Pro	PIO	nis	Gly 1990	AIA	ınr	val	AIA	1995	HIS	Asp	ser
Pro	Asp 2000	Thr	Trp	Lys	Asn	Thr 2005		Gly	Asn	Thr	His 2010	ГÀа	Thr	Ala
Val	Val 2015	Thr	Leu	Pro	Asn	Gly 2020	Gln	Gly	Thr	Arg	Asn 2025	Val	Glu	Val

Pro	Val 2030	Lys	Val	Tyr	Pro	Val 2035	Ala	Asn	Ala	Lys	Ala 2040	Pro	Ser	Arg
Asp	Val 2045	ГÀз	Gly	Gln	Asn	Leu 2050	Thr	Asn	Gly	Thr	Asp 2055	Ala	Met	Asn
Tyr	Ile 2060	Thr	Phe	Asp	Pro	Asn 2065	Thr	Asn	Thr	Asn	Gly 2070	Ile	Thr	Ala
Ala	Trp 2075	Ala	Asn	Arg	Gln	Gln 2080	Pro	Asn	Asn	Gln	Gln 2085	Ala	Gly	Val
Gln	His 2090	Leu	Asn	Val	Asp	Val 2095	Thr	Tyr	Pro	Gly	Ile 2100	Ser	Ala	Ala
ГÀа	Arg 2105	Val	Pro	Val	Thr	Val 2110	Asn	Val	Tyr	Gln	Phe 2115	Glu	Phe	Pro
Gln	Thr 2120	Thr	Tyr	Thr	Thr	Thr 2125	Val	Gly	Gly	Thr	Leu 2130	Ala	Ser	Gly
Thr	Gln 2135	Ala	Ser	Gly	Tyr	Ala 2140	His	Met	Gln	Asn	Ala 2145	Thr	Gly	Leu
Pro	Thr 2150	Asp	Gly	Phe	Thr	Tyr 2155	Lys	Trp	Asn	Arg	Asp 2160	Thr	Thr	Gly
Thr	Asn 2165	Asp	Ala	Asn	Trp	Ser 2170	Ala	Met	Asn	Lys	Pro 2175	Asn	Val	Ala
Lys	Val 2180	Val	Asn	Ala	Lys	Tyr 2185	Asp	Val	Ile	Tyr	Asn 2190	Gly	His	Thr
Phe	Ala 2195	Thr	Ser	Leu	Pro	Ala 2200	Lys	Phe	Val	Val	Lys 2205	Asp	Val	Gln
Pro	Ala 2210	Lys	Pro	Thr	Val	Thr 2215	Glu	Thr	Ala	Ala	Gly 2220	Ala	Ile	Thr
Ile	Ala 2225	Pro	Gly	Ala	Asn	Gln 2230	Thr	Val	Asn	Thr	His 2235	Ala	Gly	Asn
Val	Thr 2240	Thr	Tyr	Ala	Asp	Lys 2245	Leu	Val	Ile	Lys	Arg 2250	Asn	Gly	Asn
Val	Val 2255	Thr	Thr	Phe	Thr	Arg 2260	Arg	Asn	Asn	Thr	Ser 2265	Pro	Trp	Val
ГЛа	Glu 2270	Ala	Ser	Ala	Ala	Thr 2275	Val	Ala	Gly	Ile	Ala 2280	Gly	Thr	Asn
Asn	Gly 2285	Ile	Thr	Val	Ala	Ala 2290	Gly	Thr	Phe	Asn	Pro 2295	Ala	Asp	Thr
Ile	Gln 2300	Val	Val	Ala	Thr	Gln 2305	Gly	Ser	Gly	Glu	Thr 2310	Val	Ser	Asp
Glu	Gln 2315	Arg	Ser	Asp	Asp	Phe 2320	Thr	Val	Val	Ala	Pro 2325	Gln	Pro	Asn
Gln	Ala 2330	Thr	Thr	Lys	Ile	Trp 2335	Gln	Asn	Gly	His	Ile 2340	Asp	Ile	Thr
Pro	Asn 2345	Asn	Pro	Ser	Gly	His 2350	Leu	Ile	Asn	Pro	Thr 2355	Gln	Ala	Met
Asp	Ile 2360	Ala	Tyr	Thr	Glu	Lys 2365	Val	Gly	Asn	Gly	Ala 2370	Glu	His	Ser
Lys	Thr 2375	Ile	Asn	Val	Val	Arg 2380	Gly	Gln	Asn	Asn	Gln 2385	Trp	Thr	Ile
Ala	Asn 2390	Lys	Pro	Asp	Tyr	Val 2395	Thr	Leu	Asp	Ala	Gln 2400	Thr	Gly	Lys

Val															
Ser Thr	Val			Asn	Ala	Asn			Lys	Pro	Asn		Ser	Ile	Thr
2435	Ile			Lys	Ala	Gly		_	His	Ser	Val		Ser	Asn	Pro
Asn Ala   Val Gin Val Ala   Asn   Lys Arg Thr Ala Thr   11e Lys Asn   2475	Ser			Thr	Ala	Pro			His	Thr	Val		Thr	Thr	Glu
2465   2470   2475   2476   2476   2480   2480   2481   2485	Ile			Asp	Tyr	Gly			Val	Thr	Ala		Glu	Ile	Asn
2480   2485   2490	Asn			Gln	Val	Ala			Arg	Thr	Ala		Ile	Lys	Asn
2500	Gly			Met	Pro	Thr		Leu	Ala	Gly	Gly		Thr	Thr	Thr
2510	Ile			Thr	Val	Thr			Asp	Gly	Ser		Glu	Glu	Val
Pro Gly   2540   Thr Ile Thr Gln Tyr   2545   Thr Glu Ala Met His   2550   Asn Ala Gln   2555	Gln			Ile	Phe	Thr			Asp	Lys	Arg		Leu	Ile	Thr
2540	Ala			His	Leu	Asp			Val	Ser	Thr		Gly	Lys	ГЛа
2555	Pro			Ile	Thr	Gln			Asn	Ala	Met		Asn	Ala	Gln
2570	Gln			Asn	Thr	Ala			Glu	Ala	Gln		Val	Ile	Asn
2585	Asn			Ala	Thr	Pro		Gln	Val	Ser	Asp		Leu	Thr	Гла
2600       2605       2610         Leu Gln Ser Ser Val Asn Gln 2620       Val Pro Ser Thr Ala Gly Met Thr 2625         Gln Gln Gln Ser Ile Asp Asn Tyr 2635       Asn Ala Lys Lys Arg 2640       Glu Ala Glu 2640         Thr Glu Ile Thr Ala Ala Gln 2650       Arg Val Ile Asp Asn 2655       Gly Asp Ala 2655         Thr Ala Gln Gln Ile Ser Asp 2660       Glu Lys His Arg Val 2670       Asp Asn Ala 2670         Leu Thr Ala Leu Asn Gln Ala 2680       Lys His Asp Leu Thr 2685       Ala Asp Thr 2685         His Ala 2690       Leu Glu Gln Ala Val 2695       Gln Gln Leu Asn Arg 2700       Thr Gly Thr 2705         Thr Thr Gly Lys Lys Pro Ala 2710       Ser Ile Thr Ala Tyr Asn Asn Ser 2715         Ile Arg Ala Leu Gln Ser Asp 2725       Leu Thr Ser Ala Lys 2730       Asn Ser Ala 2730         Asn Ala Ile Ile Gln Lys Pro Ile Arg Thr Val Gln Glu Val Gln	Val			Ala	Gln	Thr			Asp	Gln	Ala	_	Ala	Leu	Leu
2615	Gln		_	Glu	Asp	Asn			Leu	Val	Thr		Lys	Asn	Asn
Thr Glu Ile Thr Ala Ala Gln Arg Val Ile Asp Asn Gly Asp Ala 2655  Thr Ala Gln Gln Ile Ser Asp 2665  Thr Ala Leu Asn Gln Ala Lys His Arg Val Asp Asn Ala 2670  Leu Thr Ala Leu Asn Gln Ala Lys His Asp Leu Thr 2685  His Ala Leu Glu Gln Ala Val Gln Gln Leu Asn Arg 2700  Thr Thr Gly Lys Lys Pro Ala Ser Ile Thr Ala Tyr Asn Asn Ser 2715  Ile Arg Ala Leu Gln Ser Asp 2725  Asn Ala Ile Ile Gln Lys Pro Ile Arg Thr Val Gln Glu Val Gln	Leu			Ser	Val	Asn			Pro	Ser	Thr		Gly	Met	Thr
Thr Ala Gln Gln Ile Ser Asp Glu Lys His Arg Val Asp Asn Ala 2660  Leu Thr Ala Leu Asn Gln Ala Lys His Asp Leu Thr 2675  His Ala Leu Glu Gln Ala Val Gln Gln Leu Asn Arg 2700  Thr Thr Gly Lys Lys Pro Ala Ser Ile Thr Ala Tyr Asn Asn Ser 2715  Ile Arg Ala Leu Gln Ser Asp Leu Thr Ser Ala Lys Asn Ser Asn Ser 2720  Asn Ala Ile Ile Gln Lys Pro Ile Arg Thr Val Gln Glu Val Gln	Gln			Ile	Asp	Asn		Asn	Ala	Lys	Lys		Glu	Ala	Glu
Leu Thr Ala Leu Asn Gln Ala Lys His Asp Leu Thr 2675  His Ala Leu Glu Gln Ala Val 2695  Thr Thr Gly Lys Lys Pro Ala 2710  Ser Ile Thr Ala Tyr Asn Asn Ser 2710  Asn Ala Ile Ile Gln Lys Pro Ile Arg Thr Val Gln Glu Val Gln  Glu Val Gln  Glu Val Gln  Glu Val Gln  Glu Val Gln  Glu Val Gln	Thr			Thr	Ala	Ala			Val	Ile	Asp		Gly	Asp	Ala
2675  2680  2685  His Ala Leu Glu Gln Ala Val Gln Gln Leu Asn Arg 2700  Thr Thr Gly Lys Lys Pro Ala Ser Ile Thr Ala Tyr 2715  Ile Arg Ala Leu Gln Ser Asp 2725  Leu Thr Ser Ala Lys Asn Ser 2730  Asn Ala Ile Ile Gln Lys Pro Ile Arg Thr Val Gln Glu Val Gln	Thr			Gln	Ile	Ser	_		Lys	His	Arg		Asp	Asn	Ala
2690 2695 2700  Thr Thr Gly Lys Lys Pro Ala Ser Ile Thr Ala Tyr Asn Asn Ser 2715  Ile Arg Ala Leu Gln Ser Asp Leu Thr Ser Ala Lys Asn Ser Ala 2720  Asn Ala Ile Ile Gln Lys Pro Ile Arg Thr Val Gln Glu Val Gln			Ala	Leu	Asn			-	His	Asp				Asp	Thr
2705 2710 2715  Ile Arg Ala Leu Gln Ser Asp Leu Thr Ser Ala Lys Asn Ser Ala 2720 2725 2730  Asn Ala Ile Ile Gln Lys Pro Ile Arg Thr Val Gln Glu Val Gln	His			Glu	Gln	Ala			Gln	Leu	Asn	_	Thr	Gly	Thr
2720 2725 2730  Asn Ala Ile Ile Gln Lys Pro Ile Arg Thr Val Gln Glu Val Gln	Thr			Lys	Lys	Pro			Ile	Thr	Ala		Asn	Asn	Ser
	Ile	_		Leu	Gln	Ser	-		Thr	Ser	Ala	-	Asn	Ser	Ala
2735 2740 2745	Asn			Ile	Gln	Lys			Arg	Thr	Val		Glu	Val	Gln
Ser Ala Leu Thr Asn Val Asn Arg Val Asn Glu Arg Leu Thr Gln 2750 2755 2760	Ser			Thr	Asn	Val		Arg	Val	Asn	Glu	_	Leu	Thr	Gln
Ala Ile Asn Gln Leu Val Pro Leu Ala Asp Asn Ser Ala Leu Lys 2765 2770 2775	Ala	Ile	Asn	Gln	Leu	Val			Ala	Asp	Asn	Ser	Ala	Leu	ГЛа
2.12		_,					_ , , ,					_ , , _			

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	278	)				2785					2790			
Tł	nr Asp 279		Met	Thr	Gln	Ser 2800		Ile	Gln	Ala	Tyr 2805		Asn	Ala
ГΖ	e Arg 281		Gly	Gln	Thr	Glu 2815		Thr	Asn	Ala	Gln 2820		Val	Ile
As	en Asn 282		Asp	Ala	Thr	Asp 2830		Gln	Ile	Ala	Ala 2835	Glu	Lys	Thr
ЬΣ	/s Val 284			Lys			Ser		Lys		Ala 2850	Ile	Ala	Gly
Le	eu Thr 285		Asp			Pro 2860		Gln	Thr	Ala	Lys 2865		Gln	Leu
G.	ln Asn 287	_		Asp		Pro 2875		Ser	Thr	Thr	Gly 2880		Thr	Ser
A.	la Ser 288		Ala	Ala	Phe	Asn 2890	Glu	Lys	Leu	Ser	Ala 2895		Arg	Thr
ĿΣ	zs Ile 290		Glu	Ile	Asp	Arg 2905		Leu	Ala	Ser	His 2910		Asp	Val
A.	la Thr 291		_	Gln		Val 2920		Ala	Ala	Asn	Ala 2925	Ala	Lys	Ser
A.	la Leu 293		Gln	Ala	Arg	Asn 2935		Leu	Thr	Val	Asp 2940		Ala	Pro
Le	eu Glu 294			Lys		Gln 2950		Gln	His	Ser	Ile 2955	Asp	Thr	Gln
Tł	nr Ser 296		Thr		Met		Gln	Asp				Ala	Tyr	Asn
A.	La Lys 297	Leu					Asn		Ile	Gln	Gln	Ile	Asn	Gln
Vá	al Leu 299	Ala	Gly				Val			Ile			Asn	Thr
Se	er Thr	Ala				Lys	Ser	Asp		Asp	His		Arg	Gln
A.	300 La Leu	Thr					Pro				Ala	_	Thr	Gln
Le	302 eu Glu			Ile			Pro						Met	Thr
	303 nr Ala	5				3040					3045	_		
	305	)				3055			_		3060			
G.	ln Lys 306		. Thr	Glu	Ile	Asn 3070		Val	Leu	Asn	Gly 3075		Pro	Thr
Vá	al Gln 308		Ile	Asn	Asp	Lys 3085	Val	Thr	Glu	Ala	Asn 3090		Ala	Lys
As	sp Gln 309		. Asn	Thr	Ala	Arg 3100		Gly	Leu	Thr	Leu 3105	_	Arg	Gln
Pı	o Ala 311		Thr	Thr	Leu	His 3115	_	Ala	Ser	Asn	Leu 3120	Asn	Gln	Ala
G.	ln Gln 312		. Asn	Phe	Thr	Gln 3130	Gln	Ile	Asn	Ala	Ala 3135	Gln	Asn	His
A.	la Ala 314		Glu	Thr	Ile	Lys 3145		Asn	Ile	Thr	Ala 3150	Leu	Asn	Thr
A.	La Met	Thr	Lys	Leu	Lys	Asp		Val	Ala	Asp	Asn	Asn	Thr	Ile
	315	5				3160					3165			

ГЛа	Ser 3170	Asp	Gln	Asn	Tyr	Thr 3175	Asp	Ala	Thr	Pro	Ala 3180	Asn	Lys	Gln
Ala	Tyr 3185	Asp	Asn	Ala	Val	Asn 3190	Ala	Ala	Lys	Gly	Val 3195	Ile	Gly	Glu
Thr	Thr 3200	Asn	Pro	Thr	Met	Asp 3205	Val	Asn	Thr	Val	Asn 3210	Gln	Lys	Ala
Ala	Ser 3215	Val	ГЛа	Ser	Thr	Lys 3220	Asp	Ala	Leu	Asp	Gly 3225	Gln	Gln	Asn
Leu	Gln 3230	Arg	Ala	Lys	Thr	Glu 3235	Ala	Thr	Asn	Ala	Ile 3240	Thr	His	Ala
Ser	Asp 3245	Leu	Asn	Gln	Ala	Gln 3250	Lys	Asn	Ala	Leu	Thr 3255	Gln	Gln	Val
Asn	Ser 3260	Ala	Gln	Asn	Val	Gln 3265	Ala	Val	Asn	Asp	Ile 3270	ГЛа	Gln	Thr
Thr	Gln 3275	Ser	Leu	Asn	Thr	Ala 3280	Met	Thr	Gly	Leu	Lys 3285	Arg	Gly	Val
Ala	Asn 3290	His	Asn	Gln	Val	Val 3295	Gln	Ser	Asp	Asn	Tyr 3300	Val	Asn	Ala
Asp	Thr 3305	Asn	ГЛа	Lys	Asn	Asp 3310	Tyr	Asn	Asn	Ala	Tyr 3315	Asn	His	Ala
Asn	Asp 3320	Ile	Ile	Asn	Gly	Asn 3325	Ala	Gln	His	Pro	Val 3330	Ile	Thr	Pro
Ser	Asp 3335	Val	Asn	Asn	Ala	Leu 3340	Ser	Asn	Val	Thr	Ser 3345	Lys	Glu	His
Ala	Leu 3350	Asn	Gly	Glu	Ala	Lys 3355	Leu	Asn	Ala	Ala	3360 Tàs	Gln	Glu	Ala
Asn	Thr 3365	Ala	Leu	Gly	His	Leu 3370	Asn	Asn	Leu	Asn	Asn 3375	Ala	Gln	Arg
Gln	Asn 3380	Leu	Gln	Ser	Gln	Ile 3385	Asn	Gly	Ala	His	Gln 3390	Ile	Asp	Ala
Val	Asn 3395	Thr	Ile	Lys	Gln	Asn 3400	Ala	Thr	Asn	Leu	Asn 3405	Ser	Ala	Met
Gly	Asn 3410	Leu	Arg	Gln	Ala	Val 3415	Ala	Asp	Lys	Asp	Gln 3420	Val	ГÀа	Arg
Thr	Glu 3425	Asp	Tyr	Ala	Asp	Ala 3430	Asp	Thr	Ala	ГÀа	Gln 3435	Asn	Ala	Tyr
Asn	Ser 3440	Ala	Val	Ser	Ser	Ala 3445	Glu	Thr	Ile	Ile	Asn 3450	Gln	Thr	Thr
Asn	Pro 3455	Thr	Met	Ser	Val	Asp 3460	Asp	Val	Asn	Arg	Ala 3465	Thr	Ser	Ala
Val	Thr 3470	Ser	Asn	Lys	Asn	Ala 3475	Leu	Asn	Gly	Tyr	Glu 3480	ГЛа	Leu	Ala
Gln	Ser 3485	Lys	Thr	Asp	Ala	Ala 3490	Arg	Ala	Ile	Asp	Ala 3495	Leu	Pro	His
Leu	Asn 3500	Asn	Ala	Gln	Lys	Ala 3505	Asp	Val	Lys	Ser	Lys 3510	Ile	Asn	Ala
Ala	Ser 3515	Asn	Ile	Ala	Gly	Val 3520	Asn	Thr	Val	Lys	Gln 3525	Gln	Gly	Thr
Asp	Leu 3530	Asn	Thr	Ala	Met	Gly 3535	Asn	Leu	Gln	Gly	Ala 3540	Ile	Asn	Asp

Glu	Gln 3545	Thr	Thr	Leu	Asn	Ser 3550	Gln	Asn	Tyr	Gln	Asp 3555	Ala	Thr	Pro
Ser	Lys 3560	Lys	Thr	Ala	Tyr	Thr 3565	Asn	Ala	Val	Gln	Ala 3570		rys	Asp
Ile	Leu 3575	Asn	Lys	Ser	Asn	Gly 3580	Gln	Asn	Lys	Thr	Lys 3585	Asp	Gln	Val
Thr	Glu 3590	Ala	Met	Asn	Gln	Val 3595	Asn	Ser	Ala	ГÀа	Asn 3600		Leu	Asp
Gly	Thr 3605	Arg	Leu	Leu	Asp	Gln 3610	Ala	Lys	Gln	Thr	Ala 3615	_	Gln	Gln
Leu	Asn 3620	Asn	Met	Thr	His	Leu 3625	Thr	Thr	Ala	Gln	Lys		Asn	Leu
Thr	Asn 3635	Gln	Ile	Asn	Ser	Gly 3640	Thr	Thr	Val	Ala	Gly 3645	Val	Gln	Thr
Val	Gln 3650	Ser	Asn	Ala	Asn	Thr 3655	Leu	Asp	Gln	Ala	Met 3660	Asn	Thr	Leu
Arg	Gln 3665	Ser	Ile	Ala	Asn	Lys 3670	Asp	Ala	Thr	Lys	Ala 3675	Ser	Glu	Asp
Tyr	Val 3680	Asp	Ala	Asn	Asn	Asp 3685	Lys	Gln	Thr	Ala	Tyr 3690	Asn	Asn	Ala
Val	Ala 3695	Ala	Ala	Glu	Thr	Ile 3700	Ile	Asn	Ala	Asn	Ser 3705	Asn	Pro	Glu
Met	Asn 3710	Pro	Ser	Thr	Ile	Thr 3715	Gln	Lys	Ala	Glu	Gln 3720	Val	Asn	Ser
Ser	Lys 3725	Thr	Ala	Leu	Asn	Gly 3730	Asp	Glu	Asn	Leu	Ala 3735	Ala	Ala	Lys
Gln	Asn 3740	Ala	Lys	Thr	Tyr	Leu 3745	Asn	Thr	Leu	Thr	Ser 3750	Ile	Thr	Asp
Ala	Gln 3755	Lys	Asn	Asn	Leu	Ile 3760	Ser	Gln	Ile	Thr	Ser 3765	Ala	Thr	Arg
Val	Ser 3770	Gly	Val	Asp	Thr	Val 3775	Lys	Gln	Asn	Ala	Gln 3780	His	Leu	Asp
Gln	Ala 3785	Met	Ala	Ser	Leu	Gln 3790	Asn	Gly	Ile	Asn	Asn 3795	Glu	Ser	Gln
Val	Lys 3800	Ser	Ser	Glu	Lys	Tyr 3805	Arg	Asp	Ala	Asp	Thr 3810	Asn	Lys	Gln
Gln	Glu 3815	Tyr	Asp	Asn	Ala	Ile 3820	Thr	Ala	Ala	Lys	Ala 3825	Ile	Leu	Asn
ГÀа	Ser 3830	Thr	Gly	Pro	Asn	Thr 3835	Ala	Gln	Asn	Ala	Val 3840	Glu	Ala	Ala
Leu	Gln 3845	Arg	Val	Asn	Asn	Ala 3850	Lys	Asp	Ala	Leu	Asn 3855	Gly	Asp	Ala
Lys	Leu 3860	Ile	Ala	Ala	Gln	Asn 3865	Ala	Ala	Lys	Gln	His 3870	Leu	Gly	Thr
Leu	Thr 3875	His	Ile	Thr	Thr	Ala 3880	Gln	Arg	Asn	Asp	Leu 3885		Asn	Gln
Ile	Ser 3890	Gln	Ala	Thr	Asn	Leu 3895	Ala	Gly	Val	Glu	Ser 3900	Val	Lys	Gln
Asn	Ala 3905	Asn	Ser	Leu	Asp	Gly 3910	Ala	Met	Gly	Asn	Leu 3915	Gln	Thr	Ala
Ile		Asp	Lys	Ser	Gly		Leu	Ala	Ser	Gln	Asn	Phe	Leu	Asp

	3920					3925					3930			
Ala	Asp 3935		Gln	Lys	Arg	Asn 3940		Tyr	Asn	Gln	Ala 3945		Ser	Ala
Ala	Glu 3950		Ile	Leu	Asn	Lys 3955	Gln		Gly		Asn 3960	Thr	Ala	Lys
Thr	Ala 3965		Glu	Gln	Ala	Leu 3970	Asn	Asn	Val	Asn	Asn 3975	Ala	Lys	His
Ala	Leu 3980			Thr		Asn 3985	Leu	Asn	Asn	Ala	Ъув 3990	Gln	Ala	Ala
Ile	Thr 3995	Ala	Ile	Asn	Gly	Ala 4000	Ser	Asp	Leu	Asn	Gln 4005	Lys	Gln	ГÀа
Asp	Ala 4010		Lys	Ala	Gln	Ala 4015	Asn	Gly	Ala	Gln	Arg 4020		Ser	Asn
Ala	Gln 4025	_	Val	Gln	His	Asn 4030	Ala	Thr	Glu	Leu	Asn 4035	Thr	Ala	Met
Gly	Thr 4040		Lys	His	Ala	Ile 4045	Ala		Lys		Asn 4050	Thr	Leu	Ala
Ser	Ser 4055	_	Tyr	Val	Asn	Ala 4060	Asp	Ser	Thr	Lys	Gln 4065	Asn	Ala	Tyr
Thr	Thr 4070		Val	Thr	Asn	Ala 4075	Glu	His	Ile	Ile	Ser 4080	Gly	Thr	Pro
Thr	Val 4085					Ser 4090	Glu	Val	Thr	Ala	Ala 4095	Ala	Asn	Gln
Val	Asn 4100			Lys		Glu 4105		Asn				Arg	Leu	Arg
Glu	Ala 4115	_	Gln	Asn		Asn 4120		Ala				Leu	Thr	Gln
Leu	Asn 4130			Gln		Ala 4135			Lys		Gln 4140	Val	Gly	Gln
Ala	Asn 4145	Arg					Gln	Thr	Val	Gln		Asn	Gly	Gln
Ala	Leu 4160	Asn					Gly					Ile	Ala	Asn
Glu	Thr 4175			Lys				Asn				Ala	Ser	Pro
Asn	Asn 4190			Thr	Tyr		Ser	Ala	Val	Ser		Ala	Lys	Gly
Ile	Ile 4205												Ala	Ile
Thr	Gln 4220	Ala	Thr	Thr	Gln		Asn	Asn	Ala	Lys			Leu	Asn
Gly	Ala	Glu	Asn	Leu	Arg	Asn	Ala	Gln	Asn	Thr	Ala	-	Gln	Asn
Leu	4235 Asn	Thr	Leu	Ser	His			Asn	Asn	Gln	4245 Lys		Ala	Ile
Ser	4250 Ser		Ile	Asp	Arg	4255 Ala		His	Val	Ser	4260 Glu	Val	Thr	Ala
	4265					4270	Ī				4275			
Thr	Lys 4280	Asn	Ala	Ala	Thr	Glu 4285	ьeu	Asn	Thr	GIn	Met 4290	GIY	Asn	ьeu
Glu	Gln 4295	Ala	Ile	His	Asp	Gln 4300	Asn	Thr	Val	Lys	Gln 4305	Ser	Val	ГÀЗ

Phe	Thr 4310	Asp	Ala	Asp	Lys	Ala 4315	Lys	Arg	Asp	Ala	Tyr 4320	Thr	Asn	Ala
Val	Ser 4325	Arg	Ala	Glu	Ala	Ile 4330	Leu	Asn	Lys	Thr	Gln 4335	Gly	Ala	Asn
Thr	Ser 4340	ГЛа	Gln	Asp	Val	Glu 4345	Ala	Ala	Ile	Gln	Asn 4350	Val	Ser	Ser
Ala	Lys 4355	Asn	Ala	Leu	Asn	Gly 4360	Asp	Gln	Asn	Val	Thr 4365	Asn	Ala	Lys
Asn	Ala 4370	Ala	Lys	Asn	Ala	Leu 4375	Asn	Asn	Leu	Thr	Ser 4380	Ile	Asn	Asn
Ala	Gln 4385	Lys	Arg	Asp	Leu	Thr 4390	Thr	Lys	Ile	Asp	Gln 4395	Ala	Thr	Thr
Val	Ala 4400	Gly	Val	Glu	Ala	Val 4405	Ser	Asn	Thr	Ser	Thr 4410	Gln	Leu	Asn
Thr	Ala 4415	Met	Ala	Asn	Leu	Gln 4420	Asn	Gly	Ile	Asn	Asp 4425	ГЛа	Thr	Asn
Thr	Leu 4430	Ala	Ser	Glu	Asn	Tyr 4435	His	Asp	Ala	Asp	Ser 4440	Asp	Lys	Lys
Thr	Ala 4445	Tyr	Thr	Gln	Ala	Val 4450	Thr	Asn	Ala	Glu	Asn 4455	Ile	Leu	Asn
Lys	Asn 4460	Ser	Gly	Ser	Asn	Leu 4465	Asp	ГÀа	Thr	Ala	Val 4470	Glu	Asn	Ala
Leu	Ser 4475	Gln	Val	Ala	Asn	Ala 4480	Lys	Gly	Ala	Leu	Asn 4485	Gly	Asn	His
Asn	Leu 4490	Glu	Gln	Ala	Lys	Ser 4495	Asn	Ala	Asn	Thr	Thr 4500	Ile	Asn	Gly
Leu	Gln 4505	His	Leu	Thr	Thr	Ala 4510	Gln	Lys	Asp	Lys	Leu 4515	Lys	Gln	Gln
Val	Gln 4520	Gln	Ala	Gln	Asn	Val 4525	Ala	Gly	Val	Asp	Thr 4530	Val	Lys	Ser
Ser	Ala 4535	Asn	Thr	Leu	Asn	Gly 4540	Ala	Met	Gly	Thr	Leu 4545	Arg	Asn	Ser
Ile	Gln 4550	Asp	Asn	Thr	Ala	Thr 4555	ГÀа	Asn	Gly	Gln	Asn 4560	Tyr	Leu	Asp
Ala	Thr 4565	Glu	Arg	Asn	Lys	Thr 4570	Asn	Tyr	Asn	Asn	Ala 4575	Val	Asp	Ser
Ala	Asn 4580	Gly	Val	Ile	Asn	Ala 4585	Thr	Ser	Asn	Pro	Asn 4590	Met	Asp	Ala
Asn	Ala 4595	Ile	Asn	Gln	Ile	Ala 4600	Thr	Gln	Val	Thr	Ser 4605	Thr	ГÀа	Asn
Ala	Leu 4610	Asp	Gly	Thr	His	Asn 4615	Leu	Thr	Gln	Ala	Lys 4620	Gln	Thr	Ala
Thr	Asn 4625	Ala	Ile	Asp	Gly	Ala 4630	Thr	Asn	Leu	Asn	Lys 4635	Ala	Gln	Lys
Asp	Ala 4640	Leu	ГЛа	Ala	Gln	Val 4645	Thr	Ser	Ala	Gln	Arg 4650	Val	Ala	Asn
Val	Thr 4655	Ser	Ile	Gln	Gln	Thr 4660	Ala	Asn	Glu	Leu	Asn 4665	Thr	Ala	Met
Gly	Gln 4670	Leu	Gln	His	Gly	Ile 4675	Asp	Asp	Glu	Asn	Ala 4680	Thr	Lys	Gln

Thr	Gln 4685		Tyr	Arg	Asp	Ala 4690		Gln	Ser	ГÀв	Lys 4695	Thr	Ala	Tyr
Asp	Gln 4700	Ala	Val	Ala	Ala	Ala 4705		Ala	Ile	Leu	Asn 4710	Lys	Gln	Thr
Gly	Ser 4715	Asn	Ser	Asp	Lys	Ala 4720	Ala	Val	Asp	Arg	Ala 4725	Leu	Gln	Gln
Val	Thr 4730	Ser	Thr	Lys	Asp	Ala 4735		Asn	Gly	Asp	Ala 4740	Lys	Leu	Ala
Glu	Ala 4745	Lys	Ala	Ala	Ala	Lys 4750	Gln	Asn	Leu	Gly	Thr 4755	Leu	Asn	His
Ile	Thr 4760	Asn	Ala	Gln	Arg	Thr 4765		Leu	Glu	Gly	Gln 4770	Ile	Asn	Gln
Ala	Thr 4775	Thr	Val	Asp	Gly	Val 4780	Asn	Thr	Val	Lys	Thr 4785	Asn	Ala	Asn
Thr	Leu 4790	Asp	Gly	Ala	Met	Asn 4795		Leu	Gln	Gly	Ser 4800	Ile	Asn	Aap
Lys	Asp 4805	Ala	Thr	Leu	Arg	Asn 4810	Gln	Asn	Tyr	Leu	Asp 4815	Ala	Asp	Glu
Ser	Lys 4820	Arg	Asn	Ala	Tyr	Thr 4825		Ala	Val	Thr	Ala 4830	Ala	Glu	Gly
Ile	Leu 4835	Asn	Lys	Gln	Thr	Gly 4840	Gly	Asn	Thr	Ser	Lys 4845	Ala	Asp	Val
Asp	Asn 4850	Ala	Leu	Asn	Ala	Val 4855		Arg	Ala	Lys	Ala 4860	Ala	Leu	Asn
Gly	Ala 4865	Asp	Asn	Leu	Arg	Asn 4870	Ala	Lys	Thr	Ser	Ala 4875	Thr	Asn	Thr
Ile	Asp 4880	Gly	Leu	Pro	Asn	Leu 4885		Gln	Leu	Gln	Lys 4890	Asp	Asn	Leu
Lys	His 4895	Gln	Val	Glu	Gln	Ala 4900	Gln	Asn	Val	Ala	Gly 4905	Val	Asn	Gly
Val	Lys 4910	Asp	Lys	Gly	Asn	Thr 4915		Asn	Thr	Ala	Met 4920	Gly	Ala	Leu
Arg	Thr 4925	Ser	Ile	Gln	Asn	Asp 4930	Asn	Thr	Thr	Lys	Thr 4935	Ser	Gln	Asn
Tyr	Leu 4940	Asp	Ala	Ser	Asp	Ser 4945		Lys	Asn	Asn	Tyr 4950	Asn	Thr	Ala
Val	Asn 4955	Asn	Ala	Asn	Gly	Val 4960	Ile	Asn	Ala	Thr	Asn 4965	Asn	Pro	Asn
Met	Asp 4970		Asn	Ala	Ile	Asn 4975		Met	Ala	Asn	Gln 4980	Val	Asn	Thr
Thr	Lys 4985		Ala	Leu	Asn	Gly 4990		Gln	Asn	Leu	Ala 4995	Gln	Ala	Lys
Thr	Asn 5000	Ala	Thr	Asn	Thr	Ile 5005	Asn	Asn	Ala	His	Asp 5010	Leu	Asn	Gln
Lys	Gln 5015	Lys	Asp	Ala	Leu	Lys 5020		Gln	Val	Asn	Asn 5025	Ala	Gln	Arg
Val	Ser 5030	_	Ala	Asn	Asn	Val 5035		His	Thr	Ala	Thr 5040	Glu	Leu	Asn
Ser	Ala 5045	Met	Thr	Ala	Leu	Lys		Ala	Ile	Ala	Asp 5055	Lys	Glu	Arg
Thr		Ala	Ser	Gly	Asn	Tyr	Val	Asn	Ala	Asp	Gln	Glu	Lys	Arg

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	5060					5065					5070			
Gl	n Ala 5075	_	Asp	Ser	Lys	Val 5080		Asn	Ala	Glu	Asn 5085	Ile	Ile	Ser
Gl	y Thr 5090		Asn	Ala	Thr	Leu 5095		Val	Asn	Asp	Val 5100	Asn	Ser	Ala
Al	a Ser 5105		Val	Asn	Ala	Ala 5110	_	Thr	Ala	Leu	Asn 5115	Gly	Asp	Asn
As	n Leu 5120	_	Val	Ala	ГЛа	Glu 5125		Ala	Asn	Asn	Thr 5130	Ile	Asp	Gly
Le	u Ala 5135		Leu	Asn	Asn	Ala 5140		-		-	Leu 5145	Lys	Glu	Gln
Va	l Gln 5150		Ala	Thr	Thr	Leu 5155		Gly	Val		Thr 5160	Val	Lys	Asn
Se	r Ser 5165		Thr	Leu	Asn	Thr 5170		Met	Lys	Gly	Leu 5175	Arg	Asp	Ser
Il	e Ala 5180		Glu	Ala	Thr	Ile 5185	_	Ala	Gly		Asn 5190	Tyr	Thr	Asp
Al	a Ser 5195	Pro	Asn	Asn	Arg		Glu	Tyr	Asp			Val	Thr	Ala
Al	a Lys 5210	Ala	Ile	Ile	Asn		Thr	Ser	Asn	Pro		Met	Glu	Pro
As	n Thr 5225	Ile	Thr	Gln	Val		Ser	Gln	Val		Thr		Glu	Gln
Al	a Leu	Asn	Gly	Ala	Arg	Asn	Leu	Ala	Gln	Ala			Thr	Ala
Ly	5240 s Asn	Asn	Leu	Asn	Asn		Thr	Ser	Ile		5250 Asn	Ala	Gln	Lys
As	5255 p Ala		Thr	Arq	Ser	5260 Ile		Gly	Ala		5265 Thr	Val	Ala	Gly
	5270 1 Asn					5275					5280			
	5285					5290					5295			
Hi	s Ser 5300					Ile 5305					Gln 5310	Thr	Lys	Gln
Th	r Gln 5315					Ala 5320					Lув 5325	Ser	Ala	Tyr
As	p Gln 5330					Ala 5335					Thr 5340		Ala	Ser
Gl	y Gln 5345		Val	Asp	Lys	Ala 5350	Ala	Val	Glu	Gln	Ala 5355	Leu	Gln	Asn
Va	1 Asn 5360		Thr	ГХа	Thr	Ala 5365	Leu	Asn	Gly	Asp	Ala 5370		Leu	Asn
Gl	u Ala 5375	-	Ala	Ala	Ala	5380 Lys		Thr	Leu	Gly	Thr 5385	Leu	Thr	His
Il	e Asn 5390	Asn	Ala	Gln	Arg		Ala	Leu	Asp	Asn		Ile	Thr	Gln
Al	a Thr	Asn	Val	Glu	Gly	Val		Thr	Val	Lys	Ala	Lys	Ala	Gln
G1	5405 n Leu		Glv	Ala	Met	5410 Glv	G] n	Leu	Gl u	Thr	5415 Ser	IJe	Ara	Asp
	5420					5425					5430			
Ly	s Asp 5435		Thr	Leu	Gln	Ser 5440	Gln	Asn	Tyr	Gln	Asp 5445	Ala	Asp	Asp

Ala	Lys 5450	Arg	Thr	Ala	Tyr	Ser 5455	Gln	Ala	Val	Asn	Ala 5460	Ala	Ala	Thr
Ile	Leu 5465	Asn	Lys	Thr	Ala	Gly 5470	Gly	Asn	Thr	Pro	Lys 5475	Ala	Asp	Val
Glu	Arg 5480	Ala	Met	Gln	Ala	Val 5485	Thr	Gln	Ala	Asn	Thr 5490	Ala	Leu	Asn
Gly	Ile 5495	Gln	Asn	Leu	Asp	Arg 5500	Ala	Lys	Gln	Ala	Ala 5505	Asn	Thr	Ala
Ile	Thr 5510	Asn	Ala	Ser	Asp	Leu 5515	Asn	Thr	Lys	Gln	Lys 5520	Glu	Ala	Leu
Lys	Ala 5525	Gln	Val	Thr	Ser	Ala 5530	Gly	Arg	Val	Ser	Ala 5535	Ala	Asn	Gly
Val	Glu 5540	His	Thr	Ala	Thr	Glu 5545	Leu	Asn	Thr	Ala	Met 5550	Thr	Ala	Leu
Lys	Arg 5555	Ala	Ile	Ala	Asp	Lys 5560	Ala	Glu	Thr	Lys	Ala 5565	Ser	Gly	Asn
Tyr	Val 5570	Asn	Ala	Asp	Ala	Asn 5575	Lys	Arg	Gln	Ala	Tyr 5580	Asp	Glu	Lys
Val	Thr 5585	Ala	Ala	Glu	Asn	Ile 5590	Val	Ser	Gly	Thr	Pro 5595	Thr	Pro	Thr
Leu	Thr 5600	Pro	Ala	Asp	Val	Thr 5605	Asn	Ala	Ala	Thr	Gln 5610	Val	Thr	Asn
Ala	Lys 5615	Thr	Gln	Leu	Asn	Gly 5620	Asn	His	Asn	Leu	Glu 5625	Val	Ala	Lys
Gln	Asn 5630	Ala	Asn	Thr	Ala	Ile 5635	Asp	Gly	Leu	Thr	Ser 5640	Leu	Asn	Gly
Pro	Gln 5645	Lys	Ala	Lys	Leu	Lys 5650	Glu	Gln	Val	Gly	Gln 5655	Ala	Thr	Thr
Leu	Pro 5660	Asn	Val	Gln	Thr	Val 5665	Arg	Asp	Asn	Ala	Gln 5670	Thr	Leu	Asn
Thr	Ala 5675	Met	Lys	Gly	Leu	Arg 5680	Asp	Ser	Ile	Ala	Asn 5685	Glu	Ala	Thr
Ile	Lys 5690	Ala	Gly	Gln	Asn	Tyr 5695	Thr	Asp	Ala	Ser	Gln 5700	Asn	Lys	Gln
Thr	Asp 5705	Tyr	Asn	Ser	Ala	Val 5710	Thr	Ala	Ala	Lys	Ala 5715	Ile	Ile	Gly
Gln	Thr 5720	Thr	Ser	Pro	Ser	Met 5725	Asn	Ala	Gln	Glu	Ile 5730	Asn	Gln	Ala
Lys	Asp 5735	Gln	Val	Thr	Ala	Lys 5740	Gln	Gln	Ala	Leu	Asn 5745	Gly	Gln	Glu
Asn	Leu 5750	Arg	Thr	Ala	Gln	Thr 5755	Asn	Ala	Lys	Gln	His 5760	Leu	Asn	Gly
Leu	Ser 5765	Asp	Leu	Thr	Asp	Ala 5770	Gln	Lys	Asp	Ala	Val 5775	Lys	Arg	Gln
Ile	Glu 5780	Gly	Ala	Thr	His	Val 5785	Asn	Glu	Val	Thr	Gln 5790	Ala	Gln	Asn
Asn	Ala 5795	Asp	Ala	Leu	Asn	Thr 5800	Ala	Met	Thr	Asn	Leu 5805	Lys	Asn	Gly
Ile	Gln 5810	Asp	Gln	Asn	Thr	Ile 5815	Lys	Gln	Gly	Val	Asn 5820	Phe	Thr	Asp

Ala	Asp 5825	Glu	Ala	Lys	Arg	Asn 5830		Tyr	Thr	Asn	Ala 5835		Thr	Gln
Ala	Glu 5840	Gln	Ile	Leu	Asn	Lys 5845		Gln	Gly	Pro	Asn 5850		Ser	Lys
Asp	Gly 5855	Val	Glu	Thr	Ala	Leu 5860		Asn	Val	Gln	Arg 5865	Ala	Lys	Asn
Glu	Leu 5870	Asn	Gly	Asn	Gln	Asn 5875		Ala	Asn	Ala	Lys 5880		Thr	Ala
Lys	Asn 5885	Ala	Leu	Asn	Asn	Leu 5890		Ser	Ile	Asn	Asn 5895	Ala	Gln	Lys
Glu	Ala 5900	Leu	Lys	Ser	Gln	Ile 5905		Gly	Ala	Thr	Thr 5910		Ala	Gly
Val	Asn 5915	Gln	Val	Ser	Thr	Thr 5920		Ser	Glu	Leu	Asn 5925	Thr	Ala	Met
Ser	Asn 5930	Leu	Gln	Asn	Gly	Ile 5935		Asp	Glu	Ala	Ala 5940	Thr	Lys	Ala
Ala	Gln 5945	Lys	Tyr	Thr	Asp	Ala 5950		Arg	Glu	ГÀа	Gln 5955	Thr	Ala	Tyr
Asn	Asp 5960	Ala	Val	Thr	Ala	Ala 5965		Thr	Leu	Leu	Asp 5970	ГÀа	Thr	Ala
Gly	Ser 5975	Asn	Asp	Asn	Lys	Ala 5980		Val	Glu	Gln	Ala 5985	Leu	Gln	Arg
Val	Asn 5990	Thr	Ala	Lys	Thr	Ala 5995		Asn	Gly	Asp	Glu 6000	Arg	Leu	Asn
Glu	Ala 6005	ГÀв	Asn	Thr	Ala	Lys 6010		Gln	Val	Ala	Thr 6015	Met	Ser	His
Leu	Thr 6020	Asp	Ala	Gln	ГÀв	Ala 6025		Leu	Thr	Ser	Gln 6030	Ile	Glu	Ser
Gly	Thr 6035	Thr	Val	Ala	Gly	Val 6040		Gly	Ile	Gln	Ala 6045	Asn	Ala	Gly
Thr	Leu 6050	Asp	Gln	Ala	Met	Asn 6055		Leu	Arg	Gln	Ser 6060	Ile	Ala	Ser
Lys	Asp 6065	Ala	Thr	Lys	Ser	Ser 6070		Asp	Tyr	Gln	Asp 6075	Ala	Asn	Ala
Asp	Leu 6080	Gln	Asn	Ala	Tyr	Asn 6085	Asp	Ala	Val	Thr	Asn 6090	Ala	Glu	Gly
	Ile 6095	Ser	Ala	Thr		Asn 6100		Glu	Met	Asn	Pro 6105		Thr	Ile
Asn	Gln 6110	_	Ala	Ser	Gln	Val 6115		Ser	Ala	ГÀа	Ser 6120	Ala	Leu	Asn
Gly	Asp 6125	Glu	Lys	Leu	Ala	Ala 6130		Lys	Gln	Thr	Ala 6135	ГÀа	Ser	Asp
Ile	Gly 6140	Arg	Leu	Thr	Asp	Leu 6145		Asn	Ala	Gln	Arg 6150	Thr	Ala	Ala
Asn	Ala 6155	Glu	Val	Asp	Gln	Ala 6160		Asn	Leu	Ala	Ala 6165	Val	Thr	Ala
Ala	Lys 6170	Asn	Lys	Ala	Thr	Ser 6175	Leu	Asn	Thr	Ala	Met 6180	Gly	Asn	Leu
ГÀа	His 6185	Ala	Leu	Ala	Glu	Lys 6190	Asp	Asn	Thr	Lys	Arg 6195	Ser	Val	Asn
Tyr	Thr	Asp	Ala	Asp	Gln	Pro	Lys	Gln	Gln	Ala	Tyr	Asp	Thr	Ala

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_		6200					6205					6210			
V		Thr 6215		Ala	Glu	Ala	Ile 6220		Asn	Ala		Gly 6225	Ser	Asn	Ala
A		Glu 6230	Thr	Gln	Val	Gln	Ala 6235		Leu	Asn	Gln	Leu 6240	Asn	Gln	Ala
L		Asn 6245	Asp	Leu	Asn	Gly	Asp 6250		Lys			Gln 6255	Ala	Lys	Glu
S		Ala 6260		Arg	Ala	Leu	Ala 6265		Tyr	Ser	Asn	Leu 6270	Asn	Asn	Ala
G:		Ser 6275	Thr	Ala	Ala	Ile	Ser 6280	Gln	Ile	Asp	Asn	Ala 6285	Thr	Thr	Val
A.		Gly 6290	Val	Thr	Ala	Ala	Gln 6295	Asn	Thr	Ala	Asn	Glu 6300	Leu	Asn	Thr
A.		Met 6305	Gly	Gln	Leu	Gln	Asn 6310		Ile	Asn	Asp	Gln 6315	Asn	Thr	Val
L		Gln 6320	Gln	Val	Asn	Phe	Thr 6325		Ala			Gly 6330	Lys	Lys	Asp
A.		Tyr 6335		Asn	Ala	Val	Thr 6340		Ala	Gln	Gly	Ile 6345	Leu	Asp	ГЛа
A.		His 6350	Gly	Gln	Asn	Met	Thr 6355	_	Ala	Gln	Val	Glu 6360	Ala	Ala	Leu
A		Gln 6365	Val	Thr	Thr	Ala	Lys 6370		Ala	Leu	Asn	Gly 6375	Asp	Ala	Asn
V		Arg 6380		Ala	Lys	Ser	Asp 6385		Lys	Ala	Asn	Leu 6390	Gly	Thr	Leu
Tl		His 6395	Leu	Asn	Asn	Ala	Gln 6400	_	Gln	Asp	Leu	Thr 6405	Ser	Gln	Ile
G:		Gly 6410		Thr	Thr	Val	Asn 6415		Val	Asn	Gly	Val 6420	Lys	Thr	Lys
A.		Gln 6425		Leu	Asp	Gly	Ala 6430	Met	Gln	Arg	Leu	Gln 6435		Ala	Ile
A.		Asn 6440	Lys	Asp	Gln	Thr	Lys 6445		Ser	Glu	Asn	Tyr 6450		Asp	Ala
A		Pro 6455	Thr	Lys	Lys	Thr	Ala 6460		Asp	Asn	Ala	Ile 6465	Thr	Gln	Ala
G.		Ser 6470	-			_	Asp 6475		_			_	_	Lys	Gln
A.		Val 6485	Glu	Gln	Ala	Ile	Gln 6490	Ser	Val	Thr	Ser	Thr 6495	Glu	Asn	Ala
L		Asn 6500	Gly	Asp	Ala	Asn	Leu 6505	Gln	Arg	Ala	Lys	Thr 6510	Glu	Ala	Ile
G:		Ala 6515	Ile	Asp	Asn	Leu	Thr 6520	His	Leu	Asn	Thr	Pro 6525	Gln	Lys	Thr
A.		Leu 6530	-	Gln	Gln	Val	Asn 6535	Ala	Ala	Gln	Arg	Val 6540	Ser	Gly	Val
Tl		Asp 6545	Leu	Lys	Asn	Ser	Ala 6550	Thr	Ser	Leu	Asn	Asn 6555	Ala	Met	Asp
G:		Leu 6560	Lys	Gln	Ala	Ile	Ala 6565	Asp	His	Asp	Thr	Ile 6570	Val	Ala	Ser
G:	ly i		Tyr	Thr	Asn	Ala	Ser 6580	Pro	Asp	Lys	Gln		Ala	Tyr	Thr
		00/0					0,500					0,000			

Asp	Ala 6590	Tyr	Asn	Ala	Ala	Lys 6595	Asn	Ile	Val	Asn	Gly 6600	Ser	Pro	Asn
Val	Ile 6605	Thr	Asn	Ala	Ala	Asp 6610	Val	Thr	Ala	Ala	Thr 6615	Gln	Arg	Val
Asn	Asn 6620	Ala	Glu	Thr	Gly	Leu 6625	Asn	Gly	Asp	Thr	Asn 6630	Leu	Ala	Thr
Ala	Lys 6635	Gln	Gln	Ala	ГЛа	Asp 6640	Ala	Leu	Arg	Gln	Met 6645	Thr	His	Leu
Ser	Asp 6650	Ala	Gln	Lys	Gln	Ser 6655	Ile	Thr	Gly	Gln	Ile 6660	Asp	Ser	Ala
Thr	Gln 6665	Val	Thr	Gly	Val	Gln 6670	Ser	Val	Lys	Asp	Asn 6675	Ala	Thr	Asn
Leu	Asp	Asn	Ala	Met	Asn	Gln 6685	Leu	Arg	Asn	Ser	Ile 6690	Ala	Asn	Lys
Aap	Asp 6695	Val	ГÀа	Ala	Ser	Gln 6700	Pro	Tyr	Val	Asp	Ala 6705	Asp	Arg	Aap
ГÀв	Gln 6710	Asn	Ala	Tyr	Asn	Thr 6715	Ala	Val	Thr	Asn	Ala 6720	Glu	Asn	Ile
Ile	Asn 6725	Ala	Thr	Ser	Gln	Pro 6730	Thr	Leu	Asp	Pro	Ser 6735	Ala	Val	Thr
Gln	Ala 6740	Ala	Asn	Gln	Val	Ser 6745	Thr	Asn	Lys	Thr	Ala 6750	Leu	Asn	Gly
Ala	Gln 6755	Asn	Leu	Ala	Asn	Lys 6760	Lys	Gln	Glu	Thr	Thr 6765	Ala	Asn	Ile
Asn	Gln 6770	Leu	Ser	His	Leu	Asn 6775	Asn	Ala	Gln	Lys	Gln 6780	Asp	Leu	Asn
Thr	Gln 6785	Val	Thr	Asn	Ala	Pro 6790	Asn	Ile	Ser	Thr	Val 6795	Asn	Gln	Val
ГÀв	Thr 6800	Lys	Ala	Glu	Gln	Leu 6805	Asp	Gln	Ala	Met	Glu 6810	Arg	Leu	Ile
Asn	Gly 6815	Ile	Gln	Asp	Lys	Asp 6820	Gln	Val	Lys	Gln	Ser 6825	Val	Asn	Phe
Thr	Asp 6830	Ala	Asp	Pro	Glu	Lys 6835	Gln	Thr	Ala	Tyr	Asn 6840	Asn	Ala	Val
Thr	Ala 6845	Ala	Glu	Asn	Ile	Ile 6850	Asn	Gln	Ala	Asn	Gly 6855	Thr	Asn	Ala
Asn	Gln 6860	Ser	Gln	Val	Glu	Ala 6865	Ala	Leu	Ser	Thr	Val 6870	Thr	Thr	Thr
ГÀа	Gln 6875	Ala	Leu	Asn	Gly	Asp 6880	Arg	ГЛа	Val	Thr	Asp 6885	Ala	ГÀа	Asn
Asn	Ala 6890	Asn	Gln	Thr	Leu	Ser 6895	Thr	Leu	Asp	Asn	Leu 6900	Asn	Asn	Ala
Gln	Lys 6905	Gly	Ala	Val	Thr	Gly 6910	Asn	Ile	Asn	Gln	Ala 6915	His	Thr	Val
Ala	Glu 6920	Val	Thr	Gln	Ala	Ile 6925	Gln	Thr	Ala	Gln	Glu 6930	Leu	Asn	Thr
Ala	Met 6935	Gly	Asn	Leu	Lys	Asn 6940	Ser	Leu	Asn	Asp	Lys 6945	Asp	Thr	Thr
Leu	Gly 6950	Ser	Gln	Asn	Phe	Ala 6955	Asp	Ala	Asp	Pro	Glu 6960	Lys	Lys	Asn

Ala	Tyr 6965	Asn	Glu	Ala	Val	His 6970	Asn	Ala	Glu	Asn	Ile 6975	Leu	Asn	ГÀв
Ser	Thr 6980	Gly	Thr	Asn	Val	Pro 6985	Lys	Asp	Gln	Val	Glu 6990	Ala	Ala	Met
Asn	Gln 6995	Val	Asn	Ala	Thr	Lys 7000	Ala	Ala	Leu	Asn	Gly 7005	Thr	Gln	Asn
Leu	Glu 7010	Lys	Ala	Lys	Gln	His 7015	Ala	Asn	Thr	Ala	Ile 7020	Asp	Gly	Leu
Ser	His 7025	Leu	Thr	Asn	Ala	Gln 7030	Lys	Glu	Ala	Leu	Lys 7035	Gln	Leu	Val
Gln	Gln 7040	Ser	Thr	Thr	Val	Ala 7045	Glu	Ala	Gln	Gly	Asn 7050	Glu	Gln	Lys
Ala	Asn 7055	Asn	Val	Asp	Ala	Ala 7060	Met	Asp	Lys	Leu	Arg 7065	Gln	Ser	Ile
Ala	Asp 7070	Asn	Ala	Thr	Thr	Lys 7075	Gln	Asn	Gln	Asn	Tyr 7080	Thr	Asp	Ala
Ser	Gln 7085	Asn	Lys	Lys	Asp	Ala 7090	Tyr	Asn	Asn	Ala	Val 7095	Thr	Thr	Ala
Gln	Gly 7100	Ile	Ile	Asp	Gln	Thr 7105	Thr	Ser	Pro	Thr	Leu 7110	Asp	Pro	Thr
Val	Ile 7115	Asn	Gln	Ala	Ala	Gly 7120	Gln	Val	Ser	Thr	Thr 7125	Lys	Asn	Ala
Leu	Asn 7130	Gly	Asn	Glu	Asn	Leu 7135	Glu	Ala	Ala	Lys	Gln 7140	Gln	Ala	Ser
Gln	Ser 7145	Leu	Gly	Ser	Leu	Asp 7150	Asn	Leu	Asn	Asn	Ala 7155	Gln	Lys	Gln
Thr	Val 7160	Thr	Asp	Gln	Ile	Asn 7165	Gly	Ala	His	Thr	Val 7170	Asp	Glu	Ala
Asn	Gln 7175	Ile	Lys	Gln	Asn	Ala 7180	Gln	Asn	Leu	Asn	Thr 7185	Ala	Met	Gly
Asn	Leu 7190	Lys	Gln	Ala	Ile	Ala 7195	Asp	Lys	Asp	Ala	Thr 7200	Lys	Ala	Thr
Val	Asn 7205	Phe	Thr	Asp	Ala	Asp 7210	Gln	Ala	Lys	Gln	Gln 7215	Ala	Tyr	Asn
Thr	Ala 7220	Val	Thr	Asn	Ala	Glu 7225	Asn	Ile	Ser	Lys	Ala 7230	Asn	Gly	Asn
Ala	Thr 7235	Gln	Ala	Glu	Val	Glu 7240	Gln	Ala	Ile	Lys	Gln 7245	Val	Asn	Ala
Ala	Lys 7250	Gln	Ala	Leu	Asn	Gly 7255	Asn	Ala	Asn	Val	Gln 7260	His	Ala	ГЛа
Asp	Glu 7265	Ala	Thr	Ala	Leu	Ile 7270	Asn	Ser	Ser	Asn	Asp 7275	Leu	Asn	Gln
Ala	Gln 7280	Lys	Asp	Ala	Leu	Lys 7285	Gln	Gln	Val	Gln	Asn 7290	Ala	Thr	Thr
Val	Ala 7295	Gly	Val	Asn	Asn	Val 7300	Lys	Gln	Thr	Ala	Gln 7305	Glu	Leu	Asn
Asn	Ala 7310	Met	Thr	Gln	Leu	Lys 7315	Gln	Gly	Ile	Ala	Asp 7320	Lys	Glu	Gln
Thr	Lys 7325	Ala	Asp	Gly	Asn	Phe 7330	Val	Asn	Ala	Asp	Pro 7335	Asp	Lys	Gln
Asn	Ala	Tyr	Asn	Gln	Ala	Val	Ala	Lys	Ala	Glu	Ala	Leu	Ile	Ser

	7340					7345					7350			
Ala	Thr 7355		Asp	Val	Val	Val 7360		Pro	Ser	Glu	Ile 7365		Ala	Ala
Leu	Asn 7370		Val	Thr	Gln	Ala 7375		Asn	Asp	Leu	Asn 7380	_	Asn	Thr
Asn	Leu 7385	Ala	Thr	Ala	Lys	Gln 7390		Val	Gln	His	Ala 7395	Ile	Asp	Gln
Leu	Pro 7400	Asn	Leu	Asn	Gln	Ala 7405	Gln		Asp		Tyr 7410	Ser	Lys	Gln
Ile	Thr 7415	Gln	Ala	Thr	Leu	Val 7420		Asn	Val	Asn	Ala 7425	Ile	Gln	Gln
Ala	Ala 7430	Thr	Thr	Leu	Asn	Asp 7435	Ala	Met	Thr	Gln	Leu 7440	rys	Gln	Gly
Ile	Ala 7445		Lys	Ala	Gln	Ile 7450		Gly			Asn 7455	Tyr	His	Asp
Ala	Asp 7460			ГХа		Thr 7465		Tyr			Ala 7470	Val	Thr	Lys
Ala	Glu 7475	Glu	Leu	Leu	Lys	Gln 7480		Thr	Asn	Pro	Thr 7485	Met	Asp	Pro
Asn	Thr 7490	Ile	Gln	Gln	Ala	Leu 7495		Lys			Asp 7500	Thr	Asn	Gln
Ala	Leu 7505	Asn		Asn		Lys 7510	Leu		Asp		Lys 7515	Gln	Asp	Ala
Lys	Thr 7520	Thr	Leu		Thr	Leu 7525	_		Leu	Asn	Asp 7530	Ala	Gln	Lys
Gln			Thr	Thr	Gln	Val 7540			Ala	Pro		Ile	Ala	Thr
Val		Asn	Val		Gln	Asn						Asn	Ala	Met
Thr					Ala	Leu 7570	Gln		Lys	Thr		Thr	Leu	Asn
Ser		Asn	Phe	Thr	Asp	Ala 7585	Asp	Gln	Ala		Lys	Asp	Ala	Tyr
Thr		Ala	Val	Ser	His	7585 Ala 7600	Glu		Ile		,550		Ala	Asn
Gly	Ser					Thr	Glu	Val	Glu	Gln			Gln	Arg
Val						7615 Ala					Asp	Asn	Val	Gln
Arg		Lys	Asp	Ala	Ala	7630 Lys	Gln	Val	Ile	Thr		Ala	Asn	Asp
Leu	7640 Asn	Gln	Ala	Gln	Lys	7645 Asp	Ala	Leu	Lys	Gln	7650 Gln	Val	Asp	Ala
	7655				-	7660			-		7665		-	
	7670					Val 7675					7680			
Asp	Leu 7685	Asn	Gln	Ala	Met	Thr 7690	Gln	Leu	Lys	Gln	Gly 7695	Ile	Ala	Asp
Lys	Asp 7700	Gln	Thr	Lys	Ala	Asn 7705	Gly	Asn	Phe	Val	Asn 7710	Ala	Asp	Thr
Asp	Lys 7715	Gln	Asn	Ala	Tyr	Asn 7720	Asn	Ala	Val	Ala	His 7725	Ala	Glu	Gln

Ile	Ile 7730	Ser	Gly	Thr	Pro	Asn 7735	Ala	Asn	Val	Asp	Pro 7740	Gln	Gln	Val
Ala	Gln 7745	Ala	Leu	Gln	Gln	Val 7750	Asn	Gln	Ala	Lys	Gly 7755	Asp	Leu	Asn
Gly	Asn 7760	His	Asn	Leu	Gln	Val 7765	Ala	Lys	Asp	Asn	Ala 7770	Asn	Thr	Ala
Ile	Asp 7775	Gln	Leu	Pro	Asn	Leu 7780	Asn	Gln	Pro	Gln	Lys 7785	Thr	Ala	Leu
Lys	Asp 7790	Gln	Val	Ser	His	Ala 7795	Glu	Leu	Val	Thr	Gly 7800	Val	Asn	Ala
Ile	Lys 7805	Gln	Asn	Ala	Asp	Ala 7810	Leu	Asn	Asn	Ala	Met 7815	Gly	Thr	Leu
Lys	Gln 7820	Gln	Ile	Gln	Ala	Asn 7825	Ser	Gln	Val	Pro	Gln 7830	Ser	Val	Asp
Phe	Thr 7835	Gln	Ala	Asp	Gln	Asp 7840	Lys	Gln	Gln	Ala	Tyr 7845	Asn	Asn	Ala
Ala	Asn 7850	Gln	Ala	Gln	Gln	Ile 7855	Ala	Asn	Gly	Ile	Pro 7860	Thr	Pro	Val
Leu	Thr 7865	Pro	Asp	Thr	Val	Thr 7870	Gln	Ala	Val	Thr	Thr 7875	Met	Asn	Gln
Ala	Lys 7880	Asp	Ala	Leu	Asn	Gly 7885	Asp	Glu	Lys	Leu	Ala 7890	Gln	Ala	Lys
Gln	Glu 7895	Ala	Leu	Ala	Asn	Leu 7900	Asp	Thr	Leu	Arg	Asp 7905	Leu	Asn	Gln
Pro	Gln 7910	Arg	Asp	Ala	Leu	Arg 7915	Asn	Gln	Ile	Asn	Gln 7920	Ala	Gln	Ala
Leu	Ala 7925	Thr	Val	Glu	Gln	Thr 7930	Lys	Gln	Asn	Ala	Gln 7935	Asn	Val	Asn
Thr	Ala 7940	Met	Ser	Asn	Leu	Lys 7945	Gln	Gly	Ile	Ala	Asn 7950	Lys	Asp	Thr
Val	Lys 7955	Ala	Ser	Glu	Asn	Tyr 7960	His	Asp	Ala	Asp	Ala 7965	Asp	rys	Gln
Thr	Ala 7970	Tyr	Thr	Asn	Ala	Val 7975	Ser	Gln	Ala	Glu	Gly 7980	Ile	Ile	Asn
Gln	Thr 7985	Thr	Asn	Pro	Thr	Leu 7990	Asn	Pro	Asp	Glu	Ile 7995	Thr	Arg	Ala
Leu	Thr 8000	Gln	Val	Thr	Asp	Ala 8005	Lys	Asn	Gly	Leu	Asn 8010	Gly	Glu	Ala
ГЛа	Leu 8015	Ala	Thr	Glu	Lys	Gln 8020	Asn	Ala	Lys	Asp	Ala 8025	Val	Ser	Gly
Met	Thr 8030	His	Leu	Asn	Asp	Ala 8035	Gln	Lys	Gln	Ala	Leu 8040	Lys	Gly	Gln
Ile	Asp 8045	Gln	Ser	Pro	Glu	Ile 8050	Ala	Thr	Val	Asn	Gln 8055	Val	Lys	Gln
Thr	Ala 8060	Thr	Ser	Leu	Asp	Gln 8065	Ala	Met	Asp	Gln	Leu 8070	Ser	Gln	Ala
Ile	Asn 8075	Asp	Lys	Ala	Gln	Thr 8080	Leu	Ala	Asp	Gly	Asn 8085	Tyr	Leu	Asn
Ala	Asp 8090	Pro	Asp	Lys	Gln	Asn 8095	Ala	Tyr	Lys	Gln	Ala 8100	Val	Ala	Lys

Ala	Glu 8105	Ala	Leu	Leu	Asn	Lys 8110	Gln	Ser	Gly	Thr	Asn 8115	Glu	Val	Gln
Ala	Gln 8120	Val	Glu	Ser	Ile	Thr 8125	Asn	Glu	Val	Asn	Ala 8130	Ala	Lys	Gln
Ala	Leu 8135	Asn	Gly	Asn	Asp	Asn 8140	Leu	Ala	Asn	Ala	Lys 8145	Gln	Gln	Ala
Lys	Gln 8150	Gln	Leu	Ala	Asn	Leu 8155	Thr	His	Leu	Asn	Asp 8160	Ala	Gln	Lys
Gln	Ser 8165	Phe	Glu	Ser	Gln	Ile 8170	Thr	Gln	Ala	Pro	Leu 8175	Val	Thr	Asp
Val	Thr 8180	Thr	Ile	Asn	Gln	Lys 8185	Ala	Gln	Thr	Leu	Asp 8190	His	Ala	Met
Glu	Leu 8195	Leu	Arg	Asn	Ser	Val 8200	Ala	Asp	Asn	Gln	Thr 8205	Thr	Leu	Ala
Ser	Glu 8210	Asp	Tyr	His	Asp	Ala 8215	Thr	Ala	Gln	Arg	Gln 8220	Asn	Asp	Tyr
Asn	Gln 8225	Ala	Val	Thr	Ala	Ala 8230	Asn	Asn	Ile	Ile	Asn 8235	Gln	Thr	Thr
Ser	Pro 8240	Thr	Met	Asn	Pro	Asp 8245	Asp	Val	Asn	Gly	Ala 8250	Thr	Thr	Gln
Val	Asn 8255	Asn	Thr	Lys	Val	Ala 8260	Leu	Asp	Gly	Asp	Glu 8265	Asn	Leu	Ala
Ala	Ala 8270	Lys	Gln	Gln	Ala	Asn 8275	Asn	Arg	Leu	Asp	Gln 8280	Leu	Asp	His
Leu	Asn 8285	Asn	Ala	Gln	Lys	Gln 8290	Gln	Leu	Gln	Ser	Gln 8295	Ile	Thr	Gln
Ser	Ser 8300	Asp	Ile	Ala	Ala	Val 8305	Asn	Gly	His	Lys	Gln 8310	Thr	Ala	Glu
Ser	Leu 8315	Asn	Thr	Ala	Met	Gly 8320	Asn	Leu	Ile	Asn	Ala 8325	Ile	Ala	Asp
His	Gln 8330	Ala	Val	Glu	Gln	Arg 8335	Gly	Asn	Phe	Ile	Asn 8340	Ala	Asp	Thr
Asp	Lys 8345	Gln	Thr	Ala	Tyr	Asn 8350		Ala	Val	Asn	Glu 8355	Ala	Ala	Ala
Met	Ile 8360	Asn	Lys	Gln	Thr	Gly 8365	Gln	Asn	Ala	Asn	Gln 8370	Thr	Glu	Val
Glu	Gln 8375	Ala	Ile	Thr	Lys	Val 8380	Gln	Thr	Thr	Leu	Gln 8385	Ala	Leu	Asn
Gly	Asp 8390	His	Asn	Leu	Gln	Val 8395	Ala	Lys	Thr	Asn	Ala 8400	Thr	Gln	Ala
Ile	Asp 8405	Ala	Leu	Thr	Ser	Leu 8410	Asn	Asp	Pro	Gln	Lys 8415	Thr	Ala	Leu
Lys	Asp 8420	Gln	Val	Thr	Ala	Ala 8425	Thr	Leu	Val	Thr	Ala 8430	Val	His	Gln
Ile	Glu 8435	Gln	Asn	Ala	Asn	Thr 8440		Asn	Gln	Ala	Met 8445	His	Gly	Leu
Arg	Gln 8450	Ser	Ile	Gln	Asp	Asn 8455		Ala	Thr	Lys	Ala 8460	Asn	Ser	Lys
Tyr	Ile 8465	Asn	Glu	Asp	Gln	Pro 8470	Glu	Gln	Gln	Asn	Tyr 8475	Asp	Gln	Ala
Val	Gln	Ala	Ala	Asn	Asn	Ile	Ile	Asn	Glu	Gln	Thr	Ala	Thr	Leu

_														
	8480					8485					8490			
Asp	Asn 8495		Ala	Ile	Asn	Gln 8500		Ala	Thr	Thr	Val 8505	Asn	Thr	Thr
Lys	Ala 8510		Leu	His	Gly	Asp 8515	Val	Lys	Leu	Gln	Asn 8520	Asp	Lys	Asp
His	Ala 8525	_	Gln	Thr	Val	Ser 8530	Gln	Leu	Ala	His	Leu 8535	Asn	Asn	Ala
Gln	Lys 8540		Met	Glu	_	Thr 8545	Leu	Ile	Asp	Ser	Glu 8550	Thr	Thr	Arg
Thr	Ala 8555		Lys	Gln	Asp	Leu 8560	Thr	Glu	Ala	Gln	Ala 8565	Leu	Asp	Gln
Leu	Met 8570	_	Ala	Leu	Gln	Gln 8575	Ser	Ile	Ala	Asp	Lys 8580	Asp	Ala	Thr
Arg	Ala 8585		Ser	Ala	Tyr	Val 8590	Asn	Ala	Glu	Pro	Asn 8595	Lys	Lys	Gln
Ser	Tyr 8600	_	Glu	Ala	Val	Gln 8605		Ala	Glu	Ser	Ile 8610	Ile	Ala	Gly
Leu	Asn 8615	Asn	Pro	Thr	Ile	Asn 8620						Ser	Ala	Thr
Gln	Ala 8630	Val	Ile	Ser	Ser	Lys	Asn	Ala	Leu	Asp		Val	Glu	Arg
Leu	Ala	Gln	Asp	Lys	Gln	Thr	Ala	Gly	Asn		Leu	Asn	His	Leu
Asp	8645 Gln	Leu	Thr	Pro	Ala			Gln		Leu		Asn	Gln	Ile
Asn	8660 Asn	Ala	Thr	Thr	Arg	_	Glu	Val	Ala	Gln	8670 Lys	Leu	Thr	Glu
Ala	8675 Gln		Leu	Asn	Gln	8680 Ala	Met	Glu	Ala	Leu	8685 Arq	Asn	Ser	Ile
	8690					8695					8700			
	Asp 8705					8710		_		-	8715			
Asp	Lys 8720					Ala 8725					Val 8730	Gln	Asn	Ala
Lys	Asp 8735		Ile	Asn		Thr 8740				Thr	Leu 8745	Asp	Lys	Ala
Gln	Val 8750					Gln 8755							Asp	Asn
Leu	His 8765		Asp	Gln	Lys	Leu 8770		Asp	Asp	Lys	Gln 8775		Ala	Val
Thr	Asp 8780		Asn	Gln	Leu	Asn 8785	Gly	Leu	Asn	Asn	Pro 8790		Arg	Gln
Ala	Leu 8795		Ser	Gln	Ile	Asn 8800	Asn	Ala	Ala	Thr	Arg 8805	Gly	Glu	Val
Ala	Gln 8810	Lys	Leu	Ala	Glu		Lys	Ala	Leu	Asp		Ala	Met	Gln
Ala	Leu	Arg	Asn	Ser	Ile	Gln	Asp	Gln	Gln	Gln	Thr	Glu	Ser	Gly
Ser	8825 Lys		Ile	Agn	G] u	8830 Asp	Lvs	Pro	G] n	Lvs	8835 Asp	Ala	Tvr	G] n
201	8840		110		CIU	8845	-y 5	110	2111	275	8850		-1-	0111
Ala	Ala 8855		Gln	Asn	Ala	Eys	_	Leu	Ile	Asn	Gln 8865	Thr	Gly	Asn

Pro	Thr 8870	Leu	Asp	Lys	Ser	Gln 8875	Val	Glu	Gln	Leu	Thr 8880	Gln	Ala	Val
Thr	Thr 8885	Ala	Lys	Asp	Asn	Leu 8890	His	Gly	Asp	Gln	Lys 8895	Leu	Ala	Arg
Asp	Gln 8900	Gln	Gln	Ala	Val	Thr 8905	Thr	Val	Asn	Ala	Leu 8910	Pro	Asn	Leu
Asn	His 8915	Ala	Gln	Gln	Gln	Ala 8920	Leu	Thr	Asp	Ala	Ile 8925	Asn	Ala	Ala
Pro	Thr 8930	Arg	Thr	Glu	Val	Ala 8935	Gln	His	Val	Gln	Thr 8940	Ala	Thr	Glu
Leu	Asp 8945	His	Ala	Met	Glu	Thr 8950	Leu	Lys	Asn	Lys	Val 8955	Asp	Gln	Val
Asn	Thr 8960	Asp	Lys	Ala	Gln	Pro 8965	Asn	Tyr	Thr	Glu	Ala 8970	Ser	Thr	Asp
Lys	Lys 8975	Glu	Ala	Val	Asp	Gln 8980	Ala	Leu	Gln	Ala	Ala 8985	Glu	Ser	Ile
Thr	Asp 8990	Pro	Thr	Asn	Gly	Ser 8995	Asn	Ala	Asn	Lys	Asp 9000	Ala	Val	Asp
Gln	Val 9005	Leu	Thr	Lys	Leu	Gln 9010	Glu	Lys	Glu	Asn	Glu 9015	Leu	Asn	Gly
Asn	Glu 9020	Arg	Val	Ala	Glu	Ala 9025	Lys	Thr	Gln	Ala	9030 Lys	Gln	Thr	Ile
Asp	Gln 9035	Leu	Thr	His	Leu	Asn 9040	Ala	Asp	Gln	Ile	Ala 9045	Thr	Ala	Lys
Gln	Asn 9050	Ile	Asp	Gln	Ala	Thr 9055	Lys	Leu	Gln	Pro	Ile 9060	Ala	Glu	Leu
Val	Asp 9065	Gln	Ala	Thr	Gln	Leu 9070	Asn	Gln	Ser	Met	Asp 9075	Gln	Leu	Gln
Gln	Ala 9080	Val	Asn	Glu	His	Ala 9085	Asn	Val	Glu	Gln	Thr 9090	Val	Asp	Tyr
Thr	Gln 9095	Ala	Asp	Ser	Asp	100 100	Gln	Asn	Ala	Tyr	105 9105	Gln	Ala	Ile
Ala	Asp 9110	Ala	Glu	Asn	Val	Leu 9115	Lys	Gln	Asn	Ala	Asn 9120	Lys	Gln	Gln
Val	Asp 9125	Gln	Ala	Leu	Gln	Asn 9130	Ile	Leu	Asn	Ala	135 9135	Gln	Ala	Leu
Asn	Gly 9140	Asp	Glu	Arg	Val	Ala 9145	Leu	Ala	Lys	Thr	Asn 9150	Gly	Lys	His
Asp	Ile 9155	Asp	Gln	Leu	Asn	Ala 9160	Leu	Asn	Asn	Ala	Gln 9165	Gln	Asp	Gly
Phe	Lys 9170	Gly	Arg	Ile	Asp	Gln 9175	Ser	Asn	Asp	Leu	Asn 9180	Gln	Ile	Gln
Gln	Ile 9185	Val	Asp	Glu	Ala	Lys 9190	Ala	Leu	Asn	Arg	Ala 9195	Met	Asp	Gln
Leu	Ser 9200	Gln	Glu	Ile	Thr	Asp 9205	Asn	Glu	Gly	Arg	Thr 9210	Lys	Gly	Ser
Thr	Asn 9215	Tyr	Val	Asn	Ala	Asp 9220	Thr	Gln	Val	Lys	Gln 9225	Val	Tyr	Asp
Glu	Thr 9230	Val	Asp	Lys	Ala	Lys 9235	Gln	Ala	Leu	Asp	Lys 9240	Ser	Thr	Gly

Gln	Asn 9245	Leu	Thr	Ala	Lys	Gln 9250	Val	Ile	Lys	Leu	Asn 9255	Asp	Ala	Val
Thr	Ala 9260	Ala	Lys	Lys	Ala	Leu 9265	Asn	Gly	Glu	Glu	Arg 9270	Leu	Asn	Asn
Arg	Lys 9275	Ala	Glu	Ala	Leu	Gln 9280	Arg	Leu	Asp	Gln	Leu 9285	Thr	His	Leu
Asn	Asn 9290	Ala	Gln	Arg	Gln	Leu 9295	Ala	Ile	Gln	Gln	Ile 9300	Asn	Asn	Ala
Glu	Thr 9305	Leu	Asn	Lys	Ala	Ser 9310	Arg	Ala	Ile	Asn	Arg 9315	Ala	Thr	Lys
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Pro	Glu	Phe	Asp	Asp 245	Leu	ГЛа	Arg	Thr	Asp 250	Ala	Ser	Gln	Val	Asp 255	Gly
Gln	Asp	Asp	Thr 260	Ser	Glu	Asp	Asp	Asp 265	Asn	Glu	Ser	Glu	Lys 270	Ala	Asp
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Gln	Glu 370	Ser	Lys	Gln	Asp	Val 375	Gln	Tyr	Glu	Ile	380 Lys	Asp	Leu	Ile	Gln
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Asp	Gln	Tyr	Lys 420	Leu	Phe	Tyr	Lys	Lys 425	Gln	Asp	Leu	Ser	Lys 430	Ser	Phe
Asp	Ala	Thr 435	Phe	Thr	Leu	Leu	Ile 440	Asp	Ala	Ser	Ala	Ser 445	Met	His	Asp
Lys	Met 450	Ala	Glu	Thr	Lys	Lys 455	Gly	Val	Val	Leu	Phe 460	His	Glu	Thr	Leu
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Phe	Asp	Ser	Asp	Glu 485	His	Ala	Gln	Pro	Asn 490	Ile	Ile	Asn	Glu	Ile 495	Ile
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Ser	Glu 530	Arg	Leu	Met	Arg	Arg 535	Asn	Gln	His	Gln	Arg 540	Phe	Leu	Ile	Val
Phe 545	Ser	Asp	Gly	Glu	Pro 550	Ser	Ala	Phe	Asn	Tyr 555	Ser	Gln	Asp	Gly	Ile 560
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Asp	Ala	Ala	Thr	Thr 85	Asn	Ala	Gln	Val	Glu 90	Ala	Ile	Lys	Thr	Lys 95	Ala
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Ala	Leu	Glu 115	Glu	Phe	Asp	Glu	Val 120	Val	Gln	Ala	Gln	Ile 125	Asp	Gln	Ala
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Lys	Pro	Leu 35	Asp	Lys	Ser	Ser	Ser 40	Ser	Leu	His	His	Gly 45	Tyr	Ser	Lys
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Leu 65	Ser	Ser	Leu	Thr	Phe 70	Asn	Lys	Asn	Gln	Asn 75	Ile	Ser	Tyr	Lys	Asp 80
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Gln	Val	Pro	Tyr	Thr 165	Ile	Thr	Val	Asn	Gly 170	Thr	Ser	Gln	Asn	Ile 175	Leu
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Val	Asp 210	Leu	Arg	Leu	Ser	Lys 215	Gln	Ala	Lys	Tyr	Thr 220	Val	Asn	Phe	Lys
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Tyr	Ser	Ile 275	Asn	Leu	Asn	Gly	Thr 280	Ser	Thr	Asn	Ile	Leu 285	Ser	Asn	Leu
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Tyr	Thr	Val 435	His	Phe	Lys	Asn	Gly 440	Thr	Lys	Lys	Val	Ile 445	Asn	Ile	Asn
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Ser	Arg 50	Thr	Val	Glu	Glu	Tyr 55	Lys	Lys	Ser	Leu	Asp 60	Asp	Leu	Ile	Trp
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Arg	Pro	Asn	Asn	Ile 245	Pro	Val	Leu	Glu	Asp 250	Glu	Lys	Gln	Glu	Glu 255	Lys
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Leu	Leu 210	Ser	Glu	Ala	Lys	Lys 215	Leu	Asn	Glu	Ser	Gln 220	Ala	Pro	Lys	Ala
Asp 225	Asn	Lys	Phe	Asn	Lув 230	Glu	Gln	Gln	Asn	Ala 235	Phe	Tyr	Glu	Ile	Leu 240

His Leu Pro Asn Leu Asn Glu Glu Gln Arg Asn Gly Phe Ile Gln Ser 250 Leu Lys Asp Asp Pro Ser Gln Ser Ala Asn Leu Leu Ala Glu Ala Lys Lys Leu Asn Asp Ala Gln Ala Pro Lys Ala Asp Asn Lys Phe Asn Lys 280 Glu Gln Gln Asn Ala Phe Tyr Glu Ile Leu His Leu Pro Asn Leu Thr Glu Glu Gln Arg Asn Gly Phe Ile Gln Ser Leu Lys Asp Asp Pro Ser Val Ser Lys Glu Ile Leu Ala Glu Ala Lys Lys Leu Asn Asp Ala Gln Ala Pro Lys Glu Glu Asp Asn Asn Lys Pro Gly Lys Glu Asp Gly Asn 345 Lys Pro Gly Lys Glu Asp Asn Asn Lys Pro Gly Lys Glu Asp Asn Lys 360 Lys Pro Gly Lys Glu Asp Asn Asn Lys Pro Gly Lys Glu Asp Asn Asn 375 Lys Pro Gly Lys Glu Asp Gly Asn Lys Pro Gly Lys Glu Asp Asn Lys 395 Lys Pro Gly Lys Glu Asp Asn Asn Lys Pro Gly Lys Glu Asp Gly Asn 405 410 Lys Pro Gly Lys Glu Asp Gly Asn Gly Val His Val Val Lys Pro Gly 425 Asp Thr Val Asn Asp Ile Ala Lys Ala Asn Gly Thr Thr Ala Asp Lys Ile Ala Ala Asp Asn Lys Leu Ala Asp Lys Asn Met Ile Lys Pro Gly 455 Gln Glu Leu Val Val Asp Lys Lys Gln Pro Ala Asn His Ala Asp Ala Asn Lys Ala Gln Ala Leu Pro Glu Thr Gly Glu Glu Asn Pro Phe Ile 490 Gly Thr Thr Val Phe Gly Gly Leu Ser Leu Ala Leu Gly Ala Ala Leu Leu Ala Gly Arg Arg Arg Glu Leu 515 <210> SEQ ID NO 34 <211> LENGTH: 291 <212> TYPE: PRT <213 > ORGANISM: Staphylococcus sp. <400> SEQUENCE: 34 Ala Gln His Asp Glu Ala Lys Lys Asn Ala Phe Tyr Gln Val Leu Asn 10 Met Pro Asn Leu Asn Ala Asp Gln Arg Asn Gly Phe Ile Gln Ser Leu 25 Lys Ala Ala Pro Ser Gln Ser Ala Asn Val Leu Gly Glu Ala Gln Lys Leu Asn Asp Ser Gln Ala Pro Lys Ala Asp Ala Gln Gln Asn Asn Phe Asn Lys Asp Lys Lys Ser Ala Phe Tyr Glu Ile Leu Asn Met Pro Asn

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100	105	110	an.
115	120	Asn Lys Glu Lys Lys A 125	sii
Ala Phe Tyr Glu Ile 130	Leu Asn Met Pro Asn 135	Leu Asn Glu Glu Gln A 140	rg
Asn Gly Phe Ile Gln 145	Ser Leu Lys Ala Ala 150	Pro Ser Gln Ser Ala A 155 1	sn 60
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His Leu Pro Asn Leu 195	Asn Glu Glu Gln Arg 200	Asn Gly Phe Ile Gln S	er
	Ser Gln Ser Ala Asn 215	Leu Leu Ala Glu Ala L 220	λa
		Asp Asn Lys Phe Asn L	уs 40
Glu Lys Lys Asn Ala	Phe Tyr Glu Ile Leu	His Leu Pro Asn Leu T	
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# 117

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Lys	Ser	Glu 675	Leu	Gly											

## 1-77. (canceled)

- **78**. A recombinant nucleic acid molecule comprising a nucleic acid sequence encoding a *Staphylococcus aureus* Protein A (SpA) D domain segment having (a) at least one amino acid substitution that disrupts Fc binding and (b) at least one amino acid substitution that disrupts VH3 binding and (c) an amino acid sequence that is at least 70% identical to the amino acid sequence of SEQ ID NO: 2, wherein the amino acid substitutions correspond to amino acids at position 9, 10, 36 and/or 37 of SEQ ID NO: 2.
- **79**. The nucleic acid molecule of claim **78**, wherein the SpA D domain segment is at least 98% identical to the amino acid sequence of SEQ ID NO: 2.
- **80**. The nucleic acid molecule of claim **78**, further comprising a nucleic acid sequence encoding one or more SpA E domain, A domain, B domain or C domain.
- **81**. The nucleic acid molecule of claim **78**, further comprising a nucleic acid sequence encoding a non-Protein A segment.
- **82**. The nucleic acid molecule of claim **78**, further comprising a nucleic acid sequence encoding a second antigen segment, wherein the second antigen segment is a staphylococcal antigen segment selected from Emp, EsxA, EsxB, EsaC, Eap, Ebh, EsaB, Coa, vWbp, vWh, Hla, SdrC, SdrD, SdrE, IsdA, IsdB, IsdC, CIfA, CIfB, and/or SasF segment.
- **83**. The nucleic acid molecule of claim **78**, wherein the substitutions at amino acids corresponding to positions 9 and 10 are with a glycine residue and the substitutions at amino acids corresponding to positions 36 and 37 are with a serine residue.
- **84**. The nucleic acid molecule of claim **78**, wherein the substitutions at amino acids corresponding to positions 9 and 10 are with a lysine residue and the substitutions at amino acids corresponding to positions 36 and 37 are with an alanine residue.
- **85**. A host cell comprising the nucleic acid molecule of claim **78**.
- **86**. A method of making an isolated polypeptide comprising expressing the polypeptide in a Fc host cell having the recombinant nucleic acid molecule of claim **78**.
- **87**. The method of claim **86**, wherein the host cell is a bacterial cell or eukaryotic cell.

- 88. The method of claim 86, wherein the method comprises culturing the host cell.
- **89**. The method of claim **88**, wherein the host cell is induced to express the polypeptide.
- **90**. The method of claim **86**, wherein the wherein the substitutions at amino acids corresponding to positions 9 and 10 are with a glycine residue and the substitutions at amino acids corresponding to positions 36 and 37 are with a serine residue.
- **91**. The method of claim **86**, wherein the substitutions at amino acids corresponding to positions 9 and 10 are with a lysine residue and the substitutions at amino acids corresponding to positions 36 and 37 are with a alanine residue.
- **92.** A method of treating a staphylococcal infection in a subject comprising administering a composition comprising an isolated polypeptide comprising a *Staphylococcus aureus* Protein A (SpA) D domain segment having (a) at least one amino acid substitution that disrupts Fc binding and (b) at least one amino acid substitution that disrupts VH3 binding and (c) an amino acid sequence that is at least 70% identical to the amino acid sequence of SEQ ID NO: 2.
- **93**. The method of claim **92**, wherein the SpA D domain is at least 98% identical to SEQ ID NO: 2.
- **94**. The method of claim **92**, wherein the composition comprises one or more SpA E domain, A domain, B domain or C domain.
- **95**. The method of claim **92**, wherein the composition comprises a second antigen segment selected from staphylococcal Emp, EsxA, EsxB, EsaC, Eap, Ebh, EsaB, Coa, vWbp, vWh, Hla, SdrC, SdrD, SdrE, IsdA, IsdB, IsdC, CIfA, CIfB, and SasF segment.
- **96**. The method of claim **92**, wherein the wherein the substitutions at amino acids corresponding to positions 9 and 10 are with a glycine residue and the substitutions at amino acids corresponding to positions 36 and 37 are with a serine residue.
- **97**. The method of claim **92**, wherein the wherein the substitutions at amino acids corresponding to positions 9 and 10 are with a lysine residue and the substitutions at amino acids corresponding to positions 36 and 37 are with a alanine residue.

\* \* \* \* \*