

CME Review

Skin as the target for allergy prevention and treatment



Andreina Marques-Mejias, MD, PhD^{*,†}; Irene Bartha, MD^{*,†}; Christina E. Ciaccio, MD^{‡,§};
 R. Sharon Chinthrajah, MD^{||}; Susan Chan, MBBS, MD^{*,†,¶}; Gurjit K. Khurana Hershey, MD, PhD^{#,**};
 Jessica W. Hui-Beckman, MD^{††}; Laurie Kost, MS in Epidemiology, MS in Biological Sciences^{||};
 Gideon Lack, MD, MBChB^{*,†,¶}; Janice A. Layhadi, PhD^{‡‡}; Donald Y.M. Leung, MD, PhD^{††};
 Hannah F. Marshall, MBChB[†]; Kari C. Nadeau, MD, PhD^{§§}; Suzana Radulovic, MD^{*,†,¶};
 Reena Rajcoomar, RN, BScN, HBSc^{||}; Mohamed H. Shamji, PhD^{§§}; Sayantani Sindher, MD^{||};
 Helen A. Brough, MA (Hons), MSc, PhD, MBBS^{*,†,¶}

^{*} Department of Women and Children's Health (Paediatric Allergy), School of Life Course Sciences, Faculty of Life Sciences and Medicine, King's College London, London, United Kingdom

[†] Children's Allergy Service, Evelina London, Guy's and St Thomas', NHS Foundation Trust, London, United Kingdom

[‡] Department of Pediatrics, The University of Chicago, Chicago, Illinois

[§] Department of Medicine, The University of Chicago, Chicago, Illinois

^{||} Department of Medicine, and Sean N Parker Center for Allergy and Asthma Research, Stanford University, Palo Alto, California

[¶] Peter Gorer Department of Immunobiology, School of Immunology & Microbial Sciences, King's College London, London, United Kingdom

[#] Division of Asthma Research, Cincinnati Children's Medical Center, Cincinnati, Ohio

^{**} Department of Pediatrics, University of Cincinnati College of Medicine, Cincinnati, Ohio

^{††} Department of Pediatrics, National Jewish Health, Denver, Colorado

^{§§} National Heart and Lung Institute, Imperial College London, London, United Kingdom

^{‡‡} Department of Environmental Health, Harvard T.H. Chan School of Public Health, Harvard University, Boston, Massachusetts

Key Messages

- Disruption of the skin barrier is associated with development of atopic sensitization.
- Atopic dermatitis is associated with immune dysregulation, which can both cause epithelial disruption and be a result of epithelial disruption.
- Both the gut and skin microbiome have been shown to have effects on skin epithelial barrier function.
- Barrier-based approaches are valuable for disease prevention; however, Th2 cytokine targeted immune-based treatments and immune abnormalities should be addressed in order to improve and resolve active atopic dermatitis and prevent food allergy development.
- Newer treatments for atopic dermatitis that are being explored include pro and prebiotics, filaggrin replacement therapies, and skin-targeted gene therapy.

ARTICLE INFO

Article history:

Received for publication August 21, 2023.

Received in revised form November 27, 2023.

Accepted for publication December 26, 2023.

ABSTRACT

The fact that genetic and environmental factors could trigger disruption of the epithelial barrier and subsequently initiate a T_H2 inflammatory cascade conversely proposes that protecting the same barrier and promoting adequate interactions with other organs, such as the gut, may be crucial for lowering the risk and preventing atopic diseases, particularly, food allergies. In this review, we provide an overview of structural characteristics that support the epithelial barrier hypothesis in patients with atopic dermatitis, including the most relevant filaggrin gene mutations, the recent discovery of the role of the transient receptor potential vanilloid 1, and the role involvement of the microbiome in healthy and damaged skin. We present experimental and human studies that support the mechanisms of allergen penetration, particularly the dual allergen exposure and the outside-in, inside-out, and outside-inside-outside hypotheses. We discuss classic skin-targeted therapies for food allergy prevention, including moisturizers, steroids, and topical calcineurin inhibitors, along with pioneering trials proposed to change their current use (Prevention of Allergy via Cutaneous Intervention and Stopping Eczema and Allergy). We provide an overview of the novel therapies that enhance the skin barrier, such as probiotics and prebiotics topical application, read-through drugs, direct and indirect FLG replacement, and interleukin and janus kinases inhibitors. Last, we

Address correspondence to: Helen A. Brough, MA (Hons), MSc, PhD, MBBS, Children's Allergy Service, Evelina Children's Hospital, Guy's and St. Thomas' Hospital NHS Foundation Trust, Second Floor, Stairwell B, South Wing, London. SE1 7EH, UK. E-mail: helen.1.brough@kcl.ac.uk.

<https://doi.org/10.1016/j.anaai.2023.12.030>

1081-1206/© 2024 American College of Allergy, Asthma & Immunology. Published by Elsevier Inc. This is an open access article under the CC BY license

(<http://creativecommons.org/licenses/by/4.0/>)

discuss the newer strategies for preventing and treating food allergies in the form of epicutaneous immunotherapy and the experimental use of single-dose of adeno-associated virus vector gene immunotherapy.

© 2024 American College of Allergy, Asthma & Immunology. Published by Elsevier Inc. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>)

Instructions

Credit can now be obtained, free for a limited time, by reading the review article and completing all activity components. Please note the instructions listed below:

- Review the target audience, learning objectives and all disclosures.
- Complete the pre-test.
- Read the article and reflect on all content as to how it may be applicable to your practice.
- Complete the post-test/evaluation and claim credit earned. At this time, physicians will have earned up to 1.0 *AMA PRA Category 1 Credit*[™]. The minimum passing score on the post-test is 70%.

Overall Purpose

Participants will be able to demonstrate increased knowledge of the clinical treatment of allergy/asthma/immunology and be able to apply new information to their own practices.

Learning Objectives

At the conclusion of this activity, participants should be able to:

- Specify the importance of the epithelial barrier in the development of food allergies.
- Categorize classic and novel skin-targeted therapies for food allergy prevention.

Release Date: August 1, 2024

Expiration Date: July 31, 2026

Target Audience

Physicians involved in providing patient care in the field of allergy/asthma/immunology.

Accreditation

The American College of Allergy, Asthma & Immunology (ACAAI) is accredited by the Accreditation Council for Continuing Medical Education (ACCME) to provide continuing medical education for physicians.

Designation

The American College of Allergy, Asthma & Immunology (ACAAI) designates this journal-based CME activity for a maximum of 1.0 *AMA PRA Category 1 Credit*[™]. Physicians should claim only the credit commensurate with the extent of their participation in the activity.

Disclosure Statement

As required by the Accreditation Council for Continuing Medical Education (ACCME) and in accordance with the American College of Allergy, Asthma and Immunology (ACAAI) policy, all individuals in a position to control or influence the content of an activity must disclose **all** financial relationships with any ineligible company that have occurred within the past **24 months**. An Ineligible Company as an entity whose primary business is producing, marketing, selling, re-selling, or distributing health care products used by or on patients. Examples 1 of such organizations include:

- Advertising, marketing, or communication firms whose clients are ineligible companies.
- Bio-medical startups that have begun a governmental regulatory approval process.
- Compounding pharmacies that manufacture proprietary compounds.
- Device manufacturers or distributors; diagnostic labs that sell proprietary products.
- Growers, distributors, manufacturers or sellers of medical foods and dietary supplements.
- Manufacturers of health-related wearable products.
- Pharmaceutical companies or distributors.
- Pharmacy benefit managers.
- Reagent manufacturers or sellers.

The ACCME does not consider providers of clinical service directly to patients to be commercial interests. For more information, visit www.accme.org. All identified relevant relationships must be mitigated and the educational content thoroughly vetted for fair balance, scientific objectivity, and appropriateness of patient care recommendations. It is required that disclosure of or absence of relevant financial relationships be provided to the learners prior to the start of the activity.

Learners must also be informed when off-label, experimental/investigational uses of drugs or devices are discussed in an educational activity or included in related materials.

Disclosure in no way implies that the information presented is biased or of lesser quality. It is incumbent upon course participants to be aware of these factors in interpreting the program contents and evaluating recommendations. Moreover, expressed views do not necessarily reflect the opinions of the ACAAI. **All identified relevant financial relationships have been mitigated.**

Planners:

- Anna Nowak-Wegrzyn, MD, Researcher: DBV Technologies; Speaker: Genentech, Novartis.
- Kurt Shulenberg, MA, has no relevant financial relationships with ineligible companies to disclose.

Authors:

- Andreina Marques-Mejias, MD, PhD, has no relevant financial relationships with ineligible companies to disclose.
- Helen Annaruth Brough, PhD, MSc, MA (Hon) MD, BS, FRCPH, Speaker: DBV Technologies.

Recognition of Commercial Support: This activity has not received external commercial support.

Copyright Statement: © 2015–2024 ACAAI. All rights reserved.

CME Inquiries: Contact the American College of Allergy, Asthma & Immunology at education@acaai.org or 847-427-1200.

Introduction

The IgE-mediated allergic diseases (also called atopic diseases) such as asthma, atopic dermatitis (AD), allergic rhinitis (AR), and food allergy (FA) have been increasing in the last few decades.¹ Although their prevalence varies between studies because of differences in methodology and demographics, the prevalence of different atopic conditions in the United States is estimated to be approximately 10% to 30% for AR, 4% for asthma, 10% to 20% for AD, and 11% for FA.^{2–4} These diseases have been found to be highly heritable. In this sense, it is estimated that between 35% and 95% of patients are diagnosed with asthma, 33% to 91% of those with AR, 71% to 84% of those with AD, and 81% of those with FA have a parent who shares the same diagnosis.^{5,6} Although the high heritability of these diseases indicates the critical role of genetics, genetics alone cannot fully explain the rapid growth in their prevalence. Environmental factors such as air pollution and climate change are thought to play an important role in the increasing prevalence.^{5,7} One systematic review and meta-analysis of 55 papers found a significant association between air pollutants and increased risk of allergic diseases.⁷

Atopic diseases seem to have natural disease progression with AD being the first to manifest, followed by FA, AR, or allergic asthma, suggesting a common underlying mechanism and perhaps different manifestations of the same disease. This natural progression has been termed the atopic march.⁸ It is currently hypothesized that disruption of the epithelial barrier is the first step in initiating an inflammatory allergic pathway.⁹ This has been termed the epithelial barrier hypothesis (Fig 1). Both genetic and environmental factors have been found to mediate epithelial barrier dysfunction.

There is growing evidence that environmental peanut exposure through the skin can result in peanut sensitization and allergy.¹⁰ A study in highly atopic children with an impaired epidermal barrier found an exposure-response relationship between peanut protein levels in household dust and peanut sensitization and likely allergy.¹¹ Other foods, such as cow's milk, egg, walnut, and fish, have also been found in dust.¹¹ In this review, we expand topics to understand the importance of skin barrier function in the development of FA and how to target the skin barrier for FA prevention and treatment.

Skin Barrier Function and Dysfunction in Atopy

In addition to providing protection from external threats, the skin helps the internal milieu by protecting the body from water loss and maintaining homeostasis. The human epidermis is

composed of 5 layers, which are formed through keratinocyte differentiation to create the stratum basale, stratum spinosum, stratum granulosum, stratum lucidum, and stratum corneum (SC).¹² The SC is the outermost layer and contributes the most to skin barrier function, as important proteins and lipids in the SC provide structural integrity. This cornified layer is composed of flaggrin and other epidermal differentiation complex (EDC) genes, including loricrin and involucrin. Tight junction proteins in the stratum granulosum provide additional barrier protection and contribute to water retention. Epidermal ceramides, including acylceramides, are also important for skin barrier function. Together, the components of the SC are necessary to maintain skin integrity, pH, and moisturization.^{12,13}

Transepidermal water loss (TEWL) quantifies the amount of water that moves from the dermis and epidermis through the SC to the skin surface. In healthy skin, the SC functions as an efficient barrier minimizing water loss. However, in diseases such as AD, this barrier is damaged, resulting in significantly larger water loss, which serves as a measure of epithelial barrier integrity (Fig 2).¹⁴ Consequently, TEWL is frequently used to evaluate the severity of AD and the response to treatment.¹⁰ It is also notable that TEWL, particularly after skin tape stripping, can differentiate between diverse AD endotypes with and without peanut allergy (PA).¹⁵

Recent studies have revealed that distinct skin endotypes are found in individuals with AD but without FA, AD, and FA and FA without AD.¹⁶ The skin barrier profile in these groups is distinct in the protein and ceramide expression and *Staphylococcus aureus* abundance. This reveals the important role of the skin barrier in atopy and that these endotypes have distinct findings in the SC epidermal layer. Moreover, FLG loss-of-function mutations have been well studied and are the strongest genetic risk factor for AD. Other defects, such as tight junction polymorphisms in claudin-1 and Notch deficiency, have also been described to interfere with keratinocyte differentiation.¹³ Disruption of keratinocyte differentiation allows for allergens and irritants to enter through the damaged skin, leading to an increase in proinflammatory cytokine expression.¹³

Environmental Irritants

Skin irritants including detergents and pollutants lead to reduced FLG expression and skin barrier damage.^{12,13} Surfactants affect tight junctions potentially increasing risk of allergic reactions.¹⁷ Some

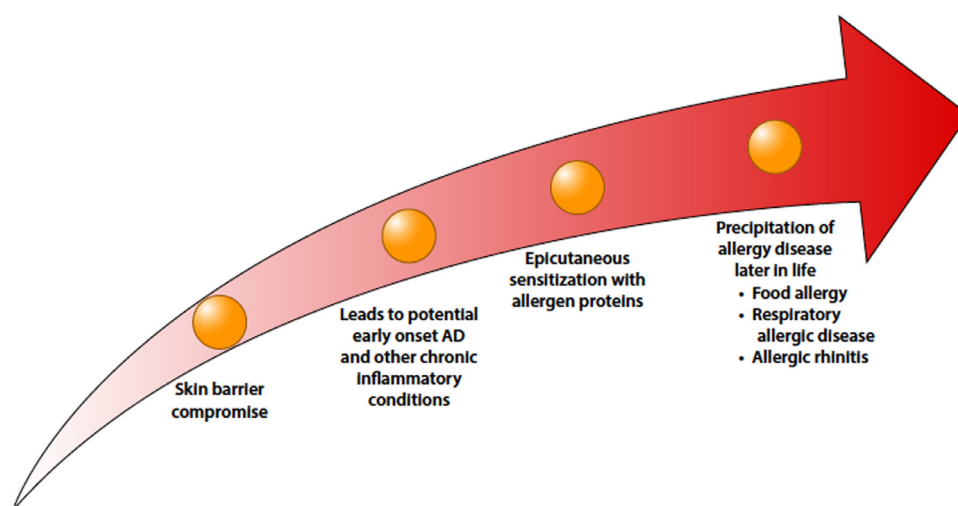


Figure 1. The sequence of events after skin barrier compromise early in life. AD, atopic dermatitis.

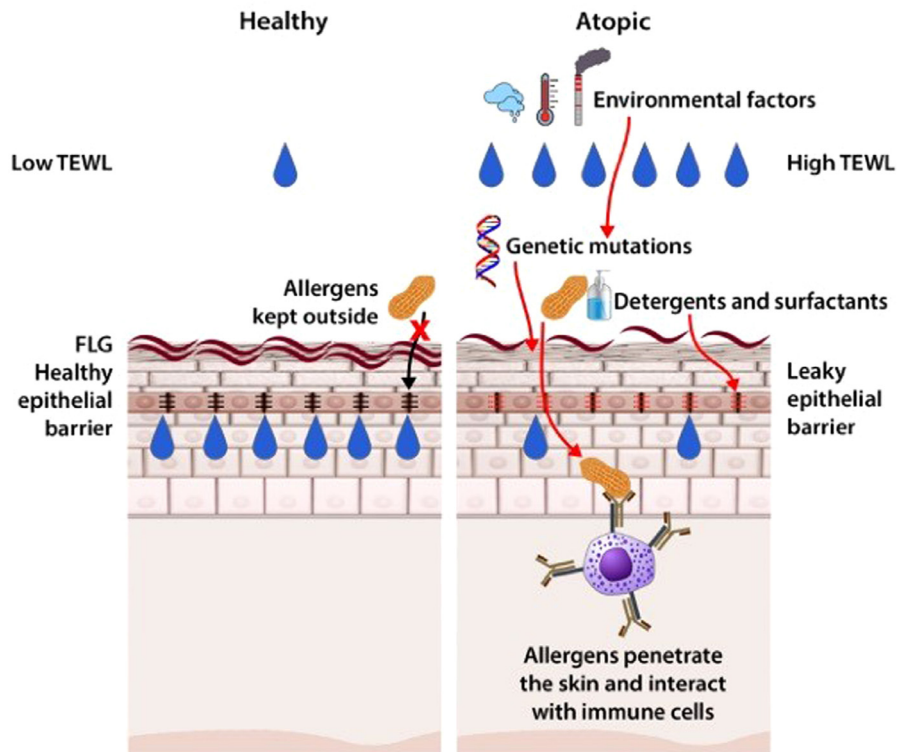


Figure 2. TEWL findings and factors contributing to epithelial barrier dysfunction in AD against healthy skin. AD, atopic dermatitis; TEWL, transepidermal water loss.

compounds found frequently in cleaning products, such as sodium dodecyl sulfate and sodium dodecylbenzene sulfonate, damage the epithelium even at dilutions of 1:100,000.¹⁷ Concurrently, pollution inhibits EDC genes.¹⁸ In addition, environmental factors are critical in early childhood, as babies begin their exposure to the world. Season, month, and time of birth can indirectly measure changes experienced during pregnancy and early infancy, such as temperature, humidity, sun exposure, and outdoor physical activity¹⁹; for example, in children born during the fall and winter, there is a higher incidence of AD.²⁰ Furthermore, low humidity triggers FLG proteolysis, increasing the skin barrier disfunction.²¹

More recently, the role of transient receptor potential vanilloid 1 has been studied as a temperature-sensing ion channel expressed in epidermal keratinocytes. Low temperature causes skin barrier dysfunction by inhibiting EDC genes and increasing proinflammatory cytokines through transient receptor potential vanilloid 1, which may be a contributing factor to worsening AD during winter.²² Thus, there are many components of the skin barrier that contribute to its important defense mechanisms, and genetic and environmental factors can lead to impaired function.

Mechanisms of Allergen Penetration

The Dual Allergen Exposure Hypothesis

The dual allergen exposure hypothesis proposes that the route of exposure to food allergens determines whether an individual develops tolerance or sensitization to the allergen. Early oral exposure to food allergens is thought to induce tolerance, whereas exposure to food and potentially respiratory allergens through the skin can lead to sensitization and the development of food and respiratory allergies (Fig 3).^{23–26}

The application of peanut oil to inflamed skin during infancy is a risk factor for the development of PA.²⁷ There is also a dose-response relationship in infants with atopy between environmental peanut exposure (measured by household peanut consumption) and the

development of PA, but this relationship no longer existed in infants who ate peanut in their first year.²⁸

The Outside-In, Inside-Out, and Outside-Inside-Outside Hypothesis

The AD is characterized by both epidermal barrier dysfunction and immune dysregulation, with skewing toward a T_H2 profile. However, whether the epidermal barrier abnormality leads to the immunologic abnormalities (the outside-in hypothesis) or the immunologic mechanisms exacerbate barrier integrity (the inside-out hypothesis) is unclear. Indeed, both mechanisms may have a role to play. Both an impaired skin barrier and immune dysregulation (T_H2 deviation) begin in early infancy. Studies have revealed that there is overexpression of interleukin (IL)-4 and IL-13 in AD skin. In the presence of these cytokines, keratinocytes exhibit reduced FLG gene expression. The interaction between these 2 theories may explain the mechanism for the outside-inside-outside hypothesis.²⁹

Role of Skin Barrier Dysfunction in Allergy Development

The integrity of the SC is maintained by a careful balance of lipids including ceramide, free fatty acids, and cholesterol.^{30,31} Atopic skin is associated with alterations in several SC lipids including altered lipid organization, a decrease in total lipids, and altered ratios of ceramides, free fatty acids, and cholesterol.^{32–34} In atopic skin, lipids are less densely organized, which could be due to the shift toward short- as opposed to long-chain ceramides.³⁴ Similar findings have also been found with free fatty acids, with a significant decrease in long-chain fatty acid (≥ 24 carbon atoms) and an increase in the short-chain fatty acid, such as C16 (palmitic acid) and C18 (octadecanoic acid) in atopic skin.³⁵

In addition to the lipid profile of the skin, the role of filaggrin and its breakdown product such as natural moisturizing factor are critical for maintaining skin barrier function. Null mutations

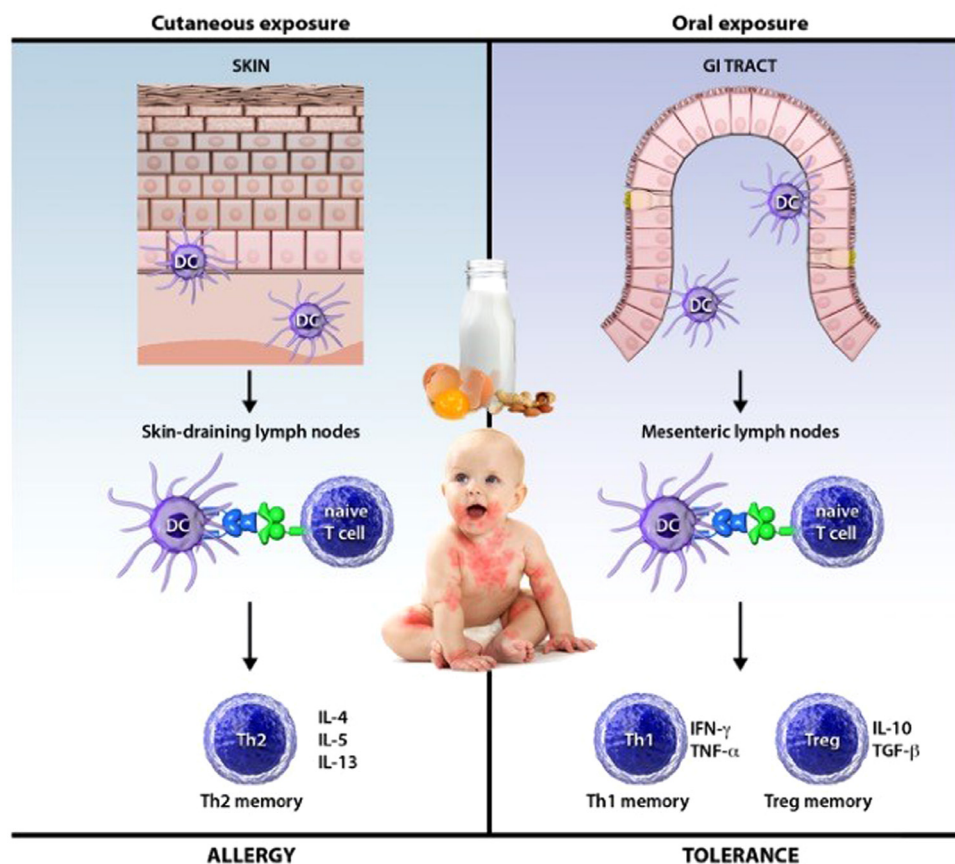


Figure 3. Dual-allergen exposure: increasing evidence suggests that early life allergen exposure through the skin causes T cell deviation toward a T_H2 allergenic type and subsequent food allergy, whereas early oral exposure causes T cell deviation toward tolerogenic T_H1 and Treg subtypes (dual allergen exposure hypothesis). Adapted from: Brough HA, Nadeau KC, Sindher SB, Alkotob SS, Chan S, Bahnson HT, et al. Epicutaneous sensitization in the development of food allergy: What is the evidence and how can this be prevented? *Allergy*. 2020;75(9):2185–205. GI, gastrointestinal.

in the FLG gene were first identified as a risk factor for AD in an Irish cohort, in which they found the FLG polymorphisms FLG R501X and 2282del4.³² FLG null mutations still hold the strongest genetic predisposition for skin barrier defects. FLG is translated as the precursor proprotein profilaggrin, which can be broken down to form filaggrin monomers. Filaggrin monomers induce the aggregation of skin keratin filaments resulting in the formation of large macrofibrils,³⁶ which play key roles in enhancing the strength and flexibility of the SC.³⁷ Furthermore, these filaggrin monomers can undergo further proteolysis leading to the generation of trans-urocanic acid (UCA) and pyrrolidone carboxylic acid, which are components of natural moisturizing factor.³⁸ Therefore, filaggrin provides both mechanical strength and hydration to the SC, and disruption of this pathway results in impaired hydration, increased acidification, and abnormal lipid content, distribution, and organization in the skin.

The FLG genetic variants were found to have a highly significant association with asthma occurring in the context of AD.³² The disrupted epithelial barrier facilitates the entry of environmental substances through the skin promoting a T_H2 inflammatory cascade.³⁹ Although there is an increased risk of FA in children with AD, a significant number of children who develop FAs do not have AD, but may only have impaired skin barrier function.⁴⁰ Furthermore, FLG loss-of-function mutations were found to have a significant association with PA even after controlling for co-existing AD.⁴¹ This was not replicated in children without skin barrier dysfunction, suggesting that the skin barrier plays a key role. Household peanut exposure has been associated with peanut sensitization and PA, if the child had FLG null mutation⁴² and AD.^{11,42}

Role of the Skin Microbiome in Allergy Prevention

In healthy skin, there is a stringent selection process governing the microbiome. This is secondary to the interaction of environmental exposure, host factors, and metabolism products. The characteristic dysbiosis present in AD implies an immune system imbalance and skin barrier malfunction.¹⁵ In this sense, environmental factors and genetics could disturb these interactions and trigger harmful bacteria overgrowth.¹⁵

S aureus, in particular, is widespread in subjects with AD, in which it tends to be more virulent through varied mechanisms; a compromised barrier is more prone to colonization because of the redistribution of fibronectin in the SC.^{15,43} Furthermore, patients with FLG mutations have higher skin pH due to lower levels of FLG-degradation products and have an enhanced tendency to generate biofilms, all contributing to successful colonization and avoidance of the immune system.⁴³ Overall, *S aureus* proliferation has been linked with AD severity, progression, and persistence.^{43,44}

Recent research has shed light on the intricate relationship between *S aureus* overgrowth in AD and the development of FAs.^{10,45–47} Its presence in the skin at any point hinders the natural occurrence of tolerance to hen's egg and interrupts the process of developing tolerance to peanut.⁴⁴ Moreover, it has been associated with higher levels of IgE in cow's milk, hen's egg, and peanut independent of the AD severity.⁴⁴ The virulence of *S aureus* through the production of Staphylococcus enterotoxin B is also recognized as an amplifier of allergic responses to food allergens.^{48,49}

Malassezia spp is an abundant part of the healthy skin microbiome. Despite that, the overgrowth of specific species such as *Malassezia furfur*, *Malassezia sympodialis*, and *Malassezia globosa* has been

related to AD and its severity. The distribution of skin lesions in the head and neck suggests an association with these microorganisms.⁵⁰ Furthermore, the reactivity to pathogenic *Malassezia* in patients with AD can be identified through specific IgE levels, skin prick, and patch tests. Selander et al⁴⁸ recognized that *Malassezia sympodialis* induces cysteinyl leukotrienes in IgE-sensitized bone marrow-derived cells, secondarily activating mast cells and preserving inflammation in patients with AD.

The previous evidence emphasizes that skin dysbiosis plays a fundamental role in AD severity and progression, potentiating allergic responses and magnifying the risk of food sensitization and FA persistence (Fig 4). Current evidence also supports a role for the gut microbiome in AD; however, because of the co-evolution with humans, commensal gut bacteria play a key role in preventing several chronic inflammatory diseases, including AD, through interactions with the epithelial barrier and mucosal immune system that may have systemic outcomes.

Although viral infections in the skin of patients with AD may become disseminated and thereafter worsen AD control, there are no data on whether this further increases predisposition toward atopy.

Gut Microbiome and Skin Barrier Dysfunction

Numerous studies have identified microbial differences in patients with AD when compared with healthy individuals, and infants with decreased gut microbial diversity are particularly susceptible to the development of AD.^{51–56} Cohort studies have identified several key bacteria differentially expressed in the gut in persons with AD, including *Bifidobacterium* (in infants) and *Clostridium sp* (in adults), that metabolize indigestible complex polysaccharides into

many essential nutrients, including the short-chain fatty acids, butyrate and propionate, and the neurotransmitters, Gamma-aminobutyric acid (GABA) and acetylcholine. Butyrate enhances epithelial barrier function thereby decreasing the permeability of the intestinal barrier.^{57,58} In the relative absence of butyrate in the gut, the epithelial barrier loses integrity allowing for the penetration of undigested food, toxins, and certain microbes, termed “pathobionts,” into the systemic circulation. A strong T_H2 response is in turn initiated in the skin causing significant tissue damage.⁵⁹ Furthermore, butyrate is essential for the proper development of the mucosal immune system inducing development of regulatory T cells.⁵⁷ Finally, GABA, the most common inhibitory neurotransmitter, plays an essential role in itch restriction, whereas acetylcholine plays a critical role in both the development and maintenance of the skin epithelium.⁵⁹

Although the role of the gut microbiome in AD is compelling, whether manipulation of intestinal microbes with probiotics will affect AD remains less clear. Although conditioned and supported by very low-quality evidence, the World Allergy Organization has determined that there is a likely net benefit from using *Lactobacillus rhamnosus* GG supplementation during pregnancy to prevent high-risk infants from developing AD.⁶⁰ Several studies evaluating the treatment of AD with probiotics have failed to find consistent benefit; however, this lack of efficacy is likely due to the fact that the bacteria used in commercially available probiotics more closely resemble that in infants' guts.⁶¹ Butyrate-producing, anaerobic bacteria that more closely resemble that which is found in adolescents and adults have yet to be encapsulated in a form that resists oxygen degradation. Although development of effective probiotics is ongoing, immune-regulating prebiotics, such as natural fibers, and post-biotic therapies, such as GABA and butyrate, may soon provide a steroid-sparing alternative treatment for AD.⁶¹

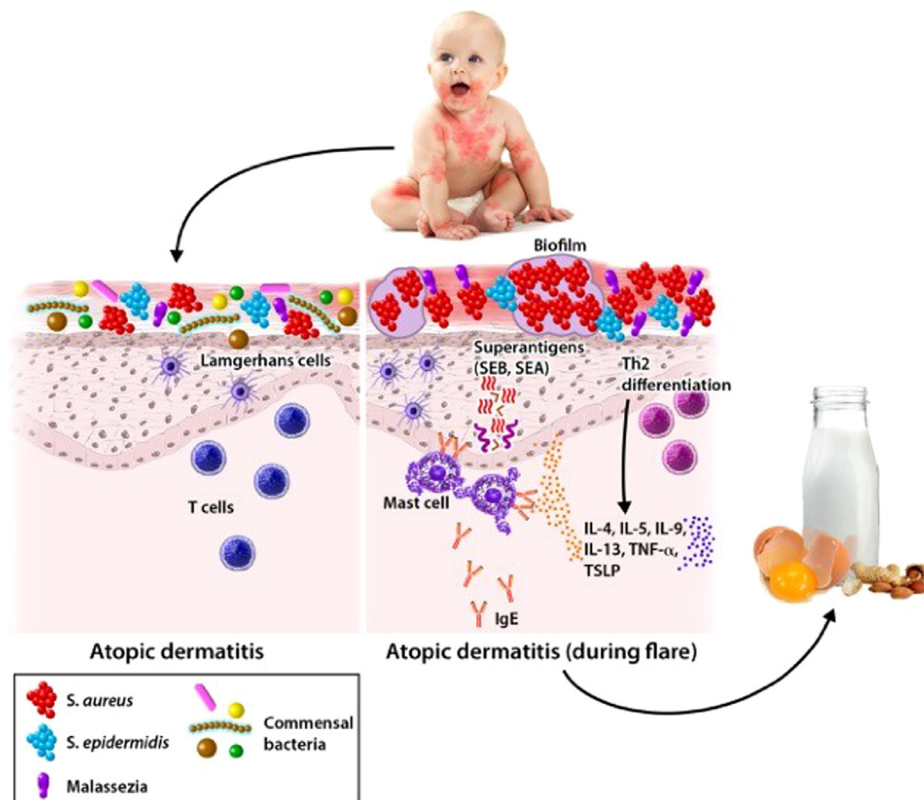


Figure 4. Microbiome dysbiosis in AD. Flares are characterized by less microbial diversity, increased harmful bacteria (*Staphylococcus aureus*) and fungi (*Malassezia*), and further wall disruption. *S aureus* superantigens activate IgE-mediated mast cells leading to T_H2 responses. The sustained colonization by *S aureus* has been related to increased production of serum-specific IgE to cow's milk, egg, and peanut. AD, atopic dermatitis.

Skin-Targeted Therapies for Atopic Dermatitis and Food Allergy Prevention

Strengthening Skin Barrier Function: Moisturizing Agents

The type of topical therapy that provides the best skin barrier protection has been evaluated in several studies including non-lipid emollients, emollients containing at least 1 ceramide, and a combination of emollients and topical steroids.¹⁰ Petrolatum-based emollients are currently considered the gold standard ointment-based emollient for the management of AD.⁶² Nevertheless, in some patients, petrolatum-based emollients can exacerbate AD. It is thought that trilipid creams, which mimic the skin's natural pH and lipid composition (3:1:1 ratio of ceramides, cholesterol, and free fatty acids), may be most effective in maintaining skin integrity.^{63,64} However, these are not available in many countries. Trilipid emollients have been reported to be more effective than paraffin-based emollients in reducing TEWL and total and specific IgE levels when compared with petrolatum-based emollients.⁶⁵

Emollients for the Prevention of Atopic Dermatitis

Several studies have evaluated emollients as a preventive strategy against AD. In a Cochrane review evaluating the effect of skin care interventions (including emollient application and bathing practices) on the prevention of AD and FA in infants,⁶⁶ the authors concluded that “This review found that skin care interventions such as emollients probably do not influence the development or time to onset of eczema in healthy term infants by age 1 to 2 years and probably increase the risk of skin infection.”⁶⁶ Factors such as frequent infant bathing before the onset of AD and bath emollients (baths with oil- or emulsifier-based additive) have been found to increase the risk of AD.^{67,68}

Conversely, a systematic review and meta-analysis of only preventative emollient therapy (not bathing practices) found that the prophylactic application of emollients initiated in early infancy may prevent AD, especially in high-risk populations and when used continuously up until the point of AD assessment.⁶⁹ Since the release of these meta-analyses, the STOP-AD (Short-term Topical Application to Prevent Atopic Dermatitis) trial has found a significant reduction in AD cumulative incidence by 1 year of age in high-risk infants who had a ceramide-based emollient applied in the first 2 months of life and then discontinued.⁷⁰

Emollients for the Prevention of Food Allergy

There are contradictory results regarding the use of preventative emollient therapy to prevent FA. The Prevention of Eczema By a Barrier Lipid Equilibrium Strategy pilot study revealed a reduction in investigator-observed AD and food sensitization (by skin prick test) at 12 months with a trilipid emollient therapy in the per-protocol group.⁷¹ However, several studies have found no significant reduction of the incidence of AD, such as the Barrier Enhancement for Eczema Prevention and the study Preventing AD and ALLergies in Children studies. The Barrier Enhancement for Eczema Prevention study also found a higher rate of skin infections in the intervention group and higher trend toward increased FA.⁷² Furthermore, in a retrospective review of skin emollient applications in participants of the Enquiring About Tolerance study, a dose-dependent association was found between frequency of application of emollients (predominantly olive oil) and the development of food sensitization allergy.⁶⁴ It should be noted that in a study evaluating emollient for the treatment of AD, 65% of these contained contact allergens.⁷³

Further studies are needed to reach definite conclusions. The Stopping Eczema and ALLergy study (NCT03742414⁷⁴), among others, is a randomized, controlled, parallel-design, open-label phase 2 clinical study that compares the effect of early proactive skin care with

petrolatum-based emollients vs trilipid-based emollients, against standard-of-care AD therapy, to reduce occurrence and severity of AD in children and prevent FA at 3 years of age.

Proactive Steroid Use and Topical Immunomodulators

Barrier-based approaches are valuable for disease prevention; however, immune-based treatments targeting T_H2 cytokines and immune abnormalities probably need to also be addressed to improve and resolve active AD and prevent FA.^{62,75} Proactive topical anti-inflammatory therapy is defined as a combination of long-term anti-inflammatory treatment applied initially daily to achieve control and then usually down-dose to 2 to 3 times per week for 16 weeks to maintain control in previously affected areas of the skin. This therapy can be done using topical corticosteroids and/or with topical calcineurin inhibitors (TCIs) in combination with an emollient (liberal use) with scheduled appointments to review AD control and monitor for any adverse effects.⁷⁵⁻⁷⁸ (Table 1).

In a retrospective analysis of infants presenting to a Japanese tertiary center, the use of proactive topical steroids within 4 months duration of moderate-severe AD vs commencing after 4 months duration of moderate-severe AD resulted in 2-fold reduction by 24 months.⁷⁹ The same group subsequently performed a randomized controlled trial using proactive topical steroids in children with mild AD within 7 to 13 weeks of age including nonaffected areas in the intervention arm and revealed a 25% reduction in egg allergy by 6 months of age. However, it also revealed a significant reduction in weight and length in the intervention arm.⁷⁹ This highlights important safety concerns with regard to use of potent topical steroids to the whole body (even areas without visible AD) for future studies.

Alternative anti-inflammatory topical therapies include TCIs licensed to treat AD in children 2 years of age and older in Europe and in the United States, tacrolimus 0.03% ointment and pimecrolimus 1% cream. However, in Canada, the latter has also been approved for 3 months of age and older. The use of these treatments in children younger than 2 years of age is off label, although, very common.⁷⁷ Proactive therapy with topical corticosteroid and TCI against reactive treatment has proven to prolong the interval between flares and to benefit from a lower barrier disruption.^{75,77} However, it has 2 potential problematic aspects to consider: the lack of knowledge regarding long-term safety and adverse effects in children younger than 2 years of age and the uncertain duration of this regimen.⁷⁵ There are, however, 10-year safety data revealing no increased risk of cancer with the use of tacrolimus 0.03% and 0.1% ointment in children with initiated therapy before 2 years⁸⁰ and a recent systematic review revealing little to no risk of cancer in pediatric or adult patients treated with TCI.⁸¹

Probiotics and Prebiotics Topical Application for the Prevention of Atopic Dermatitis and Food Allergy

Multiple factors influenced microbial diversity; however, to date, the immunomodulatory effect of topical application of “biotics” is still inconclusive.⁷⁷ Prebiotic mixtures have been found to be beneficial in preventing AD,⁷⁶ and probiotics containing lactobacillus mixtures, among others, also found to improve AD in some studies, and in others made little or no difference.^{76,82} This may be due to differences in the strains of probiotics, the characteristics of the host, the timing of the application, and other risk factors, including air pollution, climate, psychosocial factors, and diet.

To date, we still have little evidence regarding the role probiotics have in FA prevention; however, this route presents an interesting and promising alternative approach to address the gut-skin axis in patients with AD. Future research should focus on this to reach definitive conclusions.

Table 1
Comparison of Skin-Targeted Therapies for Food Allergy Prevention

Variables	Emollients/moisturizers	Topical corticosteroids ^{1,2}						Topical immunomodulators		
		Group I	Group II	Group III	Group IV	Group V	Groups VI-VII	Tacrolimus		Pimecrolimus
								0.03% Low dose	0.1% High dose	
Age restrictions (children < 2 y old)	No	Under Specialist Supervision		Yes				Off-label use	No	Off-label use
Skin application	Body and face	Body	Body and face	Body and face				Body and face		Body and face
Application schedule	BD	OD						OD or BD		
Proactive therapy for prevention of eczema flares ^{3,4,5}	NA	No	Yes	Yes ^{3,a}	No		Yes		No ⁴ /Yes ⁵	
Short-term treatment	Yes	Yes						Yes		
Long-term treatment ^{4,5,6}	Yes	No	No	No	Yes	No		Yes	Yes	
Benefits preventing FA	Inconclusive ⁷	Inconclusive		Yes ^{8,b}		Inconclusive		Inconclusive		Inconclusive
Common adverse effects	May contain allergenic plant proteins	Skin atrophy		Telangiectasia		Stretch marks		Skin atrophy		Site burning
	Skin infections	Stretch marks		Growth restriction in infancy		Stretch marks		Irritation		Skin infections

Abbreviations: AD, atopic dermatitis; BD, twice daily; FA, food allergy; NA, non-applicable; PACI, Prevention of Allergy via Cutaneous Intervention.

^aIn Reference 3: Fluticasone propionate 0.05% is considered medium potency in the United States (Reference 2). However, the same topical corticosteroid is considered a potent corticosteroid in the United Kingdom (Available at www.medicines.org.uk/emc/files/pil.9364.pdf).

^bThe PACI study: Betamethasone was used in this study to evaluate whether an enhanced early skin treatment for AD in infants reduces food allergy. Results revealed that the enhanced treatment significantly reduced hen's egg allergy compared with the conventional treatment. Please note that betamethasone valerate is considered strong (0.12% cream: Reference 1) and/or medium potency in the United States (0.1% cream: References 2 and 7), but in the United Kingdom is considered a potent topical corticosteroid (available at <https://www.medicines.org.uk/emc/product/929/smpc#about-medicine>).

References for Table 1:

- Adapted from: Saeki H, Ohya Y, Furuta J, Arakawa H, Ichiyama S, Katsunuma T, et al. English Version of Clinical Practice Guidelines for the Management of Atopic Dermatitis 2021. *J Dermatol*. 2022;49(10):e315-e375. 10.1111/1346-8138.16527. Epub 2022 Aug 22. PMID: 35996152.
- Adapted from: Eichenfield LF, Tom WL, Berger TG, Krol A, Paller AS, Schwarzenberger K, et al. Guidelines of care for the management of atopic dermatitis: section 2. Management and treatment of atopic dermatitis with topical therapies. *J Am Acad Dermatol*. 2014;71(1):116-32. 10.1016/j.jaad.2014.03.023. Epub 2014 May 9. PMID: 24813302; PMCID: PMC4326095.
- Adapted from: Berth-Jones J, Damstra RJ, Golsch S, Livden JK, Van Hooteghem O, Allegra F, et al. Multinational Study Group. Twice weekly fluticasone propionate added to emollient maintenance treatment to reduce risk of relapse in atopic dermatitis: randomised, double blind, parallel group study. *BMJ*. 2003;326(7403):1367. 10.1136/bmj.326.7403.1367. PMID: 12816824; PMCID: PMC162129.
- Adapted from: Wollenberg A, Kinberger M, Arents B, Aszodi N, Avila Valle G, Barbarot S, et al. European guideline (EuroGuiDerm) on atopic eczema - part II: non-systemic treatments and treatment recommendations for special AE patient populations. *J Eur Acad Dermatol Venereol*. 2022;36(11):1904-1926. 10.1111/jdv.18429. Epub 2022 Sep 3. PMID: 36056736.
- Adapted from: Chu DK, Chu AWL, Rayner DG, Guyatt GH, Yepes-Nuñez JJ, Gomez-Escobar L, et al. Topical treatments for atopic dermatitis (eczema): systematic review and network meta-analysis of randomized trials. *J Allergy Clin Immunol*. 2023;S0091-6749(23)01113-2. 10.1016/j.jaci.2023.08.030. Epub ahead of print. PMID: 37678572.
- Adapted from: Wollenberg A, Barbarot S, Bieber T, Christen-Zaech S, Deleuran M, Fink-Wagner A, et al. European Dermatology Forum (EDF), the European Academy of Dermatology and Venereology (EADV), the European Academy of Allergy and Clinical Immunology (EAACI), the European Task Force on Atopic Dermatitis (ETFAD), European Federation of Allergy and Airways Diseases Patients' Associations (EFA), the European Society for Dermatology and Psychiatry (ESDaP), the European Society of Pediatric Dermatology (ESPD), Global Allergy and Asthma European Network (GA2LEN) and the European Union of Medical Specialists (UEMS). Consensus-based European guidelines for treatment of atopic eczema (atopic dermatitis) in adults and children: part I. *J Eur Acad Dermatol Venereol*. 2018;32(5):657-682. 10.1111/jdv.14891. Erratum in: *J Eur Acad Dermatol Venereol*. 2019;33(7):1436. PMID: 29676534.
- Adapted from: Kelleher MM, Phillips R, Brown SJ, Cro S, Cornelius V, Carlsen KCL, et al. Skin care interventions in infants for preventing eczema and food allergy. *Cochrane Database Syst Rev*. 2022;11(11):CD013534.
- Adapted from: Yamamoto-Hanada K, Kobayashi T, Mikami M, Williams HC, Saito H, Saito-Abe M, et al. PACI Study Collaborators. Enhanced early skin treatment for atopic dermatitis in infants reduces food allergy. *J Allergy Clin Immunol*. 2023;152(1):126-135. 10.1016/j.jaci.2023.03.008. Epub 2023 Mar 22. PMID: 36963619.

Novel Skin-Targeted Therapies

The currently available data have propelled the skin as a potential immunomodulatory organ for treating FAs. A better understanding of the progression of AD has facilitated and triggered the surge of innovative treatments directed toward pathologic pathways. In this sense, since Palmer et al³² revealed that FLG loss-of-function mutations represented the most critical genetic risk linked to AD, hundreds of mutations have been identified.⁸³

Potential therapies involving FLG expression include the following: "read-through" drugs, direct replacement of FLG, indirect replacement by topical application of FLG metabolites, and inhibition of cytokine-mediated FLG down-regulation which includes interleukin and janus kinases inhibitors.⁸⁴ The "read-through" drugs manage gene expression by acting over regulatory elements, inhibiting a determined mutation, and subsequently enabling the adequate reading frame, which results in the generation of full-length FLG protein.⁸⁵ These drugs are patented but not yet available.⁸⁶ The topical application of FLG metabolites such as UCA and pyrrolidone carboxylic acid is considered an indirect FLG replacement that could preserve the permeability of the barrier function.⁸⁵ Particularly, the use of 5% cis-UCA cream⁸⁷ has been

confirmed as an efficient and safe option for patients with mild-to-moderate AD.

FLG down-regulation is currently understood to be secondary to uncontrolled expression of IL-4 and IL-13 (T_H2) and IL-22 (T_H22). Other cytokines involved in FLG deficiency are IL-20, IL-24, IL-25, IL-31, and IL-33, but the corresponding mechanisms still need to be understood. Conversely, the up-regulation of FLG expression is increased by activating a ligand-activated transcription factor, an aryl hydrocarbon receptor.^{84,88,89}

Furthermore, treatment strategies that are novel and targeted toward the skin, such as epicutaneous immunotherapy (Table 2) and skin-targeted gene therapy, aim to manage and prevent complications of allergic diseases.

Skin-Targeted Gene Therapy

Skin-targeted gene therapy is an effective and convenient treatment option for the prevention and treatment of FA which is achieved by delivering the therapeutic protein or the allergen in the form of plasmid DNA in vivo to modulate allergic immune responses. Adenovirus remains to be the most extensively used gene transfer

Table 2
Clinical Trials Using EPIT for the Treatment of FA

Title	NCT	Phase	PMID(s)	Finding	Size
Efficacy and Safety of Several Doses of Viaskin Peanut in Adults and Children With Peanut Allergy (VIPES)	NCT01675882	2b	29136445	Increase in reaction threshold in those who received peanut EPIT (250 mg Viaskin [VP250]) compared with placebo after 52 wk of treatment	221 individuals with peanut allergy between 6 and 55 y old across 22 centers in the United States and Europe
Peanut Epicutaneous Phase II Immunotherapy Clinical Trial	NCT01904604	2	28091362 33290772	Peanut EPIT was associated with significant desensitization and immunologic changes after 52 wk of treatment in those who received 100 mg (VP100) or VP250 doses, compared with placebo EPIT, with the highest product adherence and response observed within the younger children. Open-label extension revealed VP250 was well tolerated, and desensitization persisted between weeks 52 and 130, though treatment success was predominantly observed in the younger cohort. EPIT increased IgG4 but no change in IgE or basophil activation.	75 individuals with peanut allergy between 4 and 25 y old recruited across 5 clinical CoFAR sites in the United States.
Efficacy and Safety of Viaskin Peanut in Children With IgE-Mediated Peanut Allergy (PEPITES)	NCT02636699	3	30794314	After 12 mo of treatment, 25.3% of peanut EPIT-treated participants and 13.6% of placebo-treated participants reached an eliciting dose of ≥ 1000 mg peanut.	356 individuals with peanut allergy between 4 and 11 years of age recruited across 31 different sites in the United States, Canada, Australia, and Europe
Follow-up of the PEPITES Study to Evaluate Long-term Efficacy and Safety of Viaskin Peanut in Children (PEOPLE)	NCT03013517	3	32659313	After 24 mo of open-label extension of peanut EPIT, 51.8% of participants reached an eliciting dose of ≥ 1000 mg peanut in comparison to 40.4% who reached this does before the open-label extension.	213 individuals with peanut allergy who received peanut EPIT in the PEPITES trial ^a
Safety Study of Viaskin Peanut to Treat Peanut Allergy (REALISE)	NCT02916446	3	4848381	VP250 was well tolerated in children with peanut allergy, although local skin reactions were reported in all the children receiving VP250 and 83.8% of those on placebo.	399 individuals with peanut allergy between 4 and 11 years of age recruited across 32 sites in the United States and Canada
Safety and Efficacy Study of Viaskin Peanut in Peanut-allergic Young Children 1-3 Years of Age (EPITOPE)	NCT03211247	3	37163622	After 12 mo of treatment, 67% of VP250-treated participants and 33.5% of placebo-treated participants passed a peanut OFC to ≥ 1000 mg peanut. Adverse reactions were found in all VP250 participants and 99.2% of placebo-treated participants.	414 individuals with peanut allergy between 1 and 4 years old recruited across 51 sites in the United States, Canada, Australia, and Europe

Abbreviations: CoFAR, Consortium of Food Allergy Research; EPIT, epicutaneous immunotherapy; FA, food allergy; OFC, oral food challenge.

^aOnly 141 received OFCs and are included in the data.²

References for Table 2:

- Wollenberg, A. et al. Consensus-Based European Guidelines for Treatment of Atopic Eczema (Atopic Dermatitis) in Adults and Children: Part I. *J Eur Acad Dermatol Venereol.* 2018;32:657-682.
- Fleischer, D. M. et al. Long-Term, Open-Label Extension Study of the Efficacy and Safety of Epicutaneous Immunotherapy for Peanut Allergy in Children: People 3-Year Results. *J Allergy Clin Immunol.* 2020;146:863-874.

vector and has been found to be highly tolerable and efficient. One such example includes a recent study in murine model to assess the potential benefit of single-dose of adeno-associated virus (AAV) vector gene immunotherapy to treat FA.^{90,91} In this study, AAV vectors expressing ovalbumin (OVA) was injected before or after epicutaneous sensitization with OVA. The study revealed that mice treated with AAV-OVA vector were protected from allergy sensitization and had significant reduction in anaphylaxis. Furthermore, AAV gene immunotherapy resulted in induction of OVA-specific T regulatory cells, induction of IL-10, and reduction in IL-13. In addition, while a phase 1, randomized, placebo-controlled study is currently investigating the safety, tolerability, and immune response of intradermal administration of plasmid DNA encoding peanut allergens (ASP0892),⁹⁰ it has been proposed that plasmid DNA vaccination can activate natural killer cells that produce interferon gamma, influencing skewing of T helper cell responses.⁹² Further studies are warranted to further investigate the potential of gene therapies for the treatment of FAs.

Conclusion

The importance of skin in our biologic defense, its critical role in preventing environmental agents (eg, microbes and allergens) from penetrating the human body, and its response to microbial pathogens are well established. Skin barrier disruption and dysfunction is the initial step in the development of AD, the most common, complex, chronic inflammatory skin condition.

Despite the long-recognized association between AD and the subsequent development of other allergic diseases, the mechanistic link

between AD and allergic disease pathogenesis still needs to be fully understood. It is also evident that allergic disease, particularly FA, may trigger or exacerbate AD, so the relationship is complex and results in a feed-forward loop of allergic disease progression.⁹³ Given the central role of AD, the skin barrier, and skin host immune responses in allergic disease pathogenesis, it is not surprising that there is increasing interest in interventions targeting the skin, including enhancing the skin barrier and optimizing the skin biome. Studies focused on strengthening the skin barrier using emollients to prevent AD and FA have been contradictory rather than conclusive. Recent studies using topical probiotics have shown promise in preventing and treating AD through the modulation of host immune responses. However, there have also been conflicting results regarding the clinical effects of probiotics in patients with AD.

Dysregulation of neuroimmune circuits plays a critical role in the pathophysiology of AD, causing inflammation, pruritus, pain, and barrier dysfunction. Sensory nerves can also be activated by environmental or endogenous trigger factors, and on stimulation, sensory nerve endings release neuromediators, which contribute to barrier dysfunction and inflammation propagating itch. Thus, neuroimmune circuits may be key targets to control pruritus in AD, which would reduce the itch-scratch cycle, which we know increases TSLP and mast cell expansion in the gut.⁹⁴ Given the complex environmental and host factors that promote AD and allergic disease, future research is needed that is multi- and cross-disciplinary and brings together leaders in allergy, immunology, dermatology, microbiology, neurology, neuroimmunology, genetics, environmental health, nutrition and gastrointestinal health, and epidemiology to work as seamless teams. Understanding how the immune system, neuroimmune

circuits, environmental factors, genetics, and microbiome coordinate pathologic mechanisms that underlie these common disorders will be necessary to advance the field.

Disclosures

Dr Ciaccio receives research grant support from the National Institutes of Health (NIH), Food Allergy Research & Education (FARE), and Paul and Mary Yovovich and has served as a medical consultant/advisor for Genentech, Novartis, Siolta, Clostrabio, and FARE. Dr Chinthrajah reports receiving grants from National Institute of Allergy and Infectious Diseases (NIAID), CoFAR, Regeneron, Stanford Maternal and Child Health Research Institute, and FARE and is an advisory board member at Alladapt Therapeutics, Novartis, Genentech, Allergenix, Intrommune Therapeutics, and IgGenix. Dr Chan reports receiving grant from NIAID and NIH. Prof Leung reports receiving grants from Genentech, Incyte Corporation, and Sanofi-Genzyme; nonfinancial support from Aslan Pharmaceuticals; and personal fees from Leo Pharmaceuticals. Dr Marshall reports receiving research grant support from NIH. Prof Nadeau reports receiving grants from NIAID; National Heart, Lung, and Blood Institute (NHLBI); National Institute of Environmental Health Sciences (NIEHS); and FARE; receiving stock options from IgGenix, Seed Health, ClostraBio, Cour, and Alladapt; serving as an advisor at Cour Pharma; serving as a consultant for Excellergy, Red tree ventures, Before Brands, Alladapt, Cour, Latitude, Regeneron, and IgGenix; serving as a co-founder of Before Brands, Alladapt, Latitude, and IgGenix; serving as National Scientific Committee member at Immune Tolerance Network (ITN) and NIH clinical research centers; and having patents including, “Mixed allergen composition and methods for using the same,” “Granulocyte-based methods for detecting and monitoring immune system disorders,” and “Methods and Assays for Detecting and Quantifying Pure Subpopulations of White Blood Cells in Immune System Disorders.” Dr Radulovic reports receiving grant from NIAID and NIH. Prof Lack reports receiving grant from NIAID/NIH; having personal fees and stock options from DBV Technologies; having stock options from Mission MightyMe; and serving as a scientific consultant/advisor for Novartis, Sanofi-Genzyme, Regeneron, ALK-Abello, Reckitt Mead Johnson, and Lurie Children’s Hospital. Dr Sindher receives research grant support from NIH, FARE, CoFAR, DBV, AIMMUNE, and Regeneron and has served as an advisor for Genentech. Prof Brough reports receiving grant from NIAID and NIH and receiving speaker honoraria from DBV Technologies, GlaxoSmithKline, and Sanofi. The remaining authors have no conflicts of interest to report.

Funding

The authors have no funding sources to report.

References

- Nadeau K. *Approach to the patient with allergic or immunologic disease*. 235. Goldman-Cecil Med; 2019. 1634–1638.e1.
- Genuneit J, Standl M. Epidemiology of allergy: natural course and risk factors of allergic diseases. *Handb Exp Pharmacol*. 2022;268:21–27.
- Gupta RS, Warren CM, Smith BM, Jiang J, Blumenstock JA, Davis MM, et al. Prevalence and severity of food allergies among US adults. *JAMA Netw Open*. 2019;2(1):e185630.
- Aldakheel FM. Allergic diseases: a comprehensive review on risk factors, immunological mechanisms, link with COVID-19, potential treatments, and role of allergen bioinformatics. *Int J Environ Res Public Health*. 2021;18(22):12105.
- Guo H, An J, Yu Z. Identifying shared risk genes for asthma, hay fever, and eczema by multi-trait and multiomic association analyses. *Front Genet*. 2020;11:270.
- Johansson E, Biagini Myers JM, Martin LJ, He H, Ryan P, LeMasters GK, et al. Identification of two early life eczema and non-eczema phenotypes with high risk for asthma development. *Clin Exp Allergy*. 2019;49(6):829–837.
- Wang H, Li XB, Chu XJ, Cao NW, Wu H, Huang RG, et al. Ambient air pollutants increase the risk of immunoglobulin E-mediated allergic diseases: a systematic review and meta-analysis. *Environ Sci Pollut Res Int*. 2022;29(33):49534–49552.
- Paller AS, Spergel JM, Mina-Osorio P, Irvine AD. The atopic march and atopic morbidity: many trajectories, many pathways. *J Allergy Clin Immunol*. 2019;143(1):46–55.
- Kucuksezer UC, Ozdemir C, Yazici D, Pat Y, Mitamura Y, Li M, et al. The epithelial barrier theory: development and exacerbation of allergic and other chronic inflammatory diseases. *Asia Pac Allergy*. 2023;13(1):28–39.
- Brough HA, Nadeau KC, Sindher SB, Alkotob SS, Chan S, Bahnson HT, et al. Epicutaneous sensitization in the development of food allergy: what is the evidence and how can this be prevented? *Allergy*. 2020;75(9):2185–2205.
- Brough HA, Liu AH, Sicherer S, Makinson K, Douiri A, Brown SJ, et al. Atopic dermatitis increases the effect of exposure to peanut antigen in dust on peanut sensitization and likely peanut allergy. *J Allergy Clin Immunol*. 2015;135(1):164–170.
- Goleva E, Berdyshev E, Leung DY. Epithelial barrier repair and prevention of allergy. *J Clin Invest*. 2019;129(4):1463–1474.
- Leung DYM, Berdyshev E, Goleva E. Cutaneous barrier dysfunction in allergic diseases. *J Allergy Clin Immunol*. 2020;145(6):1485–1497.
- Fluhr JW, Feingold KR, Elias PM. Transepidermal water loss reflects permeability barrier status: validation in human and rodent in vivo and ex vivo models. *Exp Dermatol*. 2006;15(7):483–492.
- Leung DYM, Calatroni A, Zaramela LS, LeBeau PK, Dyjack N, Brar K, et al. The nonlesional skin surface distinguishes atopic dermatitis with food allergy as a unique endotype. *Sci Transl Med*. 2019;11(480):eaav2685.
- Hui-Beckman JW, Goleva E, Berdyshev E, Leung DYM. Endotypes of atopic dermatitis and food allergy. *J Allergy Clin Immunol*. 2023;151(1):26–28.
- Xian M, Wawrzyniak P, Rückert B, Duan S, Meng Y, Sokolowska M, et al. Anionic surfactants and commercial detergents decrease tight junction barrier integrity in human keratinocytes. *J Allergy Clin Immunol*. 2016;138(3):890–893.e9.
- Kim BE, Kim J, Goleva E, Berdyshev E, Lee J, Vang KA, et al. Particulate matter causes skin barrier dysfunction. *JCI Insight*. 2021;6(5):e145185.
- Tsuchida A, Itazawa T, Matsumura K, Yokomichi H, Yamagata Z, Adachi Y, et al. Season of birth and atopic dermatitis in early infancy: results from the Japan Environment and Children’s Study. *BMC Pediatr*. 2023;23(1):78.
- Hui-Beckman JW, Goleva E, Leung DYM, Kim BE. The impact of temperature on the skin barrier and atopic dermatitis. *Ann Allergy Asthma Immunol*. 2023;131(6):713–719.
- Kim BE, Leung DYM. Significance of skin barrier dysfunction in atopic dermatitis. *Allergy Asthma Immunol Res*. 2018;10(3):207–215.
- Kim BE, Hui-Beckman J, Lyubchenko T, Hall CF, Fallahi S, Brull A, et al. Transient receptor potential vanilloid 1 plays a major role in low temperature-mediated skin barrier dysfunction. *J Allergy Clin Immunol*. 2022;150(2):362–372.e7.
- Du Toit G, Roberts G, Sayre PH, Bahnson HT, Radulovic S, Santos AF, et al. Randomized trial of peanut consumption in infants at risk for peanut allergy. *N Engl J Med*. 2015;372(9):803–813.
- Perkin MR, Logan K, Marrs T, Radulovic S, Craven J, Flohr C, et al. Enquiring About Tolerance (EAT) study: feasibility of an early allergenic food introduction regimen. *J Allergy Clin Immunol*. 2016;137(5):1477–1486.e8.
- Skjerven HO, Lie A, Vettukattil R, Rehinder EM, LeBlanc M, Asarnoj A, et al. Early food intervention and skin emollients to prevent food allergy in young children (PreventADALL): a factorial, multicentre, cluster-randomised trial. *Lancet*. 2022;399(10344):2398–2411.
- Scarpone R, Kimkool P, Ierodiakonou D, Leonardi-Bee J, Garcia-Larsen V, Perkin MR, et al. Timing of allergenic food introduction and risk of immunoglobulin E-mediated food allergy: a systematic review and meta-analysis. *JAMA Pediatr*. 2023;177(5):489–497.
- Lack G, Fox D, Northstone K, Golding J. Avon Longitudinal Study of Parents and Children Study Team. Factors associated with the development of peanut allergy in childhood. *N Engl J Med*. 2003;348(11):977–985.
- Fox AT, Sasieni P, du Toit G, Syed H, Lack G. Household peanut consumption as a risk factor for the development of peanut allergy. *J Allergy Clin Immunol*. 2009;123(2):417–423.
- Elias PM, Hatano Y, Williams ML. Basis for the barrier abnormality in atopic dermatitis: outside-inside-outside pathogenic mechanisms. *J Allergy Clin Immunol*. 2008;121(6):1337–1343.
- Feingold KR, Elias PM. Role of lipids in the formation and maintenance of the cutaneous permeability barrier. *Biochim Biophys Acta*. 2014;1841(3):280–294.
- van Smeden J, Bouwstra JA. Stratum corneum lipids: their role for the skin barrier function in healthy subjects and atopic dermatitis patients. *Curr Probl Dermatol*. 2016;49:8–26.
- Palmer CN, Irvine AD, Terron-Kwiatkowski A, Zhao Y, Liao H, Lee SP, et al. Common loss-of-function variants of the epidermal barrier protein filaggrin are a major predisposing factor for atopic dermatitis. *Nat Genet*. 2006;38(4):441–446.
- Sandilands A, Sutherland C, Irvine AD, McLean WH. Filaggrin in the frontline: role in skin barrier function and disease. *J Cell Sci*. 2009;122(9):1285–1294.
- Janssens M, van Smeden J, Gooris GS, Bras W, Portale G, Caspers PJ, et al. Increase in short-chain ceramides correlates with an altered lipid organization and decreased barrier function in atopic eczema patients. *J Lipid Res*. 2012;53(12):2755–2766.
- van Smeden J, Janssens M, Gooris G, Bouwstra J. The important role of stratum corneum lipids for the cutaneous barrier function. *Biochim Biophys Acta (BBA) Mol Cell Biol Lipids*. 2014;1841(3):295–313.
- Mack JW, Steven AC, Steinert PM. The mechanism of interaction of filaggrin with intermediate filaments. The ionic zipper hypothesis. *J Mol Biol*. 1993;232(1):50–66.
- McAleer MA, Irvine AD. The multifunctional role of filaggrin in allergic skin disease. *J Allergy Clin Immunol*. 2013;131(2):280–291.
- Moosbrugger-Martinez V, Leprince C, Mechin MC, Simon M, Blunder S, Gruber R, et al. Revisiting the roles of filaggrin in atopic dermatitis. *Int J Mol Sci*. 2022;23(10):5318.

39. Celebi Sozener Z, Özbey Yücel Ü, Altiner S, Ozdel Oztürk B, Cerci P, Türk M, et al. The external exposome and allergies: from the perspective of the epithelial barrier hypothesis. *Front Allergy*. 2022;3: 887672.
40. Flohr C, Perkin M, Logan K, Marrs T, Radulovic S, Campbell LE, et al. Atopic dermatitis and disease severity are the main risk factors for food sensitization in exclusively breastfed infants. *J Invest Dermatol*. 2014;134(2):345–350.
41. Brown SJ, Asai Y, Cordell HJ, Campbell LE, Zhao Y, Liao H, et al. Loss-of-function variants in the filaggrin gene are a significant risk factor for peanut allergy. *J Allergy Clin Immunol*. 2011;127(3):661–667.
42. Brough HA, Simpson A, Makinson K, Hankinson J, Brown S, Douiri A, et al. Peanut allergy: effect of environmental peanut exposure in children with filaggrin loss-of-function mutations. *J Allergy Clin Immunol*. 2014;134(4): 867–75.e1.
43. Koh LF, Ong RY, Common JE. Skin microbiome of atopic dermatitis. *Allergol Int*. 2022;71(1):31–39.
44. Tsilochriou O, du Toit G, Sayre PH, Roberts G, Lawson K, Sever ML, et al. Association of *Staphylococcus aureus* colonization with food allergy occurs independently of eczema severity. *J Allergy Clin Immunol*. 2019;144(2):494–503.
45. Alexander H, Paller AS, Traidl-Hoffmann C, Beck LA, De Benedetto A, Dhar S, et al. The role of bacterial skin infections in atopic dermatitis: expert statement and review from the International Eczema Council Skin Infection Group. *Br J Dermatol*. 2020;182(6):1331–1342.
46. Berin MC, Sampson HA. Mucosal immunology of food allergy. *Curr Biol*. 2013;23(9):R389–R400.
47. Williams MR, Gallo RL. The role of the skin microbiome in atopic dermatitis. *Curr Allergy Asthma Rep*. 2015;15(11):65.
48. Selander C, Engblom C, Nilsson G, Scheynius A, Andersson CL. TLR2/MyD88-dependent and -independent activation of mast cell IgE responses by the skin commensal yeast *Malassezia sympodialis*. *J Immunol*. 2009;182(7):4208–4216.
49. Forbes-Blom E, Camberis M, Prout M, Tang SC, Le Gros G. *Staphylococcus*-derived superantigen enhances peanut induced Th2 responses in the skin. *Clin Exp Allergy*. 2012;42(2):305–314.
50. Tamagawa-Mineoka R, Katoh N. Atopic dermatitis: identification and management of complicating factors. *Int J Mol Sci*. 2020;21(8):2671.
51. Alam MJ, Xie L, Yap YA, Marques FZ, Robert R. Manipulating microbiota to treat atopic dermatitis: functions and therapies. *Pathogens*. 2022;11(6):642.
52. Abrahamsson TR, Jakobsson HE, Andersson AF, Björkstén B, Engstrand L, Jenmalm MC. Low diversity of the gut microbiota in infants with atopic eczema. *J Allergy Clin Immunol*. 2012;129(2):434–440.
53. Penders J, Stobberingh EE, Thijs C, Adams H, Vink C, van Ree R, et al. Molecular fingerprinting of the intestinal microbiota of infants in whom atopic eczema was or was not developing. *Clin Exp Allergy*. 2006;36(12):1602–1608.
54. Lee E, Lee SY, Kang MJ, Kim K, Won S, Kim BJ, et al. Clostridia in the gut and onset of atopic dermatitis via eosinophilic inflammation. *Ann Allergy Asthma Immunol*. 2016;117(1):91–92.e1.
55. Kirjavainen PV, Arvola T, Salminen SJ, Isolauri E. Aberrant composition of gut microbiota of allergic infants: a target of bifidobacterial therapy at weaning? *Gut*. 2002;51(1):51–55.
56. Nylund L, Nermes M, Isolauri E, Salminen S, de Vos WM, Satokari R. Severity of atopic disease inversely correlates with intestinal microbiota diversity and butyrate-producing bacteria. *Allergy*. 2015;70(2):241–244.
57. Kim JE, Kim HS. Microbiome of the skin and gut in atopic dermatitis (AD): understanding the pathophysiology and finding novel management strategies. *J Clin Med*. 2019;8(4):444.
58. De Pessemier B, Grine L, Debaere M, Maes A, Paetzold B, Callewaert C. Gut-skin axis: current knowledge of the interrelationship between microbial dysbiosis and skin conditions. *Microorganisms*. 2021;9(2):353.
59. Schleimer RP, Berdnikovs S. Etiology of epithelial barrier dysfunction in patients with type 2 inflammatory diseases. *J Allergy Clin Immunol*. 2017;139(6):1752–1761.
60. Fiocchi A, Pawankar R, Cuello-García C, Ahn K, Al-Hammadi S, Agarwal A, et al. World Allergy Organization-McMaster University guidelines for allergic disease prevention (GLAD-P): probiotics. *World Allergy Organ J*. 2015;8(1):4.
61. Rusu E, Enache G, Cursaru R, Alexescu A, Radu R, Onila O, et al. Probiotics and probiotics in atopic dermatitis. *Exp Ther Med*. 2019;18(2):926–931.
62. Brough HA, Lanser BJ, Sindher SB, Teng JMC, Leung DYM, Venter C, et al. Early intervention and prevention of allergic diseases. *Allergy*. 2022;77(2):416–441.
63. Luger T, Amagai M, Dreno B, Dagnelie MA, Liao W, Kabashima K, et al. Atopic dermatitis: role of the skin barrier, environment, microbiome, and therapeutic agents. *J Dermatol Sci*. 2021;102(3):142–157.
64. Perkin MR, Logan K, Marrs T, Radulovic S, Craven J, Boyle RJ, et al. Association of frequent moisturizer use in early infancy with the development of food allergy. *J Allergy Clin Immunol*. 2021;147(3): 967–976.e1.
65. Sindher S, Alkotob SS, Shojinaga MN, Brough HA, Bahnson HT, Chan S, et al. Pilot study measuring transepidermal water loss (TEWL) in children suggests trilipid cream is more effective than a paraffin-based emollient. *Allergy*. 2020;75(10):2662–2664.
66. Kelleher MM, Cro S, Cornelius V, Lodrup Carlsen KC, Skjervén HO, Rehbinder EM, et al. Skin care interventions in infants for preventing eczema and food allergy. *Cochrane Database Syst Rev*. 2021;2(2): Cd013534.
67. Marrs T, Perkin MR, Logan K, Craven J, Radulovic S, McLean WHI, et al. Bathing frequency is associated with skin barrier dysfunction and atopic dermatitis at three months of age. *J Allergy Clin Immunol Pract*. 2020;8(8):2820–2822.
68. O'Connor C, Livingstone V, OBH J, Irvine AD, Boylan G, Murray D. Early emollient bathing is associated with subsequent atopic dermatitis in an unselected birth cohort study. *Pediatr Allergy Immunol*. 2023;34(7):e13998.
69. Zhong Y, Samuel M, van Bever H, Tham EH. Emollients in infancy to prevent atopic dermatitis: a systematic review and meta-analysis. *Allergy*. 2022;77(6):1685–1699.
70. Ni Chaoimh C, Lad D, Nico C, Puppels GJ, Wong X, Common JE, et al. Early initiation of short-term emollient use for the prevention of atopic dermatitis in high-risk infants-The STOP-AD randomised controlled trial. *Allergy*. 2023;78(4):984–994.
71. Lowe A, Su J, Tang M, Lodge C, Matheson M, Allen KJ, et al. PEBBLES study protocol: a randomised controlled trial to prevent atopic dermatitis, food allergy and sensitisation in infants with a family history of allergic disease using a skin barrier improvement strategy. *BMJ Open*. 2019;9(3): e024594.
72. Chalmers JR, Haines RH, Bradshaw LE, Montgomery AA, Thomas KS, Brown SJ, et al. Daily emollient during infancy for prevention of eczema: the BEEP randomised controlled trial. *Lancet*. 2020;395(10228):962–972.
73. Ryczaj K, Dumycz K, Spiewak R, Feleszko W. Contact allergens in moisturizers in preventative emollient therapy - a systematic review. *Clin Transl Allergy*. 2022;12(6):e12150.
74. Sindher SB. SEAL Study (Seal, Stopping Eczema and Allergy Study). Full Text View. 2021. <https://ClinicalTrials.gov/NCT03742414>.
75. Frølund AS, Thyssen JP, Deleuran M, Vestergaard C. Appraisal of proactive topical therapy in atopic dermatitis: pros and cons. *Am J Clin Dermatol*. 2021;22(6):775–783.
76. Wollenberg A, Barbarot S, Bieber T, Christen-Zaech S, Deleuran M, Fink-Wagner A, et al. Consensus-based European guidelines for treatment of atopic eczema (atopic dermatitis) in adults and children: part 1. *J Eur Acad Dermatol Venereol*. 2018;32(5):657–682.
77. Wollenberg A, Christen-Zäch S, Taieb A, Paul C, Thyssen JP, de Bruin-Weller M, et al. ETFAD/EADV Eczema task force 2020 position paper on diagnosis and treatment of atopic dermatitis in adults and children. *J Eur Acad Dermatol Venereol*. 2020;34(12):2717–2744.
78. Butala S, Paller AS. Optimizing topical management of atopic dermatitis. *Ann Allergy Asthma Immunol*. 2022;128(5):488–504.
79. Yamamoto-Hanada K, Kobayashi T, Williams HC, Mikami M, Saito-Abe M, Morita K, et al. Early aggressive intervention for infantile atopic dermatitis to prevent development of food allergy: a multicenter, investigator-blinded, randomized, parallel group controlled trial (PACI Study)-protocol for a randomized controlled trial. *Clin Transl Allergy*. 2018;8:47.
80. Paller AS, Fölster-Holst R, Chen SC, Diepgen TL, Elmets C, Margolis DJ, et al. No evidence of increased cancer incidence in children using topical tacrolimus for atopic dermatitis. *J Am Acad Dermatol*. 2020;83(2):375–381.
81. Devasenapathy N, Chu A, Wong M, Srivastava A, Ceccacci R, Lin C, et al. Cancer risk with topical calcineurin inhibitors, pimecrolimus and tacrolimus, for atopic dermatitis: a systematic review and meta-analysis. *Lancet Child Adolesc Health*. 2023;7(1):13–25.
82. Makrgeorgou A, Leonardi-Bee J, Bath-Hextall FJ, Murrell DF, Tang ML, Roberts A, et al. Probiotics for treating eczema. *Cochrane Database Syst Rev*. 2018;11(11): Cd006135.
83. Gupta J, Margolis DJ. Filaggrin gene mutations with special reference to atopic dermatitis. *Curr Treat Options Allergy*. 2020;7(3):403–413.
84. Dębińska A. New treatments for atopic dermatitis targeting skin barrier repair via the regulation of FLG expression. *J Clin Med*. 2021;10(11):2506.
85. Dębińska A, Sozańska B. Epicutaneous sensitization and food allergy: preventive strategies targeting skin barrier repair-facts and challenges. *Nutrients*. 2023;15(5):1070.
86. Nagel-Wolfrum K, Möller F, Penner I, Baasov T, Wolfrum U. Targeting nonsense mutations in diseases with translational read-through-inducing drugs (TRIDs). *BioDrugs*. 2016;30(2):49–74.
87. Peltonen JM, Pyllkänen L, Jansén CT, Volanen I, Lehtinen T, Laihia JK, et al. Three randomised phase I/IIa trials of 5% cis-urocanic acid emulsion cream in healthy adult subjects and in patients with atopic dermatitis. *Acta Derm Venereol*. 2014;94(4):415–420.
88. Furue M. Regulation of filaggrin, loricrin, and involucrin by IL-4, IL-13, IL-17A, IL-22, AHR, and NRF2: pathogenic implications in atopic dermatitis. *Int J Mol Sci*. 2020;21(15):5382.
89. Brunner PM, Guttman-Yassky E, Leung DY. The immunology of atopic dermatitis and its reversibility with broad-spectrum and targeted therapies. *J Allergy Clin Immunol*. 2017;139(4s):S65–S76.
90. Immunomic Therapeutics and Astellas Pharma announce exclusive licensing agreement for LAMP-Vax platform to prevent and treat allergies. Immunomix. Accessed November 10, 2023. Available at: <https://www.immunomix.com/immunomic-therapeutics-and-astellas-pharma-announce-exclusive-licensing-agreement-for-lamp-vax-platform-to-prevent-and-treat-allergies/>.
91. Gonzalez-Visiedo M, Li X, Munoz-Melero M, Kulis MD, Daniell H, Markusic DM. Single-dose AAV vector gene immunotherapy to treat food allergy. *Mol Ther Methods Clin Dev*. 2022;26:309–322.
92. Rettman P, Blunt MD, Fulton RJ, Vallejo AF, Bastidas-Legarda LY, España-Serrano L, et al. Peptide: MHC-based DNA vaccination strategy to activate natural killer cells by targeting killer cell immunoglobulin-like receptors. *J Immunother Cancer*. 2021;9(5): e001912.
93. Papapostolou N, Xepapadaki P, Gregoriou S, Makris M. Atopic dermatitis and food allergy: A complex interplay what we know and what we would like to learn. *J Clin Med*. 2022;11(14):4232.
94. Mack MR, Kim BS. The itch-scratch cycle: a neuroimmune perspective. *Trends Immunol*. 2018;39(12):980–991.