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(54) **COMPOSITIONS AND METHODS RELATED TO PROTEIN A (SPA) VARIANTS**

(58) **Field of Classification Search**

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(57) **ABSTRACT**

The present invention concerns methods and compositions for treating or preventing a bacterial infection, particularly infection by a *Staphylococcus* bacterium. The invention provides methods and compositions for stimulating an immune response against the bacteria. In certain embodiments, the methods and compositions involve a non-toxicogenic Protein A (SpA) variant.

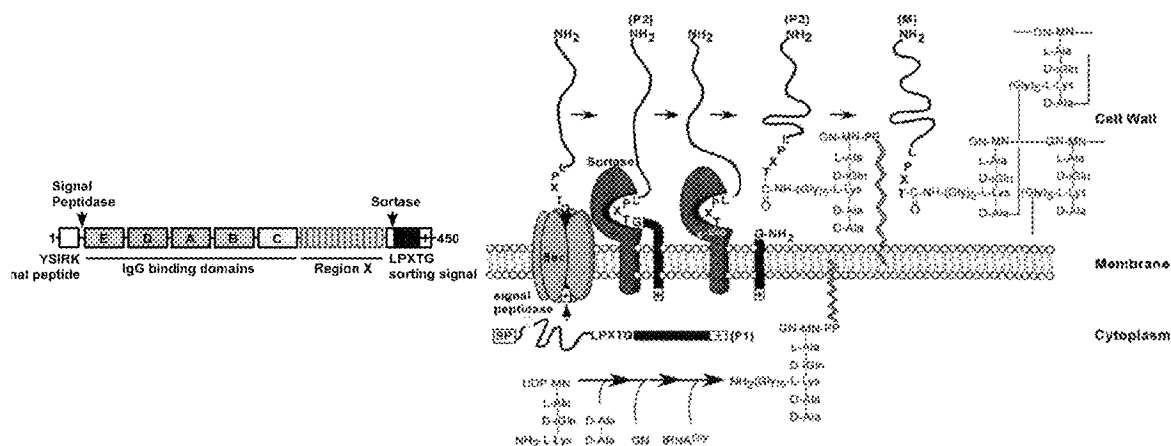
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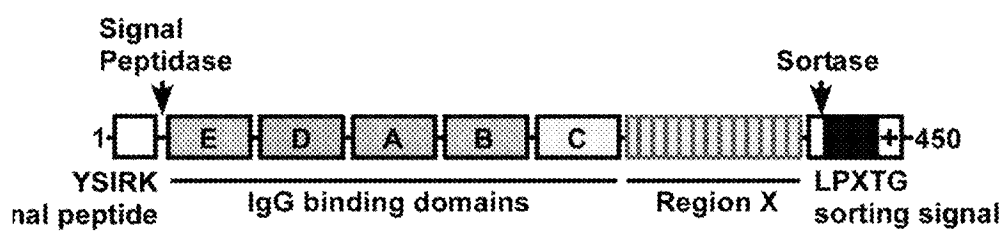


FIG. 1A

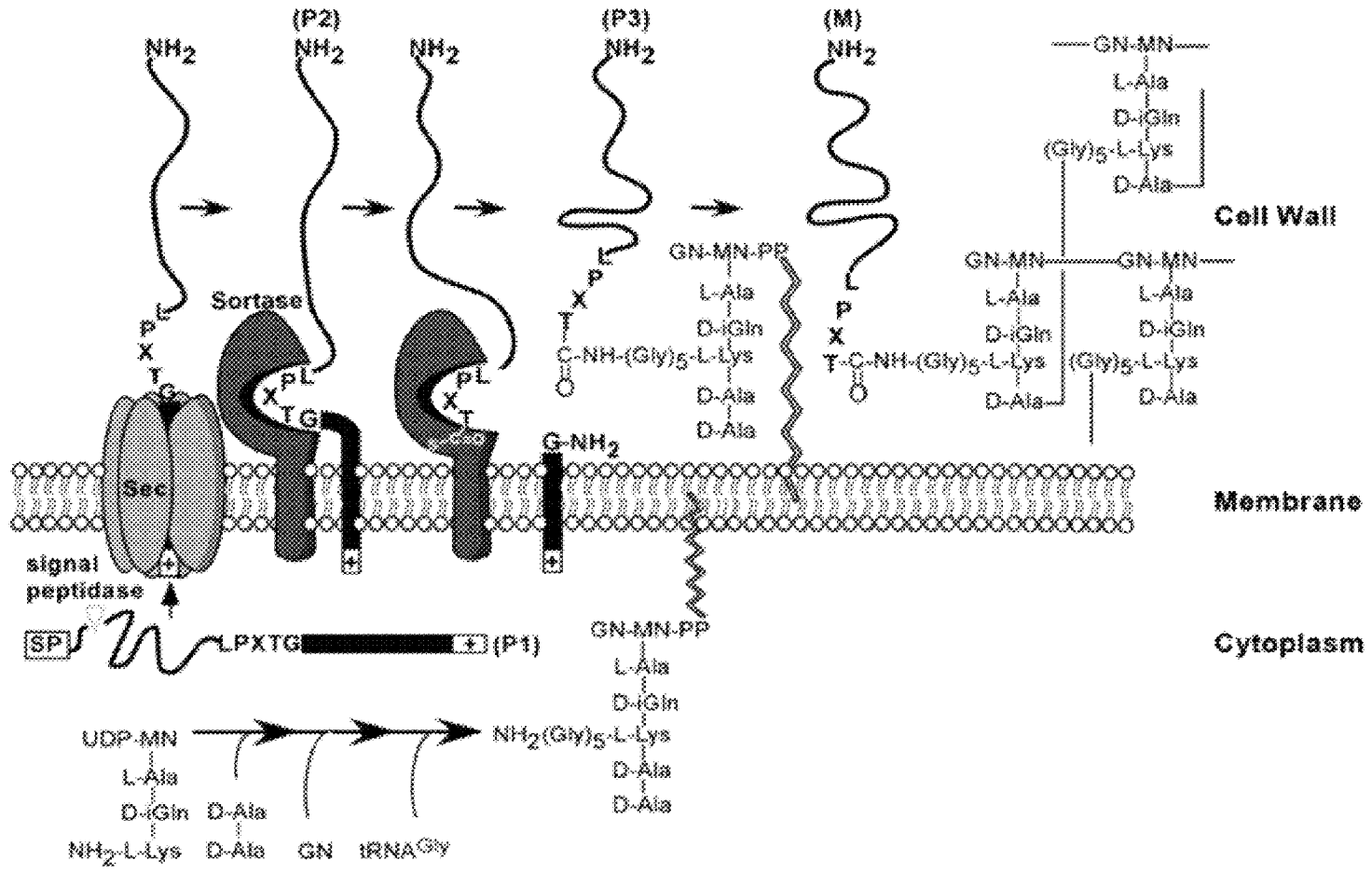


FIG. 1B

A

	5	10	15	20	25	30	35	40	45	50	55	Identity (%)				
SpA Domains	D::ADAQONNF	NKDOQ	SAFYE	ILNMP	NLNEA	RN	IQ	LK	PS	ST	VL	G	AKK	NESQ	APK	—
E::	-----QH	DEA..	N...Q	V.....	..AD					.A.			.Q.	.D.		75
A::	-----	..E..	N....E					.A.	L.	S		86
C::	-----K.	..E..	N....	..HL.	..T.E					.K.	I.	ADA.		77
B::	-----K.	..E..	N....	..HL.	..E					.A.	L.	ADA.		72

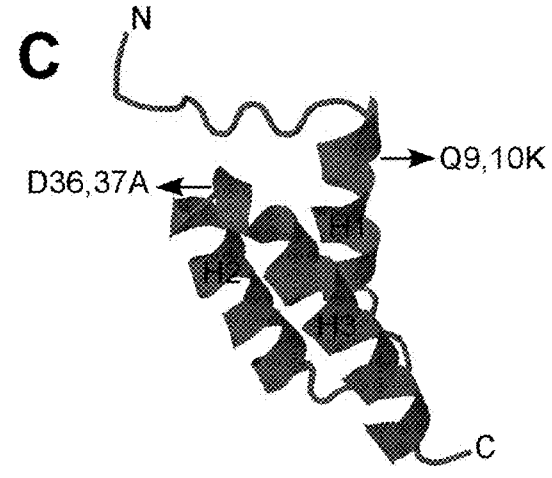
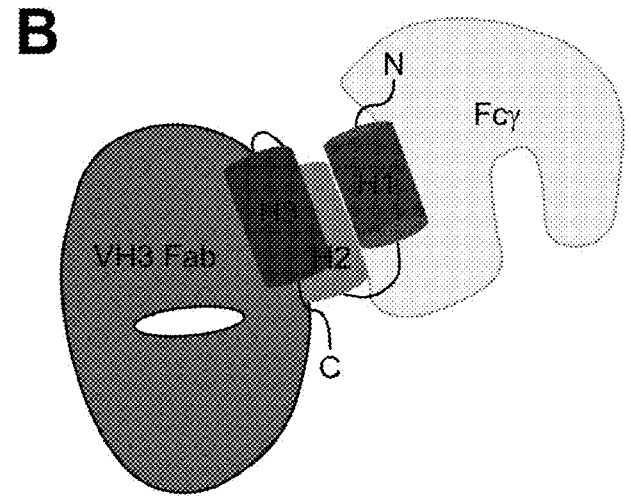


FIG. 2

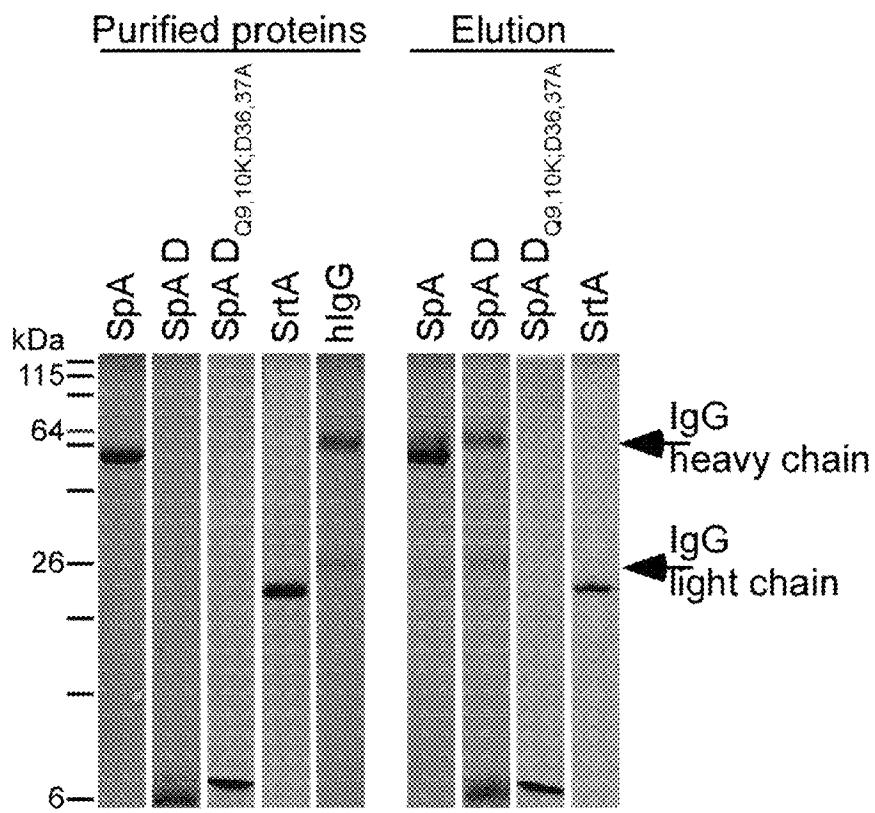


FIG. 3

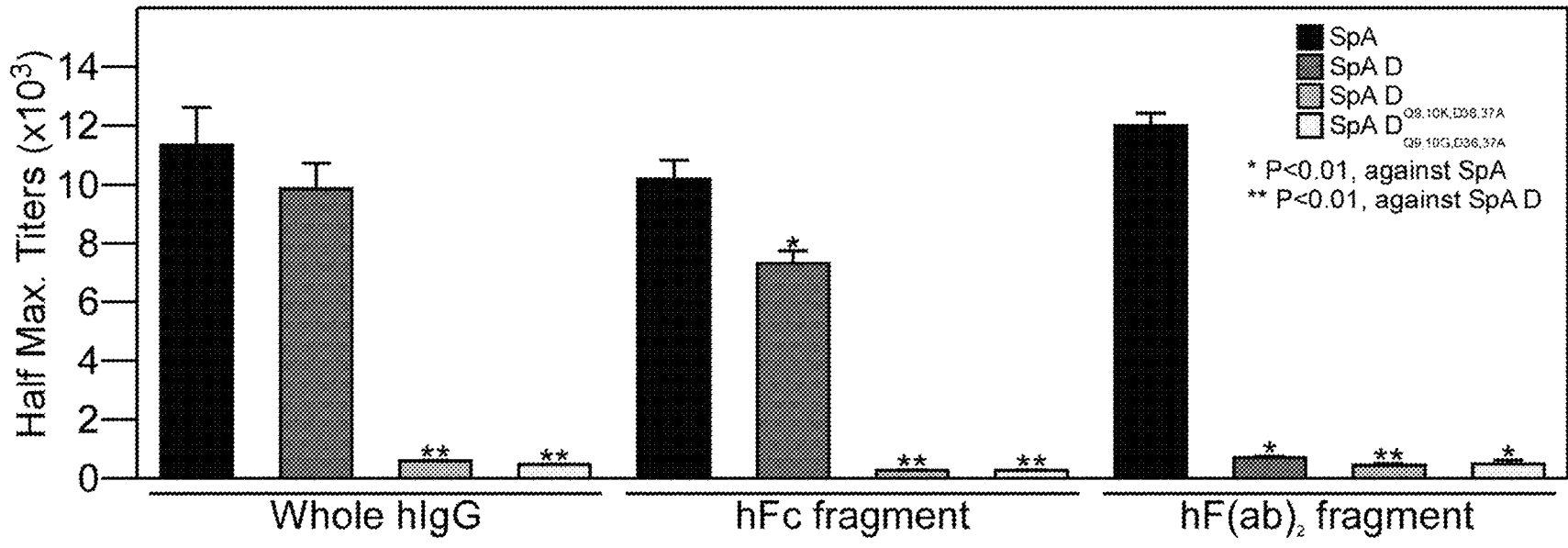


FIG. 4

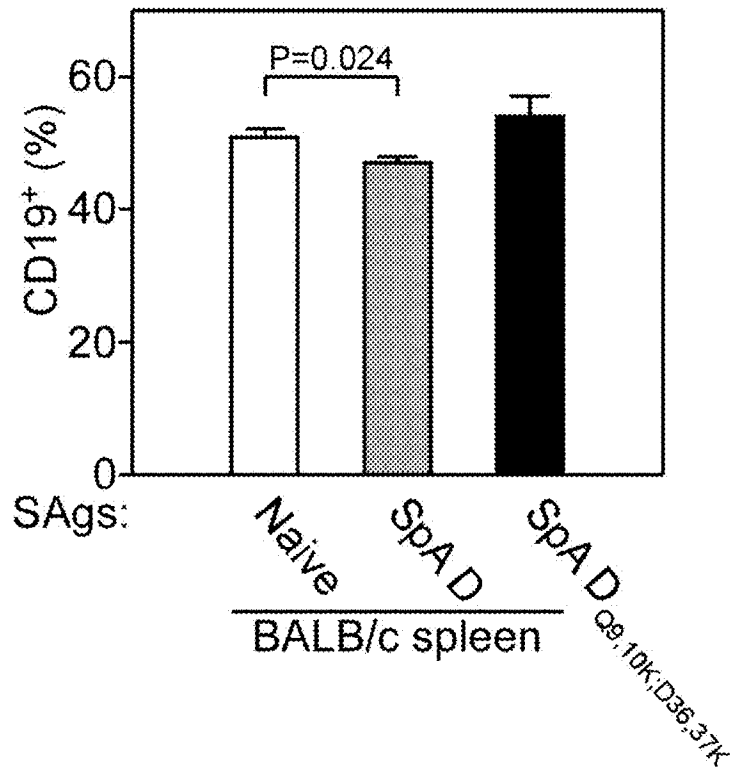


FIG. 5

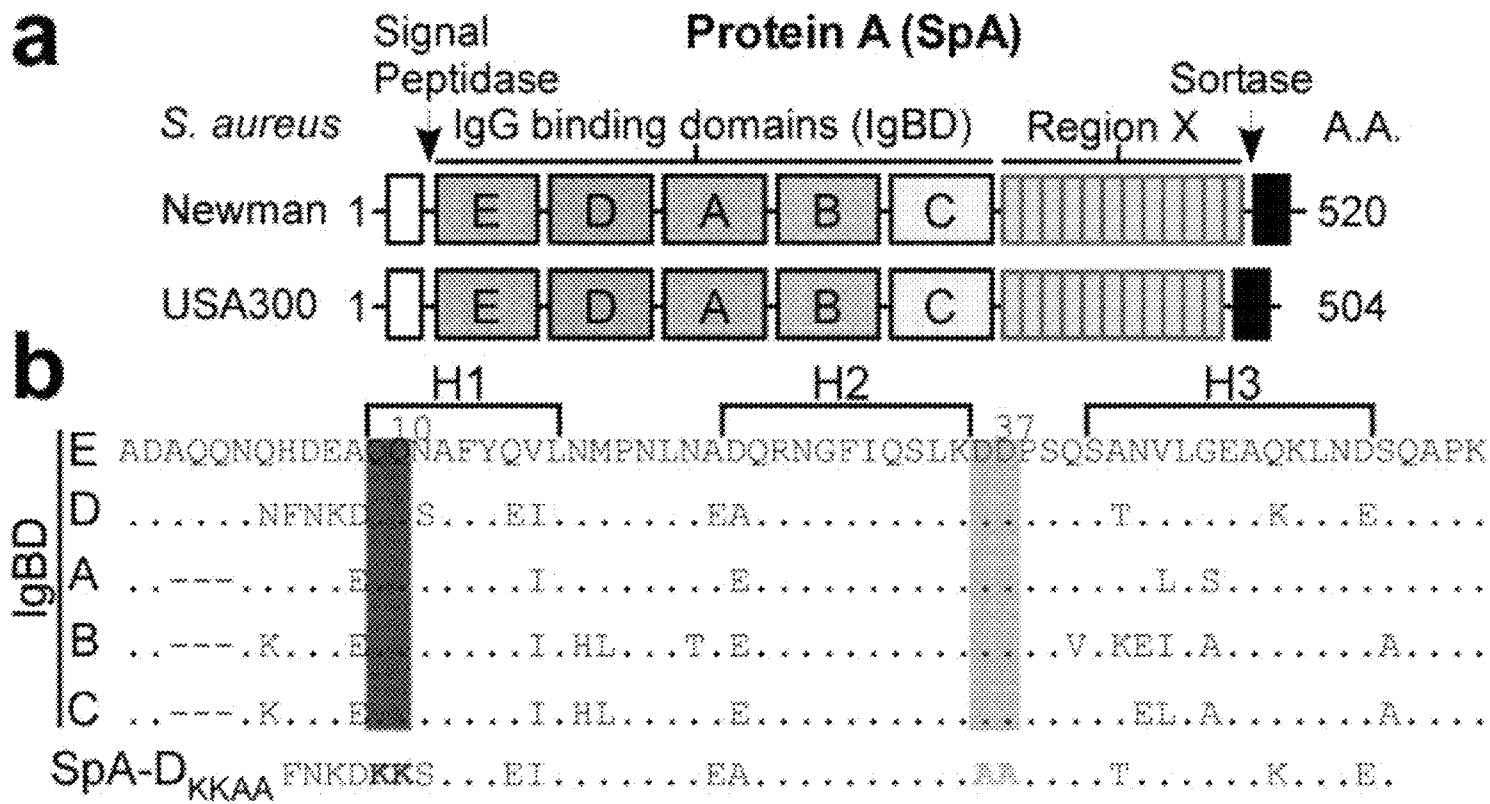


FIG. 6

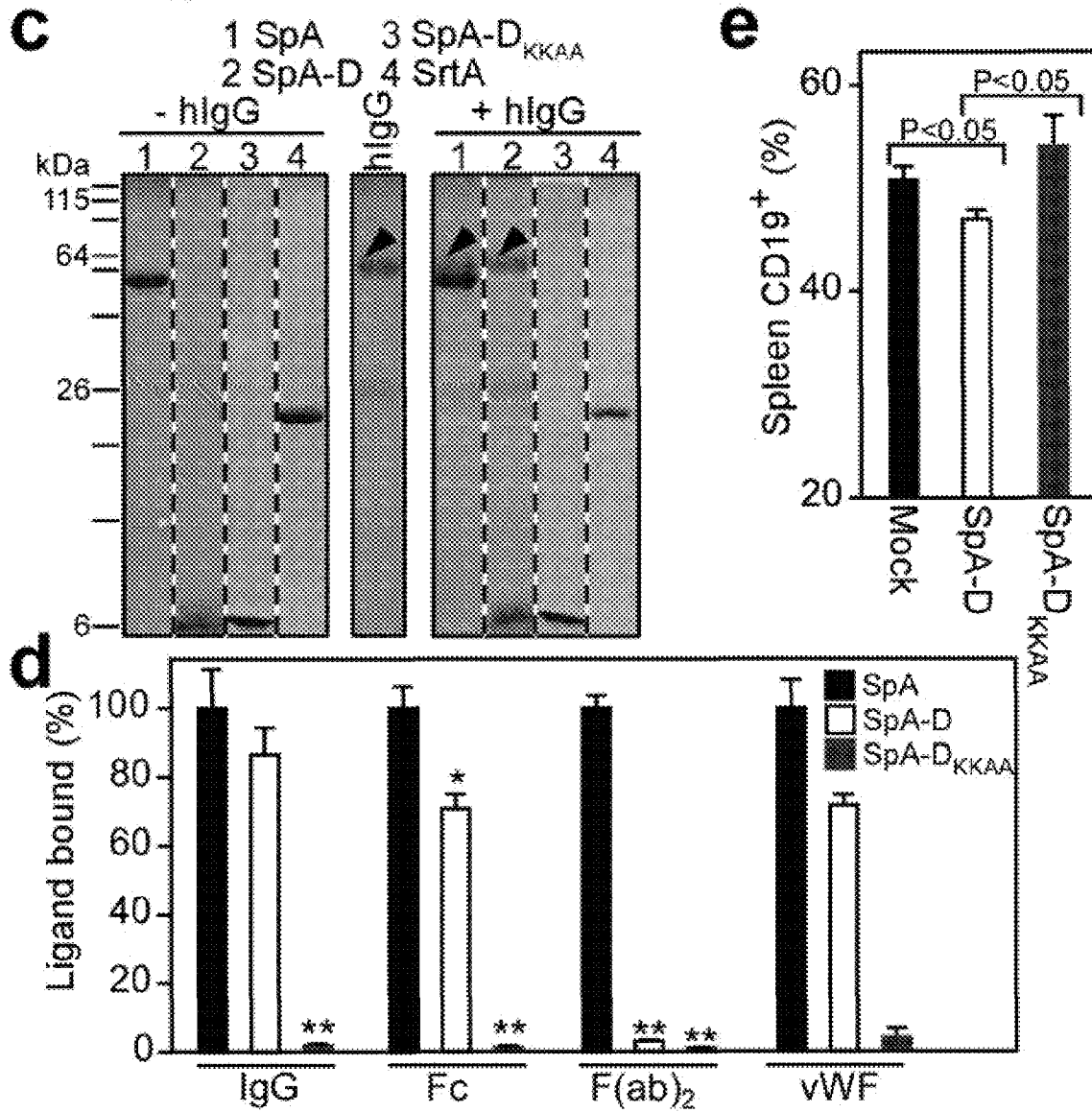


FIG. 6 (continued)

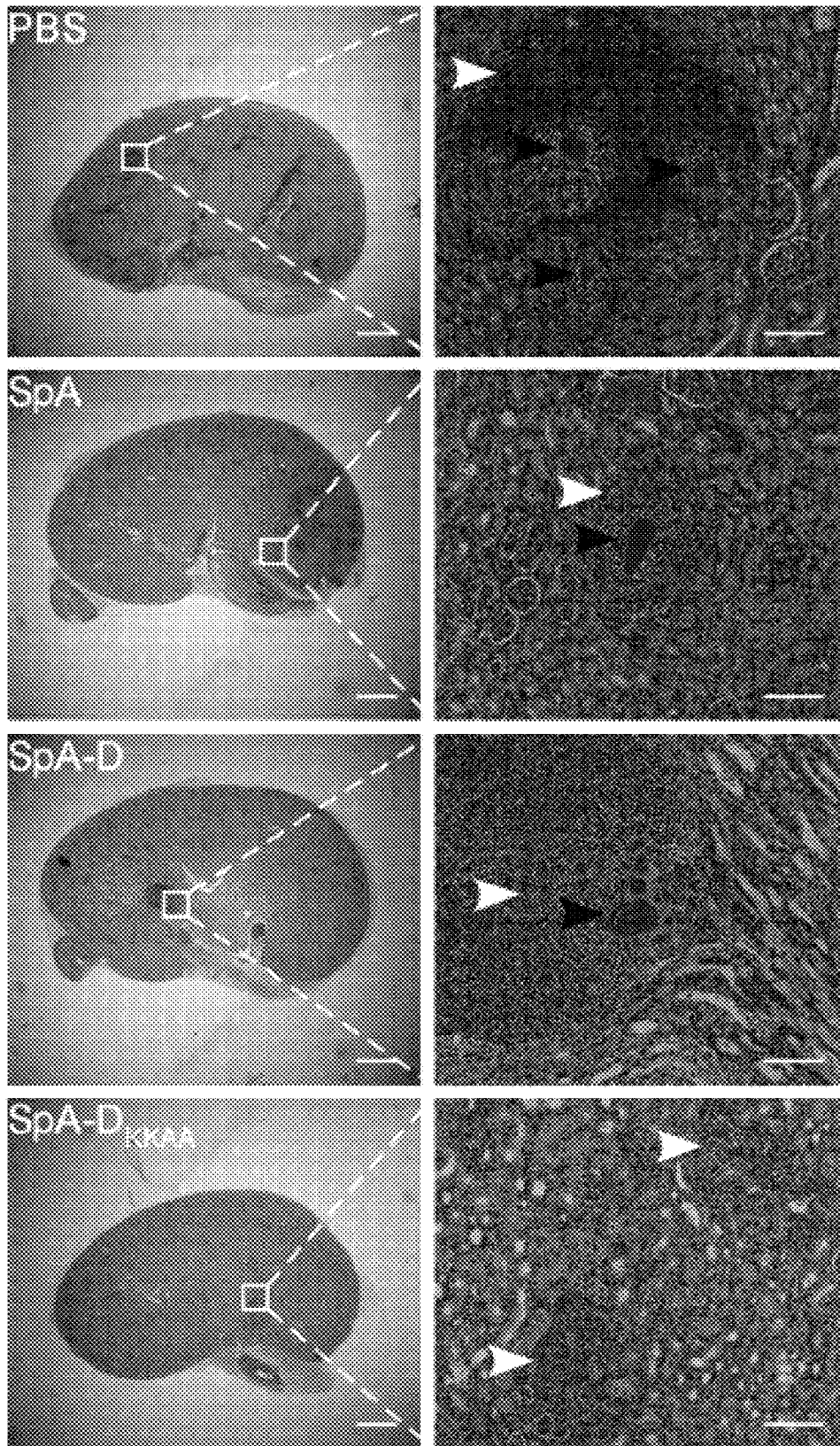


FIG. 7

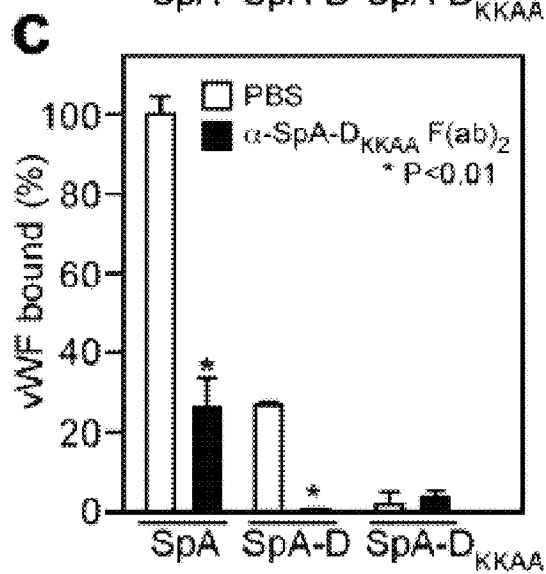
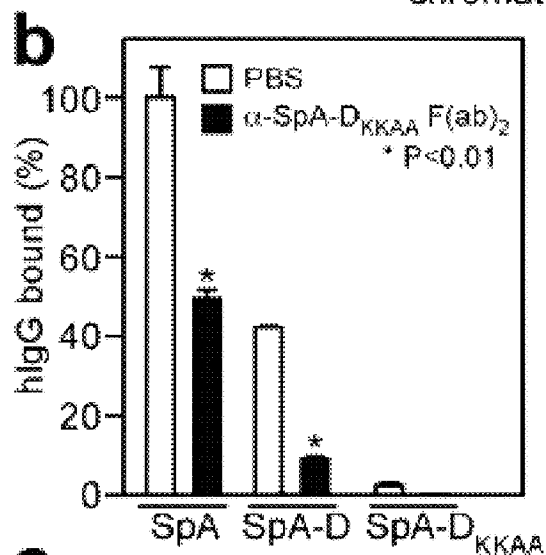
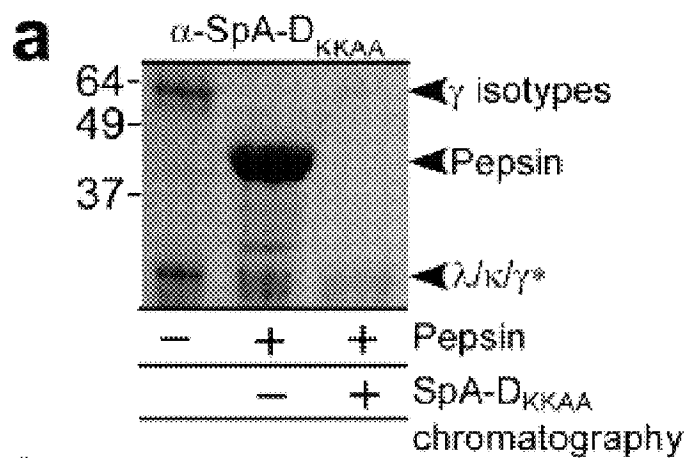


FIG. 8

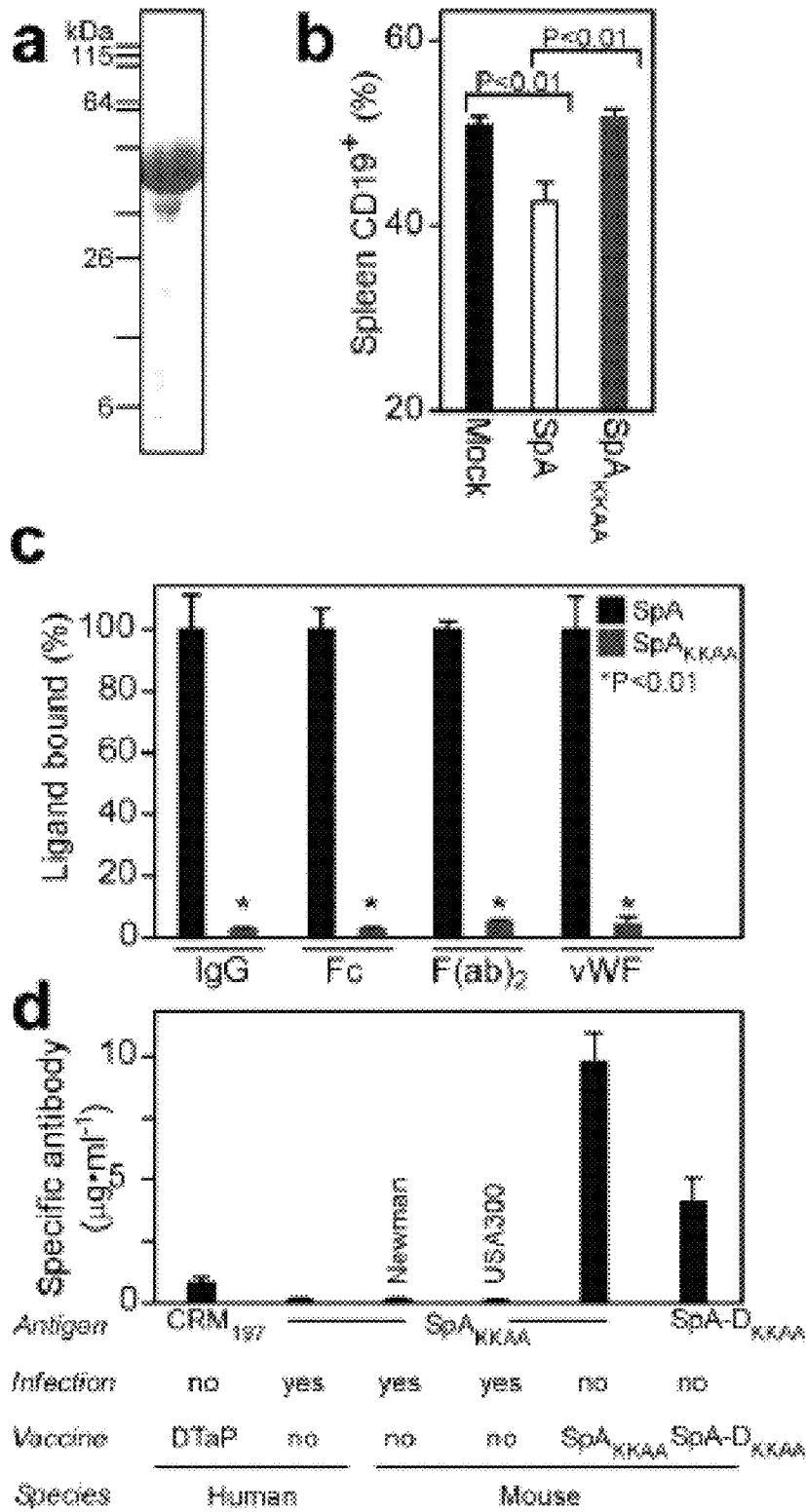


FIG. 9

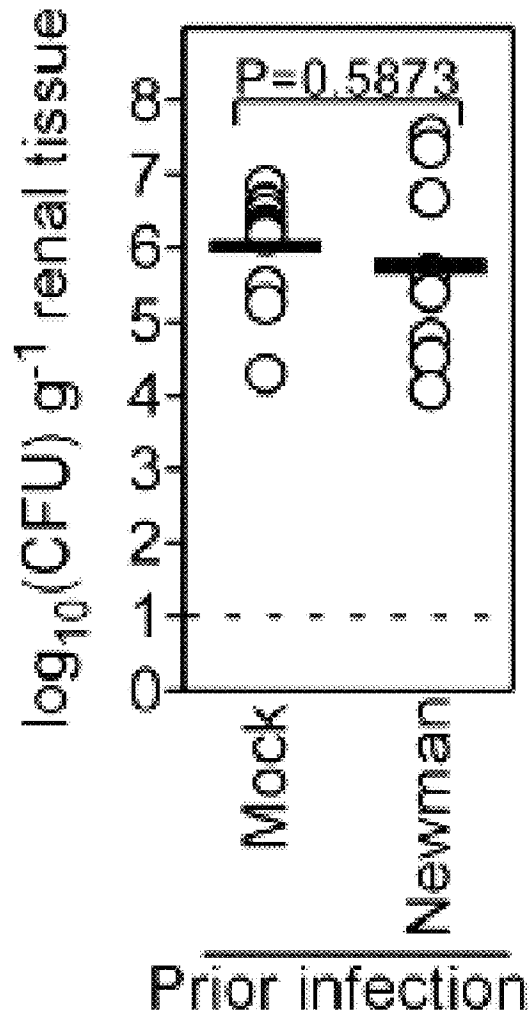


FIG. 10

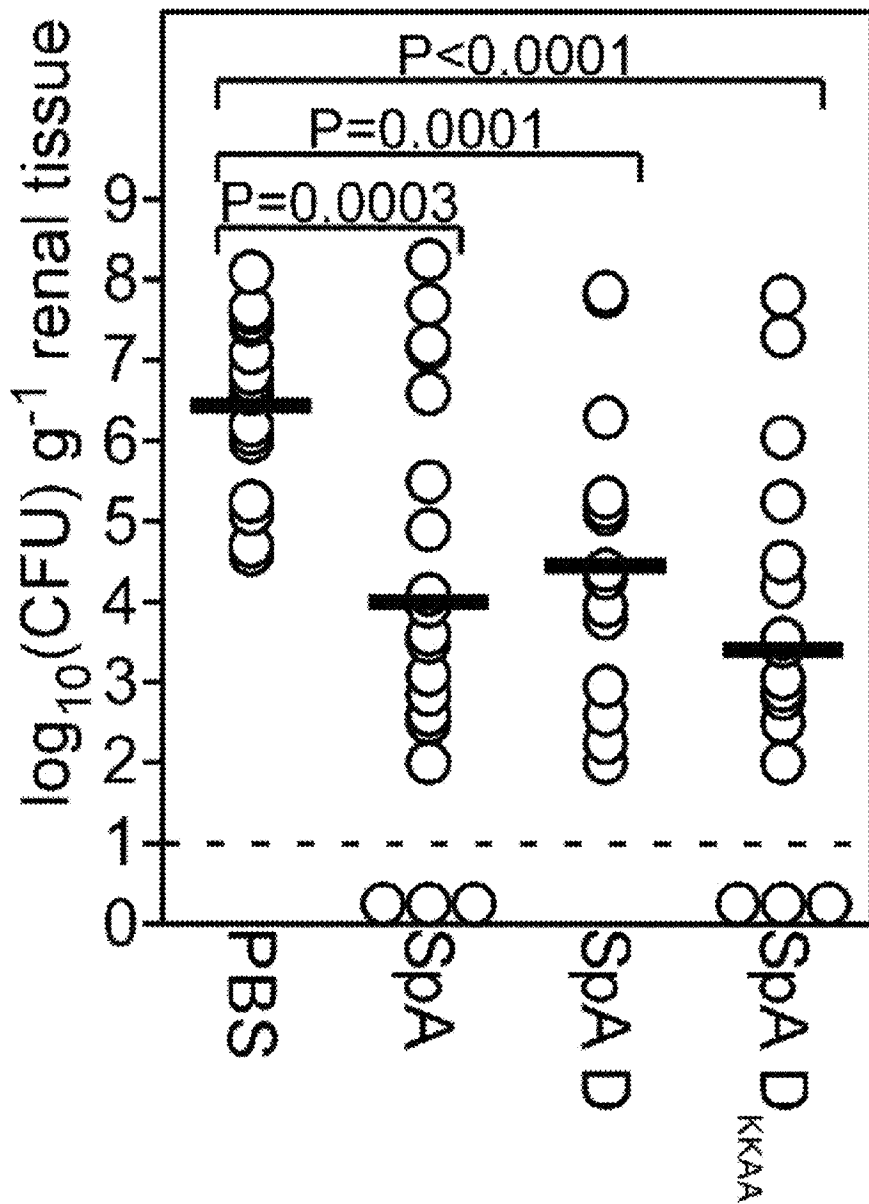


FIG. 11

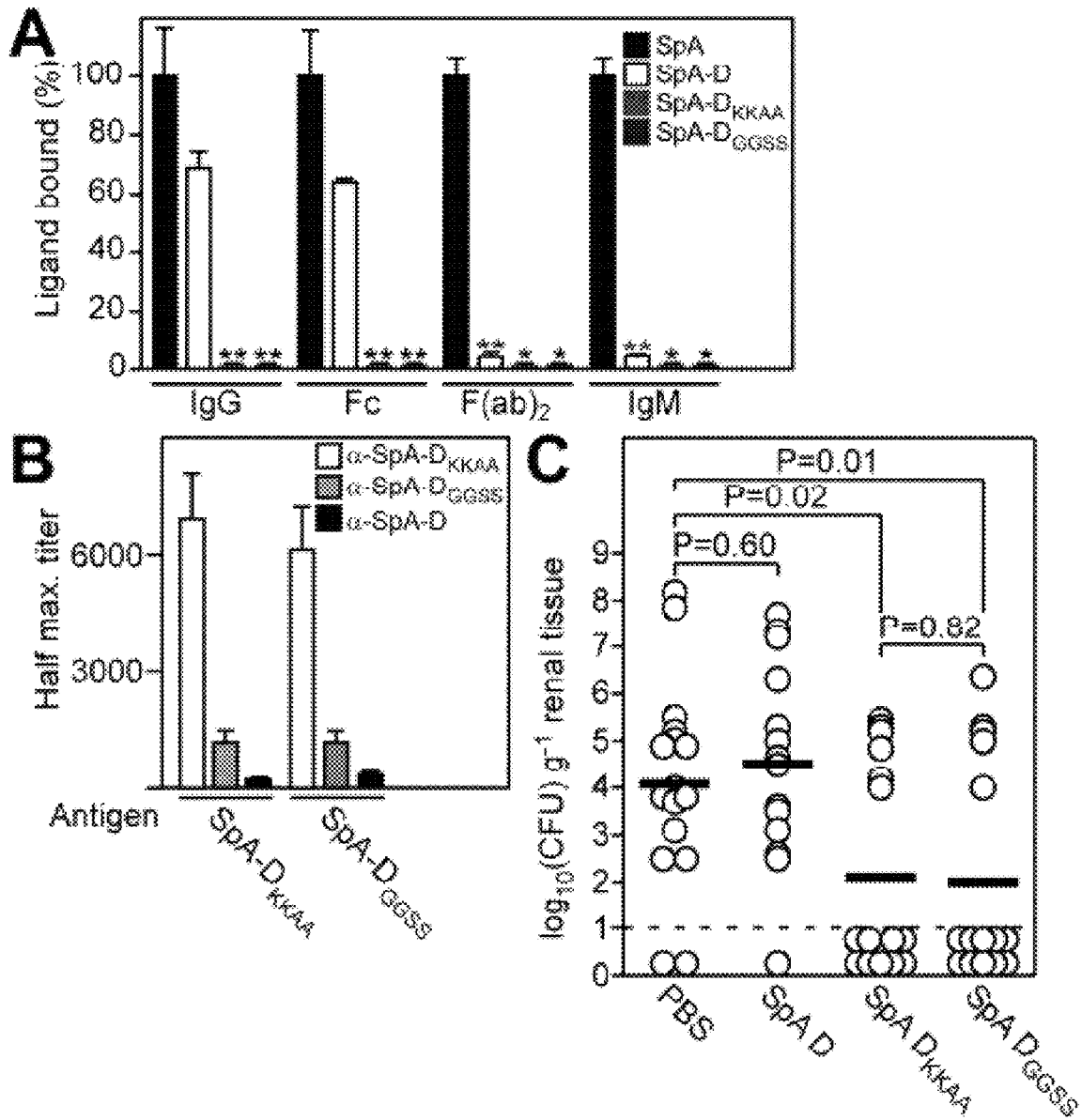


FIG. 12

COMPOSITIONS AND METHODS RELATED TO PROTEIN A (SPA) VARIANTS

This application is a continuation of U.S. patent application Ser. No. 16/661,155, filed Oct. 23, 2019, which is a continuation of U.S. patent application Ser. No. 15/702,037 filed Sep. 12, 2017, which is a continuation of U.S. patent application Ser. No. 15/060,861, filed Mar. 4, 2016, which is a continuation of U.S. patent application Ser. No. 14/466,514, filed Aug. 22, 2014, now U.S. Pat. No. 9,315,554, which is a continuation of U.S. patent application Ser. No. 13/807,598, filed Mar. 19, 2013, now U.S. Pat. No. 8,821,894, which is a national phase application under 35 U.S.C. § 371 of International Application No. PCT/US2011/042845, filed Jul. 1, 2011, which claims the benefit of priority to U.S. Provisional Patent Application Ser. No. 61/361,218 filed Jul. 2, 2010, and 61/370,725 filed Aug. 4, 2010. The entire contents of each of the above-referenced disclosures are specifically incorporated herein by reference without disclaimer.

This invention was made with government support under A1057153, A1052474, and GM007281 awarded by the National Institutes of Health. The government has certain rights in the invention.

BACKGROUND OF THE INVENTION

I. Field of the Invention

The present invention relates generally to the fields of immunology, microbiology, and pathology. More particularly, it concerns methods and compositions involving bacterial Protein A variants, which can be used to invoke an immune response against the bacteria.

II. Background

The number of both community acquired and hospital acquired infections have increased over recent years with the increased use of intravascular devices. Hospital acquired (nosocomial) infections are a major cause of morbidity and mortality, more particularly in the United States, where it affects more than 2 million patients annually. The most frequent infections are urinary tract infections (33% of the infections), followed by pneumonia (15.5%), surgical site infections (14.8%) and primary bloodstream infections (13%) (Emorl and Gaynes, 1993).

The major nosocomial pathogens include *Staphylococcus aureus*, coagulase-negative Staphylococci (mostly *Staphylococcus epidermidis*), *enterococcus* spp., *Escherichia coli* and *Pseudomonas aeruginosa*. Although these pathogens cause approximately the same number of infections, the severity of the disorders they can produce combined with the frequency of antibiotic resistant isolates balance this ranking towards *S. aureus* and *S. epidermidis* as being the most significant nosocomial pathogens.

Staphylococci can cause a wide variety of diseases in humans and other animals through either toxin production or invasion. Staphylococcal toxins are also a common cause of food poisoning, as the bacteria can grow in improperly-stored food.

Staphylococcus epidermidis is a normal skin commensal which is also an important opportunistic pathogen responsible for infections of impaired medical devices and infections at sites of surgery. Medical devices infected by *S. epidermidis* include cardiac pacemakers, cerebrospinal fluid

shunts, continuous ambulatory peritoneal dialysis catheters, orthopedic devices and prosthetic heart valves.

Staphylococcus aureus is the most common cause of nosocomial infections with a significant morbidity and mortality. It is the cause of some cases of osteomyelitis, endocarditis, septic arthritis, pneumonia, abscesses, and toxic shock syndrome. *S. aureus* can survive on dry surfaces, increasing the chance of transmission. Any *S. aureus* infection can cause the staphylococcal scalded skin syndrome, a cutaneous reaction to exotoxin absorbed into the bloodstream. It can also cause a type of septicemia called pyaemia that can be life-threatening. Problematically, Methicillin-resistant *Staphylococcus aureus* (MRSA) has become a major cause of hospital-acquired infections.

S. aureus and *S. epidermidis* infections are typically treated with antibiotics, with penicillin being the drug of choice, whereas vancomycin is used for methicillin resistant isolates. The percentage of staphylococcal strains exhibiting wide-spectrum resistance to antibiotics has become increasingly prevalent, posing a threat for effective antimicrobial therapy. In addition, the recent emergence of vancomycin resistant *S. aureus* strain has aroused fear that MRSA strains are emerging and spreading for which no effective therapy is available.

An alternative to antibiotic treatment for staphylococcal infections is under investigation that uses antibodies directed against staphylococcal antigens. This therapy involves administration of polyclonal antisera (WO00/15238, WO00/12132) or treatment with monoclonal antibodies against lipoteichoic acid (WO98/57994).

An alternative approach would be the use of active vaccination to generate an immune response against staphylococci. The *S. aureus* genome has been sequenced and many of the coding sequences have been identified (WO02/094868, EP0786519), which can lead to the identification of potential antigens. The same is true for *S. epidermidis* (WO01/34809). As a refinement of this approach, others have identified proteins that are recognized by hyperimmune sera from patients who have suffered staphylococcal infection (WO01/98499, WO02/059148).

S. aureus secretes a plethora of virulence factors into the extracellular milieu (Archer, 1998; Dinges et al., 2000; Foster, 2005; Shaw et al., 2004; Sibbald et al., 2006). Like most secreted proteins, these virulence factors are translocated by the Sec machinery across the plasma membrane. Proteins secreted by the Sec machinery bear an N-terminal leader peptide that is removed by leader peptidase once the pre-protein is engaged in the Sec translocon (Dalbey and Wickner, 1985; van Wely et al., 2001). Recent genome analysis suggests that Actinobacteria and members of the Firmicutes encode an additional secretion system that recognizes a subset of proteins in a Sec-independent manner (Pallen, 2002). ESAT-6 (early secreted antigen target 6 kDa) and CFP-10 (culture filtrate antigen 10 kDa) of *Mycobacterium tuberculosis* represent the first substrates of this novel secretion system termed ESX-1 or Ssm in *M. tuberculosis* (Andersen et al., 1995; Hsu et al., 2003; Pym et al., 2003; Stanley et al., 2003). In *S. aureus*, two ESAT-6 like factors designated EsxA and EsxB are secreted by the Ess pathway (ESAT-6 secretion system) (Burts et al., 2005).

The first generation of vaccines targeted against *S. aureus* or against the exoproteins it produces have met with limited success (Lee, 1996). There remains a need to develop effective vaccines against staphylococcal infections. Additional compositions for treating staphylococcal infections are also needed.

SUMMARY OF THE INVENTION

Protein A (SpA)(SEQ ID NO:33), a cell wall anchored surface protein of *Staphylococcus aureus*, provides for bacterial evasion from innate and adaptive immune responses. Protein A binds immunoglobulins at their Fc portion, interacts with the VH3 domain of B cell receptors inappropriately stimulating B cell proliferation and apoptosis, binds to von Willebrand factor A1 domains to activate intracellular clotting, and also binds to the TNF Receptor-1 to contribute to the pathogenesis of staphylococcal pneumonia. Due to the fact that Protein A captures immunoglobulin and displays toxic attributes, the possibility that this surface molecule may function as a vaccine in humans has not been rigorously pursued. Here the inventors demonstrate that Protein A variants no longer able to bind to immunoglobulins, which are thereby removed of their toxigenic potential, i.e., are non-toxicogenic, stimulate humoral immune responses that protect against staphylococcal disease.

In certain embodiments the SpA variant is a full length SpA variant comprising a variant A, B, C, D, and/or E domain. In certain aspects, the SpA variant comprises or consists of the amino acid sequence that is 80, 90, 95, 98, 99, or 100% identical to the amino acid sequence of SEQ ID NO:34. In other embodiments the SpA variant comprises a segment of SpA. The SpA segment can comprise at least or at most 1, 2, 3, 4, 5 or more IgG binding domains. The IgG domains can be at least or at most 1, 2, 3, 4, 5 or more variant A, B, C, D, or E domains. In certain aspects the SpA variant comprises at least or at most 1, 2, 3, 4, 5, or more variant A domains. In a further aspect the SpA variant comprises at least or at most 1, 2, 3, 4, 5, or more variant B domains. In still a further aspect the SpA variant comprises at least or at most 1, 2, 3, 4, 5, or more variant C domains. In yet a further aspect the SpA variant comprises at least or at most 1, 2, 3, 4, 5, or more variant D domains. In certain aspects the SpA variant comprises at least or at most 1, 2, 3, 4, 5, or more variant E domains. In a further aspect the SpA variant comprises a combination of A, B, C, D, and E domains in various combinations and permutations. The combinations can include all or part of a SpA signal peptide segment, a SpA region X segment, and/or a SpA sorting signal segment. In other aspects the SpA variant does not include a SpA signal peptide segment, a SpA region X segment, and/or a SpA sorting signal segment. In certain aspects a variant A domain comprises a substitution at position(s) 7, 8, 34, and/or 35 of SEQ ID NO:4. In another aspect a variant B domain comprises a substitution at position(s) 7, 8, 34, and/or 35 of SEQ ID NO:6. In still another aspect a variant C domain comprises a substitution at position(s) 7, 8, 34, and/or 35 of SEQ ID NO:5. In certain aspects a variant D domain comprises a substitution at position(s) 9, 10, 36, and/or 37 of SEQ ID NO:2. In a further aspect a variant E domain comprises a substitution at position(s) 6, 7, 33, and/or 34 of SEQ ID NO:3.

In certain aspects, an SpA domain D variant or its equivalent can comprise a mutation at position 9 and 36; 9 and 37; 9 and 10; 36 and 37; 10 and 36; 10 and 37; 9, 36, and 37; 10, 36, and 37; 9, 10 and 36; or 9, 10 and 37 of SEQ ID NO:2. In a further aspect, analogous mutations can be included in one or more of domains A, B, C, or E.

In further aspects, the amino acid glutamine (Q) at position 9 of SEQ ID NO:2 (or its analogous amino acid in other SpA domains) can be replaced with an alanine (A), an asparagine (N), an aspartic acid (D), a cysteine (C), a glutamic acid (E), a phenylalanine (F), a glycine (G), a histidine (H), an isoleucine (I), a lysine (K), a leucine (L), a

methionine (M), a proline (P), a serine (S), a threonine (T), a valine (V), a tryptophane (W), or a tyrosine (Y). In some aspects the glutamine at position 9 can be substituted with an arginine (R). In a further aspect, the glutamine at position 9 of SEQ ID NO:2, or its equivalent, can be substituted with a lysine or a glycine. Any 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more of the substitutions can be explicitly excluded.

In another aspect, the amino acid glutamine (Q) at position 10 of SEQ ID NO:2 (or its analogous amino acid in other SpA domains) can be replaced with an alanine (A), an asparagine (N), an aspartic acid (D), a cysteine (C), a glutamic acid (E), a phenylalanine (F), a glycine (G), a histidine (H), an isoleucine (I), a lysine (K), a leucine (L), a methionine (M), a proline (P), a serine (S), a threonine (T), a valine (V), a tryptophane (W), or a tyrosine (Y). In some aspects the glutamine at position 10 can be substituted with an arginine (R). In a further aspect, the glutamine at position 10 of SEQ ID NO:2, or its equivalent, can be substituted with a lysine or a glycine. Any 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more of the substitutions can be explicitly excluded.

In certain aspects, the aspartic acid (D) at position 36 of SEQ ID NO:2 (or its analogous amino acid in other SpA domains) can be replaced with an alanine (A), an asparagine (N), an arginine (R), a cysteine (C), a phenylalanine (F), a glycine (G), a histidine (H), an isoleucine (I), a lysine (K), a leucine (L), a methionine (M), a proline (P), a glutamine (Q), a serine (S), a threonine (T), a valine (V), a tryptophane (W), or a tyrosine (Y). In some aspects the aspartic acid at position 36 can be substituted with a glutamic acid (E). In certain aspects, an aspartic acid at position 36 of SEQ ID NO:2, or its equivalent, can be substituted with an alanine or a serine. Any 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more of the substitutions can be explicitly excluded.

In another aspect, the aspartic acid (D) at position 37 of SEQ ID NO:2 (or its analogous amino acid in other SpA domains) can be replaced with an alanine (A), an asparagine (N), an arginine (R), a cysteine (C), a phenylalanine (F), a glycine (G), a histidine (H), an isoleucine (I), a lysine (K), a leucine (L), a methionine (M), a proline (P), a glutamine (Q), a serine (S), a threonine (T), a valine (V), a tryptophane (W), or a tyrosine (Y). In some aspects the aspartic acid at position 37 can be substituted with a glutamic acid (E). In certain aspects, an aspartic acid at position 37 of SEQ ID NO:2, or its equivalent, can be substituted with an alanine or a serine. Any 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more of the substitutions can be explicitly excluded.

In a particular embodiment the amino acid at position 9 of SEQ ID NO:2 (or an analogous amino acid in another SpA domain) is replaced by an alanine (A), a glycine (G), an isoleucine (I), a leucine (L), a proline (P), a serine (S), or a valine (V). In certain aspects the amino acid at position 9 of SEQ ID NO:2 is replaced by a glycine. In a further aspect the amino acid at position 9 of SEQ ID NO:2 is replaced by a lysine.

In a particular embodiment the amino acid at position 10 of SEQ ID NO:2 (or an analogous amino acid in another SpA domain) is replaced by an alanine (A), a glycine (G), an isoleucine (I), a leucine (L), a proline (P), a serine (S), or a valine (V). In certain aspects the amino acid at position 10 of SEQ ID NO:2 is replaced by a glycine. In a further aspect the amino acid at position 10 of SEQ ID NO:2 is replaced by a lysine.

In a particular embodiment the amino acid at position 36 of SEQ ID NO:2 (or an analogous amino acid in another SpA domain) is replaced by an alanine (A), a glycine (G), an isoleucine (I), a leucine (L), a proline (P), a serine (S), or a valine (V). In certain aspects the amino acid at position 36

of SEQ ID NO:2 is replaced by a serine. In a further aspect the amino acid at position 36 of SEQ ID NO:2 is replaced by an alanine.

In a particular embodiment the amino acid at position 37 of SEQ ID NO:2 (or an analogous amino acid in another SpA domain) is replaced by an alanine (A), a glycine (G), an isoleucine (I), a leucine (L), a proline (P), a serine (S), or a valine (V). In certain aspects the amino acid at position 37 of SEQ ID NO:2 is replaced by a serine. In a further aspect the amino acid at position 37 of SEQ ID NO:2 is replaced by an alanine.

In certain aspects the SpA variant includes a substitution of (a) one or more amino acid substitution in an IgG Fc binding sub-domain of SpA domain A, B, C, D, and/or E that disrupts or decreases binding to IgG Fc, and (b) one or more amino acid substitution in a V_H3 binding sub-domain of SpA domain A, B, C, D, and/or E that disrupts or decreases binding to V_H3 . In still further aspects the amino acid sequence of a SpA variant comprises an amino acid sequence that is at least 50%, 60%, 70%, 80%, 90%, 95%, or 100% identical, including all values and ranges there between, to the amino acid sequence of SEQ ID NOs:2-6.

In a further aspect the SpA variant includes (a) one or more amino acid substitution in an IgG Fc binding sub-domain of SpA domain D, or at a corresponding amino acid position in other IgG domains, that disrupts or decreases binding to IgG Fc, and (b) one or more amino acid substitution in a V_H3 binding sub-domain of SpA domain D, or at a corresponding amino acid position in other IgG domains, that disrupts or decreases binding to V_H3 . In certain aspects amino acid residue F5, Q9, Q10, S11, F13, Y14, L17, N28, I31, and/or K35 (SEQ ID NO:2, QQNNFNKDDQSSAFYEILNMPNLNEAQRNG-FIQLSKDDPSQSTNVLGEAKKLNES) of the IgG Fc binding sub-domain of domain D are modified or substituted. In certain aspects amino acid residue Q26, G29, F30, S33, D36, D37, Q40, N43, and/or E47 (SEQ ID NO:2) of the V_H3 binding sub-domain of domain D are modified or substituted such that binding to Fc or V_H3 is attenuated. In further aspects corresponding modifications or substitutions can be engineered in corresponding positions of the domain A, B, C, and/or E. Corresponding positions are defined by alignment of the domain D amino acid sequence with one or more of the amino acid sequences from other IgG binding domains of SpA, for example see FIG. 2A. In certain aspects the amino acid substitution can be any of the other 20 amino acids. In a further aspect conservative amino acid substitutions can be specifically excluded from possible amino acid substitutions. In other aspects only non-conservative substitutions are included. In any event, any substitution or combination of substitutions that reduces the binding of the domain such that SpA toxicity is significantly reduced is contemplated. The significance of the reduction in binding refers to a variant that produces minimal to no toxicity when introduced into a subject and can be assessed using in vitro methods described herein.

In certain embodiments, a variant SpA comprises at least one or at most 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, or more variant SpA domain D peptides. In certain aspects 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, or 19 or more amino acid residues of the variant SpA are substituted or modified—including but not limited to amino acids F5, Q9, Q10, S11, F13, Y14, L17, N28, I31, and/or K35 (SEQ ID NO:2) of the IgG Fc binding sub-domain of domain D and amino acid residue Q26, G29, F30, S33, D36, D37, Q40, N43, and/or E47 (SEQ ID NO:2) of the V_H3 binding sub-domain of domain D. In one aspect of the invention glutamine residues

at position 9 and/or 10 of SEQ ID NO:2 (or corresponding positions in other domains) are mutated. In another aspect, aspartic acid residues 36 and/or 37 of SEQ ID NO:2 (or corresponding positions in other domains) are mutated. In a further aspect, glutamine 9 and 10, and aspartic acid residues 36 and 37 are mutated. Purified non-toxicogenic SpA or SpA-D mutants/variants described herein are no longer able to significantly bind (i.e., demonstrate attenuated or disrupted binding affinity) Fcγ or F(ab)₂ V_H3 and also do not stimulate B cell apoptosis. These non-toxicogenic Protein A variants can be used as subunit vaccines and raise humoral immune responses and confer protective immunity against *S. aureus* challenge. Compared to wild-type full-length Protein A or the wild-type SpA-domain D, immunization with SpA-D variants resulted in an increase in Protein A specific antibody. Using a mouse model of staphylococcal challenge and abscess formation, it was observed that immunization with the non-toxicogenic Protein A variants generated significant protection from staphylococcal infection and abscess formation. As virtually all *S. aureus* strains express Protein A, immunization of humans with the non-toxicogenic Protein A variants can neutralize this virulence factor and thereby establish protective immunity. In certain aspects the protective immunity protects or ameliorates infection by drug resistant strains of *Staphylococcus*, such as USA300 and other MRSA strains.

Embodiments include the use of Protein A variants in methods and compositions for the treatment of bacterial and/or staphylococcal infection. This application also provides an immunogenic composition comprising a Protein A variant or immunogenic fragment thereof. In certain aspects, the immunogenic fragment is a Protein A domain D segment. Furthermore, the present invention provides methods and compositions that can be used to treat (e.g., limiting staphylococcal abscess formation and/or persistence in a subject) or prevent bacterial infection. In some cases, methods for stimulating an immune response involve administering to the subject an effective amount of a composition including or encoding all or part of a Protein A variant polypeptide or antigen, and in certain aspects other bacterial proteins. Other bacterial proteins include, but are not limited to (i) a secreted virulence factor, and/or a cell surface protein or peptide, or (ii) a recombinant nucleic acid molecule encoding a secreted virulence factor, and/or a cell surface protein or peptide.

In other aspects, the subject can be administered all or part of a Protein A variant, such as a variant Protein A domain D segment. The polypeptide of the invention can be formulated in a pharmaceutically acceptable composition. The composition can further comprise one or more of at least or at most 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, or 19 additional staphylococcal antigen or immunogenic fragment thereof (e.g., Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla (e.g., H35 mutants), IsdC, SasF, vWbp, or vWh). Additional staphylococcal antigens that can be used in combination with a Protein A variant include, but are not limited to 52 kDa vitronectin binding protein (WO 01/60852), Aaa (GenBank CAC80837), Aap (GenBank accession AJ249487), Ant (GenBank accession NP_372518), autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg2+

TABLE 1-continued

SpA and staphylococcal antigen combinations.

ClfA	+	+	+	+	+	+	+	+	+
ClfB		+	+	+	+	+	+	+	+
Coa			+	+	+	+	+	+	+
Hla				+	+	+	+	+	+
Hla _{H35A}					+	+	+	+	+
IsdC						+	+	+	+
SasF							+	+	+
vWbp								+	+
vWh									+
ClfB		+	+	+	+	+	+	+	+
Coa			+	+	+	+	+	+	+
Hla				+	+	+	+	+	+
Hla _{H35A}					+	+	+	+	+
IsdC						+	+	+	+
SasF							+	+	+
vWbp								+	+
vWh									+
Coa		+	+	+	+	+	+	+	+
Hla			+	+	+	+	+	+	+
Hla _{H35A}				+	+	+	+	+	+
IsdC					+	+	+	+	+
SasF							+	+	+
vWbp								+	+
vWh									+
Hla _{H35A}					+	+	+	+	+
IsdC						+	+	+	+
SasF							+	+	+
vWbp								+	+
vWh									+
IsdC						+	+	+	+
SasF							+	+	+
vWbp								+	+
vWh									+
SasF							+	+	+
vWbp								+	+
vWh									+
vWbp								+	+
vWh									+
vWh									+

In still further aspects, the isolated Protein A variant is multimerized, e.g., dimerized or a linear fusion of two or more polypeptides or peptide segments. In certain aspects of the invention, a composition comprises multimers or concatamers of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 or more isolated cell surface proteins or segments thereof. Concatamers are linear polypeptides having one or more repeating peptide units. SpA polypeptides or fragments can be consecutive or separated by a spacer or other peptide sequences, e.g., one or more additional bacterial peptide. In a further aspect, the other polypeptides or peptides contained in the multimer or concatamer can include, but are not limited to 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 of Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, vWh or immunogenic fragments thereof. Additional staphylococcal antigens that can be used in combination with a Protein A variant include, but are not limited to 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg2+ transporter, MHC II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/Fig (WO 00/12689), SdrH

(WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein.

The term "Protein A variant" or "SpA variant" refers to polypeptides that include a SpA IgG domain having two or more amino acid substitutions that disrupt binding to Fc and V_H3. In certain aspect, a SpA variant includes a variant domain D peptide, as well as variants of SpA polypeptides and segments thereof that are non-toxicogenic and stimulate an immune response against *staphylococcus* bacteria Protein A and/or bacteria expressing such.

Embodiments of the present invention include methods for eliciting an immune response against a *staphylococcus* bacterium or staphylococci in a subject comprising providing to the subject an effective amount of a Protein A variant or a segment thereof. In certain aspects, the methods for eliciting an immune response against a *staphylococcus* bacterium or staphylococci in a subject comprising providing to the subject an effective amount of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or more secreted proteins and/or cell surface proteins or segments/fragments thereof. A secreted protein or cell surface protein includes, but is not limited to Eap, Ebh, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, and/or vWh proteins and immunogenic fragments thereof. Additional staphylococcal antigens that can be used in combination with a Protein A variant include, but are not limited to 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase,

autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg²⁺ transporter, MHC II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein.

Embodiments of the invention include compositions that include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to Protein A, or a second protein or peptide that is a secreted bacterial protein or a bacterial cell surface protein. In a further embodiment of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a Protein A domain D polypeptide (SEQ ID NO:2), domain E (SEQ ID NO:3), domain A (SEQ ID NO:4), domain C (SEQ ID NO:5), domain B (SEQ ID NO:6), or a nucleic acid sequence encoding a Protein A domain D, domain E, domain A, domain C, or domain B polypeptide. In certain aspects a Protein A polypeptide segment will have an amino acid sequence of SEQ ID NO:8. Similarity or identity, with identity being preferred, is known in the art and a number of different programs can be used to identify whether a protein (or nucleic acid) has sequence identity or similarity to a known sequence. Sequence identity and/or similarity is determined using standard techniques known in the art, including, but not limited to, the local sequence identity algorithm of Smith & Waterman (1981), by the sequence identity alignment algorithm of Needleman & Wunsch (1970), by the search for similarity method of Pearson & Lipman (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Drive, Madison, Wis.), the Best Fit sequence program described by Devereux et al. (1984), preferably using the default settings, or by inspection. Preferably, percent identity is calculated by using alignment tools known to and readily ascertainable to those of skill in the art. Percent identity is essentially the number of identical amino acids divided by the total number of amino acids compared times one hundred.

Still further embodiments include methods for stimulating in a subject a protective or therapeutic immune response against a *staphylococcus* bacterium comprising administering to the subject an effective amount of a composition including (i) a SpA variant, e.g., a variant SpA domain D polypeptide or peptide thereof; or, (ii) a nucleic acid molecule encoding such a SpA variant polypeptide or peptide thereof, or (iii) administering a SpA variant domain D polypeptide with any combination or permutation of bacterial proteins described herein. In a preferred embodiment the composition is not a *staphylococcus* bacterium. In certain aspects the subject is a human or a cow. In a further aspect the composition is formulated in a pharmaceutically acceptable formulation. The staphylococci may be *Staphylococcus aureus*.

Yet still further embodiments include vaccines comprising a pharmaceutically acceptable composition having an

isolated SpA variant polypeptide, or any other combination or permutation of protein(s) or peptide(s) described herein, wherein the composition is capable of stimulating an immune response against a *staphylococcus* bacterium. The vaccine may comprise an isolated SpA variant polypeptide, or any other combination or permutation of protein(s) or peptide(s) described. In certain aspects of the invention the isolated SpA variant polypeptide, or any other combination or permutation of protein(s) or peptide(s) described are multimerized, e.g., dimerized or concatamerized. In a further aspect, the vaccine composition is contaminated by less than about 10, 9, 8, 7, 6, 5, 4, 3, 2, 1, 0.5, 0.25, 0.05% (or any range derivable therein) of other Staphylococcal proteins. A composition may further comprise an isolated non-SpA polypeptide. Typically the vaccine comprises an adjuvant. In certain aspects a protein or peptide of the invention is linked (covalently or non-covalently) to the adjuvant, preferably the adjuvant is chemically conjugated to the protein.

In still yet further embodiments, a vaccine composition is a pharmaceutically acceptable composition having a recombinant nucleic acid encoding all or part of a SpA variant polypeptide, or any other combination or permutation of protein(s) or peptide(s) described herein, wherein the composition is capable of stimulating an immune response against a *staphylococcus* bacteria. The vaccine composition may comprise a recombinant nucleic acid encoding all or part of a SpA variant polypeptide, or any other combination or permutation of protein(s) or peptide(s) described herein. In certain embodiments the recombinant nucleic acid contains a heterologous promoter. Preferably the recombinant nucleic acid is a vector. More preferably the vector is a plasmid or a viral vector. In some aspects the vaccine includes a recombinant, non-*staphylococcus* bacterium containing the nucleic acid. The recombinant non-staphylococci may be *Salmonella* or another gram-positive bacteria. The vaccine may comprise a pharmaceutically acceptable excipient, more preferably an adjuvant.

Still further embodiments include methods for stimulating in a subject a protective or therapeutic immune response against a *staphylococcus* bacterium comprising administering to the subject an effective amount of a composition of a SpA variant polypeptide or segment/fragment thereof and further comprising one or more of a Eap, Ebh, Fmp, FsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, or vWh protein or peptide thereof. In a preferred embodiment the composition comprises a non-*staphylococcus* bacterium. In a further aspect the composition is formulated in a pharmaceutically acceptable formulation. The staphylococci for which a subject is being treated may be *Staphylococcus aureus*. Methods of the invention also include SpA variant compositions that contain 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or more secreted virulence factors and/or cell surface proteins, such as Eap, Ebh, Emp, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, or vWh in various combinations. In certain aspects a vaccine formulation includes Eap, Ebh, Emp, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, and vWh. In certain aspects an antigen combination can include (1) a SpA variant and IsdA; (2) SpA variant and ClfB; (3) SpA variant and SdrD; (4) SpA variant and Hla or Hla variant; (5) SpA variant and ClfB, SdrD, and Hla or Hla variant; (6) SpA variant, IsdA, SdrD, and Hla or Hla variant; (7) SpA variant, IsdA, ClfB, and Hla or Hla variant; (8) SpA variant, IsdA, ClfB, and SdrD; (9) SpA variant, IsdA, ClfB, SdrD and Hla or Hla variant; (10)

SpA variant, IsdA, ClfB, and SdrD; (11) SpA variant, IsdA, SdrD, and Hla or Hla variant; (12) SpA variant, IsdA, and Hla or Hla variant; (13) SpA variant, IsdA, ClfB, and Hla or Hla variant; (14) SpA variant, ClfB, and SdrD; (15) SpA variant, ClfB, and Hla or Hla variant; or (16) SpA variant, SdrD, and Hla or Hla variant.

In certain aspects, a bacterium delivering a composition of the invention will be limited or attenuated with respect to prolonged or persistent growth or abscess formation. In yet a further aspect, SpA variant(s) can be overexpressed in an attenuated bacterium to further enhance or supplement an immune response or vaccine formulation.

The term "EsxA protein" refers to a protein that includes isolated wild-type EsxA polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria EsxA proteins.

The term "EsxB protein" refers to a protein that includes isolated wild-type EsxB polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria EsxB proteins.

The term "SdrD protein" refers to a protein that includes isolated wild-type SdrD polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria SdrD proteins.

The term "SdrE protein" refers to a protein that includes isolated wild-type SdrE polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria SdrE proteins.

The term "IsdA protein" refers to a protein that includes isolated wild-type IsdA polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria IsdA proteins.

The term "IsdB protein" refers to a protein that includes isolated wild-type IsdB polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria IsdB proteins.

The term "Eap protein" refers to a protein that includes isolated wild-type Eap polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria Eap proteins.

The term "Ebh protein" refers to a protein that includes isolated wild-type Ebh polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria Ebh proteins.

The term "Emp protein" refers to a protein that includes isolated wild-type Emp polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria Emp proteins.

The term "EsaB protein" refers to a protein that includes isolated wild-type EsaB polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria EsaB proteins.

The term "EsaC protein" refers to a protein that includes isolated wild-type EsaC polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria EsaC proteins.

The term "SdrC protein" refers to a protein that includes isolated wild-type SdrC polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria SdrC proteins.

The term "ClfA protein" refers to a protein that includes isolated wild-type ClfA polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria ClfA proteins.

The term "ClfB protein" refers to a protein that includes isolated wild-type ClfB polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria ClfB proteins.

The term "Coa protein" refers to a protein that includes isolated wild-type Coa polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria Coa proteins.

The term "Hla protein" refers to a protein that includes isolated wild-type Hla polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria Hla proteins.

The term "IsdC protein" refers to a protein that includes isolated wild-type IsdC polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria IsdC proteins.

The term "SasF protein" refers to a protein that includes isolated wild-type SasF polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria SasF proteins.

The term "vWbp protein" refers to a protein that includes isolated wild-type vWbp (von Willebrand factor binding protein) polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria vWbp proteins.

The term "vWh protein" refers to a protein that includes isolated wild-type vWh (von Willebrand factor binding protein homolog) polypeptides from *staphylococcus* bacteria and segments thereof, as well as variants that stimulate an immune response against *staphylococcus* bacteria vWh proteins.

An immune response refers to a humoral response, a cellular response, or both a humoral and cellular response in an organism. An immune response can be measured by assays that include, but are not limited to, assays measuring the presence or amount of antibodies that specifically recognize a protein or cell surface protein, assays measuring T-cell activation or proliferation, and/or assays that measure modulation in terms of activity or expression of one or more cytokines.

In still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an EsxA protein. In certain aspects the EsxA protein will have all or part of the amino acid sequence of SEQ ID NO:11.

In still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an EsxB protein. In certain aspects the EsxB protein will have all or part of the amino acid sequence of SEQ ID NO:12.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an SdrD protein. In certain aspects the SdrD protein will have all or part of the amino acid sequence of SEQ ID NO:13.

In further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an SdrE protein. In certain aspects the SdrE protein will have all or part of the amino acid sequence of SEQ ID NO:14.

In still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an LsdA protein. In certain aspects the LsdA protein will have all or part of the amino acid sequence of SEQ ID NO:15.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an LsdB protein. In certain aspects the LsdB protein will have all or part of the amino acid sequence of SEQ ID NO:16.

Embodiments of the invention include compositions that include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a EsaB protein. In certain aspects the EsaB protein will have all or part of the amino acid sequence of SEQ ID NO:17.

In a further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a ClfB protein. In certain aspects the ClfB protein will have all or part of the amino acid sequence of SEQ ID NO:18.

In still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an LsdC protein. In certain aspects the LsdC protein will have all or part of the amino acid sequence of SEQ ID NO:19.

In yet further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a SasF protein. In certain aspects the SasF protein will have all or part of the amino acid sequence of SEQ ID NO:20.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a SdrC protein. In certain aspects the SdrC protein will have all or part of the amino acid sequence of SEQ ID NO:21.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a ClfA protein. In certain aspects the ClfA protein will have all or part of the amino acid sequence of SEQ ID NO:22.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an Eap protein. In certain aspects the Eap protein will have all or part of the amino acid sequence of SEQ ID NO:23.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an Ebh protein. In certain aspects the Ebh protein will have all or part of the amino acid sequence of SEQ ID NO:24.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an Emp protein. In certain aspects the Emp protein will have all or part of the amino acid sequence of SEQ ID NO:25.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to an EsaC protein. In certain aspects the EsaC protein will have all or part of the amino acid sequence of SEQ ID NO:26. Sequence of EsaC polypeptides can be found in the protein databases and include, but are not limited to accession numbers ZP_02760162 (GI:168727885), NP_645081.1 (GI:21281993), and NP_370813.1 (GI:15923279), each of which is incorporated herein by reference as of the priority date of this application.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a Coa protein. In certain aspects the Coa protein will have all or part of the amino acid sequence of SEQ ID NO:27.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a Hla protein. In certain aspects the Hla protein will have all or part of the amino acid sequence of SEQ ID NO:28.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a vWa protein. In certain aspects the vWa protein will have all or part of the amino acid sequence of SEQ ID NO:29.

In yet still further embodiments of the invention a composition may include a polypeptide, peptide, or protein that is or is at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% identical or similar to a vWbp protein. In certain aspects the vWbp protein will have all or part of the amino acid sequence of SEQ ID NO:32.

In certain aspects, a polypeptide or segment/fragment can have a sequence that is at least 85%, at least 90%, at least 95%, at least 98%, or at least 99% or more identical to the amino acid sequence of the reference polypeptide. The term "similarity" refers to a polypeptide that has a sequence that has a certain percentage of amino acids that are either identical with the reference polypeptide or constitute conservative substitutions with the reference polypeptides.

The polypeptides described herein may include 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, or more variant amino acids within at least, or at most 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 101, 102, 103, 104, 105, 106, 107, 108, 109, 110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 125, 126, 127, 128, 129, 130,

131, 132, 133, 134, 135, 136, 137, 138, 139, 140, 141, 142, 143, 144, 145, 146, 147, 148, 149, 150, 151, 152, 153, 154, 155, 156, 157, 158, 159, 160, 161, 162, 163, 164, 165, 166, 167, 168, 169, 170, 171, 172, 173, 174, 175, 176, 177, 178, 179, 180, 181, 182, 183, 184, 185, 186, 187, 188, 189, 190, 191, 192, 193, 194, 195, 196, 197, 198, 199, 200, 201, 202, 203, 204, 205, 206, 207, 208, 209, 210, 211, 212, 213, 214, 215, 216, 217, 218, 219, 220, 221, 222, 223, 224, 225, 226, 227, 228, 229, 230, 231, 232, 233, 234, 235, 236, 237, 238, 239, 240, 241, 242, 243, 244, 245, 246, 247, 248, 249, 250, 300, 400, 500, 550, 1000 or more contiguous amino acids, or any range derivable therein, of SEQ ID NO:2-30, or SEQ ID NO:32-34.

A polypeptide segment as described herein may include 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 101, 102, 103, 104, 105, 106, 107, 108, 109, 110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 125, 126, 127, 128, 129, 130, 131, 132, 133, 134, 135, 136, 137, 138, 139, 140, 141, 142, 143, 144, 145, 146, 147, 148, 149, 150, 151, 152, 153, 154, 155, 156, 157, 158, 159, 160, 161, 162, 163, 164, 165, 166, 167, 168, 169, 170, 171, 172, 173, 174, 175, 176, 177, 178, 179, 180, 181, 182, 183, 184, 185, 186, 187, 188, 189, 190, 191, 192, 193, 194, 195, 196, 197, 198, 199, 200, 201, 202, 203, 204, 205, 206, 207, 208, 209, 210, 211, 212, 213, 214, 215, 216, 217, 218, 219, 220, 221, 222, 223, 224, 225, 226, 227, 228, 229, 230, 231, 232, 233, 234, 235, 236, 237, 238, 239, 240, 241, 242, 243, 244, 245, 246, 247, 248, 249, 250, 300, 400, 500, 550, 1000 or more contiguous amino acids, or any range derivable therein, of SEQ ID NO:2-30, or SEQ ID NO:33-34.

The compositions may be formulated in a pharmaceutically acceptable composition. In certain aspects of the invention the *staphylococcus* bacterium is an *S. aureus* bacterium.

In further aspects, a composition may be administered more than one time to the subject, and may be administered 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20 or more times. The administration of the compositions include, but is not limited to oral, parenteral, subcutaneous, intramuscular, intravenous, or various combinations thereof, including inhalation or aspiration.

In still further embodiments, a composition comprises a recombinant nucleic acid molecule encoding a polypeptide described herein or segments/fragments thereof. Typically a recombinant nucleic acid molecule encoding a polypeptide described herein contains a heterologous promoter. In certain aspects, a recombinant nucleic acid molecule of the invention is a vector, in still other aspects the vector is a plasmid. In certain embodiments the vector is a viral vector. In certain aspects a composition includes a recombinant, non-*staphylococcus* bacterium containing or expressing a polypeptide described herein. In particular aspects the recombinant non-*staphylococcus* bacteria is *Salmonella* or another gram-positive bacteria. A composition is typically administered to mammals, such as human subjects, but administration to other animals that are capable of eliciting an immune response is contemplated. In further aspects the *staphylococcus* bacterium containing or expressing the polypeptide is *Staphylococcus aureus*. In further embodiments the immune response is a protective immune response.

In further embodiments a composition comprises a recombinant nucleic acid molecule encoding all or part of

one or more of a Eap, Ehb, Emp, EsaB, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, SpA, vWbp, or vWh protein or peptide or variant thereof. Additional staphylococcal antigens that can be used in combination with the polypeptides described herein include, but are not limited to 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg2+ transporter, MHC II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF(WO 00/12689), SdrG/Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein. In particular aspects, a bacteria is a recombinant non-*staphylococcus* bacteria, such as a *Salmonella* or other gram-positive bacteria.

Compositions of the invention are typically administered to human subjects, but administration to other animals that are capable of eliciting an immune response to a *staphylococcus* bacterium is contemplated, particularly cattle, horses, goats, sheep and other domestic animals, i.e., mammals.

In certain aspects the *staphylococcus* bacterium is a *Staphylococcus aureus*. In further embodiments the immune response is a protective immune response. In still further aspects, the methods and compositions of the invention can be used to prevent, ameliorate, reduce, or treat infection of tissues or glands, e.g., mammary glands, particularly mastitis and other infections. Other methods include, but are not limited to prophylactically reducing bacterial burden in a subject not exhibiting signs of infection, particularly those subjects suspected of or at risk of being colonized by a target bacteria, e.g., patients that are or will be at risk or susceptible to infection during a hospital stay, treatment, and/or recovery.

Any embodiment discussed with respect to one aspect of the invention applies to other aspects of the invention as well. In particular, any embodiment discussed in the context of a SpA variant polypeptide or peptide or nucleic acid may be implemented with respect to other antigens, such as Eap, Ehb, Emp, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, vWh, 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase, autolysin amidase, Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg2+ transporter, MHC II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF(WO 00/12689), SdrG/Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein (or nucleic acids), and vice versa. It is also understood that any one or

more of Eap, Ebh, Emp, EsaC, EsxA, EsxB, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, Coa, Hla, IsdC, SasF, vWbp, vWh, 52 kDa vitronectin binding protein (WO 01/60852), Aaa, Aap, Ant, autolysin glucosaminidase, autolysin amidase. Cna, collagen binding protein (U.S. Pat. No. 6,288,214), EFB (FIB), Elastin binding protein (EbpS), EPB, FbpA, fibrinogen binding protein (U.S. Pat. No. 6,008,341), Fibronectin binding protein (U.S. Pat. No. 5,840,846), FnbA, FnbB, GehD (US 2002/0169288), HarA, HBP, Immunodominant ABC transporter, IsaA/PisA, laminin receptor, Lipase GehD, MAP, Mg²⁺ transporter, MHC II analogue (U.S. Pat. No. 5,648,240), MRPII, Npase, RNA III activating protein (RAP), SasA, SasB, SasC, SasD, SasK, SBI, SdrF (WO 00/12689), SdrG/Fig (WO 00/12689), SdrH (WO 00/12689), SEA exotoxins (WO 00/02523), SEB exotoxins (WO 00/02523), SitC and Ni ABC transporter, SitC/MntC/saliva binding protein (U.S. Pat. No. 5,801,234), SsaA, SSP-1, SSP-2, and/or Vitronectin binding protein can be specifically excluded from a claimed composition.

Embodiments of the invention include compositions that contain or do not contain a bacterium. A composition may or may not include an attenuated or viable or intact staphylococcal bacterium. In certain aspects, the composition comprises a bacterium that is not a staphylococcal bacterium or does not contain staphylococcal bacteria. In certain embodiments a bacterial composition comprises an isolated or recombinantly expressed staphylococcal Protein A variant or a nucleotide encoding the same. The composition may be or include a recombinantly engineered *staphylococcus* bacterium that has been altered in a way that comprises specifically altering the bacterium with respect to a secreted virulence factor or cell surface protein. For example, the bacteria may be recombinantly modified to express more of the virulence factor or cell surface protein than it would express if unmodified.

The term "isolated" can refer to a nucleic acid or polypeptide that is substantially free of cellular material, bacterial material, viral material, or culture medium (when produced by recombinant DNA techniques) of their source of origin, or chemical precursors or other chemicals (when chemically synthesized). Moreover, an isolated compound refers to one that can be administered to a subject as an isolated compound; in other words, the compound may not simply be considered "isolated" if it is adhered to a column or embedded in an agarose gel. Moreover, an "isolated nucleic acid fragment" or "isolated peptide" is a nucleic acid or protein fragment that is not naturally occurring as a fragment and/or is not typically in the functional state.

Moieties of the invention, such as polypeptides, peptides, antigens, or immunogens, may be conjugated or linked covalently or noncovalently to other moieties such as adjuvants, proteins, peptides, supports, fluorescence moieties, or labels. The term "conjugate" or "immunoconjugate" is broadly used to define the operative association of one moiety with another agent and is not intended to refer solely to any type of operative association, and is particularly not limited to chemical "conjugation." Recombinant fusion proteins are particularly contemplated. Compositions of the invention may further comprise an adjuvant or a pharmaceutically acceptable excipient. An adjuvant may be covalently or non-covalently coupled to a polypeptide or peptide of the invention. In certain aspects, the adjuvant is chemically conjugated to a protein, polypeptide, or peptide.

The term "providing" is used according to its ordinary meaning to indicate "to supply or furnish for use." In some embodiments, the protein is provided directly by administering the protein, while in other embodiments, the protein

is effectively provided by administering a nucleic acid that encodes the protein. In certain aspects the invention contemplates compositions comprising various combinations of nucleic acid, antigens, peptides, and/or epitopes.

The subject will have (e.g., are diagnosed with a staphylococcal infection), will be suspected of having, or will be at risk of developing a staphylococcal infection. Compositions of the present invention include immunogenic compositions wherein the antigen(s) or epitope(s) are contained in an amount effective to achieve the intended purpose. More specifically, an effective amount means an amount of active ingredients necessary to stimulate or elicit an immune response, or provide resistance to, amelioration of, or mitigation of infection. In more specific aspects, an effective amount prevents, alleviates or ameliorates symptoms of disease or infection, or prolongs the survival of the subject being treated. Determination of the effective amount is well within the capability of those skilled in the art, especially in light of the detailed disclosure provided herein. For any preparation used in the methods of the invention, an effective amount or dose can be estimated initially from in vitro studies, cell culture, and/or animal model assays. For example, a dose can be formulated in animal models to achieve a desired immune response or circulating antibody concentration or titer. Such information can be used to more accurately determine useful doses in humans.

The embodiments in the Example section are understood to be embodiments of the invention that are applicable to all aspects of the invention.

The use of the term "or" in the claims is used to mean "and/or" unless explicitly indicated to refer to alternatives only or the alternatives are mutually exclusive, although the disclosure supports a definition that refers to only alternatives and "and/or." It is also contemplated that anything listed using the term "or" may also be specifically excluded.

Throughout this application, the term "about" is used to indicate that a value includes the standard deviation of error for the device or method being employed to determine the value.

Following long-standing patent law, the words "a" and "an," when used in conjunction with the word "comprising" in the claims or specification, denotes one or more, unless specifically noted.

Other objects, features and advantages of the present invention will become apparent from the following detailed description. It should be understood, however, that the detailed description and the specific examples, while indicating specific embodiments of the invention, are given by way of illustration only, since various changes and modifications within the spirit and scope of the invention will become apparent to those skilled in the art from this detailed description.

DESCRIPTION OF THE DRAWINGS

So that the matter in which the above-recited features, advantages and objects of the invention as well as others which will become clear are attained and can be understood in detail, more particular descriptions and certain embodiments of the invention briefly summarized above are illustrated in the appended drawings. These drawings form a part of the specification. It is to be noted, however, that the appended drawings illustrate certain embodiments of the invention and therefore are not to be considered limiting in their scope.

FIGS. 1A-1B. (FIG. 1A) Primary structure of the Protein A precursor with an N-terminal YSIRK motif signal peptide,

five immunoglobulin binding domains as tandem repeats designated E, D, A, B, C, region X, and the LPXTG sorting signal. (FIG. 1B) Following synthesis of the Protein A precursor, staphylococci secrete this product via the Sec pathway, and sortase A cleaves the LPXTG sorting signal between the T and G residues. Nucleophilic attack of the amino group within lipid II at the sortase-Protein A thioester-linked intermediate forms the amide bond that links Protein A to the cell wall envelope and enables its display on the bacterial surface.

FIG. 2. Three dimensional model of the molecular interactions between the SpA-domain D of Protein A, the VH3 Fab domain of the B cell receptor, and of the Fc γ domain of immunoglobulin. The model is derived from two crystal structures (Graillie et al., 2000 and Gouda et al., 1992) that revealed side chain residues involved in the formation of ionic bonds that enable these complexes. Gln-9 and Gln-10 of SpA-D promote binding to Fc γ , whereas Asp-36 and Asp-37 enable complex formation with VH3 Fab.

FIG. 3. Left panel—Coomassie Blue stained SDS-PAGE reveals the migrational position of purified His-tagged SpA, SpA-D, SpA-D_{Q9,10K;D36,37A}, human IgG, and sortase A (SrtA), a control protein. Right panel—Coomassie Blue stained SDS-PAGE to reveal the elution of Protein A immunoglobulin complexes eluted following affinity chromatography of human IgG on Ni-NTA columns pre-charged with His-tagged SpA, SpA-D, SpA-D_{Q9,10K;D36,37A} or SrtA.

FIG. 4. ELISA assays to quantify human immunoglobulin (hIgG), human F(ab)₂ IgG fragments and human Fc fragments of immunoglobulin (hFc). Plates were coated with equal amounts of His-tagged SpA, SpA-D, SpA-D_{Q9,10K;D36,37A} or SrtA. hIgG-HRP, F(ab)₂-HRP and hFc-HRP were added onto the plates and incubated for an hour. Absorbance at 450 nm was recorded and plotted to determine the half maximal titers.

FIG. 5. Purified SpA-D, SpA-D_{Q9,10K;D36,37A} or a PBS mock control were injected into the peritoneum of mice and analyzed for their ability to reduce the B cell population in the spleen of experimental BALB/c mice. Animals were killed 4 hours following injection, their spleen removed, tissue homogenized and stained with CD19 antibodies directed against B cells. The number of B cells was quantified by FACS sorting.

FIG. 6 Generation of a non-toxicogenic protein A vaccine. a, Translational protein A (SpA) product of *S. aureus* Newman and USA300 LAC with an N-terminal signal peptide (white box), five immunoglobulin binding domains (IgBDs designated E, D, A, B and C), variable region X and C-terminal sorting signal (black box). b, Amino acid sequence of the five IgBDs as well as nontoxicogenic SpA-D_{KKAA}, with the positions of triple α -helical bundles (H1, H2 and H3) as well as glutamine (Q) 9, 10 and aspartate (D) 36, 37 indicated. c, Coomassie Blue-stained SDS-PAGE of SpA, SpA-D, SpA-D_{KKAA} or SrtA purified on Ni-NTA sepharose in the presence or absence of human immunoglobulin (hIgG). d, ELISA examining the association of immobilized SpA, SpA-D or SpA-D_{KKAA} with human IgG as well as its Fc or F(ab)₂ fragments and von Willebrand factor (vWF). e, CD19+B lymphocytes in splenic tissue of BALB/c mice that had been mock immunized or treated with SpA-D or SpA-D_{KKAA} were quantified by FACS.

FIG. 7 Non-toxicogenic protein A vaccine prevents abscess formation. Histopathology of renal tissue isolated during necropsy of BALB/c mice that had been mock immunized (PBS) or vaccinated with SpA, SpA-D as well as SpA-D_{KKAA} and challenged with *S. aureus* Newman. Thin sectioned tissues were stained with hematoxylin-eosin. White

arrows identify polymorphonuclear leukocyte (PMN) infiltrates. Dark arrows identify staphylococcal abscess communities.

FIG. 8 Antibodies raised by the non-toxicogenic protein A vaccine block the B cell superantigen function of SpA. a, Rabbit antibodies raised against SpA-D_{KKAA} were purified on a matrix with immobilized antigen and analyzed by Coomassie Blue-stained SDS-PAGE. Antibodies were cleaved with pepsin and F(ab)₂ fragments were purified by a second round of affinity chromatography on SpA-D_{KKAA} matrix. b, SpA-D_{KKAA} specific F(ab)₂ interfere with the binding of SpA or SpA-D to human immunoglobulin (hIgG) or, c, to von Willebrand Factor (vWF).

FIG. 9 Full-length non-toxicogenic protein A generates improved immune responses. a, Full-length SpA_{KKAA} was purified on Ni-NTA sepharose and analyzed by Coomassie-Blue stained SDS-PAGE. b, CD19+B lymphocytes in splenic tissue of BALB/c mice that had been mock immunized or treated with SpA or SpA_{KKAA} were quantified by FACS. c, ELISA examining the association of immobilized SpA or SpA_{KKAA} with human IgG as well as its Fc or F(ab)₂ fragments or von Willebrand factor (vWF). d, Human or mouse serum antibody titers to diphtheria toxoid (CRM197) and non-toxicogenic SpA_{KKAA} or SpA-D_{KKAA}. Human volunteers with a history of DTaP immunization and staphylococcal infection (n=16) as well as mice (n=20) that had been infected with *S. aureus* Newman or USA 300 LAC or immunized with SpA_{KKAA} or SpA-D_{KKAA} were examined by quantitative dot blot.

FIG. 10 Staphylococcal infection does not generate protective immunity. BALB/c mice (n=20) were infected with *S. aureus* Newman or mock challenged (PBS) for thirty days and infection cleared with chloramphenicol treatment. Both cohorts of animals were then challenged with *S. aureus* Newman and bacterial load (CFU) in kidney tissue homogenate analyzed following necropsy on day 4.

FIG. 11 Comparison of abscess formation in mice treated with PBS, SpA, SpA-D, and SpA-D_{KKAA}.

FIG. 12 (A) ELISA examining the association of immobilized SpA, SpA-D, SpA-DKKAA or SpA-DGGSS with human IgG as well as its Fc or F(ab)₂ fragments and IgM. Statistical significance of SpA-DKKAA and SpA-DGGSS binding to each ligand was compared against SpA-D; SpA-D binding was compared against SpA (n=3); *signifies P<0.05; **signifies P<0.01. (B) ELISA examining the level of cross-reactive antibodies of hyper-immune sera samples collected from actively immunized mice (n=5) with SpA-D, SpA-DKKAA and SpA-DGGSS. (C) Abscess formation in mice treated with PBS, SpA-D, SpA-D_{KKAA} and SpA-D_{GGSS}.

DETAILED DESCRIPTION

Staphylococcus aureus is a commensal of the human skin and nares, and the leading cause of bloodstream, skin and soft tissue infections (Klevens et al., 2007). Recent dramatic increases in the mortality of staphylococcal diseases are attributed to the spread of methicillin-resistant *S. aureus* (MRSA) strains often not susceptible to antibiotics (Kennedy et al., 2008). In a large retrospective study, the incidence of MRSA infections was 4.6% of all hospital admissions in the United States (Klevens et al., 2007). The annual health care costs for 94,300 MRSA infected individuals in the United States exceed \$2.4 billion (Klevens et al., 2007). The current MRSA epidemic has precipitated a public health crisis that needs to be addressed by development of a

preventive vaccine (Boucher and Corey, 2008). To date, an FDA licensed vaccine that prevents *S. aureus* diseases is not available.

The inventors describe here the use of Protein A, a cell wall anchored surface protein of staphylococci, for the generation of variants that can serve as subunit vaccines. The pathogenesis of staphylococcal infections is initiated as bacteria invade the skin or blood stream via trauma, surgical wounds, or medical devices (Lowy, 1998). Although the invading pathogen may be phagocytosed and killed, staphylococci can also escape innate immune defenses and seed infections in organ tissues, inducing inflammatory responses that attract macrophages, neutrophils, and other phagocytes (Lowy, 1998). The responsive invasion of immune cells to the site of infection is accompanied by liquefaction necrosis as the host seeks to prevent staphylococcal spread and allow for removal of necrotic tissue debris (Lam et al., 1963). Such lesions can be observed by microscopy as hypercellular areas containing necrotic tissue, leukocytes, and a central nidus of bacteria (Lam et al., 1963). Unless staphylococcal abscesses are surgically drained and treated with antibiotics, disseminated infection and septicemia produce a lethal outcome (Sheagren, 1984).

II. STAPHYLOCOCCAL ANTIGENS

A. Staphylococcal Protein A (SpA)

All *Staphylococcus aureus* strains express the structural gene for Protein A (*spa*) (Jensen, 1958; Said-Salim et al., 2003), a well characterized virulence factor whose cell wall anchored surface protein product (SpA) encompasses five highly homologous immunoglobulin binding domains designated E, D, A, B, and C (Sjodahl, 1977). These domains display ~ 80% identity at the amino acid level, are 56 to 61 residues in length, and are organized as tandem repeats (Uhlen et al., 1984). SpA is synthesized as a precursor protein with an N-terminal YSIRK/GS signal peptide and a C-terminal LPXTG motif sorting signal (DeDent et al., 2008; Schneewind et al., 1992). Cell wall anchored Protein A is displayed in great abundance on the staphylococcal surface (DeDent et al., 2007; Sjoquist et al., 1972). Each of its immunoglobulin binding domains is composed of anti-parallel α -helices that assemble into a three helix bundle and bind the Fc domain of immunoglobulin G (IgG) (Deisenhofer, 1981; Deisenhofer et al., 1978), the VH3 heavy chain (Fab) of IgM (i.e., the B cell receptor) (Graille et al., 2000), the von Willebrand factor at its A1 domain [vWF A1 is a ligand for platelets] (O'Seaghda et al., 2006) and the tumor necrosis factor α (TNF- α) receptor I (TNFRI) (Gomez et al., 2006), which is displayed on surfaces of airway epithelia (Gomez et al., 2004; Gomez et al., 2007).

SpA impedes neutrophil phagocytosis of staphylococci through its attribute of binding the Fc component of IgG (Jensen, 1958; Uhlen et al., 1984). Moreover, SpA is able to activate intravascular clotting via its binding to von Willebrand factor A1 domains (Hartleib et al., 2000). Plasma proteins such as fibrinogen and fibronectin act as bridges between staphylococci (C1fA and C1fB) and the platelet integrin GPIIb/IIIa (O'Brien et al., 2002), an activity that is supplemented through Protein A association with vWF A1, which allows staphylococci to capture platelets via the GPIIb- α platelet receptor (Foster, 2005; O'Seaghda et al., 2006). SpA also binds TNFRI and this interaction contributes to the pathogenesis of staphylococcal pneumonia (Gomez et al., 2004). SpA activates proinflammatory signaling through TNFR1 mediated activation of TRAF2, the p38/c-Jun kinase, mitogen activate protein kinase (MAPK)

and the Rel-transcription factor NF-KB. SpA binding further induces TNFR1 shedding, an activity that appears to require the TNF-converting enzyme (TACE)(Gomez et al., 2007). All of the aforementioned SpA activities are mediated through its five IgG binding domains and can be perturbed by the same amino acid substitutions, initially defined by their requirement for the interaction between Protein A and human IgG1 (Cedergren et al., 1993).

SpA also functions as a B cell superantigen by capturing the Fab region of VH3 bearing IgM, the B cell receptor (Gomez et al., 2007; Goodyear et al., 2003; Goodyear and Silverman, 2004; Roben et al., 1995). Following intravenous challenge, staphylococcal Protein A (SpA) mutations show a reduction in staphylococcal load in organ tissues and dramatically diminished ability to form abscesses (described herein). During infection with wildtype *S. aureus*, abscesses are formed within forty-eight hours and are detectable by light microscopy of hematoxylin-eosin stained, thin-sectioned kidney tissue, initially marked by an influx of polymorphonuclear leukocytes (PMNs). On day 5 of infection, abscesses increase in size and enclosed a central population of staphylococci, surrounded by a layer of eosinophilic, amorphous material and a large cuff of PMNs. Histopathology revealed massive necrosis of PMNs in proximity to the staphylococcal nidus at the center of abscess lesions as well as a mantle of healthy phagocytes. The inventors also observed a rim of necrotic PMNs at the periphery of abscess lesions, bordering the eosinophilic pseudocapsule that separated healthy renal tissue from the infectious lesion. Staphylococcal variants lacking Protein A are unable to establish the histopathology features of abscesses and are cleared during infection.

In previous studies, Cedergren et al. (1993) engineered five individual substitutions in the Fc fragment binding sub-domain of the B domain of SpA, L17D, N28A, I31A and K35A. These authors created these proteins to test data gathered from a three dimensional structure of a complex between one domain of SpA and Fc₁. Cedergren et al. determined the effects of these mutations on stability and binding, but did not contemplate use of such substitutions for the production of a vaccine antigen.

Brown et al. (1998) describe studies designed to engineer new proteins based on SpA that allow the use of more favorable elution conditions when used as affinity ligands. The mutations studied included single mutations of Q13A, Q14H, N15A, N15H, F17H, Y18F, L21H, N32H, or K39H. Brown et al. report that Q13A, N15A, N15H, and N32H substitutions made little difference to the dissociation constant values and that the Y18F substitution resulted in a 2 fold decrease in binding affinity as compared to wild type SpA. Brown et al. also report that L21H and F17H substitutions decrease the binding affinity by five-fold and a hundred-fold respectively. The authors also studied analogous substitutions in two tandem domains. Thus, the Brown et al. studies were directed to generating a SpA with a more favorable elution profile, hence the use of His substitutions to provide a pH sensitive alteration in the binding affinity. Brown et al. is silent on the use of SpA as a vaccine antigen.

Graille et al. (2000) describe a crystal structure of domain D of SpA and the Fab fragment of a human IgM antibody. Graille et al. define by analysis of a crystal structure the D domain amino acid residues that interact with the Fab fragment as residues Q26, G29, F30, Q32, S33, D36, D37, Q40, N43, E47, or L51, as well as the amino acid residues that form the interface between the domain D sub-domains. Graille et al. define the molecular interactions of these two

proteins, but is silent in regard to any use of substitutions in the interacting residues in producing a vaccine antigen.

O'Seaghdha et al. (2006) describe studies directed at elucidating which sub-domain of domain D binds vWF. The authors generated single mutations in either the Fc or V_{H3} binding sub-domains, i.e., amino acid residues F5A, Q9A, Q10A, F13A, Y14A, L17A, N28A, I31A, K35A, G29A, F30A, S33A, D36A, D37A, Q40A, E47A, or Q32A. The authors discovered that vWF binds the same sub-domain that binds Fc. O'Seaghdha et al. define the sub-domain of domain D responsible for binding vWF, but is silent in regard to any use of substitutions in the interacting residues in producing a vaccine antigen.

Gomez et al. (2006) describe the identification of residues responsible for activation of the TNFR1 by using single mutations of F5A, F13A, Y14A, L17A, N21A, I31A, Q32A, and K35A. Gomez et al. is silent in regard to any use of substitutions in the interacting residues in producing a vaccine antigen.

Recombinant affinity tagged Protein A, a polypeptide encompassing the five IgG domains (EDCAB) (Sjodahl, 1977) but lacking the C-terminal Region X (Guss et al., 1984), was purified from recombinant *E. coli* and used as a vaccine antigen (Stranger-Jones et al., 2006). Because of the attributes of SpA in binding the Fc portion of IgG, a specific humoral immune response to Protein A could not be measured (Stranger-Jones et al., 2006). The inventors have overcome this obstacle through the generation of SpA-DQ9, 10K:D36,37A. BALB/c mice immunized with recombinant Protein A (SpA) displayed significant protection against intravenous challenge with *S. aureus* strains: a 2.951 log reduction in staphylococcal load as compared to the wild-type (P>0.005; Student's t-test) (Stranger-Jones et al., 2006). SpA specific antibodies may cause phagocytic clearance prior to abscess formation and/or impact the formation of the aforementioned eosinophilic barrier in abscesses that separate staphylococcal communities from immune cells since these do not form during infection with Protein A mutant strains. Each of the five SpA domains (i.e., domains formed from three helix bundles designated E, D, A, B, and C) exerts similar binding properties (Jansson et al., 1998). The solution and crystal structure of the domain D has been solved both with and without the Fe and VH3 (Fab) ligands, which bind Protein A in a non-competitive manner at distinct sites (Graille et al., 2000). Mutations in residues known to be involved in IgG binding (FS, Q9, Q10, S11, F13, Y14, L17, N28, I31 and K35) are also required for vWF A1 and TNFR1 binding (Cedergren et al., 1993; Gomez et al., 2006; O'Seaghdha et al., 2006), whereas residues important for the VH3 interaction (Q26, G29, F30, 533, D36, D37, Q40, N43, E47) appear to have no impact on the other binding activities (Graille et al., 2000; Jansson et al., 1998). SpA specifically targets a subset of B cells that express VH3 family related IgM on their surface, i.e., VH3 type B cell receptors (Roben et al., 1995). Upon interaction with SpA, these B cells proliferate and commit to apoptosis, leading to preferential and prolonged deletion of innate-like B lymphocytes (i.e., marginal zone B cells and follicular B2 cells)(Goodyear et al., 2003; Goodyear et al., 2004).

Molecular basis of Protein A surface display and function. Protein A is synthesized as a precursor in the bacterial cytoplasm and secreted via its YSIRK signal peptide at the cross wall, i.e. the cell division septum of staphylococci (FIG. 1) (DeDent et al., 2007; DeDent et al., 2008). Following cleavage of the C-terminal LPXTG sorting signal, Protein A is anchored to bacterial peptidoglycan cross-bridges by sortase A (Mazmanian et al., 1999; Schneewind

et al., 1995; Mazmanian et al., 2000). Protein A is the most abundant surface protein of staphylococci; the molecule is expressed by virtually all *S. aureus* strains (Cespedes et al., 2005; Kennedy et al., 2008; Said-Salim et al., 2003). Staphylococci turn over 15-20% of their cell wall per division cycle (Navarre and Schneewind, 1999). Murine hydrolases cleave the glycan strands and wall peptides of peptidoglycan, thereby releasing Protein A with its attached C-terminal cell wall disaccharide tetrapeptide into the extracellular medium (Ton-That et al., 1999). Thus, by physiological design, Protein A is both anchored to the cell wall and displayed on the bacterial surface but also released into surrounding tissues during host infection (Marraffini et al., 2006).

Protein A captures immunoglobulins on the bacterial surface and this biochemical activity enables staphylococcal escape from host innate and acquired immune responses (Jensen, 1958; Goodyear et al., 2004). Interestingly, region X of Protein A (Guss et al., 1984), a repeat domain that tethers the IgG binding domains to the LPXTG sorting signal/cell wall anchor, is perhaps the most variable portion of the staphylococcal genome (Said-Salim, 2003; Schneewind et al., 1992). Each of the five immunoglobulin binding domains of Protein A (SpA), formed from three helix bundles and designated E, D, A, B, and C, exerts similar structural and functional properties (Sjodahl, 1977; Jansson et al., 1998). The solution and crystal structure of the domain D has been solved both with and without the Fe and V_{H3} (Fab) ligands, which bind Protein A in a non-competitive manner at distinct sites (Graille 2000).

In the crystal structure complex, the Fab interacts with helix II and helix III of domain D via a surface composed of four VH region β -strands (Graille 2000). The major axis of helix II of domain D is approximately 500 to the orientation of the strands, and the interhelical portion of domain D is most proximal to the C0 strand. The site of interaction on Fab is remote from the Ig light chain and the heavy chain constant region. The interaction involves the following domain D residues: Asp-36 of helix II, Asp-37 and Gln-40 in the loop between helix II and helix III and several other residues (Graille 2000). Both interacting surfaces are composed predominantly of polar side chains, with three negatively charged residues on domain D and two positively charged residues on the 2A2 Fab buried by the interaction, providing an overall electrostatic attraction between the two molecules. Of the five polar interactions identified between Fab and domain D, three are between side chains. A salt bridge is formed between Arg-H19 and Asp-36 and two hydrogen bonds are made between Tyr-H59 and Asp-37 and between Asn-H82a and Ser-33. Because of the conservation of Asp-36 and Asp-37 in all five IgG binding domains of Protein A, the inventors mutated these residues.

The SpA-D sites responsible for Fab binding are structurally separate from the domain surface that mediates Fc γ binding. The interaction of Fc γ with domain D primarily involves residues in helix I with lesser involvement of helix II (Gouda et al., 1992; Deisenhofer, 1981). With the exception of the Gln-32, a minor contact in both complexes, none of the residues that mediate the Fc γ interaction are involved in Fab binding. To examine the spatial relationship between these different Ig-binding sites, the SpA domains in these complexes have been superimposed to construct a model of a complex between Fab, the SpA-domain D, and the Fc γ molecule. In this ternary model, Fab and Fc γ form a sandwich about opposite faces of the helix II without evidence of steric hindrance of either interaction. These findings illustrate how, despite its small size (i.e., 56-61 aa), an SpA domain can simultaneously display both activities, explain-

ing experimental evidence that the interactions of Fab with an individual domain are noncompetitive. Residues for the interaction between SpA-D and Fc γ are Gln-9 and Gln-10.

In contrast, occupancy of the Fc portion of IgG on the domain D blocks its interaction with vWF A1 and probably also TNFR1 (O'Scaghda et al., 2006). Mutations in residues essential for IgG Fc binding (F5, Q9, Q10, S11, F13, Y14, L17, N28, 131 and K35) are also required for vWF A1 and TNFR1 binding (O'Scaghda et al., 2006; Cedergren et al., 1993; Gomez et al., 2006), whereas residues critical for the VH3 interaction (Q26, G29, F30, S33, D36, D37, Q40, N43, E47) have no impact on the binding activities of IgG Fc, vWF A1 or TNFR1 (Jansson et al., 1998; Graille et al., 2000). The Protein A immunoglobulin Fab binding activity targets a subset of B cells that express V_H3 family related IgM on their surface, i.e., these molecules function as VH3 type B cell receptors (Roben et al., 1995). Upon interaction with SpA, these B cells rapidly proliferate and then commit to apoptosis, leading to preferential and prolonged deletion of innate-like B lymphocytes (i.e., marginal zone B cells and follicular B2 cells) (Goodyear and Silverman, 2004; Goodyear and Silverman, 2003). More than 40% of circulating B cells are targeted by the Protein A interaction and the V_H3 family represents the largest family of human B cell receptors to impart protective humoral responses against pathogens (Goodyear and Silverman, 2004; Goodyear and Silverman, 2003). Thus, Protein A functions analogously to staphylococcal superantigens (Roben et al., 1995), albeit that the latter class of molecules, for example SEB, TSST-1, TSST-2, form complexes with the T cell receptor to inappropriately stimulate host immune responses and thereby precipitating characteristic disease features of staphylococcal infections (Roben et al., 1995; Tiedemann et al., 1995). Together these findings document the contributions of Protein A in establishing staphylococcal infections and in modulating host immune responses.

In sum, Protein A domains can be viewed as displaying two different interfaces for binding with host molecules and any development of Protein A based vaccines must consider the generation of variants that do not perturb host cell signaling, platelet aggregation, sequestration of immunoglobulins or the induction of B cell proliferation and apoptosis. Such Protein A variants should also be useful in analyzing vaccines for the ability of raising antibodies that block the aforementioned SpA activities and occupy the five repeat domains at their dual binding interfaces. This goal is articulated and pursued here for the first time and methods are described in detail for the generation of Protein A variants that can be used as a safe vaccine for humans. To perturb IgG Fc γ , vWF A1 and TNFR1 binding, glutamine (Q) 9 and 10 [numbering derived from the SpA domain D as described in Uhlen et al., 1984] were mutated, and generated lysine substitutions for both glutamines with the expectation that these abolish the ligand attributes at the first binding interface. To perturb IgM Fab VH3 binding, aspartate (D) 36 and 37 were mutated, each of which is required for the association with the B cell receptor. D36 and D37 were both substituted with alanine. Q9,10K and D36,37A mutations are here combined in the recombinant molecule SpA-DQ9,10K:D36,37A and tested for the binding attributes of Protein A. Further, SpA-D and SpA-DQ9,10K:D36,37A are subjected to immunization studies in mice and rabbits and analyzed for [1] the production of specific antibodies (SpA-D Ab); [2] the ability of SpA-D Ab to block the association between Protein A and its four different ligands;

and, [3] the attributes of SpA-D Ab to generate protective immunity against staphylococcal infections. (See Examples section below).

B. Staphylococcal Coagulases

Coagulases are enzymes produced by *Staphylococcus* bacteria that convert fibrinogen to fibrin. Coa and vWh activate prothrombin without proteolysis (Friedrich et al., 2003). The coagulase prothrombin complex recognizes fibrinogen as a specific substrate, converting it directly into fibrin. The crystal structure of the active complex revealed binding of the D1 and D2 domains to prothrombin and insertion of its Ile1-Val² N-terminus into the Ile¹⁶ pocket, inducing a functional active site in the zymogen through conformational change (Friedrich et al., 2003). Exosite 1 of α -thrombin, the fibrinogen recognition site, and proexosite 1 on prothrombin are blocked by the D2 of Coa (Friedrich et al., 2003). Nevertheless, association of the tetrameric (Coa prothrombin)₂ complex binds fibrinogen at a new site with high affinity (Panizzi et al., 2006). This model explains the coagulant properties and efficient fibrinogen conversion by coagulase (Panizzi et al., 2006).

Fibrinogen is a large glycoprotein (Mr~340,000), formed by three pairs of A α -, B β -, and γ -chains covalently linked to form a "dimer of trimers," where A and B designate the fibrinopeptides released by thrombin cleavage (Panizzi et al., 2006). The elongated molecule folds into three separate domains, a central fragment E that contains the N-termini of all six chains and two flanking fragments D formed mainly by the C-termini of the B β - and γ -chains. These globular domains are connected by long triple-helical structures. Coagulase-prothrombin complexes, which convert human fibrinogen to the self-polymerizing fibrin, are not targeted by circulating thrombin inhibitors (Panizzi et al., 2006). Thus, staphylococcal coagulases bypass the physiological blood coagulation pathway.

All *S. aureus* strains secrete coagulase and vWbp (Bjerketorp et al., 2004; Field and Smith, 1945). Although early work reported important contributions of coagulase to the pathogenesis of staphylococcal infections (Ekstedt and Yotis, 1960; Smith et al., 1947), more recent investigations with molecular genetics tools challenged this view by observing no virulence phenotypes with endocarditis, skin abscess and mastitis models in mice (Moreillon et al., 1995; Phonimdaeng et al., 1990). Generating isogenic variants of *S. aureus* Newman, a fully virulent clinical isolate (Duthie et al., 1952), it is described herein that coa mutants indeed display virulence defects in a lethal bacteremia and renal abscess model in mice. In the inventors experience, *S. aureus* 8325-4 is not fully virulent and it is presumed that mutational lesions in this strain may not be able to reveal virulence defects in vivo. Moreover, antibodies raised against Coa or vWbp perturb the pathogenesis of *S. aureus* Newman infections to a degree mirroring the impact of gene deletions. Coa and vWbp contribute to staphylococcal abscess formation and lethal bacteremia and may also function as protective antigens in subunit vaccines.

Biochemical studies document the biological value of antibodies against Coa and vWbp. By binding to antigen and blocking its association with clotting factors, the antibodies prevent the formation of Coa prothrombin and vWbp prothrombin complexes. Passive transfer studies revealed protection of experimental animals against staphylococcal abscess formation and lethal challenge by Coa and vWbp antibodies. Thus, Coa and vWbp neutralizing antibodies generate immune protection against staphylococcal disease.

Earlier studies revealed a requirement of coagulase for resisting phagocytosis in blood (Smith et al., 1947) and the

inventors observed a similar phenotype for Δ coa mutants in lepirudin-treated mouse blood (see Example 3 below). As vWbp displays higher affinity for human prothrombin than the mouse counterpart, it is suspected the same may be true for Δ vWbp variants in human blood. Further, expression of Coa and vWbp in abscess lesions as well as their striking distribution in the eosinophilic pseudocapsule surrounding (staphylococcal abscess communities (SACs) or the peripheral fibrin wall, suggest that secreted coagulases contribute to the establishment of these lesions. This hypothesis was tested and, indeed, Δ coa mutants were defective in the establishment of abscesses. A corresponding test, blocking Coa function with specific antibodies, produced the same effect. Consequently, it is proposed that the clotting of fibrin is a critical event in the establishment of staphylococcal abscesses that can be targeted for the development of protective vaccines. Due to their overlapping function on human prothrombin, both Coa and vWbp are considered excellent candidates for vaccine development.

C. Other Staphylococcal Antigens

Research over the past several decades identified *S. aureus* exotoxins, surface proteins and regulatory molecules as important virulence factors (Foster, 2005; Mazmanian et al., 2001; Novick, 2003). Much progress has been achieved regarding the regulation of these genes. For example, staphylococci perform a bacterial census via the secretion of auto-inducing peptides that bind to a cognate receptor at threshold concentration, thereby activating phospho-relay reactions and transcriptional activation of many of the exotoxin genes (Novick, 2003). The pathogenesis of staphylococcal infections relies on these virulence factors (secreted exotoxins, exopolysaccharides, and surface adhesins). The development of staphylococcal vaccines is hindered by the multifaceted nature of staphylococcal invasion mechanisms. It is well established that live attenuated micro-organisms are highly effective vaccines; immune responses elicited by such vaccines are often of greater magnitude and of longer duration than those produced by non-replicating immunogens. One explanation for this may be that live attenuated strains establish limited infections in the host and mimic the early stages of natural infection. Embodiments of the invention are directed to compositions and methods including variant SpA polypeptides and peptides, as well as other immunogenic extracellular proteins, polypeptides, and peptides (including both secreted and cell surface proteins or peptides) of gram positive bacteria for the use in mitigating or immunizing against infection. In particular embodiments the bacteria is a *staphylococcus* bacteria. Extracellular proteins, polypeptides, or peptides include, but are not limited to secreted and cell surface proteins of the targeted bacteria.

The human pathogen *S. aureus* secretes EsxA and EsxB, two ESAT-6 like proteins, across the bacterial envelope (Burts et al., 2005, which is incorporated herein by reference). Staphylococcal esxA and esxB are clustered with six other genes in the order of transcription: esxA esaA essA esaB essB essC esaC esxB. The acronyms esa, ess, and esx stand for ESAT-6 secretion accessory, system, and extracellular, respectively, depending whether the encoded proteins play an accessory (esa) or direct (ess) role for secretion, or are secreted (esx) in the extracellular milieu. The entire cluster of eight genes is herein referred to as the Ess cluster. EsxA, esxB, essA, essB, and essC are all required for synthesis or secretion of EsxA and EsxB. Mutants that fail to produce EsxA, EsxB, and EssC display defects in the pathogenesis of *S. aureus* murine abscesses, suggesting that this specialized secretion system may be a general strategy of human bacterial pathogenesis. Secretion of non-WXG100

substrates by the ESX-1 pathway has been reported for several antigens including EspA, EspB, Rv3483c, and Rv3615c (Fortune et al., 2005; MacGurn et al., 2005; McLaughlin et al., 2007; Xu et al., 2007). The alternate ESX-5 pathway has also been shown to secrete both WXG100 and non-WXG100 proteins in pathogenic mycobacteria (Abdallah et al., 2007; Abdallah et al., 2006).

The *Staphylococcus aureus* Ess pathway can be viewed as a secretion module equipped with specialized transport components (Ess), accessory factors (Esa) and cognate secretion substrates (Esx). EssA, EssB and EssC are required for EsxA and EsxB secretion. Because EssA, EssB and EssC are predicted to be transmembrane proteins, it is contemplated that these proteins form a secretion apparatus. Some of the proteins in the ess gene cluster may actively transport secreted substrates (acting as motor) while others may regulate transport (regulator). Regulation may be achieved, but need not be limited to, transcriptional or post-translational mechanisms for secreted polypeptides, sorting of specific substrates to defined locations (e.g., extracellular medium or host cells), or timing of secretion events during infection. At this point, it is unclear whether all secreted Esx proteins function as toxins or contribute indirectly to pathogenesis.

Staphylococci rely on surface protein mediated-adhesion to host cells or invasion of tissues as a strategy for escape from immune defenses. Furthermore, *S. aureus* utilize surface proteins to sequester iron from the host during infection. The majority of surface proteins involved in staphylococcal pathogenesis carry C-terminal sorting signals, i.e., they are covalently linked to the cell wall envelope by sortase. Further, staphylococcal strains lacking the genes required for surface protein anchoring, i.e., sortase A and B, display a dramatic defect in the virulence in several different mouse models of disease. Thus, surface protein antigens represent a validated vaccine target as the corresponding genes are essential for the development of staphylococcal disease and can be exploited in various embodiments of the invention. The sortase enzyme superfamily are Gram-positive transpeptidases responsible for anchoring surface protein virulence factors to the peptidoglycan cell wall layer. Two sortase isoforms have been identified in *Staphylococcus aureus*, SrtA and SrtB. These enzymes have been shown to recognize a LPXTG motif in substrate proteins. The SrtB isoform appears to be important in heme iron acquisition and iron homeostasis, whereas the SrtA isoform plays a critical role in the pathogenesis of Gram-positive bacteria by modulating the ability of the bacterium to adhere to host tissue via the covalent anchoring of adhesins and other proteins to the cell wall peptidoglycan. In certain embodiments the SpA variants described herein can be used in combination with other staphylococcal proteins such as Coa, Eap, Ehb, Emp, EsaC, EsaB, EsxA, EsxB, Hla, SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, IsdC, SasF, vWbp, and/or vWh proteins.

Certain aspects of the invention include methods and compositions concerning proteinaceous compositions including polypeptides, peptides, or nucleic acid encoding SpA variant(s) and other staphylococcal antigens such as other proteins transported by the Ess pathway, or sortase substrates. These proteins may be modified by deletion, insertion, and/or substitution.

The Esx polypeptides include the amino acid sequence of Esx proteins from bacteria in the *Staphylococcus* genus. The Esx sequence may be from a particular *staphylococcus* species, such as *Staphylococcus aureus*, and may be from a particular strain, such as Newman. In certain embodiments, the EsxA sequence is SAV0282 from strain Mu50 (which is

the same amino acid sequence for Newman) and can be accessed using Genbank Accession Number Q99WU4 (gil68565539), which is hereby incorporated by reference. In other embodiments, the EsxB sequence is SAV0290 from strain Mu50 (which is the same amino acid sequence for Newman) and can be accessed using Genbank Accession Number Q99WT7 (gil68565532), which is hereby incorporated by reference. In further embodiments, other polypeptides transported by the Ess pathway may be used, the sequences of which may be identified by one of skill in the art using databases and internet accessible resources.

The sortase substrate polypeptides include, but are not limited to the amino acid sequence of SdrC, SdrD, SdrE, IsdA, IsdB, ClfA, ClfB, IsdC or SasF proteins from bacteria in the *Staphylococcus* genus. The sortase substrate polypeptide sequence may be from a particular *staphylococcus* species, such as *Staphylococcus aureus*, and may be from a particular strain, such as Newman. In certain embodiments, the SdrD sequence is from strain N315 and can be accessed using Genbank Accession Number NP_373773.1 (gil15926240), which is incorporated by reference. In other embodiments, the SdrE sequence is from strain N315 and can be accessed using Genbank Accession Number NP_373774.1 (gil15926241), which is incorporated by reference. In other embodiments, the IsdA sequence is SAV1130 from strain Mu50 (which is the same amino acid sequence for Newman) and can be accessed using Genbank Accession Number NP_371654.1 (gil15924120), which is incorporated by reference. In other embodiments, the IsdB sequence is SAV1129 from strain Mu50 (which is the same amino acid sequence for Newman) and can be accessed using Genbank Accession Number NP_371653.1 (gil15924119), which is incorporated by reference. In further embodiments, other polypeptides transported by the Ess pathway or processed by sortase may be used, the sequences of which may be identified by one of skill in the art using databases and internet accessible resources.

Examples of various proteins that can be used in the context of the present invention can be identified by analysis of database submissions of bacterial genomes, including but not limited to accession numbers NC_002951 (GI:57650036 and GenBank CP000046), NC_002758 (GI:57634611 and GenBank BA000017), NC_002745 (GI:29165615 and GenBank BA000018), NC_003923 (GI:21281729 and GenBank BA000033), NC_002952 (GI:49482253 and GenBank BX571856), NC_002953 (GI:49484912 and GenBank BX571857), NC_007793 (GI:87125858 and GenBank CP000255), NC_007795 (GI:87201381 and GenBank CP000253) each of which are incorporated by reference.

As used herein, a "protein" or "polypeptide" refers to a molecule comprising at least ten amino acid residues. In some embodiments, a wild-type version of a protein or polypeptide are employed, however, in many embodiments of the invention, a modified protein or polypeptide is employed to generate an immune response. The terms described above may be used interchangeably. A "modified protein" or "modified polypeptide" or a "variant" refers to a protein or polypeptide whose chemical structure, particularly its amino acid sequence, is altered with respect to the wild-type protein or polypeptide. In some embodiments, a modified/variant protein or polypeptide has at least one modified activity or function (recognizing that proteins or polypeptides may have multiple activities or functions). It is specifically contemplated that a modified/variant protein or polypeptide may be altered with respect to one activity or function yet retain a wild-type activity or function in other respects, such as immunogenicity.

In certain embodiments the size of a protein or polypeptide (wild-type or modified) may comprise, but is not limited to, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, 200, 210, 220, 230, 240, 250, 275, 300, 325, 350, 375, 400, 425, 450, 475, 500, 525, 550, 575, 600, 625, 650, 675, 700, 725, 750, 775, 800, 825, 850, 875, 900, 925, 950, 975, 1000, 1100, 1200, 1300, 1400, 1500, 1750, 2000, 2250, 2500 amino molecules or greater, and any range derivable therein, or derivative of a corresponding amino sequence described or referenced herein. It is contemplated that polypeptides may be mutated by truncation, rendering them shorter than their corresponding wild-type form, but also they might be altered by fusing or conjugating a heterologous protein sequence with a particular function (e.g., for targeting or localization, for enhanced immunogenicity, for purification purposes, etc.).

As used herein, an "amino molecule" refers to any amino acid, amino acid derivative, or amino acid mimic known in the art. In certain embodiments, the residues of the proteinaceous molecule are sequential, without any non-amino molecule interrupting the sequence of amino molecule residues. In other embodiments, the sequence may comprise one or more non-amino molecule moieties. In particular embodiments, the sequence of residues of the proteinaceous molecule may be interrupted by one or more non-amino molecule moieties.

Accordingly, the term "proteinaceous composition" encompasses amino molecule sequences comprising at least one of the 20 common amino acids in naturally synthesized proteins, or at least one modified or unusual amino acid.

Proteinaceous compositions may be made by any technique known to those of skill in the art, including (i) the expression of proteins, polypeptides, or peptides through standard molecular biological techniques, (ii) the isolation of proteinaceous compounds from natural sources, or (iii) the chemical synthesis of proteinaceous materials. The nucleotide as well as the protein, polypeptide, and peptide sequences for various genes have been previously disclosed, and may be found in the recognized computerized databases. One such database is the National Center for Biotechnology Information's Genbank and GenPept databases (on the World Wide Web at ncbi.nlm.nih.gov/). The coding regions for these genes may be amplified and/or expressed using the techniques disclosed herein or as would be known to those of ordinary skill in the art.

Amino acid sequence variants of SpA, coagulases and other polypeptides of the invention can be substitutional, insertional, or deletion variants. A variation in a polypeptide of the invention may affect 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, or more non-contiguous or contiguous amino acids of the polypeptide, as compared to wild-type. A variant can comprise an amino acid sequence that is at least 50%, 60%, 70%, 80%, or 90%, including all values and ranges there between, identical to any sequence provided or referenced herein, e.g., SEQ ID NO:2-8 or SEQ ID NO:11-30, A variant can include 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, or more substitute amino acids. A polypeptide processed or secreted by the Ess pathway or other surface proteins (see Table 1) or sortase substrates

from any *staphylococcus* species and strain are contemplated for use in compositions and methods described herein.

Deletion variants typically lack one or more residues of the native or wild-type protein. Individual residues can be deleted or a number of contiguous amino acids can be deleted. A stop codon may be introduced (by substitution or insertion) into an encoding nucleic acid sequence to generate a truncated protein. Insertional mutants typically involve the addition of material at a non-terminal point in the polypeptide. This may include the insertion of one or more residues. Terminal additions, called fusion proteins, may also be generated. These fusion proteins include multimers or concatamers of one or more peptide or polypeptide described or referenced herein.

Substitutional variants typically contain the exchange of one amino acid for another at one or more sites within the protein, and may be designed to modulate one or more properties of the polypeptide, with or without the loss of other functions or properties. Substitutions may be conservative, that is, one amino acid is replaced with one of similar shape and charge. Conservative substitutions are well known in the art and include, for example, the changes of: alanine to serine; arginine to lysine; asparagine to glutamine or histidine; aspartate to glutamate; cysteine to serine; glutamine to asparagine; glutamate to aspartate; glycine to proline; histidine to asparagine or glutamine; isoleucine to leucine or valine; leucine to valine or isoleucine; lysine to arginine; methionine to leucine or isoleucine; phenylalanine to tyrosine, leucine or methionine; serine to threonine; threonine to serine; tryptophan to tyrosine; tyrosine to tryptophan or phenylalanine; and valine to isoleucine or leucine. Alternatively, substitutions may be non-conservative such that a function or activity of the polypeptide is affected. Non-conservative changes typically involve substituting a residue with one that is chemically dissimilar, such as a polar or charged amino acid for a nonpolar or uncharged amino acid, and vice versa.

TABLE 2

Exemplary surface proteins of <i>S. aureus</i> strains.								
SAV #	SA#	Surface	MW2	Mu50	N315	Newman	MRSA252*	MSSA476*
SAV0111	SA0107	Spa	492	450	450	520	516	492
SAV2503	SA2291	FnBPA	1015	1038	1038	741	—	1015
SAV2502	SA2290	FnBPPB	943	961	961	677	965	957
SAV0811	SA0742	ClfA	946	935	989	933	1029	928
SAV2630	SA2423	ClfB	907	877	877	913	873	905
Np	Np	Cna	1183	—	—	—	1183	1183
SAV0561	SA0519	SdrC	955	953	953	947	906	957
SAV0562	SA0520	SdrD	1347	1385	1385	1315	—	1365
SAV0563	SA0521	SdrE	1141	1141	1141	1166	1137	1141
Np	Np	Pls	—	—	—	—	—	—
SAV2654	SA2447	SasA	2275	2271	2271	2271	1351	2275
SAV2160	SA1964	SasB	686	2481	2481	2481	2222	685
	SA1577	SasC	2186	213	2186	2186	2189	2186
SAV0134	SA0129	SasD	241	241	241	241	221	241
SAV1130	SA0977	SasE/IsdA	350	350	350	350	354	350
SAV2646	SA2439	SasF	635	635	635	635	627	635
SAV2496		SasG	1371	525	927	—	—	1371
SAV0023	SA0022	SasH	772	—	772	772	786	786
SAV1731	SA1552	SasI	895	891	891	891	534	895
SAV1129	SA0976	SasJ/IsdB	645	645	645	645	652	645
	SA2381	SasK	198	211	211	—	—	197
	Np	SasL	—	232	—	—	—	—
SAV1131	SA0978	IsdC	227	227	227	227	227	227

Proteins of the invention may be recombinant, or synthesized in vitro. Alternatively, a non-recombinant or recom-

binant protein may be isolated from bacteria. It is also contemplated that a bacteria containing such a variant may be implemented in compositions and methods of the invention. Consequently, a protein need not be isolated.

The term “functionally equivalent codon” is used herein to refer to codons that encode the same amino acid, such as the six codons for arginine or serine, and also refers to codons that encode biologically equivalent amino acids (see Table 2, below).

TABLE 3

Codon Table					
Amino Acids		Codons			
Alanine	Ala A	GCA	GCC	GCG	GCU
Cysteine	Cys C	UGC	UGU		
Aspartic acid	Asp D	GAC	GAU		
Glutamic acid	Glu E	GAA	GAG		
Phenylalanine	Phe F	UUC	UUU		
Glycine	Gly G	GGA	GGC	GGG	GGU
Histidine	His H	CAC	CAU		
Isoleucine	Ile I	AUA	AUC	AUU	
Lysine	Lys K	AAA	AAG		
Leucine	Leu L	UUA	UUG	CUA	CUC
Methionine	Met M	AUG			
Asparagine	Asn N	AAC	AAU		
Proline	Pro P	CCA	CCC	CCG	CCU
Glutamine	Gln Q	CAA	CAG		
Arginine	Arg R	AGA	AGG	CGA	CGC
Serine	Ser S	AGC	AGU	UCA	UCC
Threonine	Thr T	ACA	ACC	ACG	ACU
Valine	Val V	GUA	GUC	GUG	GUU
Tryptophan	Trp W	UGG			
Tyrosine	Tyr Y	UAC	UAU		

It also will be understood that amino acid and nucleic acid sequences may include additional residues, such as additional N- or C-terminal amino acids, or 5' or 3' sequences, respectively, and yet still be essentially as set forth in one of the sequences disclosed herein, so long as the sequence

meets the criteria set forth above, including the maintenance of biological protein activity (e.g., immunogenicity) where

protein expression is concerned. The addition of terminal sequences particularly applies to nucleic acid sequences that may, for example, include various non-coding sequences flanking either of the 5' or 3' portions of the coding region.

The following is a discussion based upon changing of the amino acids of a protein to create a variant polypeptide or peptide. For example, certain amino acids may be substituted for other amino acids in a protein structure with or without appreciable loss of interactive binding capacity with structures such as, for example, antigen-binding regions of antibodies or binding sites on substrate molecules. Since it is the interactive capacity and nature of a protein that defines that protein's functional activity, certain amino acid substitutions can be made in a protein sequence, and in its underlying DNA coding sequence, and nevertheless produce a protein with a desirable property. It is thus contemplated by the inventors that various changes may be made in the DNA sequences of genes.

It is contemplated that in compositions of the invention, there is between about 0.001 mg and about 10 mg of total polypeptide, peptide, and/or protein per ml. The concentration of protein in a composition can be about, at least about or at most about 0.001, 0.010, 0.050, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1.0, 1.5, 2.0, 2.5, 3.0, 3.5, 4.0, 4.5, 5.0, 5.5, 6.0, 6.5, 7.0, 7.5, 8.0, 8.5, 9.0, 9.5, 10.0 mg/ml or more (or any range derivable therein). Of this, about, at least about, or at most about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100% may be an SpA variant or a coagulase, and may be used in combination with other peptides or polypeptides, such as other bacterial peptides and/or antigens.

The present invention contemplates the administration of variant SpA polypeptides or peptides to effect a preventative therapy or therapeutic effect against the development of a disease or condition associated with infection by a *staphylococcus* pathogen.

In certain aspects, combinations of staphylococcal antigens are used in the production of an immunogenic composition that is effective at treating or preventing staphylococcal infection. Staphylococcal infections progress through several different stages. For example, the staphylococcal life cycle involves commensal colonization, initiation of infection by accessing adjoining tissues or the bloodstream, and/or anaerobic multiplication in the blood. The interplay between *S. aureus* virulence determinants and the host defense mechanisms can induce complications such as endocarditis, metastatic abscess formation, and sepsis syndrome. Different molecules on the surface of the bacterium are involved in different steps of the infection cycle. Combinations of certain antigens can elicit an immune response which protects against multiple stages of staphylococcal infection. The effectiveness of the immune response can be measured either in animal model assays and/or using an opsonophagocytic assay.

D. Polypeptides and Polypeptide Production

The present invention describes polypeptides, peptides, and proteins and immunogenic fragments thereof for use in various embodiments of the present invention. For example, specific polypeptides are assayed for or used to elicit an immune response. In specific embodiments, all or part of the proteins of the invention can also be synthesized in solution or on a solid support in accordance with conventional

techniques. Various automatic synthesizers are commercially available and can be used in accordance with known protocols. See, for example, Stewart and Young, (1984); Tam et al., (1983); Merrifield, (1986); and Barany and Merrifield (1979), each incorporated herein by reference.

Alternatively, recombinant DNA technology may be employed wherein a nucleotide sequence which encodes a peptide of the invention is inserted into an expression vector, transformed or transfected into an appropriate host cell and cultivated under conditions suitable for expression.

One embodiment of the invention includes the use of gene transfer to cells, including microorganisms, for the production and/or presentation of polypeptides or peptides. The gene for the polypeptide or peptide of interest may be transferred into appropriate host cells followed by culture of cells under the appropriate conditions. The generation of recombinant expression vectors, and the elements included therein, are well known in the art and briefly discussed herein. Alternatively, the protein to be produced may be an endogenous protein normally synthesized by the cell that is isolated and purified.

Another embodiment of the present invention uses autologous B lymphocyte cell lines, which are transfected with a viral vector that expresses an immunogen product, and more specifically, a protein having immunogenic activity. Other examples of mammalian host cell lines include, but are not limited to Vero and HeLa cells, other B- and T-cell lines, such as CEM, 721.221, H9, Jurkat, Raji, as well as cell lines of Chinese hamster ovary, W138, BHK, COS-7, 293, HepG2, 3T3, RIN and MDCK cells. In addition, a host cell strain may be chosen that modulates the expression of the inserted sequences, or that modifies and processes the gene product in the manner desired. Such modifications (e.g., glycosylation) and processing (e.g., cleavage) of protein products may be important for the function of the protein. Different host cells have characteristic and specific mechanisms for the post-translational processing and modification of proteins. Appropriate cell lines or host systems can be chosen to ensure the correct modification and processing of the foreign protein expressed.

A number of selection systems may be used including, but not limited to HSV thymidine kinase, hypoxanthine-guanine phosphoribosyltransferase, and adenine phosphoribosyltransferase genes, in tk-, hgprt- or aprt-cells, respectively. Also, anti-metabolite resistance can be used as the basis of selection: for dhfr, which confers resistance to trimethoprim and methotrexate; gpt, which confers resistance to mycophenolic acid; neo, which confers resistance to the aminoglycoside G418; and hygromycin, which confers resistance to hygromycin.

Animal cells can be propagated in vitro in two modes: as non-anchorage-dependent cells growing in suspension throughout the bulk of the culture or as anchorage-dependent cells requiring attachment to a solid substrate for their propagation (i.e., a monolayer type of cell growth).

Non-anchorage dependent or suspension cultures from continuous established cell lines are the most widely used means of large scale production of cells and cell products. However, suspension cultured cells have limitations, such as tumorigenic potential and lower protein production than adherent cells.

Where a protein is specifically mentioned herein, it is preferably a reference to a native or recombinant protein or optionally a protein in which any signal sequence has been removed. The protein may be isolated directly from the staphylococcal strain or produced by recombinant DNA techniques. Immunogenic fragments of the protein may be

incorporated into the immunogenic composition of the invention. These are fragments comprising at least 10 amino acids, 20 amino acids, 30 amino acids, 40 amino acids, 50 amino acids, or 100 amino acids, including all values and ranges there between, taken contiguously from the amino acid sequence of the protein. In addition, such immunogenic fragments are immunologically reactive with antibodies generated against the Staphylococcal proteins or with antibodies generated by infection of a mammalian host with Staphylococci. Immunogenic fragments also include fragments that when administered at an effective dose, (either alone or as a hapten bound to a carrier), elicit a protective or therapeutic immune response against Staphylococcal infection. In certain aspects it is protective against *S. aureus* and/or *S. epidermidis* infection. Such an immunogenic fragment may include, for example, the protein lacking an N-terminal leader sequence, and/or a transmembrane domain and/or a C-terminal anchor domain. In a preferred aspect the immunogenic fragment according to the invention comprises substantially all of the extracellular domain of a protein which has at least 80% identity, at least 85% identity, at least 90% identity, at least 95% identity, or at least 97-99% identity, including all values and ranges there between, to a sequence selected segment of a polypeptide described or referenced herein.

Also included in immunogenic compositions of the invention are fusion proteins composed of one or more Staphylococcal proteins, or immunogenic fragments of staphylococcal proteins. Such fusion proteins may be made recombinantly and may comprise one portion of at least 1, 2, 3, 4, 5, or 6 staphylococcal proteins or segments. Alternatively, a fusion protein may comprise multiple portions of at least 1, 2, 3, 4 or 5 staphylococcal proteins. These may combine different Staphylococcal proteins and/or multiples of the same protein or proten fragment, or immunogenic fragments in the same protein (forming a multimer or a concatamer). Alternatively, the invention also includes individual fusion proteins of Staphylococcal proteins or immunogenic fragments thereof, as a fusion protein with heterologous sequences such as a provider of T-cell epitopes or purification tags, for example: β -galactosidase, glutathione-S-transferase, green fluorescent proteins (GFP), epitope tags such as FLAG, myc tag, poly histidine, or viral surface proteins such as influenza virus haemagglutinin, or bacterial proteins such as tetanus toxoid, diphtheria toxoid, or CRM197.

II. NUCLEIC ACIDS

In certain embodiments, the present invention concerns recombinant polynucleotides encoding the proteins, polypeptides, peptides of the invention. The nucleic acid sequences for SpA, coagulases and other bacterial proteins are included, all of which are incorporated by reference, and can be used to prepare peptides or polypeptides.

As used in this application, the term "polynucleotide" refers to a nucleic acid molecule that either is recombinant or has been isolated free of total genomic nucleic acid. Included within the term "polynucleotide" are oligonucleotides (nucleic acids of 100 residues or less in length), recombinant vectors, including, for example, plasmids, cosmids, phage, viruses, and the like. Polynucleotides include, in certain aspects, regulatory sequences, isolated substantially away from their naturally occurring genes or protein encoding sequences. Polynucleotides may be single-stranded (coding or antisense) or double-stranded, and may be RNA, DNA (genomic, cDNA or synthetic), analogs

thereof, or a combination thereof. Additional coding or non-coding sequences may, but need not, be present within a polynucleotide.

In this respect, the term "gene," "polynucleotide," or "nucleic acid" is used to refer to a nucleic acid that encodes a protein, polypeptide, or peptide (including any sequences required for proper transcription, post-translational modification, or localization). As will be understood by those in the art, this term encompasses genomic sequences, expression cassettes, cDNA sequences, and smaller engineered nucleic acid segments that express, or may be adapted to express, proteins, polypeptides, domains, peptides, fusion proteins, and mutants. A nucleic acid encoding all or part of a polypeptide may contain a contiguous nucleic acid sequence of: 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, 150, 160, 170, 180, 190, 200, 210, 220, 230, 240, 250, 260, 270, 280, 290, 300, 310, 320, 330, 340, 350, 360, 370, 380, 390, 400, 410, 420, 430, 440, 441, 450, 460, 470, 480, 490, 500, 510, 520, 530, 540, 550, 560, 570, 580, 590, 600, 610, 620, 630, 640, 650, 660, 670, 680, 690, 700, 710, 720, 730, 740, 750, 760, 770, 780, 790, 800, 810, 820, 830, 840, 850, 860, 870, 880, 890, 900, 910, 920, 930, 940, 950, 960, 970, 980, 990, 1000, 1010, 1020, 1030, 1040, 1050, 1060, 1070, 1080, 1090, 1095, 1100, 1500, 2000, 2500, 3000, 3500, 4000, 4500, 5000, 5500, 6000, 6500, 7000, 7500, 8000, 9000, 10000, or more nucleotides, nucleosides, or base pairs, including all values and ranges therebetween, of a polynucleotide encoding one or more amino acid sequence described or referenced herein. It also is contemplated that a particular polypeptide may be encoded by nucleic acids containing variations having slightly different nucleic acid sequences but, nonetheless, encode the same or substantially similar protein (see Table 3 above).

In particular embodiments, the invention concerns isolated nucleic acid segments and recombinant vectors incorporating nucleic acid sequences that encode a variant SpA or coagulase. The term "recombinant" may be used in conjunction with a polynucleotide or polypeptide and generally refers to a polypeptide or polynucleotide produced and/or manipulated in vitro or that is a replication product of such a molecule.

In other embodiments, the invention concerns isolated nucleic acid segments and recombinant vectors incorporating nucleic acid sequences that encode a variant SpA or coagulase polypeptide or peptide to generate an immune response in a subject. In various embodiments the nucleic acids of the invention may be used in genetic vaccines.

The nucleic acid segments used in the present invention can be combined with other nucleic acid sequences, such as promoters, polyadenylation signals, additional restriction enzyme sites, multiple cloning sites, other coding segments, and the like, such that their overall length may vary considerably. It is therefore contemplated that a nucleic acid fragment of almost any length may be employed, with the total length preferably being limited by the ease of preparation and use in the intended recombinant nucleic acid protocol. In some cases, a nucleic acid sequence may encode a polypeptide sequence with additional heterologous coding sequences, for example to allow for purification of the polypeptide, transport, secretion, post-translational modification, or for therapeutic benefits such as targeting or efficacy. As discussed above, a tag or other heterologous polypeptide may be added to the modified polypeptide-encoding sequence, wherein "heterologous" refers to a polypeptide that is not the same as the modified polypeptide.

In certain other embodiments, the invention concerns isolated nucleic acid segments and recombinant vectors that

include within their sequence a contiguous nucleic acid sequence from SEQ ID NO:1 (SpA domain D) or SEQ ID NO:3 (SpA) or any other nucleic acid sequences encoding coagulases or other secreted virulence factors and/or surface proteins including proteins transported by the Ess pathway, processed by sortase, or proteins incorporated herein by reference.

In certain embodiments, the present invention provides polynucleotide variants having substantial identity to the sequences disclosed herein; those comprising at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% or higher sequence identity, including all values and ranges there between, compared to a polynucleotide sequence of this invention using the methods described herein (e.g., BLAST analysis using standard parameters).

The invention also contemplates the use of polynucleotides which are complementary to all the above described polynucleotides.

A. Vectors

Polypeptides of the invention may be encoded by a nucleic acid molecule comprised in a vector. The term "vector" is used to refer to a carrier nucleic acid molecule into which a heterologous nucleic acid sequence can be inserted for introduction into a cell where it can be replicated and expressed. A nucleic acid sequence can be "heterologous," which means that it is in a context foreign to the cell in which the vector is being introduced or to the nucleic acid in which is incorporated, which includes a sequence homologous to a sequence in the cell or nucleic acid but in a position within the host cell or nucleic acid where it is ordinarily not found. Vectors include DNAs, RNAs, plasmids, cosmids, viruses (bacteriophage, animal viruses, and plant viruses), and artificial chromosomes (e.g., YACs). One of skill in the art would be well equipped to construct a vector through standard recombinant techniques (for example Sambrook et al., 2001; Ausubel et al., 1996, both incorporated herein by reference). In addition to encoding a variant SpA polypeptide the vector can encode other polypeptide sequences such as a one or more other bacterial peptide, a tag, or an immunogenicity enhancing peptide. Useful vectors encoding such fusion proteins include pIN vectors (Inouye et al., 1985), vectors encoding a stretch of histidines, and pGEX vectors, for use in generating glutathione S-transferase (GST) soluble fusion proteins for later purification and separation or cleavage.

The term "expression vector" refers to a vector containing a nucleic acid sequence coding for at least part of a gene product capable of being transcribed. In some cases, RNA molecules are then translated into a protein, polypeptide, or peptide. Expression vectors can contain a variety of "control sequences," which refer to nucleic acid sequences necessary for the transcription and possibly translation of an operably linked coding sequence in a particular host organism. In addition to control sequences that govern transcription and translation, vectors and expression vectors may contain nucleic acid sequences that serve other functions as well and are described herein.

1. Promoters and Enhancers

A "promoter" is a control sequence. The promoter is typically a region of a nucleic acid sequence at which initiation and rate of transcription are controlled. It may contain genetic elements at which regulatory proteins and molecules may bind such as RNA polymerase and other transcription factors. The phrases "operatively positioned," "operatively linked," "under control," and "under transcriptional control" mean that a promoter is in a correct functional location and/or orientation in relation to a nucleic acid

sequence to control transcriptional initiation and expression of that sequence. A promoter may or may not be used in conjunction with an "enhancer," which refers to a cis-acting regulatory sequence involved in the transcriptional activation of a nucleic acid sequence.

Naturally, it may be important to employ a promoter and/or enhancer that effectively directs the expression of the DNA segment in the cell type or organism chosen for expression. Those of skill in the art of molecular biology generally know the use of promoters, enhancers, and cell type combinations for protein expression (see Sambrook et al., 2001, incorporated herein by reference). The promoters employed may be constitutive, tissue-specific, or inducible and in certain embodiments may direct high level expression of the introduced DNA segment under specified conditions, such as large-scale production of recombinant proteins or peptides.

Various elements/promoters may be employed in the context of the present invention to regulate the expression of a gene. Examples of such inducible elements, which are regions of a nucleic acid sequence that can be activated in response to a specific stimulus, include but are not limited to Immunoglobulin Heavy Chain (Banerji et al., 1983; Gilles et al., 1983; Grosschedl et al., 1985; Atchinson et al., 1986, 1987; Imler et al., 1987; Weinberger et al., 1984; Kiledjian et al., 1988; Porton et al.; 1990), Immunoglobulin Light Chain (Queen et al., 1983; Picard et al., 1984), T Cell Receptor (Luria et al., 1987; Winoto et al., 1989; Redondo et al.; 1990), HLA DQ α and/or DQ β (Sullivan et al., 1987), β Interferon (Goodbourn et al., 1986; Fujita et al., 1987; Goodbourn et al., 1988), Interleukin-2 (Greene et al., 1989), Interleukin-2 Receptor (Greene et al., 1989; Lin et al., 1990), MHC Class II 5 (Koch et al., 1989), MHC Class II HLA-DR α (Sherman et al., 1989), β -Actin (Kawamoto et al., 1988; Ng et al.; 1989), Muscle Creatine Kinase (MCK) (Jaynes et al., 1988; Horlick et al., 1989; Johnson et al., 1989), Prealbumin (Transthyretin) (Costa et al., 1988), Elastase I (Ornitz et al., 1987), Metallothionein (MTII) (Karin et al., 1987; Culotta et al., 1989), Collagenase (Pinkert et al., 1987; Angel et al., 1987), Albumin (Pinkert et al., 1987; Tronche et al., 1989, 1990), α -Fetoprotein (Godbout et al., 1988; Campere et al., 1989), γ -Globin (Bodine et al., 1987; Perez-Stable et al., 1990), β -Globin (Trudel et al., 1987), c-fos (Cohen et al., 1987), c-Ha-Ras (Triesman, 1986; Deschamps et al., 1985), Insulin (Edlund et al., 1985), Neural Cell Adhesion Molecule (NCAM) (Hirsh et al., 1990), α 1-Antitrypsin (Latimer et al., 1990), H2B (TH2B) Histone (Hwang et al., 1990), Mouse and/or Type I Collagen (Ripe et al., 1989), Glucose-Regulated Proteins (GRP94 and GRP78) (Chang et al., 1989), Rat Growth Hormone (Larsen et al., 1986), Human Serum Amyloid A (SAA) (Edbrooke et al., 1989), Troponin I (TN I) (Yutzey et al., 1989), Platelet-Derived Growth Factor (PDGF) (Pech et al., 1989), Duchenne Muscular Dystrophy (Klamut et al., 1990), SV40 (Banerji et al., 1981; Moreau et al., 1981; Sleight et al., 1985; Firak et al., 1986; Herr et al., 1986; Imbra et al., 1986; Kadesch et al., 1986; Wang et al., 1986; Ondek et al., 1987; Kuhl et al., 1987; Schaffner et al., 1988), Polyoma (Swartzendruber et al., 1975; Vasseur et al., 1980; Katinka et al., 1980, 1981; Tyndell et al., 1981; Dandolo et al., 1983; de Villiers et al., 1984; Hen et al., 1986; Satake et al., 1988; Campbell et al., 1988), Retroviruses (Kriegler et al., 1982, 1983; Levinson et al., 1982; Kriegler et al., 1983, 1984a, b, 1988; Bosze et al., 1986; Miksicsek et al., 1986; Celander et al., 1987; Thiesen et al., 1988; Celander et al., 1988; Choi et al., 1988; Reisman et al., 1989), Papilloma Virus (Campo et al., 1983; Lusky et al.,

1983; Spandidos and Wilkie, 1983; Spalholz et al., 1985; Lusky et al., 1986; Cripe et al., 1987; Gloss et al., 1987; Hirochika et al., 1987; Stephens et al., 1987), Hepatitis B Virus (Bulla et al., 1986; Jameel et al., 1986; Shaul et al., 1987; Spandau et al., 1988; Vannice et al., 1988), Human Immunodeficiency Virus (Muesing et al., 1987; Hauber et al., 1988; Jakobovits et al., 1988; Feng et al., 1988; Takebe et al., 1988; Rosen et al., 1988; Berkhout et al., 1989; Laspia et al., 1989; Sharp et al., 1989; Braddock et al., 1989), Cytomegalovirus (CMV) IE (Weber et al., 1984; Boshart et al., 1985; Foecking et al., 1986), Gibbon Ape Leukemia Virus (Holbrook et al., 1987; Quinn et al., 1989).

Inducible elements include, but are not limited to MT II-Phorbol Ester (TFA)/Heavy metals (Palmiter et al., 1982; Haslinger et al., 1985; Searle et al., 1985; Stuart et al., 1985; Imagawa et al., 1987; Karin et al., 1987; Angel et al., 1987b; McNeall et al., 1989); MMTV (mouse mammary tumor virus)-Glucocorticoids (Huang et al., 1981; Lee et al., 1981; Majors et al., 1983; Chandler et al., 1983; Lee et al., 1984; Ponta et al., 1985; Sakai et al., 1988); β -Interferon-poly(rI) x/poly(rc) (Tavernier et al., 1983); Adenovirus 5 E2-E1A (Imperiale et al., 1984); Collagenase-Phorbol Ester (TPA) (Angel et al., 1987a); Stromelysin-Phorbol Ester (TPA) (Angel et al., 1987b); SV40-Phorbol Ester (TPA) (Angel et al., 1987b); Murine MX Gene-Interferon, Newcastle Disease Virus (Hug et al., 1988); GRP78 Gene-A23187 (Resendez et al., 1988); α -2-Macroglobulin-IL-6 (Kunz et al., 1989); Vimentin-Serum (Rittling et al., 1989); MHC Class I Gene H-2kb-Interferon (Blonar et al., 1989); HSP70-E1A/SV40 Large T Antigen (Taylor et al., 1989, 1990a, 1990b); Proliferin-Phorbol Ester/TPA (Mordacq et al., 1989); Tumor Necrosis Factor-PMA (Hensel et al., 1989); and Thyroid Stimulating Hormone a Gene-Thyroid Hormone (Chatterjee et al., 1989).

The particular promoter that is employed to control the expression of peptide or protein encoding polynucleotide of the invention is not believed to be critical, so long as it is capable of expressing the polynucleotide in a targeted cell, preferably a bacterial cell. Where a human cell is targeted, it is preferable to position the polynucleotide coding region adjacent to and under the control of a promoter that is capable of being expressed in a human cell. Generally speaking, such a promoter might include either a bacterial, human or viral promoter.

In embodiments in which a vector is administered to a subject for expression of the protein, it is contemplated that a desirable promoter for use with the vector is one that is not down-regulated by cytokines or one that is strong enough that even if down-regulated, it produces an effective amount of a variant SpA for eliciting an immune response. Non-limiting examples of these are CMV IE and RSV LTR. Tissue specific promoters can be used, particularly if expression is in cells in which expression of an antigen is desirable, such as dendritic cells or macrophages. The mammalian MHC I and MHC II promoters are examples of such tissue-specific promoters.

2. Initiation Signals and Internal Ribosome Binding Sites (IRES)

A specific initiation signal also may be required for efficient translation of coding sequences. These signals include the ATG initiation codon or adjacent sequences. Exogenous translational control signals, including the ATG initiation codon, may need to be provided. One of ordinary skill in the art would readily be capable of determining this and providing the necessary signals.

In certain embodiments of the invention, the use of internal ribosome entry sites (IRES) elements are used to

create multigene, or polycistronic, messages. IRES elements are able to bypass the ribosome scanning model of 5' E methylated Cap dependent translation and begin translation at internal sites (Pelletier and Sonenberg, 1988; Macejak and Sarnow, 1991). IRES elements can be linked to heterologous open reading frames. Multiple open reading frames can be transcribed together, each separated by an IRES, creating polycistronic messages. Multiple genes can be efficiently expressed using a single promoter/enhancer to transcribe a single message (see U.S. Pat. Nos. 5,925,565 and 5,935,819, herein incorporated by reference).

3. Selectable and Screenable Markers

In certain embodiments of the invention, cells containing a nucleic acid construct of the present invention may be identified *in vitro* or *in vivo* by encoding a screenable or selectable marker in the expression vector. When transcribed and translated, a marker confers an identifiable change to the cell permitting easy identification of cells containing the expression vector. Generally, a selectable marker is one that confers a property that allows for selection. A positive selectable marker is one in which the presence of the marker allows for its selection, while a negative selectable marker is one in which its presence prevents its selection. An example of a positive selectable marker is a drug resistance marker.

B. Host Cells

As used herein, the terms "cell," "cell line," and "cell culture" may be used interchangeably. All of these terms also include their progeny, which is any and all subsequent generations. It is understood that all progeny may not be identical due to deliberate or inadvertent mutations. In the context of expressing a heterologous nucleic acid sequence, "host cell" refers to a prokaryotic or eukaryotic cell, and it includes any transformable organism that is capable of replicating a vector or expressing a heterologous gene encoded by a vector. A host cell can, and has been, used as a recipient for vectors or viruses. A host cell may be "transfected" or "transformed," which refers to a process by which exogenous nucleic acid, such as a recombinant protein-encoding sequence, is transferred or introduced into the host cell. A transformed cell includes the primary subject cell and its progeny.

Host cells may be derived from prokaryotes or eukaryotes, including bacteria, yeast cells, insect cells, and mammalian cells for replication of the vector or expression of part or all of the nucleic acid sequence(s). Numerous cell lines and cultures are available for use as a host cell, and they can be obtained through the American Type Culture Collection (ATCC), which is an organization that serves as an archive for living cultures and genetic materials (www.atcc.org).

C. Expression Systems

Numerous expression systems exist that comprise at least a part or all of the compositions discussed above. Prokaryote- and/or eukaryote-based systems can be employed for use with the present invention to produce nucleic acid sequences, or their cognate polypeptides, proteins and peptides. Many such systems are commercially and widely available.

The insect cell/baculovirus system can produce a high level of protein expression of a heterologous nucleic acid segment, such as described in U.S. Pat. Nos. 5,871,986, 4,879,236, both herein incorporated by reference, and which can be bought, for example, under the name MAXBAC® 2.0 from INVITROGEN® and BACPACK™ BACULOVIRUS EXPRESSION SYSTEM FROM CLONTECH®.

In addition to the disclosed expression systems of the invention, other examples of expression systems include STRATAGENE®'s COMPLETE CONTROL™ Inducible Mammalian Expression System, which involves a synthetic ecdysone-inducible receptor, or its pET Expression System, an *E. coli* expression system. Another example of an inducible expression system is available from INVITROGEN®, which carries the T-REX™ (tetracycline-regulated expression) System, an inducible mammalian expression system that uses the full-length CMV promoter. INVITROGEN® also provides a yeast expression system called the *Pichia methanolica* Expression System, which is designed for high-level production of recombinant proteins in the methylotrophic yeast *Pichia methanolica*. One of skill in the art would know how to express a vector, such as an expression construct, to produce a nucleic acid sequence or its cognate polypeptide, protein, or peptide.

III. POLYSACCHARIDES

The immunogenic compositions of the invention may further comprise capsular polysaccharides including one or more of PIA (also known as PNAG) and/or *S. aureus* Type V and/or type VIII capsular polysaccharide and/or *S. epidermidis* Type I, and/or Type II and/or Type III capsular polysaccharide.

A. PIA (PNAG)

It is now clear that the various forms of staphylococcal surface polysaccharides identified as PS/A, PIA and SAA are the same chemical entity—PNAG (Maira-Litran et al., 2004). Therefore the term PIA or PNAG encompasses all these polysaccharides or oligosaccharides derived from them.

PIA is a polysaccharide intercellular adhesin and is composed of a polymer of β -(1 \rightarrow 6)-linked glucosamine substituted with N-acetyl and O-succinyl constituents. This polysaccharide is present in both *S. aureus* and *S. epidermidis* and can be isolated from either source (Joyce et al., 2003; Maira-Litran et al., 2002). For example, PNAG may be isolated from *S. aureus* strain MN8m (WO04/43407). PIA isolated from *S. epidermidis* is an integral constituent of biofilm. It is responsible for mediating cell-cell adhesion and probably also functions to shield the growing colony from the host's immune response. The polysaccharide previously known as poly-N-succinyl- β -(1 \rightarrow 6)-glucosamine (PNSG) was recently shown not to have the expected structure since the identification of N-succinylation was incorrect (Maira-Litran et al., 2002). Therefore the polysaccharide formally known as PNSG and now found to be PNAG is also encompassed by the term PIA.

PIA (or PNAG) may be of different sizes varying from over 400 kDa to between 75 and 400 kDa to between 10 and 75 kDa to oligosaccharides composed of up to 30 repeat units (of β -(1 \rightarrow 6)-linked glucosamine substituted with N-acetyl and O-succinyl constituents). Any size of PIA polysaccharide or oligosaccharide may be used in an immunogenic composition of the invention, in one aspect the polysaccharide is over 40 kDa. Sizing may be achieved by any method known in the art, for instance by microfluidization, ultrasonic irradiation or by chemical cleavage (WO 03/53462, EP497524, EP497525). In certain aspects PIA (PNAG) is at least or at most 40-400 kDa, 40-300 kDa, 50-350 kDa, 60-300 kDa, 50-250 kDa and 60-200 kDa.

PIA (PNAG) can have different degree of acetylation due to substitution on the amino groups by acetate. PIA produced in vitro is almost fully substituted on amino groups (95-100%). Alternatively, a deacetylated PIA (PNAG) can be

used having less than 60%, 50%, 40%, 30%, 20%, 10% acetylation. Use of a deacetylated PIA (PNAG) is preferred since non-acetylated epitopes of PNAG are efficient at mediating opsonic killing of Gram positive bacteria, preferably *S. aureus* and/or *S. epidermidis*. In certain aspects, the PIA (PNAG) has a size between 40 kDa and 300 kDa and is deacetylated so that less than 60%, 50%, 40%, 30% or 20% of amino groups are acetylated.

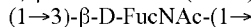
The term deacetylated PNAG (dPNAG) refers to a PNAG polysaccharide or oligosaccharide in which less than 60%, 50%, 40%, 30%, 20% or 10% of the amino groups are acetylated. In certain aspects, PNAG is deacetylated to form dPNAG by chemically treating the native polysaccharide. For example, the native PNAG is treated with a basic solution such that the pH rises to above 10. For instance the PNAG is treated with 0.1-5 M, 0.2-4 M, 0.3-3 M, 0.5-2 M, 0.75-1.5 M or 1 M NaOH, KOH or NH₄OH. Treatment is for at least 10 to 30 minutes, or 1, 2, 3, 4, 5, 10, 15 or 20 hours at a temperature of 20-100, 25-80, 30-60 or 30-50 or 35-45° C. dPNAG may be prepared as described in WO 04/43405.

The polysaccharide(s) can be conjugated or unconjugated to a carrier protein.

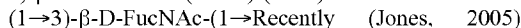
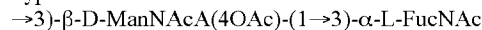
B. Type 5 and Type 8 polysaccharides from *S. aureus*

Most strains of *S. aureus* that cause infection in man contain either Type 5 or Type 8 polysaccharides. Approximately 60% of human strains are Type 8 and approximately 30% are Type 5. The structures of Type 5 and Type 8 capsular polysaccharide antigens are described in Moreau et al., (1990) and Fournier et al., (1984). Both have FucNAc in their repeat unit as well as ManNAcA which can be used to introduce a sulfhydryl group. The structures are:

Type 5

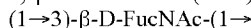
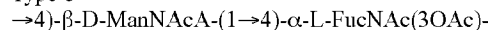


Type 8

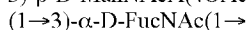


Recently (Jones, 2005) NMR spectroscopy revised the structures to:

Type 5



Type 8



Polysaccharides may be extracted from the appropriate strain of *S. aureus* using method well known to of skill in the art, See U.S. Pat. No. 6,294,177. For example, ATCC 12902 is a Type 5 *S. aureus* strain and ATCC 12605 is a Type 8 *S. aureus* strain.

Polysaccharides are of native size or alternatively may be sized, for instance by microfluidisation, ultrasonic irradiation, or by chemical treatment. The invention also covers oligosaccharides derived from the type 5 and 8 polysaccharides from *S. aureus*. The type 5 and 8 polysaccharides included in the immunogenic composition of the invention are preferably conjugated to a carrier protein as described below or are alternatively unconjugated. The immunogenic compositions of the invention alternatively contains either type 5 or type 8 polysaccharide.

C. *S. aureus* 336 Antigen

In an embodiment, the immunogenic composition of the invention comprises the *S. aureus* 336 antigen described in U.S. Pat. No. 6,294,177. The 336 antigen comprises β -linked hexosamine, contains no O-acetyl groups, and specifically binds to antibodies to *S. aureus* Type 336 deposited under ATCC 55804. In an embodiment, the 336 antigen is a polysaccharide which is of native size or

alternatively may be sized, for instance by microfluidisation, ultrasonic irradiation, or by chemical treatment. The invention also covers oligosaccharides derived from the 336 antigen. The 336 antigen can be unconjugated or conjugated to a carrier protein.

D. Type I, II and III Polysaccharides from *S. epidermidis*

Amongst the problems associated with the use of polysaccharides in vaccination, is the fact that polysaccharides per se are poor immunogens. It is preferred that the polysaccharides utilized in the invention are linked to a protein carrier which provide bystander T-cell help to improve immunogenicity. Examples of such carriers which may be conjugated to polysaccharide immunogens include the Diphtheria and Tetanus toxoids (DT, DT CRM197 and TT respectively), Keyhole Limpet Haemocyanin (KLH), and the purified protein derivative of Tuberculin (PPD), *Pseudomonas aeruginosa* exoprotein A (rEPA), protein D from *Haemophilus influenzae*, pneumolysin or fragments of any of the above. Fragments suitable for use include fragments encompassing T-helper epitopes. In particular the protein D fragment from *H. influenzae* will preferably contain the N-terminal $\frac{1}{3}$ of the protein. Protein D is an IgD-binding protein from *Haemophilus influenzae* (EP0 594 610 B1) and is a potential immunogen. In addition, staphylococcal proteins may be used as a carrier protein in the polysaccharide conjugates of the invention.

A carrier protein that would be particularly advantageous to use in the context of a staphylococcal vaccine is staphylococcal alpha toxoid. The native form may be conjugated to a polysaccharide since the process of conjugation reduces toxicity. Preferably genetically detoxified alpha toxins such as the His35Leu or His35Arg variants are used as carriers since residual toxicity is lower. Alternatively the alpha toxin is chemically detoxified by treatment with a cross-linking reagent, formaldehyde or glutaraldehyde. A genetically detoxified alpha toxin is optionally chemically detoxified, preferably by treatment with a cross-linking reagent, formaldehyde or glutaraldehyde to further reduce toxicity.

The polysaccharides may be linked to the carrier protein(s) by any known method (for example those methods described in U.S. Pat. Nos. 4,372,945, 4,474,757, and 4,356,170). Preferably, CDAP conjugation chemistry is carried out (see WO95/08348). In CDAP, the cyanating reagent 1-cyano-dimethylaminopyridinium tetrafluoroborate (CDAP) is preferably used for the synthesis of polysaccharide-protein conjugates. The cyanilation reaction can be performed under relatively mild conditions, which avoids hydrolysis of the alkaline sensitive polysaccharides. This synthesis allows direct coupling to a carrier protein.

Conjugation preferably involves producing a direct linkage between the carrier protein and polysaccharide. Optionally a spacer (such as adipic dihydride (ADH)) may be introduced between the carrier protein and the polysaccharide.

IV. IMMUNE RESPONSE AND ASSAYS

As discussed above, the invention concerns evoking or inducing an immune response in a subject against a variant SpA or coagulase peptide. In one embodiment, the immune response can protect against or treat a subject having, suspected of having, or at risk of developing an infection or related disease, particularly those related to staphylococci. One use of the immunogenic compositions of the invention is to prevent nosocomial infections by inoculating a subject prior to undergoing procedures in a hospital or other environment having an increased risk of infection.

A. Immunoassays

The present invention includes the implementation of serological assays to evaluate whether and to what extent an immune response is induced or evoked by compositions of the invention. There are many types of immunoassays that can be implemented. Immunoassays encompassed by the present invention include, but are not limited to, those described in U.S. Pat. No. 4,367,110 (double monoclonal antibody sandwich assay) and U.S. Pat. No. 4,452,901 (western blot). Other assays include immunoprecipitation of labeled ligands and immunocytochemistry, both in vitro and in vivo.

Immunoassays generally are binding assays. Certain preferred immunoassays are the various types of enzyme linked immunosorbent assays (ELISAs) and radioimmunoassays (RIA) known in the art. Immunohistochemical detection using tissue sections is also particularly useful. In one example, antibodies or antigens are immobilized on a selected surface, such as a well in a polystyrene microtiter plate, dipstick, or column support. Then, a test composition suspected of containing the desired antigen or antibody, such as a clinical sample, is added to the wells. After binding and washing to remove non specifically bound immune complexes, the bound antigen or antibody may be detected. Detection is generally achieved by the addition of another antibody, specific for the desired antigen or antibody, that is linked to a detectable label. This type of ELISA is known as a "sandwich ELISA." Detection also may be achieved by the addition of a second antibody specific for the desired antigen, followed by the addition of a third antibody that has binding affinity for the second antibody, with the third antibody being linked to a detectable label.

Competition ELISAs are also possible implementations in which test samples compete for binding with known amounts of labeled antigens or antibodies. The amount of reactive species in the unknown sample is determined by mixing the sample with the known labeled species before or during incubation with coated wells. The presence of reactive species in the sample acts to reduce the amount of labeled species available for binding to the well and thus reduces the ultimate signal. Irrespective of the format employed, ELISAs have certain features in common, such as coating, incubating or binding, washing to remove non specifically bound species, and detecting the bound immune complexes.

Antigen or antibodies may also be linked to a solid support, such as in the form of plate, beads, dipstick, membrane, or column matrix, and the sample to be analyzed is applied to the immobilized antigen or antibody. In coating a plate with either antigen or antibody, one will generally incubate the wells of the plate with a solution of the antigen or antibody, either overnight or for a specified period. The wells of the plate will then be washed to remove incompletely-adsorbed material. Any remaining available surfaces of the wells are then "coated" with a nonspecific protein that is antigenically neutral with regard to the test antisera. These include bovine serum albumin (BSA), casein, and solutions of milk powder. The coating allows for blocking of nonspecific adsorption sites on the immobilizing surface and thus reduces the background caused by nonspecific binding of antisera onto the surface.

B. Diagnosis of Bacterial Infection

In addition to the use of proteins, polypeptides, and/or peptides, as well as antibodies binding these polypeptides, proteins, and/or peptides, to treat or prevent infection as described above, the present invention contemplates the use of these polypeptides, proteins, peptides, and/or antibodies

in a variety of ways, including the detection of the presence of Staphylococci to diagnose an infection, whether in a patient or on medical equipment which may also become infected. In accordance with the invention, a preferred method of detecting the presence of infections involves the steps of obtaining a sample suspected of being infected by one or more staphylococcal bacteria species or strains, such as a sample taken from an individual, for example, from one's blood, saliva, tissues, bone, muscle, cartilage, or skin. Following isolation of the sample, diagnostic assays utilizing the polypeptides, proteins, peptides, and/or antibodies of the present invention may be carried out to detect the presence of staphylococci, and such assay techniques for determining such presence in a sample are well known to those skilled in the art and include methods such as radioimmunoassay, western blot analysis and ELISA assays. In general, in accordance with the invention, a method of diagnosing an infection is contemplated wherein a sample suspected of being infected with staphylococci has added to it the polypeptide, protein, peptide, antibody, or monoclonal antibody in accordance with the present invention, and staphylococci are indicated by antibody binding to the polypeptides, proteins, and/or peptides, or polypeptides, proteins, and/or peptides binding to the antibodies in the sample.

Accordingly, antibodies in accordance with the invention may be used for the prevention of infection from staphylococcal bacteria (i.e., passive immunization), for the treatment of an ongoing infection, or for use as research tools. The term "antibodies" as used herein includes monoclonal, polyclonal, chimeric, single chain, bispecific, simianized, and humanized or primatized antibodies as well as Fab fragments, such as those fragments which maintain the binding specificity of the antibodies, including the products of an Fab immunoglobulin expression library. Accordingly, the invention contemplates the use of single chains such as the variable heavy and light chains of the antibodies. Generation of any of these types of antibodies or antibody fragments is well known to those skilled in the art. Specific examples of the generation of an antibody to a bacterial protein can be found in U.S. Patent Application Pub. No. 20030153022, which is incorporated herein by reference in its entirety.

Any of the above described polypeptides, proteins, peptides, and/or antibodies may be labeled directly with a detectable label for identification and quantification of staphylococcal bacteria. Labels for use in immunoassays are generally known to those skilled in the art and include enzymes, radioisotopes, and fluorescent, luminescent and chromogenic substances, including colored particles such as colloidal gold or latex beads. Suitable immunoassays include enzyme-linked immunosorbent assays (ELISA).

C. Protective Immunity

In some embodiments of the invention, proteinaceous compositions confer protective immunity to a subject. Protective immunity refers to a body's ability to mount a specific immune response that protects the subject from developing a particular disease or condition that involves the agent against which there is an immune response. An immunogenically effective amount is capable of conferring protective immunity to the subject.

As used herein in the specification and in the claims section that follows, the term polypeptide or peptide refer to a stretch of amino acids covalently linked there amongst via peptide bonds. Different polypeptides have different functionalities according to the present invention. While according to one aspect, a polypeptide is derived from an immu-

nogen designed to induce an active immune response in a recipient, according to another aspect of the invention, a polypeptide is derived from an antibody which results following the elicitation of an active immune response in, for example, an animal, and which can serve to induce a passive immune response in the recipient. In both cases, however, the polypeptide is encoded by a polynucleotide according to any possible codon usage.

As used herein the phrase "immune response" or its equivalent "immunological response" refers to the development of a humoral (antibody mediated), cellular (mediated by antigen-specific T cells or their secretion products) or both humoral and cellular response directed against a protein, peptide, carbohydrate, or polypeptide of the invention in a recipient patient. Such a response can be an active response induced by administration of immunogen or a passive response induced by administration of antibody, antibody containing material, or primed T-cells. A cellular immune response is elicited by the presentation of polypeptide epitopes in association with Class I or Class II MHC molecules, to activate antigen-specific CD4 (+) T helper cells and/or CD8 (+) cytotoxic T cells. The response may also involve activation of monocytes, macrophages, NK cells, basophils, dendritic cells, astrocytes, microglia cells, eosinophils or other components of innate immunity. As used herein "active immunity" refers to any immunity conferred upon a subject by administration of an antigen.

As used herein "passive immunity" refers to any immunity conferred upon a subject without administration of an antigen to the subject. "Passive immunity" therefore includes, but is not limited to, administration of activated immune effectors including cellular mediators or protein mediators (e.g., monoclonal and/or polyclonal antibodies) of an immune response. A monoclonal or polyclonal antibody composition may be used in passive immunization for the prevention or treatment of infection by organisms that carry the antigen recognized by the antibody. An antibody composition may include antibodies that bind to a variety of antigens that may in turn be associated with various organisms. The antibody component can be a polyclonal antiserum. In certain aspects the antibody or antibodies are affinity purified from an animal or second subject that has been challenged with an antigen(s). Alternatively, an antibody mixture may be used, which is a mixture of monoclonal and/or polyclonal antibodies to antigens present in the same, related, or different microbes or organisms, such as gram-positive bacteria, gram-negative bacteria, including but not limited to *staphylococcus* bacteria.

Passive immunity may be imparted to a patient or subject by administering to the patient immunoglobulins (Ig) and/or other immune factors obtained from a donor or other non-patient source having a known immunoreactivity. In other aspects, an antigenic composition of the present invention can be administered to a subject who then acts as a source or donor for globulin, produced in response to challenge with the antigenic composition ("hyperimmune globulin"), that contains antibodies directed against *Staphylococcus* or other organism. A subject thus treated would donate plasma from which hyperimmune globulin would then be obtained, via conventional plasma-fractionation methodology, and administered to another subject in order to impart resistance against or to treat *staphylococcus* infection. Hyperimmune globulins according to the invention are particularly useful for immune-compromised individuals, for individuals undergoing invasive procedures or where time does not permit the individual to produce their own antibodies in response to vaccination. See U.S. Pat. Nos. 6,936,258,

6,770,278, 6,756,361, 5,548,066, 5,512,282, 4,338,298, and 4,748,018, each of which is incorporated herein by reference in its entirety, for exemplary methods and compositions related to passive immunity.

For purposes of this specification and the accompanying claims the terms "epitope" and "antigenic determinant" are used interchangeably to refer to a site on an antigen to which B and/or T cells respond or recognize. B-cell epitopes can be formed both from contiguous amino acids or noncontiguous amino acids juxtaposed by tertiary folding of a protein. Epitopes formed from contiguous amino acids are typically retained on exposure to denaturing solvents whereas epitopes formed by tertiary folding are typically lost on treatment with denaturing solvents. An epitope typically includes at least 3, and more usually, at least 5 or 8-10 amino acids in a unique spatial conformation. Methods of determining spatial conformation of epitopes include, for example, x-ray crystallography and 2-dimensional nuclear magnetic resonance. See, e.g., Epitope Mapping Protocols (1996). Antibodies that recognize the same epitope can be identified in a simple immunoassay showing the ability of one antibody to block the binding of another antibody to a target antigen. T-cells recognize continuous epitopes of about nine amino acids for CD8 cells or about 13-15 amino acids for CD4 cells. T cells that recognize the epitope can be identified by in vitro assays that measure antigen-dependent proliferation, as determined by ³H-thymidine incorporation by primed T cells in response to an epitope (Burke et al., 1994), by antigen-dependent killing (cytotoxic T lymphocyte assay, Tigges et al., 1996) or by cytokine secretion.

The presence of a cell-mediated immunological response can be determined by proliferation assays (CD4 (+) T cells) or CTL (cytotoxic T lymphocyte) assays. The relative contributions of humoral and cellular responses to the protective or therapeutic effect of an immunogen can be distinguished by separately isolating IgG and T-cells from an immunized syngeneic animal and measuring protective or therapeutic effect in a second subject.

As used herein and in the claims, the terms "antibody" or "immunoglobulin" are used interchangeably and refer to any of several classes of structurally related proteins that function as part of the immune response of an animal or recipient, which proteins include IgG, IgD, IgE, IgA, IgM and related proteins.

Under normal physiological conditions antibodies are found in plasma and other body fluids and in the membrane of certain cells and are produced by lymphocytes of the type denoted B cells or their functional equivalent. Antibodies of the IgG class are made up of four polypeptide chains linked together by disulfide bonds. The four chains of intact IgG molecules are two identical heavy chains referred to as H-chains and two identical light chains referred to as L-chains.

In order to produce polyclonal antibodies, a host, such as a rabbit or goat, is immunized with the antigen or antigen fragment, generally with an adjuvant and, if necessary, coupled to a carrier. Antibodies to the antigen are subsequently collected from the sera of the host. The polyclonal antibody can be affinity purified against the antigen rendering it monospecific.

Monoclonal antibodies can be produced by hyperimmunization of an appropriate donor with the antigen or ex-vivo by use of primary cultures of splenic cells or cell lines derived from spleen (Anavi, 1998; Huston et al., 1991; Johnson et al., 1991; Mernaugh et al., 1995).

As used herein and in the claims, the phrase "an immunological portion of an antibody" includes a Fab fragment of

an antibody, a Fv fragment of an antibody, a heavy chain of an antibody, a light chain of an antibody, a heterodimer consisting of a heavy chain and a light chain of an antibody, a variable fragment of a light chain of an antibody, a variable fragment of a heavy chain of an antibody, and a single chain variant of an antibody, which is also known as scFv. In addition, the term includes chimeric immunoglobulins which are the expression products of fused genes derived from different species, one of the species can be a human, in which case a chimeric immunoglobulin is said to be humanized. Typically, an immunological portion of an antibody competes with the intact antibody from which it was derived for specific binding to an antigen.

Optionally, an antibody or preferably an immunological portion of an antibody, can be chemically conjugated to, or expressed as, a fusion protein with other proteins. For purposes of this specification and the accompanying claims, all such fused proteins are included in the definition of antibodies or an immunological portion of an antibody.

As used herein the terms "immunogenic agent" or "immunogen" or "antigen" are used interchangeably to describe a molecule capable of inducing an immunological response against itself on administration to a recipient, either alone, in conjunction with an adjuvant, or presented on a display vehicle.

D. Treatment Methods

A method of the present invention includes treatment for a disease or condition caused by a *staphylococcus* pathogen. An immunogenic polypeptide of the invention can be given to induce an immune response in a person infected with *staphylococcus* or suspected of having been exposed to *staphylococcus*. Methods may be employed with respect to individuals who have tested positive for exposure to *staphylococcus* or who are deemed to be at risk for infection based on possible exposure.

In particular, the invention encompasses a method of treatment for staphylococcal infection, particularly hospital acquired nosocomial infections. The immunogenic compositions and vaccines of the invention are particularly advantageous to use in cases of elective surgery. Such patients will know the date of surgery in advance and could be inoculated in advance. The immunogenic compositions and vaccines of the invention are also advantageous to use to inoculate health care workers.

In some embodiments, the treatment is administered in the presence of adjuvants or carriers or other staphylococcal antigens. Furthermore, in some examples, treatment comprises administration of other agents commonly used against bacterial infection, such as one or more antibiotics.

The use of peptides for vaccination can require, but not necessarily, conjugation of the peptide to an immunogenic carrier protein, such as hepatitis B surface antigen, keyhole limpet hemocyanin, or bovine serum albumin. Methods for performing this conjugation are well known in the art.

V. VACCINE AND OTHER PHARMACEUTICAL COMPOSITIONS AND ADMINISTRATION

A. Vaccines

The present invention includes methods for preventing or ameliorating staphylococcal infections, particularly hospital acquired nosocomial infections. As such, the invention contemplates vaccines for use in both active and passive immunization embodiments. Immunogenic compositions, proposed to be suitable for use as a vaccine, may be prepared from immunogenic SpA polypeptide(s), such as a SpA domain D variant, or immunogenic coagulases. In other

embodiments SpA or coagulases can be used in combination with other secreted virulence proteins, surface proteins or immunogenic fragments thereof. In certain aspects, antigenic material is extensively dialyzed to remove undesired small molecular weight molecules and/or lyophilized for more ready formulation into a desired vehicle.

Other options for a protein/peptide-based vaccine involve introducing nucleic acids encoding the antigen(s) as DNA vaccines. In this regard, recent reports described construction of recombinant vaccinia viruses expressing either 10 contiguous minimal CTL epitopes (Thomson, 1996) or a combination of B cell, cytotoxic T-lymphocyte (CTL), and T-helper (Th) epitopes from several microbes (An, 1997), and successful use of such constructs to immunize mice for priming protective immune responses. Thus, there is ample evidence in the literature for successful utilization of peptides, peptide-pulsed antigen presenting cells (APCs), and peptide-encoding constructs for efficient *in vivo* priming of protective immune responses. The use of nucleic acid sequences as vaccines is exemplified in U.S. Pat. Nos. 5,958,895 and 5,620,896.

The preparation of vaccines that contain polypeptide or peptide sequence(s) as active ingredients is generally well understood in the art, as exemplified by U.S. Pat. Nos. 4,608,251; 4,601,903; 4,599,231; 4,599,230; 4,596,792; and 4,578,770, all of which are incorporated herein by reference. Typically, such vaccines are prepared as injectables either as liquid solutions or suspensions: solid forms suitable for solution in or suspension in liquid prior to injection may also be prepared. The preparation may also be emulsified. The active immunogenic ingredient is often mixed with excipients that are pharmaceutically acceptable and compatible with the active ingredient. Suitable excipients are, for example, water, saline, dextrose, glycerol, ethanol, or the like and combinations thereof. In addition, if desired, the vaccine may contain amounts of auxiliary substances such as wetting or emulsifying agents, pH buffering agents, or adjuvants that enhance the effectiveness of the vaccines. In specific embodiments, vaccines are formulated with a combination of substances, as described in U.S. Pat. Nos. 6,793,923 and 6,733,754, which are incorporated herein by reference.

Vaccines may be conventionally administered parenterally, by injection, for example, either subcutaneously or intramuscularly. Additional formulations which are suitable for other modes of administration include suppositories and, in some cases, oral formulations. For suppositories, traditional binders and carriers may include, for example, polyalkalene glycols or triglycerides: such suppositories may be formed from mixtures containing the active ingredient in the range of about 0.5% to about 10%, preferably about 1% to about 2%. Oral formulations include such normally employed excipients as, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate and the like. These compositions take the form of solutions, suspensions, tablets, pills, capsules, sustained release formulations or powders and contain about 10% to about 95% of active ingredient, preferably about 25% to about 70%.

The polypeptides and polypeptide-encoding DNA constructs may be formulated into a vaccine as neutral or salt forms. Pharmaceutically-acceptable salts include the acid addition salts (formed with the free amino groups of the peptide) and those that are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like.

Typically, vaccines are administered in a manner compatible with the dosage formulation, and in such amount as will be therapeutically effective and immunogenic. The quantity to be administered depends on the subject to be treated, including the capacity of the individual's immune system to synthesize antibodies and the degree of protection desired. Precise amounts of active ingredient required to be administered depend on the judgment of the practitioner. However, suitable dosage ranges are of the order of several hundred micrograms of active ingredient per vaccination. Suitable regimes for initial administration and booster shots are also variable, but are typified by an initial administration followed by subsequent inoculations or other administrations.

The manner of application may be varied widely. Any of the conventional methods for administration of a vaccine are applicable. These are believed to include oral application within a solid physiologically acceptable base or in a physiologically acceptable dispersion, parenterally, by injection and the like. The dosage of the vaccine will depend on the route of administration and will vary according to the size and health of the subject.

In certain instances, it will be desirable to have multiple administrations of the vaccine, e.g., 2, 3, 4, 5, 6 or more administrations. The vaccinations can be at 1, 2, 3, 4, 5, 6, 7, 8, to 5, 6, 7, 8, 9, 10, 11, 12 twelve week intervals, including all ranges there between. Periodic boosters at intervals of 1-5 years will be desirable to maintain protective levels of the antibodies. The course of the immunization may be followed by assays for antibodies against the antigens, as described in U.S. Pat. Nos. 3,791,932; 4,174,384 and 3,949,064.

1. Carriers

A given composition may vary in its immunogenicity. It is often necessary therefore to boost the host immune system, as may be achieved by coupling a peptide or polypeptide to a carrier. Exemplary and preferred carriers are keyhole limpet hemocyanin (KLH) and bovine serum albumin (BSA). Other albumins such as ovalbumin, mouse serum albumin, or rabbit serum albumin can also be used as carriers. Means for conjugating a polypeptide to a carrier protein are well known in the art and include glutaraldehyde, m-maleimidobencoyl-N-hydroxysuccinimide ester, carbodiimide, and bis-biazotized benzidine.

2. Adjuvants

The immunogenicity of polypeptide or peptide compositions can be enhanced by the use of non-specific stimulators of the immune response, known as adjuvants. Suitable adjuvants include all acceptable immunostimulatory compounds, such as cytokines, toxins, or synthetic compositions. A number of adjuvants can be used to enhance an antibody response against a variant SpA polypeptide or coagulase, or any other bacterial protein or combination contemplated herein. Adjuvants can (1) trap the antigen in the body to cause a slow release; (2) attract cells involved in the immune response to the site of administration; (3) induce proliferation or activation of immune system cells; or (4) improve the spread of the antigen throughout the subject's body.

Adjuvants include, but are not limited to, oil-in-water emulsions, water-in-oil emulsions, mineral salts, polynucleotides, and natural substances. Specific adjuvants that may be used include IL-1, IL-2, IL-4, IL-7, IL-12, γ -interferon, GM-CSF, BCG, aluminum salts, such as aluminum hydroxide or other aluminum compound, MDP compounds, such as thur-MDP and nor-MDP, CGP (MTP-PE), lipid A, and monophosphoryl lipid A (MPL). RIBI, which contains three components extracted from bacteria, MPL, trehalose dimy-

colate (TDM), and cell wall skeleton (CWS) in a 2% squalene/Tween 80 emulsion. MHC antigens may even be used. Others adjuvants or methods are exemplified in U.S. Pat. Nos. 6,814,971, 5,084,269, 6,656,462, each of which is incorporated herein by reference).

Various methods of achieving adjuvant affect for the vaccine includes use of agents such as aluminum hydroxide or phosphate (alum), commonly used as about 0.05 to about 0.1% solution in phosphate buffered saline, admixture with synthetic polymers of sugars (Carbopol®) used as an about 0.25% solution, aggregation of the protein in the vaccine by heat treatment with temperatures ranging between about 700 to about 101° C. for a 30-second to 2-minute period, respectively. Aggregation by reactivating with pepsin-treated (Fab) antibodies to albumin; mixture with bacterial cells (e.g., *C. parvum*), endotoxins or lipopolysaccharide components of Gram-negative bacteria; emulsion in physiologically acceptable oil vehicles (e.g., mannide monooleate (Aracel A)); or emulsion with a 20% solution of a perfluorocarbon (Fluosol-DA®) used as a block substitute may also be employed to produce an adjuvant effect.

Examples of and often preferred adjuvants include complete Freund's adjuvant (a non-specific stimulator of the immune response containing killed *Mycobacterium tuberculosis*), incomplete Freund's adjuvants, and aluminum hydroxide.

In some aspects, it is preferred that the adjuvant be selected to be a preferential inducer of either a Th1 or a Th2 type of response. High levels of Th1-type cytokines tend to favor the induction of cell mediated immune responses to a given antigen, while high levels of Th2-type cytokines tend to favor the induction of humoral immune responses to the antigen.

The distinction of Th1 and Th2-type immune response is not absolute. In reality an individual will support an immune response which is described as being predominantly Th1 or predominantly Th2. However, it is often convenient to consider the families of cytokines in terms of that described in murine CD4+ T cell clones by Mosmann and Coffman (Mosmann, and Coffman, 1989). Traditionally, Th1-type responses are associated with the production of the INF-7 and IL-2 cytokines by T-lymphocytes. Other cytokines often directly associated with the induction of Th1-type immune responses are not produced by T-cells, such as IL-12. In contrast, Th2-type responses are associated with the secretion of IL-4, IL-5, IL-6, IL-10.

In addition to adjuvants, it may be desirable to co-administer biologic response modifiers (BRM) to enhance immune responses. BRMs have been shown to upregulate T cell immunity or downregulate suppresser cell activity. Such BRMs include, but are not limited to, Cimetidine (CIM; 1200 mg/d) (Smith/Kline, PA); or low-dose Cyclophosphamide (CYP; 300 mg/m²) (Johnson/Mead, NJ) and cytokines such as γ -interferon, IL-2, or IL-12 or genes encoding proteins involved in immune helper functions, such as B-7.

B. Lipid Components and Moieties

In certain embodiments, the present invention concerns compositions comprising one or more lipids associated with a nucleic acid or a polypeptide/peptide. A lipid is a substance that is insoluble in water and extractable with an organic solvent. Compounds other than those specifically described herein are understood by one of skill in the art as lipids, and are encompassed by the compositions and methods of the present invention. A lipid component and a non-lipid may be attached to one another, either covalently or non-covalently.

A lipid may be a naturally occurring lipid or a synthetic lipid. However, a lipid is usually a biological substance.

Biological lipids are well known in the art, and include for example, neutral fats, phospholipids, phosphoglycerides, steroids, terpenes, lysolipids, glycosphingolipids, glucolipids, sulphatides, lipids with ether and ester-linked fatty acids and polymerizable lipids, and combinations thereof.

A nucleic acid molecule or a polypeptide/peptide, associated with a lipid may be dispersed in a solution containing a lipid, dissolved with a lipid, emulsified with a lipid, mixed with a lipid, combined with a lipid, covalently bonded to a lipid, contained as a suspension in a lipid or otherwise associated with a lipid. A lipid or lipid-poxvirus-associated composition of the present invention is not limited to any particular structure. For example, they may also simply be interspersed in a solution, possibly forming aggregates which are not uniform in either size or shape. In another example, they may be present in a bilayer structure, as micelles, or with a "collapsed" structure. In another non-limiting example, a lipofectamine(Gibco BRL)-poxvirus or Superfect (Qiagen)-poxvirus complex is also contemplated.

In certain embodiments, a composition may comprise about 1%, about 2%, about 3%, about 4%, about 5%, about 6%, about 7%, about 8%, about 9%, about 10%, about 11%, about 12%, about 13%, about 14%, about 15%, about 16%, about 17%, about 18%, about 19%, about 20%, about 21%, about 22%, about 23%, about 24%, about 25%, about 26%, about 27%, about 28%, about 29%, about 30%, about 31%, about 32%, about 33%, about 34%, about 35%, about 36%, about 37%, about 38%, about 39%, about 40%, about 41%, about 42%, about 43%, about 44%, about 45%, about 46%, about 47%, about 48%, about 49%, about 50%, about 51%, about 52%, about 53%, about 54%, about 55%, about 56%, about 57%, about 58%, about 59%, about 60%, about 61%, about 62%, about 63%, about 64%, about 65%, about 66%, about 67%, about 68%, about 69%, about 70%, about 71%, about 72%, about 73%, about 74%, about 75%, about 76%, about 77%, about 78%, about 79%, about 80%, about 81%, about 82%, about 83%, about 84%, about 85%, about 86%, about 87%, about 88%, about 89%, about 90%, about 91%, about 92%, about 93%, about 94%, about 95%, about 96%, about 97%, about 98%, about 99%, or any range therebetween, of a particular lipid, lipid type, or non-lipid component such as an adjuvant, antigen, peptide, polypeptide, sugar, nucleic acid or other material disclosed herein or as would be known to one of skill in the art. In a non-limiting example, a composition may comprise about 10% to about 20% neutral lipids, and about 33% to about 34% of a cerebroside, and about 1% cholesterol. In another non-limiting example, a liposome may comprise about 4% to about 12% terpenes, wherein about 1% of the micelle is specifically lycopene, leaving about 3% to about 11% of the liposome as comprising other terpenes; and about 10% to about 35% phosphatidyl choline, and about 1% of a non-lipid component. Thus, it is contemplated that compositions of the present invention may comprise any of the lipids, lipid types or other components in any combination or percentage range.

C. Combination Therapy

The compositions and related methods of the present invention, particularly administration of a secreted virulence factor or surface protein, including a variant SpA polypeptide or peptide, and/or other bacterial peptides or proteins to a patient/subject, may also be used in combination with the administration of traditional therapies. These include, but are not limited to, the administration of antibiotics such as streptomycin, ciprofloxacin, doxycycline, gentamycin, chloramphenicol, trimethoprim, sulfamethoxazole, ampicillin, tetracycline or various combinations of antibiotics.

In one aspect, it is contemplated that a polypeptide vaccine and/or therapy is used in conjunction with antibacterial treatment. Alternatively, the therapy may precede or follow the other agent treatment by intervals ranging from minutes to weeks. In embodiments where the other agents and/or a proteins or polynucleotides are administered separately, one would generally ensure that a significant period of time did not expire between the time of each delivery, such that the agent and antigenic composition would still be able to exert an advantageously combined effect on the subject. In such instances, it is contemplated that one may administer both modalities within about 12-24 h of each other or within about 6-12 h of each other. In some situations, it may be desirable to extend the time period for administration significantly, where several days (2, 3, 4, 5, 6 or 7) to several weeks (1, 2, 3, 4, 5, 6, 7 or 8) lapse between the respective administrations.

Various combinations may be employed, for example antibiotic therapy is "A" and the immunogenic molecule given as part of an immune therapy regime, such as an antigen, is

A/B/A B/A/B B/B/A A/A/B A/B/B B/A/A A/B/B/B B/A/B/B

B/B/B/A B/B/A/B A/A/B/B A/B/A/B A/B/B/A B/B/A/A B/A/B/A B/A/A/B A/A/A/B B/A/A/A A/B/A/A A/A/B/A

Administration of the immunogenic compositions of the present invention to a patient/subject will follow general protocols for the administration of such compounds, taking into account the toxicity, if any, of the SpA composition, or other compositions described herein. It is expected that the treatment cycles would be repeated as necessary. It also is contemplated that various standard therapies, such as hydration, may be applied in combination with the described therapy.

D. General Pharmaceutical Compositions

In some embodiments, pharmaceutical compositions are administered to a subject. Different aspects of the present invention involve administering an effective amount of a composition to a subject. In some embodiments of the present invention, staphylococcal antigens, members of the Ess pathway, including polypeptides or peptides of the Esa or Esx class, and/or members of sortase substrates may be administered to the patient to protect against infection by one or more *staphylococcus* pathogens. Alternatively, an expression vector encoding one or more such polypeptides or peptides may be given to a patient as a preventative treatment. Additionally, such compounds can be administered in combination with an antibiotic or an antibacterial. Such compositions will generally be dissolved or dispersed in a pharmaceutically acceptable carrier or aqueous medium.

In addition to the compounds formulated for parenteral administration, such as those for intravenous or intramuscular injection, other pharmaceutically acceptable forms include, e.g., tablets or other solids for oral administration; time release capsules; and any other form currently used, including creams, lotions, mouthwashes, inhalants and the like.

The active compounds of the present invention can be formulated for parenteral administration, e.g., formulated for injection via the intravenous, intramuscular, sub-cutaneous, or even intraperitoneal routes. The preparation of an aqueous composition that contains a compound or compounds that increase the expression of an MHC class I molecule will be known to those of skill in the art in light of the present disclosure. Typically, such compositions can be prepared as injectables, either as liquid solutions or suspensions; solid forms suitable for use to prepare solutions or

suspensions upon the addition of a liquid prior to injection can also be prepared; and, the preparations can also be emulsified.

Solutions of the active compounds as free base or pharmaceutically acceptable salts can be prepared in water suitably mixed with a surfactant, such as hydroxypropylcellulose. Dispersions can also be prepared in glycerol, liquid polyethylene glycols, and mixtures thereof and in oils. Under ordinary conditions of storage and use, these preparations contain a preservative to prevent the growth of microorganisms.

The pharmaceutical forms suitable for injectable use include sterile aqueous solutions or dispersions; formulations including sesame oil, peanut oil, or aqueous propylene glycol; and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions. In all cases the form must be sterile and must be fluid to the extent that it may be easily injected. It also should be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms, such as bacteria and fungi.

The proteinaceous compositions may be formulated into a neutral or salt form. Pharmaceutically acceptable salts, include the acid addition salts (formed with the free amino groups of the protein) and which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups can also be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, histidine, procaine and the like.

The carrier also can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and vegetable oils. The proper fluidity can be maintained, for example, by the use of a coating, such as lecithin, by the maintenance of the required particle size in the case of dispersion, and by the use of surfactants. The prevention of the action of microorganisms can be brought about by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

Sterile injectable solutions are prepared by incorporating the active compounds in the required amount in the appropriate solvent with various of the other ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the various sterilized active ingredients into a sterile vehicle which contains the basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum-drying and freeze-drying techniques, which yield a powder of the active ingredient, plus any additional desired ingredient from a previously sterile-filtered solution thereof.

Administration of the compositions according to the present invention will typically be via any common route. This includes, but is not limited to oral, nasal, or buccal administration. Alternatively, administration may be by orthotopic, intradermal, subcutaneous, intramuscular, intraperitoneal,

intranasal, or intravenous injection. In certain embodiments, a vaccine composition may be inhaled (e.g., U.S. Pat. No. 6,651,655, which is specifically incorporated by reference). Such compositions would normally be administered as pharmaceutically acceptable compositions that include physiologically acceptable carriers, buffers or other excipients. As used herein, the term “pharmaceutically acceptable” refers to those compounds, materials, compositions, and/or dosage forms which are, within the scope of sound medical judgment, suitable for contact with the tissues of human beings and animals without excessive toxicity, irritation, allergic response, or other problem complications commensurate with a reasonable benefit/risk ratio. The term “pharmaceutically acceptable carrier,” means a pharmaceutically acceptable material, composition or vehicle, such as a liquid or solid filler, diluent, excipient, solvent or encapsulating material, involved in carrying or transporting a chemical agent.

For parenteral administration in an aqueous solution, for example, the solution should be suitably buffered, if necessary, and the liquid diluent first rendered isotonic with sufficient saline or glucose. These particular aqueous solutions are especially suitable for intravenous, intramuscular, subcutaneous, and intraperitoneal administration. In this connection, sterile aqueous media which can be employed will be known to those of skill in the art in light of the present disclosure. For example, one dosage could be dissolved in isotonic NaCl solution and either added to hypodermoclysis fluid or injected at the proposed site of infusion, (see for example, Remington’s Pharmaceutical Sciences, 1990). Some variation in dosage will necessarily occur depending on the condition of the subject. The person responsible for administration will, in any event, determine the appropriate dose for the individual subject.

An effective amount of therapeutic or prophylactic composition is determined based on the intended goal. The term “unit dose” or “dosage” refers to physically discrete units suitable for use in a subject, each unit containing a predetermined quantity of the composition calculated to produce the desired responses discussed above in association with its administration, i.e., the appropriate route and regimen. The quantity to be administered, both according to number of treatments and unit dose, depends on the protection desired.

Precise amounts of the composition also depend on the judgment of the practitioner and are peculiar to each individual. Factors affecting dose include physical and clinical state of the subject, route of administration, intended goal of treatment (alleviation of symptoms versus cure), and potency, stability, and toxicity of the particular composition.

Upon formulation, solutions will be administered in a manner compatible with the dosage formulation and in such amount as is therapeutically or prophylactically effective. The formulations are easily administered in a variety of dosage forms, such as the type of injectable solutions described above.

E. In Vitro, Ex Vivo, or In Vivo Administration

As used herein, the term *in vitro* administration refers to manipulations performed on cells removed from or outside of a subject, including, but not limited to cells in culture. The

term *ex vivo* administration refers to cells which have been manipulated *in vitro*, and are subsequently administered to a subject. The term *in vivo* administration includes all manipulations performed within a subject.

In certain aspects of the present invention, the compositions may be administered either *in vitro*, *ex vivo*, or *in vivo*. In certain *in vitro* embodiments, autologous B-lymphocyte cell lines are incubated with a virus vector of the instant invention for 24 to 48 hours or with a variant SpA and/or cogoulase and/or any other composition described herein for two hours. The transduced cells can then be used for *in vitro* analysis, or alternatively for *ex vivo* administration. U.S. Pat. Nos. 4,690,915 and 5,199,942, both incorporated herein by reference, disclose methods for *ex vivo* manipulation of blood mononuclear cells and bone marrow cells for use in therapeutic applications.

F. Antibodies And Passive Immunization

Another aspect of the invention is a method of preparing an immunoglobulin for use in prevention or treatment of staphylococcal infection comprising the steps of immunizing a recipient or donor with the vaccine of the invention and isolating immunoglobulin from the recipient or donor. An immunoglobulin prepared by this method is a further aspect of the invention. A pharmaceutical composition comprising the immunoglobulin of the invention and a pharmaceutically acceptable carrier is a further aspect of the invention which could be used in the manufacture of a medicament for the treatment or prevention of staphylococcal disease. A method for treatment or prevention of staphylococcal infection comprising a step of administering to a patient an effective amount of the pharmaceutical preparation of the invention is a further aspect of the invention.

Inocula for polyclonal antibody production are typically prepared by dispersing the antigenic composition in a physiologically tolerable diluent such as saline or other adjuvants suitable for human use to form an aqueous composition. An immunostimulatory amount of inoculum is administered to a mammal and the inoculated mammal is then maintained for a time sufficient for the antigenic composition to induce protective antibodies.

The antibodies can be isolated to the extent desired by well known techniques such as affinity chromatography (Harlow and Lane, 1988). Antibodies can include antiserum preparations from a variety of commonly used animals, e.g. goats, primates, donkeys, swine, horses, guinea pigs, rats or man.

An immunoglobulin produced in accordance with the present invention can include whole antibodies, antibody fragments or subfragments. Antibodies can be whole immunoglobulins of any class (e.g., IgG, IgM, IgA, IgD or IgF), chimeric antibodies or hybrid antibodies with dual specificity to two or more antigens of the invention. They may also be fragments (e.g., F(ab')₂, Fab', Fab, Fv and the like) including hybrid fragments. An immunoglobulin also

includes natural, synthetic, or genetically engineered proteins that act like an antibody by binding to specific antigens to form a complex.

A vaccine of the present invention can be administered to a recipient who then acts as a source of immunoglobulin, produced in response to challenge from the specific vaccine. A subject thus treated would donate plasma from which hyperimmune globulin would be obtained via conventional plasma fractionation methodology. The hyperimmune globulin would be administered to another subject in order to impart resistance against or treat staphylococcal infection. Hyperimmune globulins of the invention are particularly useful for treatment or prevention of staphylococcal disease in infants, immune compromised individuals, or where treatment is required and there is no time for the individual to produce antibodies in response to vaccination.

An additional aspect of the invention is a pharmaceutical composition comprising two or more monoclonal antibodies (or fragments thereof, preferably human or humanised) reactive against at least two constituents of the immunogenic composition of the invention, which could be used to treat or prevent infection by Gram positive bacteria, preferably staphylococci, more preferably *S. aureus* or *S. epidermidis*. Such pharmaceutical compositions comprise monoclonal antibodies that can be whole immunoglobulins of any class, chimeric antibodies, or hybrid antibodies with specificity to two or more antigens of the invention. They may also be fragments (e.g., F(ab')₂, Fab', Fab, Fv and the like) including hybrid fragments.

Methods of making monoclonal antibodies are well known in the art and can include the fusion of splenocytes with myeloma cells (Kohler and Milstein, 1975; Harlow and Lane, 1988). Alternatively, monoclonal Fv fragments can be obtained by screening a suitable phage display library (Vaughan et al., 1998). Monoclonal antibodies may be humanized or part humanized by known methods.

VI. EXAMPLES

The following examples are given for the purpose of illustrating various embodiments of the invention and are not meant to limit the present invention in any fashion. One skilled in the art will appreciate readily that the present invention is well adapted to carry out the objects and obtain the ends and advantages mentioned, as well as those objects, ends and advantages inherent herein. The present examples, along with the methods described herein are presently representative of preferred embodiments, are exemplary, and are not intended as limitations on the scope of the invention. Changes therein and other uses which are encompassed within the spirit of the invention as defined by the scope of the claims will occur to those skilled in the art.

Example 1

Non-Toxicogenic Protein A Variants as Subunit Vaccines to Prevent *Staphylococcus Aureus* Infections

A. Results

An animal model for *S. aureus* infection BALB/c mice were infected by intravenous injection with 1×10^7 CFU of the human clinical isolate *S. aureus* Newman (Baba et al., 2007). Within 6 hours following infection, 99.999% of staphylococci disappeared from the blood stream and were distributed via the vasculature. Staphylococcal dissemina-

tion to peripheral tissues occurred rapidly, as the bacterial load in kidney and other peripheral organ tissues reached 1×10^5 CFU g⁻¹ within the first three hours. The staphylococcal load in kidney tissues increased by 1.5 log CFU within twenty-four hours. Forty-eight hours following infection, mice developed disseminated abscesses in multiple organs, detectable by light microscopy of hematoxylin-eosin stained, thin-sectioned kidney tissue. The initial abscess diameter was 524 μM (±65 μM); lesions were initially marked by an influx of polymorphonuclear leukocytes (PMNs) and harbored no discernable organization of staphylococci, most of which appeared to reside within PMNs. On day 5 of infection, abscesses increased in size and enclosed a central population of staphylococci, surrounded by a layer of eosinophilic, amorphous material and a large cuff of PMNs. Histopathology revealed massive necrosis of PMNs in proximity to the staphylococcal nidus at the center of abscess lesions as well as a mantle of healthy phagocytes. A rim of necrotic PMNs were observed at the periphery of abscess lesions, bordering eosinophilic, amorphous material that separates healthy renal tissue from lesions. Abscesses eventually reached a diameter of ≥1,524 μM on day 15 or 36. At later time intervals, the staphylococcal load was increased to 10^4 - 10^6 CFU g⁻¹ and growing abscess lesions migrated towards the organ capsule. Peripheral lesions were prone to rupture, thereby releasing necrotic material and staphylococci into the peritoneal cavity or the retroperitoneal space. These events resulted in bacteremia as well as a secondary wave of abscesses, eventually precipitating a lethal outcome.

To enumerate staphylococcal load in renal tissue, animals were killed, their kidneys excised and tissue homogenate spread on agar media for colony formation. On day 5 of infection, a mean of 1×10^6 CFU g⁻¹ renal tissue for *S. aureus* Newman was observed. To quantify abscess formation, kidneys were visually inspected, and each individual organ was given a score of one or zero. The final sum was divided by the total number of kidneys to calculate percent surface abscesses (Table 4). In addition, randomly chosen kidneys were fixed in formalin, embedded, thin sectioned, and stained with hematoxylin-eosin. For each kidney, four sagittal sections at 200 μM intervals were viewed by microscopy. The numbers of lesions were counted for each section and averaged to quantify the number of abscesses within the kidneys. *S. aureus* Newman caused 4.364 ± 0.889 abscesses per kidney, and surface abscesses were observed on 14 out of 20 kidneys (70%) (Table 4).

When examined by scanning electron microscopy, *S. aureus* Newman was located in tightly associated lawns at the center of abscesses. Staphylococci were contained by an amorphous pseudocapsule that separated bacteria from the cuff of abscesses leukocytes. No immune cells were observed in these central nests of staphylococci, however occasional red blood cells were located among the bacteria. Bacterial populations at the abscess center, designated staphylococcal abscess communities (SAC), appeared homogenous and coated by an electron-dense, granular material. The kinetics of the appearance of infectious lesions and the morphological attributes of abscesses formed by *S. aureus* Newman were similar to those observed following mouse infection with *S. aureus* USA300 (LAC), the current epidemic community-acquired methicillin-resistant *S. aureus* (CA-MRSA) clone in the United States (Diep et al., 2006).

TABLE 4

Genetic requirements for <i>S. aureus</i> Newman abscess formation in mice						
Genotype	Staphylococcal load in kidney tissue		Abscess formation in kidney tissue			
	^a log ₁₀ CFU g ⁻¹ tissue	^b Significance (P-value)	^c Reduction (log ₁₀ CFU g ⁻¹)	^d Surface abscesses (%)	^e Number of abscesses per kidney	^f Significance (P-value)
wild-type	6.141 ± 0.192	—	—	70	4.364 ± 0.889	—
AsrtA	4.095 ± 0.347	6.7 × 10 ⁻⁶	2.046	0	0.000 ± 0.000	0.0216
spa	5.137 ± 0.374	0.0144	1.004	13	0.375 ± 0.374	0.0356

^aMeans of staphylococcal load calculated as log₁₀ CFU g⁻¹ in homogenized renal tissues 5 days following infection in cohorts of fifteen BALB/c mice per challenge strain. Standard error of the means (±SEM) is indicated.

^bStatistical significance was calculated with the Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

^cReduction in bacterial load calculated as log₁₀ CFU g⁻¹.

^dAbscess formation in kidney tissues five days following infection was measured by macroscopic inspection (% positive) Histopathology of hematoxylin-eosin stained, thin sectioned kidneys from eight to ten animals; the average number of abscesses per kidney was recorded and averaged again for the final mean (±SEM).

^f Statistical significance was calculated with the Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

S. aureus Protein A (spa) mutants are avirulent and cannot form abscesses Sortase A is a transpeptidase that immobilizes nineteen surface proteins in the envelope of *S. aureus* strain Newman (Mazmanian et al., 1999; Mazmanian et al., 2000). Earlier work identified sortase A as a virulence factor in multiple animal model systems, however the contributions of this enzyme and its anchored surface proteins to abscess formation or persistence have not yet been revealed (Jonsson et al., 2002; Weiss et al., 2004). Compared to the wild-type parent (Baba et al., 2007), an isogenic srtA variant (Δ srtA) failed to form abscess lesions on either macroscopic or histopathology examination on days 2, 5, or 15. In mice infected with the srtA mutant, only 1×10⁴ CFU g⁻¹ was recovered from kidney tissue on day 5 of infection, which is a 2.046 log₁₀ CFU g⁻¹ reduction compared to the wild-type parent strain (P=6.73×10⁻⁶). A similar defect was observed for the srtA mutant of MRSA strain USA300 (data not shown). Scanning electron microscopy showed that srtA mutants were highly dispersed and often associated with leukocytes in otherwise healthy renal tissue. On day fifteen following infection, srtA mutants were cleared from renal tissues, a ≥3.5 log₁₀ CFU g⁻¹ reduction compared to the wild-type (Table 3). Thus, sortase A anchored surface proteins enable the formation of abscess lesions and the persistence of bacteria in host tissues, wherein staphylococci replicate as communities embedded in an extracellular matrix and shielded from surrounding leukocytes by an amorphous pseudocapsule.

Sortase A anchors a large spectrum of proteins with LPXTG motif sorting signals to the cell wall envelope, thereby providing for the surface display of many virulence factors (Mazmanian et al., 2002). To identify surface proteins required for staphylococcal abscess formation, *bursa aurealis* insertions were introduced in 5' coding sequences of genes that encode polypeptides with LPXTG motif proteins (Bae et al., 2004) and these mutations were transduced into *S. aureus* Newman. Mutations in the structural gene for Protein A (spa) reduced the staphylococcal load in infected mouse kidney tissues by 1.004 log₁₀ (P=0.0144). When analyzed for their ability to form abscesses in kidney tissues by histopathology, we observed that the spa mutants were unable to form abscesses as compared with the wild-type parent strain *S. aureus* Newman (wild-type *S. aureus* Newman 4.364±0.889 abscesses per kidney vs. the isogenic spa mutant with 0.375±0.374 lesions; P=0.0356).

Protein A blocks innate and adaptive immune responses. Studies identified Protein A as a critical virulence factor

during the pathogenesis of *S. aureus* infections. Earlier work demonstrated that Protein A impedes phagocytosis of staphylococci by binding the Fc component of immunoglobulin (Jensen 1958; Uhlen et al., 1984), activates platelet aggregation via the von Willebrand factor (Hartleib et al., 2000), functions as a B cell superantigen by capturing the F(ab)₂ region of VH3 bearing IgM (Roben et al., 1995), and, through its activation of TNFR1, can initiate staphylococcal pneumonia (Gomez et al., 2004). Due to the fact that Protein A captures immunoglobulin and displays toxic attributes, the possibility that this surface molecule may function as a vaccine in humans has not been rigorously pursued. The inventors demonstrate for the first time that Protein A variants no longer able to bind to immunoglobulins, vWF and TNFR-1 are removed of their toxigenic potential and are able to stimulate humoral immune responses that protect against staphylococcal disease.

Molecular basis of Protein A surface display and function. Protein A is synthesized as a precursor in the bacterial cytoplasm and secreted via its YSIRK signal peptide at the cross wall, i.e., the cell division septum of staphylococci (FIG. 1). (DeDent et al., 2007; DeDent et al., 2008). Following cleavage of the C-terminal LPXTG sorting signal, Protein A is anchored to bacterial peptidoglycan cross-bridges by sortase A (Schneewind et al., 1995; Mazmanian et al., 1999; Mazmanian et al., 2000). Protein A is the most abundant surface protein of staphylococci; the molecule is expressed by virtually all *S. aureus* strains (Said-Salim et al., 2003; Cespedes et al., 2005; Kennedy et al., 2008). Staphylococci turn over 15-20% of their cell wall per division cycle (Navarre and Schneewind 1999). Murine hydrolases cleave the glycan strands and wall peptides of peptidoglycan, thereby releasing Protein A with its attached C-terminal cell wall disaccharide tetrapeptide into the extracellular medium (Ton-That et al., 1999). Thus, by physiological design, Protein A is both anchored to the cell wall and displayed on the bacterial surface but also released into surrounding tissues during host infection (Marraffini et al., 2006).

Protein A captures immunoglobulins on the bacterial surface and this biochemical activity enables staphylococcal escape from host innate and acquired immune responses (Jensen 1958; Goodyear and Silverman 2004). Interestingly, region X of Protein A (Guss et al., 1984), a repeat domain that tethers the IgG binding domains to the LPXTG sorting signal/cell wall anchor, is perhaps the most variable portion of the staphylococcal genome (Schneewind et al., 1992; Said-Salim et al., 2003). Each of the five immunoglobulin

binding domains of Protein A (SpA), formed from three helix bundles and designated E, D, A, B, and C, exerts similar structural and functional properties (Sjodahl 1977; Jansson et al., 1998). The solution and crystal structure of domain D has been solved both with and without the Fc and V_H3 (Fab) ligands, which bind Protein A in a non-competitive manner at distinct sites (Graille et al., 2000).

In the crystal structure complex, the Fab interacts with helix II and helix III of domain D via a surface composed of four VH region β -strands (Graille et al., 2000). The major axis of helix II of domain D is approximately 500 to the orientation of the strands, and the interhelical portion of domain D is most proximal to the C0 strand. The site of interaction on Fab is remote from the Ig light chain and the heavy chain constant region. The interaction involves the following domain D residues: Asp-36 of helix II as well as Asp-37 and Gln-40 in the loop between helix II and helix III, in addition to several other residues with SpA-D (Graille et al., 2000). Both interacting surfaces are composed predominantly of polar side chains, with three negatively charged residues on domain D and two positively charged residues on the 2A2 Fab buried by the interaction, providing an overall electrostatic attraction between the two molecules. Of the five polar interactions identified between Fab and domain D, three are between side chains. A salt bridge is formed between Arg-H19 and Asp-36 and two hydrogen bonds are made between Tyr-H59 and Asp-37 and between Asn-H82a and Ser-33. Because of the conservation of Asp-36 and Asp-37 in all five IgG binding domains of Protein A, these residues were selected for mutagenesis.

The SpA-D sites responsible for Fab binding are structurally separate from the domain surface that mediates Fc γ binding. The interaction of Fc γ with domain B primarily involves residues in helix I with lesser involvement of helix II (Deisenhofer 1981; Gouda et al., 1992). With the exception of the Gln-32, a minor contact in both complexes, none of the residues that mediate the Fc γ interaction are involved in Fab binding. To examine the spatial relationship between these different Ig-binding sites, the SpA domains in these complexes have been superimposed to construct a model of a complex between Fab, the SpA-domain D, and the Fc molecule. In this ternary model, Fab and Fc γ form a sandwich about opposite faces of the helix II without evidence of steric hindrance of either interaction. These findings illustrate how, despite its small size (i.e., 56-61 aa), a SpA domain can simultaneously display both activities, explaining experimental evidence that the interactions of Fab with an individual domain are noncompetitive. Residues for the interaction between SpA-D and Fc γ are Gln-9 and Gln-10.

In contrast, occupancy of the Fc portion of IgG on the domain D blocks its interaction with vWF A1 and probably also TNFR1 (O'Seaghda et al., 2006). Mutations in residues essential for IgG Fc binding (F5, Q9, Q10, 511, F13, Y14, L17, N28, 131 and K35) are also required for vWF A1 and TNFR1 binding (Cedergren et al., 1993; Gómez et al., 2006; O'Seaghda et al. 2006), whereas residues critical for the V_H3 interaction (Q26, G29, F30, S33, D36, D37, Q40, N43, E47) have no impact on the binding activities of IgG Fc, vWF A1 or TNFR1 (Jansson et al., 1998; Graille et al., 2000). The Protein A immunoglobulin Fab binding activity targets a subset of B cells that express VH3 family related IgM on their surface, i.e. these molecules function as VH3 type B cell receptors (Roben et al., 1995). Upon interaction with SpA, these B cells rapidly proliferate and then commit to apoptosis, leading to preferential and prolonged deletion of innate-like B lymphocytes (i.e. marginal zone B cells and follicular B2 cells) (Goodyear and Silverman 2003; Good-

year and Silverman 2004). It is important to note that more than 40% of circulating B cells are targeted by the Protein A interaction and the VH3 family represents the largest family of human B cell receptors to impart protective humoral responses against pathogens (Goodyear and Silverman 2003; Goodyear and Silverman 2004). Thus, Protein A functions analogously to staphylococcal superantigens (Roben et al., 1995), albeit that the latter class of molecules, for example SEB, TSST-1, TSST-2, form complexes with the T cell receptor to inappropriately stimulate host immune responses and thereby precipitating characteristic disease features of staphylococcal infections (Roben et al., 1995; Tiedemann et al., 1995). Together these findings document the contributions of Protein A in establishing staphylococcal infections and in modulating host immune responses.

Non-toxicogenic variant of Protein A. The inventors have developed a non-toxicogenic variant of staphylococcal Protein A and, with this reagent in hand, aimed for the first time to measure the immune response of animals to Protein A immunization. Further, the inventors address whether immunization of animals with a non-toxicogenic variant of Protein A could generate immune responses that raise protective immunity against staphylococcal infection.

To perturb the IgG Fc, vWF A1 and TNFR1 binding activities of Protein A, glutamine (Q) residues 9 and 10 [the numbering here is derived from that established for the SpA domain D] were modified generating lysine or glycine substitutions for both glutamines with the expectation that these substitutions abolish the ion bonds formed between wild-type Protein A and its ligands. The added effect of the dual lysine substitutions may be that these positively charged residues institute a repellent charge for immunoglobulins. To perturb IgM Fab VH3 binding, the inventors selected the aspartate (D) residues 36 and 37 of SpA-D, each of which is required for the association of Protein A with the B cell receptor. D36 and D37 were both substituted with alanine. The Q9,10K and D36,37A mutations were combined in the recombinant molecule SpA-D_{Q9,10K;D36,37A} and examined for the binding attributes of Protein A.

In brief, the Protein A (spa) genomic sequence of *Staphylococcus aureus* N315 was PCR amplified with the primers (GCTGCACATATGGCGCAACACGATGAAGCTCAAC [5' primer] (SEQ ID NO:35) and AGTGGATCCT-TATGCTTTGTTAGCATCTGC [3' primer] (SEQ ID NO:36)), cloned into the pET15b vector (pYSJ1, codons 48-486) (Stranger-Jones, et al., 2006) and recombinant plasmid transformed into *E. coli* BL21(DE3) (Studier et al., 1990). The Protein A product derived from pYSJ1 harbors SpA residues 36-265 fused to the N-terminal His tag (MGSSHHHHHHSSGLVPRGS (SEQ ID NO:37)). Following IPTG inducible expression, recombinant N-terminal His6-tagged SpA was purified by affinity chromatography on Ni-NTA resin (Stranger-Jones et al., 2006). The domain D of SpA (SpA-D) was PCR amplified with a pair of specific primers (AACATATGTTCAACAAAGATCAACAAAGC [5' primer] (SEQ ID NO:38) and AAGGATCCAGATTCGTTAATTTTTAGC [3' primer] (SEQ ID NO:39)), sub-cloned into the pET15b vector (pHAN1, spa codons 212-261) and recombinant plasmid transformed into *E. coli* BL21(DE3) to express and purify recombinant N-terminal His6-tagged protein. To generate mutations in the SpA-D coding sequence, sets of two pairs of primers were synthesized (for D to A substitutions: CTTCAATCAAAGTCT-TAAAGCCGCCCAAGCCAAAGCACTAAC [5' primer] (SEQ ID NO:40) and GTTAGTGCTTTGGCTTGGGCGCGCTTTAAGACTTTGAATGAAG [3' primer] (SEQ ID NO:41); for Q to K substitutions CATATGTT-

CAACAAAGATAAAAAAGCGCCTTCTATGAAATC [5' primer] (SEQ ID NO:42) and GATTTCAT-AGAAGGCGCTTTTTTATCTTTGTTGAACATAIG [3' primer](SEQ ID NO:43); for Q to G substitutions CATATGTTCAACAAAGATGGAGGAAGCGCCTTC-
TATGAAATC [5' primer] (SEQ ID NO:44) and GATT-
CATAGAAGGCGCTTCTCCATCTTTGTTGAA-
CATATG' [3' primer] (SEQ ID NO:45). Primers were used for quick-change mutagenesis protocols. Following mutagenesis, DNA sequences were confirmed for each of the recombinant proteins: SpA, SpA-D and SpA-D_{Q9,10G;D36,37A} and SpA-D_{Q9,10K;D36,37A}. All proteins were purified from lysates of recombinant *E. coli* using Ni-NTA chromatography and subsequently dialyzed against PBS and stored at 4° C.

To measure binding of immunoglobulin to Protein A and its variants, 200 µg of purified protein was diluted into a 1 ml volume using column buffer (50 mM Tris-HCl, 150 mM NaCl, pH7.5) and then loaded onto a pre-equilibrated Ni-NTA column (1 ml bed volume). Columns were washed with 10 ml of column buffer. 200 µg of purified human IgG was diluted in a total volume of 1 ml column buffer and then applied to each of the columns charged with Protein A and its variants. The columns were subsequently washed with 5 ml wash buffer (10 mM imidazole in column buffer) and 5 ml column buffer. Protein samples were eluted with 2 ml elution buffer (500 mM imidazole in column buffer), fractions collected and aliquots subjected to SDS-PAGE gel electrophoresis, followed by Coomassie-Blue staining. As shown in FIG. 3, wild-type Protein A (SpA) and its SpA-domain D both retained immunoglobulin during chromatography. In contrast, the SpA-D_{Q9,10K;D36,37A} variant did not bind to immunoglobulin.

To quantify the binding of Protein A and its variants to the Fc portion of immunoglobulin and the VH3 domain of Fab, HRP conjugated human immunoglobulin G [hIgG], the Fc portion of human IgG [hFc] and the F(ab)₂ portion of human IgG [hF(ab)₂] as well as ELISA assays were used to quantify the relative amount binding to Protein A and its variants. The data in FIG. 4 demonstrate the binding of SpA and SpA-D to hIgG and hFc, whereas SpA-D_{Q9,10G;D36,37A} and SpA-D_{Q9,10K;D36,37A} displayed only background binding activities. SpA bound similar amounts of hFc and hF(ab)₂, however the binding of SpA-D to hF(ab)₂ was reduced compared to full length SpA. This result suggests that the presence of multiple IgG binding domains may cooperatively increase the ability of Protein A to bind to the B cell receptor. When compared with the reduced binding power of SpA-D for hF(ab)₂, of the two variants only SpA-D_{Q9,10K;D36,37A} displayed a significant reduction in the ability to bind the VH3 domain of immunoglobulin. To examine the toxigenic attributes of SpA-D and its variants, purified proteins were injected into mice, which were sacrificed after 4 hours to remove their spleens. Organ tissue was homogenized, capsular material removed and B cells stained with fluorescent CD19 antibodies. Following FACS analysis to quantify the abundance of B cells in splenic tissues, it was observed that SpA-D caused a 5% drop in the B cell count compared to a mock (PBS) control (FIG. 5). In contrast, SpA-D_{Q9,10K;D36,37A} did not cause a reduction in B-cell counts, indicating that the mutant molecule had lost its toxigenic attributes of stimulating B cell proliferation and death (FIG. 5). In summary, amino acid substitutions in the SpA-D residues Q9, Q10, D36, and D37 abolished the ability of Protein A domains to bind immunoglobulins or exert toxigenic functions in human and animal tissues.

Non-toxicigenic Protein A variants elicit vaccine protection. To test whether or not Protein A and its variants can function as vaccine antigens, SpA, SpA-D, SpA-D_{Q9,10K;D36,37A}, and SpA-D_{Q9,10G;D36,37A} were emulsified with complete or incomplete Freund's adjuvant and immunized 4 week old BALB/c mice on day 1 and day 11 with 50 µg of purified protein. Cohort of animals (n=5) were analyzed for humoral immune responses to immunization by bleeding the animals before (day 0) and after the immunization schedule (day 21). Table 5 indicates that immunized mice generated only a modest humoral immune response directed at wild-type Protein A or its SpA-D module, whereas the amount of antibody raised following immunization with SpA-D_{Q9,10K;D36,37A} or SpA-D_{Q9,10G;D36,37A} was increased four to five fold. Following intravenous challenge with 1×10⁷ CFU *S. aureus* Newman, animals were killed on day 4, their kidneys removed and either analyzed for staphylococcal load (by plating tissue homogenate on agar plates and enumerating colony forming units, CFU) or histopathology. As expected, mock (PBS) immunized mice (n=19) harbored 6.46 log₁₀ (±0.25) CFU in kidney tissue and infectious lesions were organized into 3.7 (±1.2) abscesses per organ (n=10)(Table 5). Immunization of animals with SpA led to a 2.51 log₁₀ CFU reduction on day 5 (P=0.0003) with 2.1 (±1.2) abscesses per organ. The latter data indicate that there was no significant reduction in abscess formation (P=0.35). Immunization with SpA-D generated similar results: a 2.03 log₁₀ CFU reduction on day 5 (P=0.0001) with 1.5 (0.8) abscesses per organ (P=0.15). In contrast, immunization with SpA-D_{Q9,10K;D36,37A} or SpA-D_{Q9,10G;D36,37A} created increased protection, with 3.07 log₁₀ and 3.03 log₁₀ CFU reduction on day 4, respectively (statistical significance P<0.0001 for both observations). Further, immunization with both SpA-D_{Q9,10K;D36,37A} and SpA-D_{Q9,10G;D36,37A} generated significant protection from staphylococcal abscess formation, as only 0.5 (±0.4) and 0.8 (±0.5) infectious lesions per organ (P=0.02 and P=0.04) were identified. Thus, immunization with non-toxicigenic Protein A variants generates increased humoral immune responses for Protein A and provides protective immunity against staphylococcal challenge. These data indicate that Protein A is an ideal candidate for a human vaccine that prevents *S. aureus* disease.

These exciting results have several implications for the design of a human vaccine. First, the generation of substitution mutations that affect the ability of the immunoglobulin binding domains of Protein A, either alone or in combination of two or more domains, can generate non-toxicigenic variants suitable for vaccine development. It seems likely that a combination of mutant IgG binding domains closely resembling the structure of Protein A can generate even better humoral immune responses as is reported here for the SpA-domain D alone. Further, a likely attribute of Protein A specific antibodies may be that the interaction of antigen binding sites with the microbial surface can neutralize the ability of staphylococci to capture immunoglobulins via their Fc portion or to stimulate the B cell receptor via the VH3 binding activities.

TABLE 5

Antigen	Bacterial load in kidney (n = number of mice)			Abscess formation in mice (n = number of mice)					
	^a log ₁₀ CFU g ⁻¹	^b Reduction	^c P value	IgG titer	^d Surface abscess	Reduction	^e Histopathology	Reduction	^f P value
Mock	6.46 ± 0.25 (n = 19)	—	—	<100	14/19 (70%)	—	3.7 ± 1.2 (n=10)	—	—
SpA	3.95 ± 0.56 (n = 20)	2.51	0.0003	1706 ± 370	10/20 (50%)	32%	2.1 ± 1.2 (n=10)	2.2	0.35
SpA-D	4.43 ± 0.41 (n = 18)	2.03	0.0001	381 ± 27	10/18 (55%)	25%	1.5 ± 0.8 (n=10)	2.2	0.15
SpA-D1	3.39 ± 0.50 (n = 19)	3.07	<0.0001	5600 ± 801	6/20 (30%)	59%	0.5 ± 0.4 (n=10)	3.2	0.02
SpA-D2	3.43 ± 0.46 (n = 19)	3.03	<0.0001	3980 ± 676	6/19 (32%)	57%	0.8 ± 0.5 (n=10)	2.9	0.04

^aMeans of staphylococcal load calculated as log₁₀ CFU g⁻¹ in homogenized renal tissues 4 days following infection in cohorts of 18 to 20 BALB/c mice. Standard error of the means (±SEM) is indicated.

^cStatistical significance was calculated with the Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

^bReduction in bacterial load calculated as log₁₀ CFU g⁻¹.

^dAbscess formation in kidney tissues four days following infection was measured by macroscopic inspection (% positive)

^eHistopathology of hematoxylin-eosin stained, thin sectioned kidneys from ten animals; the number of abscesses per kidney was recorded and averaged for the final mean (±SEM).

^fStatistical significance was calculated with the Students t-test and P-values recorded; P-values < 0.05 were deemed significant.

SpA-D1 and SpA-D2 represent SpA-D_{Q9,10K;D36,37A} and SpA-D_{Q9,10K;D36,37A}, respectively.

Vaccine protection in murine abscess, murine lethal infection, and murine pneumonia models. Three animal models have been established for the study of *S. aureus* infectious disease. These models are used here to examine the level of protective immunity provided via the generation of Protein A specific antibodies.

Murine abscess-BALB/c mice (24-day-old female, 8-10 mice per group, Charles River Laboratories, Wilmington, MA) are immunized by intramuscular injection into the hind leg with purified protein (Chang et al., 2003; Schneewind et al., 1992). Purified SpA, SpA-D or SpA-D_{Q9,10K;D36,37A} (50 µg protein) is administered on days 0 (emulsified 1:1 with complete Freund's adjuvant) and 11 (emulsified 1:1 with incomplete Freund's adjuvant). Blood samples are drawn by retroorbital bleeding on days 0, 11, and 20. Sera are examined by ELISA for IgG titers for specific SpA-D and SpA-D_{Q9,10K;D36,37A} binding activity. Immunized animals are challenged on day 21 by retroorbital injection of 100 µl of *S. aureus* Newman or *S. aureus* USA300 suspension (1×10⁷ cfu). For this, overnight cultures of *S. aureus* Newman are diluted 1:100 into fresh tryptic soy broth and grown for 3 h at 37° C. Staphylococci are centrifuged, washed twice, and diluted in PBS to yield an A₆₀₀ of 0.4 (1×10⁸ cfu per ml). Dilutions are verified experimentally by agar plating and colony formation. Mice are anesthetized by intraperitoneal injection of 80-120 mg of ketamine and 3-6 mg of xylazine per kilogram of body weight and infected by retroorbital injection. On day 5 or 15 following challenge, mice are euthanized by compressed CO₂ inhalation. Kidneys are removed and homogenized in 1% Triton X-100. Aliquots are diluted and plated on agar medium for triplicate determination of cfu. For histology, kidney tissue is incubated at room temperature in 10% formalin for 24 h. Tissues are embedded in paraffin, thin-sectioned, stained with hematoxylin-eosin, and examined by microscopy.

Murine lethal infection-BALB/c mice (24-day-old female, 8-10 mice per group, Charles River Laboratories, Wilmington, MA) are immunized by intramuscular injection

into the hind leg with purified SpA, SpA-D or SpA-D_{Q9,10K;D36,37A} (50 µg protein). Vaccine is administered on days 0 (emulsified 1:1 with complete Freund's adjuvant) and 11 (emulsified 1:1 with incomplete Freund's adjuvant). Blood samples are drawn by retroorbital bleeding on days 0, 11, and 20. Sera are examined by ELISA for IgG titers with specific SpA-D and SpA-D_{Q9,10K;D36,37A} binding activity. Immunized animals are challenged on day 21 by retroorbital injection of 100 µl of *S. aureus* Newman or *S. aureus* USA300 suspension (15×10⁷ cfu) (34). For this, overnight cultures of *S. aureus* Newman are diluted 1:100 into fresh tryptic soy broth and grown for 3 h at 37° C. Staphylococci are centrifuged, washed twice, diluted in PBS to yield an A₆₀₀ of 0.4 (1×10⁸ cfu per ml) and concentrated. Dilutions are verified experimentally by agar plating and colony formation. Mice are anesthetized by intraperitoneal injection of 80-120 mg of ketamine and 3-6 mg of xylazine per kilogram of body weight. Immunized animals are challenged on day 21 by intraperitoneal inject with 2×10¹⁰ cfu of *S. aureus* Newman or 3-10×10⁹ cfu of clinical *S. aureus* isolates. Animals are monitored for 14 days, and lethal disease is recorded.

Murine pneumonia model—*S. aureus* strains Newman or USA300 (LAC) are grown at 37° C. in tryptic soy broth/agar to OD₆₆₀ 0.5. 50-ml culture aliquots are centrifuged, washed in PBS, and suspended in 750 µl PBS for mortality studies (3-4×10⁸ CFU per 30-µl volume), or 1,250 µl PBS (2×10⁸ CFU per 30-µl volume) for bacterial load and histopathology experiments (2, 3). For lung infection, 7-wk-old C57BL/6J mice (The Jackson Laboratory) are anesthetized before inoculation of 30 µl of *S. aureus* suspension into the left nares. Animals are placed into the cage in a supine position for recovery and observed for 14 days. For active immunization, 4-wk-old mice receive 20 µg SpA-D or SpA-D_{Q9,10K;D36,37A} in CFA on day 0 via the i.m. route, followed by a boost with 20 µg SpA-D or SpA-D_{Q9,10K;D36,37A} in incomplete Freund's adjuvant (IFA) on day 10. Animals are challenged with *S. aureus* on day 21. Sera are collected

before immunization and on day 20 to assess specific antibody production. For passive immunization studies, 7-wk-old mice receive 100 μ l of either NRS (normal rabbit serum) or SpA-D-specific rabbit antisera via i.p. injection 24 h before challenge. To assess the pathological correlates of pneumonia, infected animals are killed via forced CO₂ inhalation before removal of both lungs. The right lung is homogenized for enumeration of lung bacterial load. The left lung is placed in 1% formalin and paraffin embedded, thin sectioned, stained with hematoxylin-eosin, and analyzed by microscopy.

Rabbit antibodies—Purified 200 μ g SpA-D or SpA-D_{Q9,10K;D36,37A} is used as an immunogen for the production of rabbit antisera. 200 μ g protein is emulsified with CFA for injection at day 0, followed by booster injections with 200 μ g protein emulsified with IFA on days 21 and 42. Rabbit antibody titers are determined by ELISA. Purified antibodies are obtained by affinity chromatography of rabbit serum on SpA-D or SpA-D_{Q9,10K;D36,37A} sepharose. The concentration of eluted antibodies is measured by absorbance at A₂₈₀ and specific antibody titers are determined by ELISA.

Active immunization with SpA-domain D variants.—To determine vaccine efficacy, animals are actively immunized with purified SpA-D or SpAD_{Q9,10K;D36,37A}. As a control, animals are immunized with adjuvant alone. Antibody titers against Protein A preparations are determined using SpA-D or SpA-D_{Q9,10K;D36,37A} as antigens; note that the SpA-D_{Q9,10K;D36,37A} variant cannot bind the Fc or Fab portion of IgG. Using infectious disease models described above, any reduction in bacterial load (murine abscess and pneumonia), histopathology evidence of staphylococcal disease (murine abscess and pneumonia) and protection from lethal disease (murine lethal challenge and pneumonia) is measured.

Passive immunization with affinity purified rabbit polyclonal antibodies generated against SpA-domain D variants. To determine protective immunity of Protein A specific rabbit antibodies, mice are passively immunized with 5 mg/kg of purified SpA-D or SpA-D_{Q9,10K;D36,37A} derived rabbit antibodies. Both of these antibody preparations are purified by affinity chromatography using immobilized SpA-D or SpA-D_{Q9,10K;D36,37A}. As a control, animals are passively immunized with rV10 antibodies (a plague protective antigen that has no impact on the outcome of staphylococcal infections). Antibody titers against all Protein A preparations are determined using SpA-D_{Q9,10K;D36,37A} as an antigen, as this variant cannot bind the Fc or Fab portion of IgG. Using the infectious disease models described above, the reduction in bacterial load (murine abscess and pneumonia), histopathology evidence of staphylococcal disease (murine abscess and pneumonia), and the protection from lethal disease (murine lethal challenge and pneumonia) is measured.

Example 2

Non-Toxicogenic Protein A Vaccine for Methicillin-Resistant *Staphylococcus Aureus* Infection

Clinical isolates of *S. aureus* express protein A (Shopsin et al., 1999, whose primary translational product is comprised of an N-terminal signal peptide (DeDent et al., 2008), five Ig-BDs (designated E, D, A, B and C)(Sjodahl, 1977), region X with variable repeats of an eight residue peptide (Guss et al., 1984), and C-terminal sorting signal for the cell wall anchoring of SpA (Schneewind et al., 1992; Schneewind et al., 1995) (FIG. 6). Guided by amino acid homology

(Uhlen et al., 1984), the triple α -helical bundle structure of IgBDs (Deisenhofer et al., 1978; Deisenhofer et al., 1981) and their atomic interactions with Fab V_H3 (Graille et al., 2000) or Fc γ (Gouda et al., 1998), glutamine 9 and 10 were selected as well as aspartate 36 and 37 as critical for the association of SpA with antibodies or B cell receptor, respectively. Substitutions Gln9Lys, Gln10Lys, Asp36Ala and Asp37Ala were introduced into the D domain to generate SpA-D_{KKAA} (FIG. 6). The ability of isolated SpA-D or SpA-D_{KKAA} to bind human IgG was analyzed by affinity chromatography (FIG. 6). Polyhistidine tagged SpA-D as well as full-length SpA retained human IgG on Ni-NTA, whereas SpA-D_{KKAA} and a negative control (SrtA) did not (FIG. 6). A similar result was observed with von Willebrand factor (Hartleib et al., 2000), which, along with tumor necrosis factor receptor 1 (TNFR1)(Gomez et al., 2004), can also bind protein A via glutamine 9 and 10 (FIG. 6). Human immunoglobulin encompasses 60-70% V_H3-type IgG. The inventors distinguish between Fc domain and B cell receptor activation of Igs and measured association of human Fc γ and F(ab)₂ fragments, both of which bound to full-length SpA or SpA-D, but not to SpA-D_{KKAA} (FIG. 6). Injection of SpA-D into the peritoneal cavity of mice resulted in B cell expansion followed by apoptotic collapse of CD19+ lymphocytes in spleen tissue of BALB/c mice (Goodyear and Silverman, 2003)(FIG. 6). B cell superantigen activity was not observed following injection with SpA-D_{KKAA}, and TUNEL-staining of splenic tissue failed to detect the increase in apoptotic cells that follows injection of SpA or SpA-D (FIG. 6).

Naive six week old BALB/c mice were injected with 50 μ g each of purified SpA, SpA-D or SpA-D_{KKAA} emulsified in CFA and boosted with the same antigen emulsified in IFA. In agreement with the hypothesis that SpA-D promotes the apoptotic collapse of activated clonal B cell populations, the inventors observed a ten-fold higher titer of SpA-D_{KKAA} specific antibodies following immunization of mice with the non-toxicogenic variant as compared to the B cell superantigen (SpA-D vs. SpA-D_{KKAA} P<0.0001, Table 6). Antibody titers raised by immunization with full-length SpA were higher than those elicited by SpA-D (P=0.0022), which is likely due to the larger size and reiterative domain structure of this antigen (Table 6). Nevertheless, even SpA elicited lower antibody titers than SpA-D_{KKAA} (P=0.0003), which encompasses only 50 amino acids of protein A (520 residues, SFQ ID NO:33). Immunized mice were challenged by intravenous inoculation with *S. aureus* Newman and the ability of staphylococci to seed abscesses in renal tissues was examined by necropsy four days after challenge. In homogenized renal tissue of mock (PBS/adjuvant) immunized mice, an average staphylococcal load of 6.46 log₁₀ CFU g⁻¹ was enumerated (Table 6). Immunization of mice with SpA or SpA-D led to a reduction in staphylococcal load, however SpA-D_{KKAA} vaccinated animals displayed an even greater, 3.07 log₁₀ CFU g⁻¹ reduction of *S. aureus* Newman in renal tissues (P<0.0001, Table 6). Abscess formation in kidneys was analyzed by histopathology (FIG. 7). Mock immunized animals harbored an average of 3.7 (\pm 1.2) abscesses per kidney (Table 6). Vaccination with SpA-D_{KKAA} reduced the average number of abscesses to 0.5 (\pm 0.4)(P=0.0204), whereas immunization with SpA or SpA-D did not cause a significant reduction in the number of abscess lesions (Table 6). Lesions from SpA-D_{KKAA} vaccinated animals were smaller in size, with fewer infiltrating PMNs and characteristically lacked staphylococcal abscess communities (Cheng et al., 2009)(FIG. 7). Abscesses in animals that had been immunized with SpA or SpA-D displayed the same overall structure of lesions in mock immunized animals (FIG. 7).

The inventors examined whether SpA-D_{KKAA} immunization can protect mice against MRSA strains and selected the USA300 LAC isolate for animal challenge (Diep et al., 2006). This highly virulent CA-MRSA strain spread rapidly throughout the United States, causing significant human morbidity and mortality (Kennedy et al., 2008). Compared to adjuvant control mice, SpA-D_{KKAA} immunized animals harbored a 1.07 log₁₀ CFU g⁻¹ reduction in bacterial load of infected kidney tissues. Histopathology examination of renal tissue following *S. aureus* USA300 challenge revealed that the average number of abscesses was reduced from 4.04 (±0.8) to 1.6 (±0.6) (P=0.02774). In contrast, SpA or SpA-D immunization did not cause a significant reduction in bacterial load or abscess formation (Table 6).

Rabbits were immunized with SpA-D_{KKAA} and specific antibodies were purified on SpA-D_{KKAA} affinity column followed by SDS-PAGE (FIG. 8). SpA-D_{KKAA} specific IgG was cleaved with pepsin to generate F_γ and F(ab)₂ fragments, the latter of which were purified by chromatography on SpA-D_{KKAA} column (FIG. 8). Binding of human IgG or vWF to SpA or SpA-D was perturbed by SpA-D_{KKAA} specific F(ab)₂, indicating that SpA-D_{KKAA} derived antibodies neutralize the B cell superantigen function of protein A as well as its interactions with Ig (FIG. 8).

To further improve the vaccine properties for non-toxic protein A, the inventors generated SpA_{KKAA}, which includes all five IgBDs with four amino acid substitutions—substitutions corresponding to Gln9Lys, Gln10Lys, Asp36Ala and Asp37Ala of domain D—in each of its five domains (E, D, A, B and C). Polyhistidine tagged SpA_{KKAA}

was purified by affinity chromatography and analyzed by Coomassie Blue-stained SDS-PAGE (FIG. 9). Unlike full-length SpA, SpA_{KKAA} did not bind human IgG, Fc and F(ab)₂ or vWF (FIG. 9). SpA_{KKAA} failed to display B cell superantigen activity, as injection of the variant into BALB/c mice did not cause a depletion of CD19+ B cells in splenic tissue (FIG. 9). SpA_{KKAA} vaccination generated higher specific antibody titers than SpA-D_{KKAA} immunization and provided mice with elevated protection against *S. aureus* USA300 challenge (Table 6). Four days following challenge, SpA_{KKAA} vaccinated animals harbored 3.54 log₁₀ CFU g⁻¹ fewer staphylococci in renal tissues (P=0.0001) and also caused a greater reduction in the number of abscess lesions (P=0.0109) (Table 6).

SpA_{KKAA} was used to immunize rabbits. Rabbit antibodies specific for SpA-D_{KKAA} or SpA_{KKAA} were affinity purified on matrices with immobilized cognate antigen and injected at a concentration of 5 mg kg⁻¹ body weight into the peritoneal cavity of BALB/c mice (Table 7). Twenty-four hours later, specific antibody titers were determined in serum and animals challenged by intravenous inoculation with *S. aureus* Newman. Passive transfer reduced the staphylococcal load in kidney tissues for SpA-D_{KKAA} (P=0.0016) or SpA_{KKAA} (P=0.0005) specific antibodies. On histopathology examination, both antibodies reduced the abundance of abscess lesions in the kidneys of mice challenged with *S. aureus* Newman (Table 7). Together these data reveal that vaccine protection following immunization with SpA-D_{KKAA} or SpA_{KKAA} is conferred by antibodies that neutralize protein A.

TABLE 6

Immunization of mice with protein A vaccines.						
Staphylococcal load and abscess formation in renal tissue						
Antigen	^a Log ₁₀ CFU g ⁻¹	^b P value	^c Reduction (log ₁₀ CFU g ⁻¹)	^d IgG titer	^e Number of abscesses	^b P-value
<i>S. aureus</i> Newman challenge						
Mock	6.46 ± 0.25	—	—	<100	3.7 ± 1.2	—
SpA	3.95 ± 0.56	0.0003	2.51	1,706 ± 370	2.1 ± 1.2	0.3581
SpA-D	4.43 ± 0.41	0.0001	2.03	381 ± 27	1.5 ± 0.8	0.1480
SpA D _{KKAA}	3.39 ± 0.50	<0.0001	3.07	5,600 ± 801	0.5 ± 0.4	0.0204
<i>S. aureus</i> USA300 (LAC) challenge						
Mock	7.20 ± 0.24	—	—	<100	4.0 ± 0.8	—
SpA	6.81 ± 0.26	0.2819	0.39	476 ± 60	3.3 ± 1.0	0.5969
SpA-D	6.34 ± 0.52	0.1249	0.86	358 ± 19	2.2 ± 0.6	0.0912
SpA- D _{KKAA}	6.00 ± 0.42	0.0189	1.20	3,710 ± 1147	1.6 ± 0.6	0.0277
SpA _{KKAA}	3.66 ± 0.76	0.0001	3.54	10,200 ± 2476	1.2 ± 0.5	0.0109

^aMeans of staphylococcal load calculated as log₁₀ CFU g⁻¹ in homogenized renal tissues 4 days following infection in cohorts of fifteen to twenty BALB/c mice per immunization. Representative of two independent and reproducible animal experiments is shown. Standard error of the means (±SEM) is indicated.

^bStatistical significance was calculated with the unpaired two-tailed student's t-test and p-values were recorded; p-values < 0.05 were deemed significant.

^cReduction in bacterial load calculated as log₁₀ CFU g⁻¹.

^dMeans of five randomly chosen serum, IgG titers were measured prior to staphylococcal infection by ELISA.

^eHistopathology of hematoxylin-eosin-stained thin-sectioned kidneys from ten animals; the average number of abscesses per kidney was recorded and averaged again for the final mean (±SEM).

TABLE 7

Passive immunization of mice with antibodies against protein A.						
Staphylococcal load and abscess formation in renal tissue						
^a Antigen	^b \log_{10} CFU g^{-1}	^c P value	^d Reduction (\log_{10} CFU g^{-1})	^e IgG titer	^f Number of abscesses	^g P-value
Mock	7.20 \pm 0.14	—	—	<100	4.5 \pm 0.8	—
α -SpA-D _{KKAA}	5.58 \pm 0.43	0.0016	1.57	466 \pm 114	1.9 \pm 0.7	0.0235
α -SpA _{KKAA}	5.69 \pm 0.34	0.0005	1.41	1575 \pm 152	1.6 \pm 0.5	0.0052

^aAffinity-purified antibodies were injected into the peritoneal cavity of BALB/c mice at a concentration of 5 mg \cdot kg⁻¹ twenty-four hours prior to intravenous challenge with 1×10^7 CFU *S. aureus* Newman.

^bMeans of staphylococcal load calculated as \log_{10} CFU g^{-1} in homogenized renal tissues 4 d following infection in cohorts of 15 BALB/c mice per immunization. Representative of two independent and reproducible animal experiments is shown. Standard error of the means (\pm SEM) is indicated.

^cStatistical significance was calculated with the unpaired two-tailed Student's t test and p-values recorded; p-values < 0.05 were deemed significant.

^dReduction in bacterial load calculated as \log_{10} CFU g^{-1} .

^eMeans of five randomly chosen serum IgG titers were measured prior to staphylococcal infection by ELISA.

^fHistopathology of hematoxylin-eosin-stained, thick sectioned kidneys from ten animals; the average number of abscesses per kidney was recorded and averaged again for the final mean (\pm SEM).

Following infection with virulent *S. aureus*, mice do not develop protective immunity against subsequent infection with the same strain (Buts et al., 2008)(FIG. 10). The average abundance of SpA-D_{KKAA} specific IgG in these animals was determined by dot blot as 0.20 μ g ml⁻¹ (\pm 0.04) and 0.14 μ g ml⁻¹ (\pm 0.01) for strains Newman and USA300 LAC, respectively (FIG. 9). The minimal concentration of protein A-specific IgG required for disease protection in SpA_{KKAA} or SpA-D_{KKAA} vaccinated animals (P 0.0.05 \log_{10} reduction in staphylococcal CFU g^{-1} renal tissue) was calculated as 4.05 μ g ml⁻¹ (\pm 0.88). Average serum concentration of SpA-specific IgG in adult healthy human volunteers (n=16) was 0.21 μ g ml⁻¹ (\pm 0.02). Thus, *S. aureus* infections in mice or humans are not associated with immune responses that raise significant levels of neutralizing antibodies directed against protein A, which is likely due to the B cell superantigen attributes of this molecule. In contrast, the average serum concentration of IgG specific for diphtheria toxin in human volunteers, 0.068 μ g ml⁻¹ (\pm 0.20), was within range for protective immunity against diphtheria (Behring, 1890; Lagergard et al., 1992).

Clinical *S. aureus* isolates express protein A, an essential virulence factor whose B cell superantigen activity and evasive attributes towards opsono-phagocytic clearance are absolutely required for staphylococcal abscess formation (Palmqvist et al., 2005; Cheng et al., 2009; Silverman and Goodyear, 2006). Protein A can thus be thought of as a toxin, essential for pathogenesis, whose molecular attributes must be neutralized in order to achieve protective immunity. By generating non-toxicogenic variants unable to bind Igs via Fc γ or VH3-Fab domains, the inventors measure here for the first time protein A neutralizing immune responses as a correlate for protective immunity against *S. aureus* infection. In contrast to many methicillin-sensitive strains, CA-MRSA isolate USA300 LAC is significantly more virulent (Cheng et al., 2009). For example, immunization of experimental animals with the surface protein IsdB (Kuklin et al., 2006; Stranger-Jones et al., 2006) raises antibodies that confer protection against *S. aureus* Newman (Stranger-Jones et al., 2009) but not against USA300 challenge.

The methods utilized include:

Bacterial strains and growth. *Staphylococcus aureus* strains Newman and USA300 were grown in tryptic soy broth (TSB) at 37° C. *Escherichia coli* strains DH5a and BL21 (DE3) were grown in Luria-Bertani (LB) broth with 100 μ g ml⁻¹ ampicillin at 37° C.

Rabbit Antibodies. The coding sequence for SpA was PCR-amplified with two primers, gctgcacatattggcgcaacacgatgaagctcaac (SEQ ID NO:35) and agtgatccctatgcttgagctttgtagcatctgc (SEQ ID NO:36) using *S. aureus* Newman template DNA. SpA-D was PCR-amplified with two primers, aacatattcaacaagaatcaacaagaac (SEQ ID NO:38) and aaggatccagattcgttaatttttagc (SEQ ID NO:39). The sequence for SpA-D_{KKAA} was mutagenized with two sets of primers catatgttcaacaagaataaaaaagcgcttctatgaaatc (SEQ ID NO:42) and gatttcatagaagggcgtttttttctttgtgaaatag (SEQ ID NO:43) for Q9K, Q10K as well as ctccattcaaagtct-taaagccgccccaaagcaaacactaac (SEQ ID NO:40) and gttagtctttggcttggggcgctttaaagacttgaatgaag (SEQ ID NO:41) for D36A,D37A. The sequence of SpA_{KKAA} was synthesized by Integrated DNA Technologies, Inc. PCR products were cloned into pET-15b generating N-terminal His6 tagged recombinant protein. Plasmids were transformed into BL21(DE3). Overnight cultures of transformants were diluted 1:100 into fresh media and grown at 37° C. to an OD₆₀₀ 0.5, at which point cultures were induced with 1 mM isopropyl β -D-1-thiogalactopyranoside (IPTG) and grown for an additional three hours. Bacterial cells were sedimented by centrifugation, suspended in column buffer (50 mM Tris-HCl, pH 7.5, 150 mM NaCl) and disrupted with a French pressure cell at 14,000 psi. Lysates were cleared of membrane and insoluble components by ultracentrifugation at 40,000 \times g. Proteins in the soluble lysate were subjected to nickel-nitrilotriacetic acid (Ni-NTA, Qiagen) affinity chromatography. Proteins were eluted in column buffer containing successively higher concentrations of imidazole (100-500 mM). Protein concentrations were determined by bicinchoninic acid (BCA) assay (Thermo Scientific). For antibody generation, rabbits (6 month old New-Zealand white, female, Charles River Laboratories) were immunized with 500 μ g protein emulsified in Complete Freund's Adjuvant (Difco) by subcutaneous injection. For booster immunizations, proteins emulsified in Incomplete Freund's Adjuvant and injected 24 or 48 days following the initial immunization. On day 60, rabbits were bled and serum recovered.

Purified antigen (5 mg protein) was covalently linked to HiTrap NHS-activated HP columns (GE Healthcare). Antigen-matrix was used for affinity chromatography of 10-20 ml of rabbit serum at 4° C. Charged matrix was washed with 50 column volumes of PBS, antibodies eluted with elution buffer (1 M glycine, pH 2.5, 0.5 M NaCl) and immediately

neutralized with 1M Tris-HCl, pH 8.5. Purified antibodies were dialyzed overnight against PBS at 4° C.

F(ab)₂ fragments. Affinity purified antibodies were mixed with 3 mg of pepsin at 37° C. for 30 minutes. The reaction was quenched with 1 M Tris-HCl, pH 8.5 and F(ab)₂ fragments were affinity purified with specific antigen-conjugated HiTrap NHS-activated HP columns. Purified antibodies were dialyzed overnight against PBS at 4° C., loaded onto SDS-PAGE gel and visualized with Coomassie Blue staining.

Active and passive immunization. BALB/c mice (3 week old, female, Charles River Laboratories) were immunized with 50 µg protein emulsified in Complete Freund's Adjuvant (Difco) by intramuscular injection. For booster immunizations, proteins were emulsified in Incomplete Freund's Adjuvant and injected 11 days following the initial immunization. On day 20 following immunization, 5 mice were bled to obtain sera for specific antibody titers by enzyme-linked immunosorbent assay (ELISA).

Affinity purified antibodies in PBS were injected at a concentration 5 mg kg⁻¹ of experimental animal weight into the peritoneal cavity of BALB/c mice (6 week old, female, Charles River Laboratories) 24 hours prior to challenge with *S. aureus*. Animal blood was collected via periorbital vein puncture. Blood cells were removed with heparinized microhematocrit capillary tubes (Fisher) and Z-gel serum separation micro tubes (Sarstedt) were used to collect and measure antigen specific antibody titers by ELISA.

Mouse renal abscess. Overnight cultures of *S. aureus* Newman or USA300 (LAC) were diluted 1:100 into fresh TSB and grown for 2 hours at 37° C. Staphylococci were sedimented, washed and suspended PBS at OD₆₀₀ of 0.4 (~1×10⁷ CFU ml⁻¹). Inocula were quantified by spreading sample aliquots on TSA and enumerating colonies formed. BALB/c mice (6 week old, female, Charles River Laboratories) were anesthetized via intraperitoneal injection with 100 mg ml⁻¹ ketamine and 20 mg ml⁻¹ xylazine per kilogram of body weight. Mice were infected by retro-orbital injection with 1×10⁷ CFU of *S. aureus* Newman or 5×10⁶ CFU of *S. aureus* USA300. On day 4 following challenge, mice were killed by CO₂ inhalation. Both kidneys were removed, and the staphylococcal load in one organ was analyzed by homogenizing renal tissue with PBS, 1% Triton X-100. Serial dilutions of homogenate were spread on TSA and incubated for colony formation. The remaining organ was examined by histopathology. Briefly, kidneys were fixed in 10% formalin for 24 hours at room temperature. Tissues were embedded in paraffin, thin-sectioned, stained with hematoxylin-eosin, and inspected by light microscopy to enumerate abscess lesions. All mouse experiments were performed in accordance with the institutional guidelines following experimental protocol review and approval by the Institutional Biosafety Committee (IBC) and the Institutional Animal Care and Use Committee (IACUC) at the University of Chicago.

Protein A binding. For human IgG binding, Ni-NTA affinity columns were pre-charged with 200 µg of purified proteins (SpA, SpA-D, SpA-D_{KKAA}, and SrtA) in column buffer. After washing, 200 µg of human IgG (Sigma) was loaded onto the column. Protein samples were collected from washes and elutions and subjected to SDS-PAGE gel electrophoresis, followed by Coomassie Blue staining. Purified proteins (SpA, SpA_{KKAA}, SpA-D and SpA-D_{KKAA}) were coated onto MaxiSorp ELISA plates (NUNC) in 0.1M carbonate buffer (pH 9.5) at 1 µg ml⁻¹ concentration overnight at 4° C. Plates were next blocked with 5% whole milk followed by incubation with serial dilutions of peroxidase-

conjugated human IgG, Fc or F(ab)₂ fragments for one hour. Plates were washed and developed using OptEIA ELISA reagents (BD). Reactions were quenched with 1 M phosphoric acid and A₄₅₀ readings were used to calculate half maximal titer and percent binding.

von Willebrand Factor (vWF) binding assays. Purified proteins (SpA, SpA_{KKAA}, SpA D and SpA-D_{KKAA}) were coated and blocked as described above. Plates were incubated with human vWF at 1 µg ml⁻¹ concentration for two hours, then washed and blocked with human IgG for another hour. After washing, plates were incubated with serial dilution of peroxidase-conjugated antibody directed against human vWF for one hour. Plates were washed and developed using OptEIA ELISA reagents (BD). Reactions were quenched with 1 M phosphoric acid and A₄₅₀ readings were used to calculate half maximal titer and percent binding. For inhibition assays, plates were incubated with affinity purified F(ab)₂ fragments specific for SpA-D_{KKAA} at 10 µg ml concentration for one hour prior to ligand binding assays.

Splenocyte apoptosis. Affinity purified proteins (150 µg of SpA, SpA-D, SpA_{KKAA}, and SpA-D_{KKAA}) were injected into the peritoneal cavity of BALB/c mice (6 week old, female, Charles River Laboratories). Four hours following injection, animals were killed by CO₂ inhalation. Their spleens were removed and homogenized. Cell debris were removed using cell strainer and suspended cells were transferred to ACK lysis buffer (0.15 M NH₄Cl, 10 mM KHCO₃, 0.1 mM EDTA) to lyse red blood cells. White blood cells were sedimented by centrifugation, suspended in PBS and stained with 1:250 diluted R-PE conjugated anti-CD19 monoclonal antibody (Invitrogen) on ice and in the dark for one hour. Cells were washed with 1% FBS and fixed with 4% formalin overnight at 4° C. The following day, cells were diluted in PBS and analyzed by flow cytometry. The remaining organ was examined for histopathology. Briefly, spleens were fixed in 10% formalin for 24 hours at room temperature. Tissues were embedded in paraffin, thin-sectioned, stained with the Apoptosis detection kit (Millipore), and inspected by light microscopy.

Antibody quantification. Sera were collected from healthy human volunteers or BALB/c mice that had been either infected with *S. aureus* Newman or USA300 for 30 days or that had been immunized with SpA-D_{KKAA}/SpA_{KKAA} as described above. Human/mouse IgG (Jackson Immunology Laboratory), SpA_{KKAA}, and CRM197 were blotted onto nitrocellulose membrane. Membranes were blocked with 5% whole milk, followed by incubation with either human or mouse sera. IRDye 700DX conjugated affinity purified anti-human/mouse IgG (Rockland) was used to quantify signal intensities using the Odyssey™ infrared imaging system (Li-cor). Experiments with blood from human volunteers involved protocols that were reviewed, approved and performed under regulatory supervision of The University of Chicago's Institutional Review Board (IRB).

Statistical Analysis. Two tailed Student's t tests were performed to analyze the statistical significance of renal abscess, ELISA, and B cell superantigen data.

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 Ala Ala Asn Ala Ala Gln His Asp Glu Ala Gln Gln Asn Ala Phe Tyr
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 Gln Val Leu Asn Met Pro Asn Leu Asn Ala Asp Gln Arg Asn Gly Phe
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 Thr Arg Ala Gln Gly Glu Ile Ala Ala Asn Trp Glu Gly Gln Ala Phe
 35 40 45
 Ser Arg Phe Glu Glu Gln Phe Gln Gln Leu Ser Pro Lys Val Glu Lys
 50 55 60
 Phe Ala Gln Leu Leu Glu Glu Ile Lys Gln Gln Leu Asn Ser Thr Ala
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 Asp Ala Val Gln Glu Gln Asp Gln Gln Leu Ser Asn Asn Phe Gly Leu
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 Gln

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 35 40 45
 Gly Gln Phe Ala Asn Lys Val Lys Asp Val Leu Leu Ile Met Ala Lys
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 Phe Gln Glu Glu Leu Val Gln Pro Met Ala Asp His Gln Lys Ala Ile
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 100

<210> SEQ ID NO 13
 <211> LENGTH: 1385
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 13

Met Leu Asn Arg Glu Asn Lys Thr Ala Ile Thr Arg Lys Gly Met Val
 1 5 10 15

Ser Asn Arg Leu Asn Lys Phe Ser Ile Arg Lys Tyr Thr Val Gly Thr
 20 25 30

Ala Ser Ile Leu Val Gly Thr Thr Leu Ile Phe Gly Leu Gly Asn Gln
 35 40 45

Glu Ala Lys Ala Ala Glu Ser Thr Asn Lys Glu Leu Asn Glu Ala Thr
 50 55 60

Thr Ser Ala Ser Asp Asn Gln Ser Ser Asp Lys Val Asp Met Gln Gln
 65 70 75 80

Leu Asn Gln Glu Asp Asn Thr Lys Asn Asp Asn Gln Lys Glu Met Val
 85 90 95

Ser Ser Gln Gly Asn Glu Thr Thr Ser Asn Gly Asn Lys Ser Ile Glu
 100 105 110

Lys Glu Ser Val Gln Ser Thr Thr Gly Asn Lys Val Glu Val Ser Thr
 115 120 125

Ala Lys Ser Asp Glu Gln Ala Ser Pro Lys Ser Thr Asn Glu Asp Leu
 130 135 140

Asn Thr Lys Gln Thr Ile Ser Asn Gln Glu Gly Leu Gln Pro Asp Leu
 145 150 155 160

Leu Glu Asn Lys Ser Val Val Asn Val Gln Pro Thr Asn Glu Glu Asn
 165 170 175

Lys Lys Val Asp Ala Lys Thr Glu Ser Thr Thr Leu Asn Val Lys Ser
 180 185 190

Asp Ala Ile Lys Ser Asn Ala Glu Thr Leu Val Asp Asn Asn Ser Asn
 195 200 205

Ser Asn Asn Glu Asn Asn Ala Asp Ile Ile Leu Pro Lys Ser Thr Ala
 210 215 220

Pro Lys Ser Leu Asn Thr Arg Met Arg Met Ala Ala Ile Gln Pro Asn
 225 230 235 240

Ser Thr Asp Ser Lys Asn Val Asn Asp Leu Ile Thr Ser Asn Thr Thr
 245 250 255

Leu Thr Val Val Asp Ala Asp Asn Ser Lys Thr Ile Val Pro Ala Gln
 260 265 270

Asp Tyr Leu Ser Leu Lys Ser Gln Ile Thr Val Asp Asp Lys Val Lys
 275 280 285

Ser Gly Asp Tyr Phe Thr Ile Lys Tyr Ser Asp Thr Val Gln Val Tyr
 290 295 300

Gly Leu Asn Pro Glu Asp Ile Lys Asn Ile Gly Asp Ile Lys Asp Pro
 305 310 315 320

Asn Asn Gly Glu Thr Ile Ala Thr Ala Lys His Asp Thr Ala Asn Asn
 325 330 335

Leu Ile Thr Tyr Thr Phe Thr Asp Tyr Val Asp Arg Phe Asn Ser Val
 340 345 350

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Lys Met Gly Ile Asn Tyr Ser Ile Tyr Met Asp Ala Asp Thr Ile Pro
 355 360 365

Val Asp Lys Lys Asp Val Pro Phe Ser Val Thr Ile Gly Asn Gln Ile
 370 375 380

Thr Thr Thr Thr Ala Asp Ile Thr Tyr Pro Ala Tyr Lys Glu Ala Asp
 385 390 395 400

Asn Asn Ser Ile Gly Ser Ala Phe Thr Glu Thr Val Ser His Val Gly
 405 410 415

Asn Val Glu Asp Pro Gly Tyr Tyr Asn Gln Val Val Tyr Val Asn Pro
 420 425 430

Met Asp Lys Asp Leu Lys Gly Ala Lys Leu Lys Val Glu Ala Tyr His
 435 440 445

Pro Lys Tyr Pro Thr Asn Ile Gly Gln Ile Asn Gln Asn Val Thr Asn
 450 455 460

Ile Lys Ile Tyr Arg Val Pro Glu Gly Tyr Thr Leu Asn Lys Gly Tyr
 465 470 475 480

Asp Val Asn Thr Asn Asp Leu Val Asp Val Thr Asp Glu Phe Lys Asn
 485 490 495

Lys Met Thr Tyr Gly Ser Asn Gln Ser Val Asn Leu Asp Phe Gly Asp
 500 505 510

Ile Thr Ser Ala Tyr Val Val Met Val Asn Thr Lys Phe Gln Tyr Thr
 515 520 525

Asn Ser Glu Ser Pro Thr Leu Val Gln Met Ala Thr Leu Ser Ser Thr
 530 535 540

Gly Asn Lys Ser Val Ser Thr Gly Asn Ala Leu Gly Phe Thr Asn Asn
 545 550 555 560

Gln Ser Gly Gly Ala Gly Gln Glu Val Tyr Lys Ile Gly Asn Tyr Val
 565 570 575

Trp Glu Asp Thr Asn Lys Asn Gly Val Gln Glu Leu Gly Glu Lys Gly
 580 585 590

Val Gly Asn Val Thr Val Thr Val Phe Asp Asn Asn Thr Asn Thr Lys
 595 600 605

Val Gly Glu Ala Val Thr Lys Glu Asp Gly Ser Tyr Leu Ile Pro Asn
 610 615 620

Leu Pro Asn Gly Asp Tyr Arg Val Glu Phe Ser Asn Leu Pro Lys Gly
 625 630 635 640

Tyr Glu Val Thr Pro Ser Lys Gln Gly Asn Asn Glu Glu Leu Asp Ser
 645 650 655

Asn Gly Leu Ser Ser Val Ile Thr Val Asn Gly Lys Asp Asn Leu Ser
 660 665 670

Ala Asp Leu Gly Ile Tyr Lys Pro Lys Tyr Asn Leu Gly Asp Tyr Val
 675 680 685

Trp Glu Asp Thr Asn Lys Asn Gly Ile Gln Asp Gln Asp Glu Lys Gly
 690 695 700

Ile Ser Gly Val Thr Val Thr Leu Lys Asp Glu Asn Gly Asn Val Leu
 705 710 715 720

Lys Thr Val Thr Thr Asp Ala Asp Gly Lys Tyr Lys Phe Thr Asp Leu
 725 730 735

Asp Asn Gly Asn Tyr Lys Val Glu Phe Thr Thr Pro Glu Gly Tyr Thr
 740 745 750

Pro Thr Thr Val Thr Ser Gly Ser Asp Ile Glu Lys Asp Ser Asn Gly
 755 760 765

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Leu Thr Thr Thr Gly Val Ile Asn Gly Ala Asp Asn Met Thr Leu Asp
 770 775 780

Ser Gly Phe Tyr Lys Thr Pro Lys Tyr Asn Leu Gly Asn Tyr Val Trp
 785 790 795 800

Glu Asp Thr Asn Lys Asp Gly Lys Gln Asp Ser Thr Glu Lys Gly Ile
 805 810 815

Ser Gly Val Thr Val Thr Leu Lys Asn Glu Asn Gly Glu Val Leu Gln
 820 825 830

Thr Thr Lys Thr Asp Lys Asp Gly Lys Tyr Gln Phe Thr Gly Leu Glu
 835 840 845

Asn Gly Thr Tyr Lys Val Glu Phe Glu Thr Pro Ser Gly Tyr Thr Pro
 850 855 860

Thr Gln Val Gly Ser Gly Thr Asp Glu Gly Ile Asp Ser Asn Gly Thr
 865 870 875 880

Ser Thr Thr Gly Val Ile Lys Asp Lys Asp Asn Asp Thr Ile Asp Ser
 885 890 895

Gly Phe Tyr Lys Pro Thr Tyr Asn Leu Gly Asp Tyr Val Trp Glu Asp
 900 905 910

Thr Asn Lys Asn Gly Val Gln Asp Lys Asp Glu Lys Gly Ile Ser Gly
 915 920 925

Val Thr Val Thr Leu Lys Asp Glu Asn Asp Lys Val Leu Lys Thr Val
 930 935 940

Thr Thr Asp Glu Asn Gly Lys Tyr Gln Phe Thr Asp Leu Asn Asn Gly
 945 950 955 960

Thr Tyr Lys Val Glu Phe Glu Thr Pro Ser Gly Tyr Thr Pro Thr Ser
 965 970 975

Val Thr Ser Gly Asn Asp Thr Glu Lys Asp Ser Asn Gly Leu Thr Thr
 980 985 990

Thr Gly Val Ile Lys Asp Ala Asp Asn Met Thr Leu Asp Ser Gly Phe
 995 1000 1005

Tyr Lys Thr Pro Lys Tyr Ser Leu Gly Asp Tyr Val Trp Tyr Asp
 1010 1015 1020

Ser Asn Lys Asp Gly Lys Gln Asp Ser Thr Glu Lys Gly Ile Lys
 1025 1030 1035

Asp Val Lys Val Ile Leu Leu Asn Glu Lys Gly Glu Val Ile Gly
 1040 1045 1050

Thr Thr Lys Thr Asp Glu Asn Gly Lys Tyr Arg Phe Asp Asn Leu
 1055 1060 1065

Asp Ser Gly Lys Tyr Lys Val Ile Phe Glu Lys Pro Thr Gly Leu
 1070 1075 1080

Thr Gln Thr Gly Thr Asn Thr Thr Glu Asp Asp Lys Asp Ala Asp
 1085 1090 1095

Gly Gly Glu Val Asp Val Thr Ile Thr Asp His Asp Asp Phe Thr
 1100 1105 1110

Leu Asp Asn Gly Tyr Tyr Glu Glu Glu Thr Ser Asp Ser Asp Ser
 1115 1120 1125

Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp
 1130 1135 1140

Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
 1145 1150 1155

Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp
 1160 1165 1170

Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser

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1175	1180	1185
Asp Ser Asp Ser Asp Ser	Asp Ser Asp Ser Asp Ser	Asp Ser Asp
1190	1195	1200
Ser Asp Ser Asp Ser Asp	Ser Asp Ser Asp Ser Asp	Ser Asp Ser
1205	1210	1215
Asp Ser Asp Ser Asp Ser	Asp Ser Asp Ser Asp Ser	Asp Ser Asp
1220	1225	1230
Ser Asp Ser Asp Ser Asp	Ser Asp Ser Asp Ser Asp	Ser Asp Ser
1235	1240	1245
Asp Ser Asp Ser Asp Ser	Asp Ser Asp Ser Asp Ser	Asp Ser Asp
1250	1255	1260
Ser Asp Ser Asp Ser Asp	Ser Asp Ser Asp Ser Asp	Ser Asp Ser
1265	1270	1275
Asp Ser Asp Ser Asp Ser	Asp Ser Asp Ser Asp Ser	Asp Ser Asp
1280	1285	1290
Ser Asp Ser Asp Ser Asp	Ser Asp Ser Asp Ser Asp	Ser Asp Ser
1295	1300	1305
Asp Ser Asp Ser Asp Ser	Asp Ser Asp Ser Asp Ser	Asp Ser Asp
1310	1315	1320
Ser Asp Ala Gly Lys His	Thr Pro Val Lys Pro Met	Ser Thr Thr
1325	1330	1335
Lys Asp His His Asn Lys	Ala Lys Ala Leu Pro Glu	Thr Gly Asn
1340	1345	1350
Glu Asn Ser Gly Ser Asn	Asn Ala Thr Leu Phe Gly	Gly Leu Phe
1355	1360	1365
Ala Ala Leu Gly Ser Leu	Leu Leu Phe Gly Arg Arg	Lys Lys Gln
1370	1375	1380
Asn Lys		
1385		

<210> SEQ ID NO 14
 <211> LENGTH: 1141
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 14

Met Ile Asn Arg Asp Asn Lys Lys Ala Ile Thr Lys Lys Gly Met Ile
1 5 10 15
Ser Asn Arg Leu Asn Lys Phe Ser Ile Arg Lys Tyr Thr Val Gly Thr
20 25 30
Ala Ser Ile Leu Val Gly Thr Thr Leu Ile Phe Gly Leu Gly Asn Gln
35 40 45
Glu Ala Lys Ala Ala Glu Asn Thr Ser Thr Glu Asn Ala Lys Gln Asp
50 55 60
Asp Ala Thr Thr Ser Asp Asn Lys Glu Val Val Ser Glu Thr Glu Asn
65 70 75 80
Asn Ser Thr Thr Glu Asn Asp Ser Thr Asn Pro Ile Lys Lys Glu Thr
85 90 95
Asn Thr Asp Ser Gln Pro Glu Ala Lys Glu Glu Ser Thr Thr Ser Ser
100 105 110
Thr Gln Gln Gln Gln Asn Asn Val Thr Ala Thr Thr Glu Thr Lys Pro
115 120 125
Gln Asn Ile Glu Lys Glu Asn Val Lys Pro Ser Thr Asp Lys Thr Ala
130 135 140

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Thr Glu Asp Thr Ser Val Ile Leu Glu Glu Lys Lys Ala Pro Asn Tyr
 145 150 155 160

Thr Asn Asn Asp Val Thr Thr Lys Pro Ser Thr Ser Glu Ile Gln Thr
 165 170 175

Lys Pro Thr Thr Pro Gln Glu Ser Thr Asn Ile Glu Asn Ser Gln Pro
 180 185 190

Gln Pro Thr Pro Ser Lys Val Asp Asn Gln Val Thr Asp Ala Thr Asn
 195 200 205

Pro Lys Glu Pro Val Asn Val Ser Lys Glu Glu Leu Lys Asn Asn Pro
 210 215 220

Glu Lys Leu Lys Glu Leu Val Arg Asn Asp Asn Asn Thr Asp Arg Ser
 225 230 235 240

Thr Lys Pro Val Ala Thr Ala Pro Thr Ser Val Ala Pro Lys Arg Leu
 245 250 255

Asn Ala Lys Met Arg Phe Ala Val Ala Gln Pro Ala Ala Val Ala Ser
 260 265 270

Asn Asn Val Asn Asp Leu Ile Thr Val Thr Lys Gln Thr Ile Lys Val
 275 280 285

Gly Asp Gly Lys Asp Asn Val Ala Ala Ala His Asp Gly Lys Asp Ile
 290 295 300

Glu Tyr Asp Thr Glu Phe Thr Ile Asp Asn Lys Val Lys Lys Gly Asp
 305 310 315 320

Thr Met Thr Ile Asn Tyr Asp Lys Asn Val Ile Pro Ser Asp Leu Thr
 325 330 335

Asp Lys Asn Asp Pro Ile Asp Ile Thr Asp Pro Ser Gly Glu Val Ile
 340 345 350

Ala Lys Gly Thr Phe Asp Lys Ala Thr Lys Gln Ile Thr Tyr Thr Phe
 355 360 365

Thr Asp Tyr Val Asp Lys Tyr Glu Asp Ile Lys Ala Arg Leu Thr Leu
 370 375 380

Tyr Ser Tyr Ile Asp Lys Gln Ala Val Pro Asn Glu Thr Ser Leu Asn
 385 390 395 400

Leu Thr Phe Ala Thr Ala Gly Lys Glu Thr Ser Gln Asn Val Ser Val
 405 410 415

Asp Tyr Gln Asp Pro Met Val His Gly Asp Ser Asn Ile Gln Ser Ile
 420 425 430

Phe Thr Lys Leu Asp Glu Asn Lys Gln Thr Ile Glu Gln Gln Ile Tyr
 435 440 445

Val Asn Pro Leu Lys Lys Thr Ala Thr Asn Thr Lys Val Asp Ile Ala
 450 455 460

Gly Ser Gln Val Asp Asp Tyr Gly Asn Ile Lys Leu Gly Asn Gly Ser
 465 470 475 480

Thr Ile Ile Asp Gln Asn Thr Glu Ile Lys Val Tyr Lys Val Asn Pro
 485 490 495

Asn Gln Gln Leu Pro Gln Ser Asn Arg Ile Tyr Asp Phe Ser Gln Tyr
 500 505 510

Glu Asp Val Thr Ser Gln Phe Asp Asn Lys Lys Ser Phe Ser Asn Asn
 515 520 525

Val Ala Thr Leu Asp Phe Gly Asp Ile Asn Ser Ala Tyr Ile Ile Lys
 530 535 540

Val Val Ser Lys Tyr Thr Pro Thr Ser Asp Gly Glu Leu Asp Ile Ala
 545 550 555 560

Gln Gly Thr Ser Met Arg Thr Thr Asp Lys Tyr Gly Tyr Tyr Asn Tyr

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Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
 995 1000 1005
 Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp
 1010 1015 1020
 Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
 1025 1030 1035
 Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp
 1040 1045 1050
 Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
 1055 1060 1065
 Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ala Gly
 1070 1075 1080
 Lys His Thr Pro Val Lys Pro Met Ser Thr Thr Lys Asp His His
 1085 1090 1095
 Asn Lys Ala Lys Ala Leu Pro Glu Thr Gly Ser Glu Asn Asn Gly
 1100 1105 1110
 Ser Asn Asn Ala Thr Leu Phe Gly Gly Leu Phe Ala Ala Leu Gly
 1115 1120 1125
 Ser Leu Leu Leu Phe Gly Arg Arg Lys Lys Gln Asn Lys
 1130 1135 1140

<210> SEQ ID NO 15
 <211> LENGTH: 350
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 15

Met Thr Lys His Tyr Leu Asn Ser Lys Tyr Gln Ser Glu Gln Arg Ser
 1 5 10 15
 Ser Ala Met Lys Lys Ile Thr Met Gly Thr Ala Ser Ile Ile Leu Gly
 20 25 30
 Ser Leu Val Tyr Ile Gly Ala Asp Ser Gln Gln Val Asn Ala Ala Thr
 35 40 45
 Glu Ala Thr Asn Ala Thr Asn Asn Gln Ser Thr Gln Val Ser Gln Ala
 50 55 60
 Thr Ser Gln Pro Ile Asn Phe Gln Val Gln Lys Asp Gly Ser Ser Glu
 65 70 75 80
 Lys Ser His Met Asp Asp Tyr Met Gln His Pro Gly Lys Val Ile Lys
 85 90 95
 Gln Asn Asn Lys Tyr Tyr Phe Gln Thr Val Leu Asn Asn Ala Ser Phe
 100 105 110
 Trp Lys Glu Tyr Lys Phe Tyr Asn Ala Asn Asn Gln Glu Leu Ala Thr
 115 120 125
 Thr Val Val Asn Asp Asn Lys Lys Ala Asp Thr Arg Thr Ile Asn Val
 130 135 140
 Ala Val Glu Pro Gly Tyr Lys Ser Leu Thr Thr Lys Val His Ile Val
 145 150 155 160
 Val Pro Gln Ile Asn Tyr Asn His Arg Tyr Thr Thr His Leu Glu Phe
 165 170 175
 Glu Lys Ala Ile Pro Thr Leu Ala Asp Ala Ala Lys Pro Asn Asn Val
 180 185 190
 Lys Pro Val Gln Pro Lys Pro Ala Gln Pro Lys Thr Pro Thr Glu Gln
 195 200 205
 Thr Lys Pro Val Gln Pro Lys Val Glu Lys Val Lys Pro Thr Val Thr

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His Phe Asn Asn Lys Glu Glu Lys Tyr Asp Tyr Thr Leu Met Glu Phe
 245 250 255

Ala Gln Pro Ile Tyr Asn Ser Ala Asp Lys Phe Lys Thr Glu Glu Asp
 260 265 270

Tyr Lys Ala Glu Lys Leu Leu Ala Pro Tyr Lys Lys Ala Lys Thr Leu
 275 280 285

Glu Arg Gln Val Tyr Glu Leu Asn Lys Ile Gln Asp Lys Leu Pro Glu
 290 295 300

Lys Leu Lys Ala Glu Tyr Lys Lys Lys Leu Glu Asp Thr Lys Lys Ala
 305 310 315 320

Leu Asp Glu Gln Val Lys Ser Ala Ile Thr Glu Phe Gln Asn Val Gln
 325 330 335

Pro Thr Asn Glu Lys Met Thr Asp Leu Gln Asp Thr Lys Tyr Val Val
 340 345 350

Tyr Glu Ser Val Glu Asn Asn Glu Ser Met Met Asp Thr Phe Val Lys
 355 360 365

His Pro Ile Lys Thr Gly Met Leu Asn Gly Lys Lys Tyr Met Val Met
 370 375 380

Glu Thr Thr Asn Asp Asp Tyr Trp Lys Asp Phe Met Val Glu Gly Gln
 385 390 395 400

Arg Val Arg Thr Ile Ser Lys Asp Ala Lys Asn Asn Thr Arg Thr Ile
 405 410 415

Ile Phe Pro Tyr Val Glu Gly Lys Thr Leu Tyr Asp Ala Ile Val Lys
 420 425 430

Val His Val Lys Thr Ile Asp Tyr Asp Gly Gln Tyr His Val Arg Ile
 435 440 445

Val Asp Lys Glu Ala Phe Thr Lys Ala Asn Thr Asp Lys Ser Asn Lys
 450 455 460

Lys Glu Gln Gln Asp Asn Ser Ala Lys Lys Glu Ala Thr Pro Ala Thr
 465 470 475 480

Pro Ser Lys Pro Thr Pro Ser Pro Val Glu Lys Glu Ser Gln Lys Gln
 485 490 495

Asp Ser Gln Lys Asp Asp Asn Lys Gln Leu Pro Ser Val Glu Lys Glu
 500 505 510

Asn Asp Ala Ser Ser Glu Ser Gly Lys Asp Lys Thr Pro Ala Thr Lys
 515 520 525

Pro Thr Lys Gly Glu Val Glu Ser Ser Ser Thr Thr Pro Thr Lys Val
 530 535 540

Val Ser Thr Thr Gln Asn Val Ala Lys Pro Thr Thr Ala Ser Ser Lys
 545 550 555 560

Thr Thr Lys Asp Val Val Gln Thr Ser Ala Gly Ser Ser Glu Ala Lys
 565 570 575

Asp Ser Ala Pro Leu Gln Lys Ala Asn Ile Lys Asn Thr Asn Asp Gly
 580 585 590

His Thr Gln Ser Gln Asn Asn Lys Asn Thr Gln Glu Asn Lys Ala Lys
 595 600 605

Ser Leu Pro Gln Thr Gly Glu Glu Ser Asn Lys Asp Met Thr Leu Pro
 610 615 620

Leu Met Ala Leu Leu Ala Leu Ser Ser Ile Val Ala Phe Val Leu Pro
 625 630 635 640

Arg Lys Arg Lys Asn
 645

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<210> SEQ ID NO 17
<211> LENGTH: 80
<212> TYPE: PRT
<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 17
Met Asn Gln His Val Lys Val Thr Phe Asp Phe Thr Asn Tyr Asn Tyr
1          5          10          15
Gly Thr Tyr Asp Leu Ala Val Pro Ala Tyr Leu Pro Ile Lys Asn Leu
20          25          30
Ile Ala Leu Val Leu Asp Ser Leu Asp Ile Ser Ile Phe Asp Val Asn
35          40          45
Thr Gln Ile Lys Val Met Thr Lys Gly Gln Leu Leu Val Glu Asn Asp
50          55          60
Arg Leu Ile Asp Tyr Gln Ile Ala Asp Gly Asp Ile Leu Lys Leu Leu
65          70          75          80

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<210> SEQ ID NO 18
<211> LENGTH: 877
<212> TYPE: PRT
<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 18
Met Lys Lys Arg Ile Asp Tyr Leu Ser Asn Lys Gln Asn Lys Tyr Ser
1          5          10          15
Ile Arg Arg Phe Thr Val Gly Thr Thr Ser Val Ile Val Gly Ala Thr
20          25          30
Ile Leu Phe Gly Ile Gly Asn His Gln Ala Gln Ala Ser Glu Gln Ser
35          40          45
Asn Asp Thr Thr Gln Ser Ser Lys Asn Asn Ala Ser Ala Asp Ser Glu
50          55          60
Lys Asn Asn Met Ile Glu Thr Pro Gln Leu Asn Thr Thr Ala Asn Asp
65          70          75          80
Thr Ser Asp Ile Ser Ala Asn Thr Asn Ser Ala Asn Val Asp Ser Thr
85          90          95
Thr Lys Pro Met Ser Thr Gln Thr Ser Asn Thr Thr Thr Thr Glu Pro
100         105         110
Ala Ser Thr Asn Glu Thr Pro Gln Pro Thr Ala Ile Lys Asn Gln Ala
115         120         125
Thr Ala Ala Lys Met Gln Asp Gln Thr Val Pro Gln Glu Ala Asn Ser
130         135         140
Gln Val Asp Asn Lys Thr Thr Asn Asp Ala Asn Ser Ile Ala Thr Asn
145         150         155         160
Ser Glu Leu Lys Asn Ser Gln Thr Leu Asp Leu Pro Gln Ser Ser Pro
165         170         175
Gln Thr Ile Ser Asn Ala Gln Gly Thr Ser Lys Pro Ser Val Arg Thr
180         185         190
Arg Ala Val Arg Ser Leu Ala Val Ala Glu Pro Val Val Asn Ala Ala
195         200         205
Asp Ala Lys Gly Thr Asn Val Asn Asp Lys Val Thr Ala Ser Asn Phe
210         215         220
Lys Leu Glu Lys Thr Thr Phe Asp Pro Asn Gln Ser Gly Asn Thr Phe
225         230         235         240
Met Ala Ala Asn Phe Thr Val Thr Asp Lys Val Lys Ser Gly Asp Tyr
245         250         255
Phe Thr Ala Lys Leu Pro Asp Ser Leu Thr Gly Asn Gly Asp Val Asp

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260				265				270							
Tyr	Ser	Asn	Ser	Asn	Asn	Thr	Met	Pro	Ile	Ala	Asp	Ile	Lys	Ser	Thr
		275					280					285			
Asn	Gly	Asp	Val	Val	Ala	Lys	Ala	Thr	Tyr	Asp	Ile	Leu	Thr	Lys	Thr
	290					295					300				
Tyr	Thr	Phe	Val	Phe	Thr	Asp	Tyr	Val	Asn	Asn	Lys	Glu	Asn	Ile	Asn
305					310					315					320
Gly	Gln	Phe	Ser	Leu	Pro	Leu	Phe	Thr	Asp	Arg	Ala	Lys	Ala	Pro	Lys
				325					330					335	
Ser	Gly	Thr	Tyr	Asp	Ala	Asn	Ile	Asn	Ile	Ala	Asp	Glu	Met	Phe	Asn
			340					345				350			
Asn	Lys	Ile	Thr	Tyr	Asn	Tyr	Ser	Ser	Pro	Ile	Ala	Gly	Ile	Asp	Lys
	355						360					365			
Pro	Asn	Gly	Ala	Asn	Ile	Ser	Ser	Gln	Ile	Ile	Gly	Val	Asp	Thr	Ala
	370					375					380				
Ser	Gly	Gln	Asn	Thr	Tyr	Lys	Gln	Thr	Val	Phe	Val	Asn	Pro	Lys	Gln
385					390					395					400
Arg	Val	Leu	Gly	Asn	Thr	Trp	Val	Tyr	Ile	Lys	Gly	Tyr	Gln	Asp	Lys
			405					410						415	
Ile	Glu	Glu	Ser	Ser	Gly	Lys	Val	Ser	Ala	Thr	Asp	Thr	Lys	Leu	Arg
			420					425				430			
Ile	Phe	Glu	Val	Asn	Asp	Thr	Ser	Lys	Leu	Ser	Asp	Ser	Tyr	Tyr	Ala
	435					440					445				
Asp	Pro	Asn	Asp	Ser	Asn	Leu	Lys	Glu	Val	Thr	Asp	Gln	Phe	Lys	Asn
	450				455						460				
Arg	Ile	Tyr	Tyr	Glu	His	Pro	Asn	Val	Ala	Ser	Ile	Lys	Phe	Gly	Asp
465				470						475					480
Ile	Thr	Lys	Thr	Tyr	Val	Val	Leu	Val	Glu	Gly	His	Tyr	Asp	Asn	Thr
			485					490						495	
Gly	Lys	Asn	Leu	Lys	Thr	Gln	Val	Ile	Gln	Glu	Asn	Val	Asp	Pro	Val
			500					505					510		
Thr	Asn	Arg	Asp	Tyr	Ser	Ile	Phe	Gly	Trp	Asn	Asn	Glu	Asn	Val	Val
		515					520					525			
Arg	Tyr	Gly	Gly	Gly	Ser	Ala	Asp	Gly	Asp	Ser	Ala	Val	Asn	Pro	Lys
	530					535					540				
Asp	Pro	Thr	Pro	Gly	Pro	Pro	Val	Asp	Pro	Glu	Pro	Ser	Pro	Asp	Pro
545					550					555					560
Glu	Pro	Glu	Pro	Thr	Pro	Asp	Pro	Glu	Pro	Ser	Pro	Asp	Pro	Glu	Pro
			565					570						575	
Glu	Pro	Ser	Pro	Asp	Pro	Asp	Pro	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
			580					585					590		
Gly	Ser	Asp	Ser	Asp	Ser	Gly	Ser	Asp	Ser	Asp	Ser	Glu	Ser	Asp	Ser
	595					600						605			
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Glu	Ser
	610				615							620			
Asp	Ser	Asp	Ser	Glu	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
625				630						635					640
Asp	Ser	Asp	Ser	Asp	Ser	Glu	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
			645						650					655	
Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Glu	Ser	Asp	Ser	Asp	Ser	Glu	Ser
			660					665					670		
Asp	Ser	Glu	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser	Asp	Ser
	675					680								685	

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Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
 690 695 700
 Asp Ser Asp Ser Asp Ser Asp Ser Glu Ser Asp Ser Asp Ser Asp Ser
 705 710 715 720
 Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
 725 730 735
 Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
 740 745 750
 Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
 755 760 765
 Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
 770 775 780
 Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser Asp Ser
 785 790 795 800
 Asp Ser Asp Ser Arg Val Thr Pro Pro Asn Asn Glu Gln Lys Ala Pro
 805 810 815
 Ser Asn Pro Lys Gly Glu Val Asn His Ser Asn Lys Val Ser Lys Gln
 820 825 830
 His Lys Thr Asp Ala Leu Pro Glu Thr Gly Asp Lys Ser Glu Asn Thr
 835 840 845
 Asn Ala Thr Leu Phe Gly Ala Met Met Ala Leu Leu Gly Ser Leu Leu
 850 855 860
 Leu Phe Arg Lys Arg Lys Gln Asp His Lys Glu Lys Ala
 865 870 875

<210> SEQ ID NO 19
 <211> LENGTH: 227
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 19

Met Lys Asn Ile Leu Lys Val Phe Asn Thr Thr Ile Leu Ala Leu Ile
 1 5 10 15
 Ile Ile Ile Ala Thr Phe Ser Asn Ser Ala Asn Ala Ala Asp Ser Gly
 20 25 30
 Thr Leu Asn Tyr Glu Val Tyr Lys Tyr Asn Thr Asn Asp Thr Ser Ile
 35 40 45
 Ala Asn Asp Tyr Phe Asn Lys Pro Ala Lys Tyr Ile Lys Lys Asn Gly
 50 55 60
 Lys Leu Tyr Val Gln Ile Thr Val Asn His Ser His Trp Ile Thr Gly
 65 70 75 80
 Met Ser Ile Glu Gly His Lys Glu Asn Ile Ile Ser Lys Asn Thr Ala
 85 90 95
 Lys Asp Glu Arg Thr Ser Glu Phe Glu Val Ser Lys Leu Asn Gly Lys
 100 105 110
 Ile Asp Gly Lys Ile Asp Val Tyr Ile Asp Glu Lys Val Asn Gly Lys
 115 120 125
 Pro Phe Lys Tyr Asp His His Tyr Asn Ile Thr Tyr Lys Phe Asn Gly
 130 135 140
 Pro Thr Asp Val Ala Gly Ala Asn Ala Pro Gly Lys Asp Asp Lys Asn
 145 150 155 160
 Ser Ala Ser Gly Ser Asp Lys Gly Ser Asp Gly Thr Thr Thr Gly Gln
 165 170 175
 Ser Glu Ser Asn Ser Ser Asn Lys Asp Lys Val Glu Asn Pro Gln Thr

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      180              185              190
Asn Ala Gly Thr Pro Ala Tyr Ile Tyr Ala Ile Pro Val Ala Ser Leu
  195                      200                      205

Ala Leu Leu Ile Ala Ile Thr Leu Phe Val Arg Lys Lys Ser Lys Gly
  210                      215                      220

Asn Val Glu
  225

<210> SEQ ID NO 20
<211> LENGTH: 635
<212> TYPE: PRT
<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 20

Met Ala Lys Tyr Arg Gly Lys Pro Phe Gln Leu Tyr Val Lys Leu Ser
  1      5      10      15

Cys Ser Thr Met Met Ala Ser Ser Ile Ile Leu Thr Asn Ile Leu Pro
  20      25      30

Tyr Asp Ala Gln Ala Ala Ser Glu Lys Asp Thr Glu Ile Ser Lys Glu
  35      40      45

Ile Leu Ser Lys Gln Asp Leu Leu Asp Lys Val Asp Lys Ala Ile Arg
  50      55      60

Gln Ile Glu Gln Leu Lys Gln Leu Ser Ala Ser Ser Lys Ala His Tyr
  65      70      75      80

Lys Ala Gln Leu Asn Glu Ala Lys Thr Ala Ser Gln Ile Asp Glu Ile
  85      90      95

Ile Lys Arg Ala Asn Glu Leu Asp Ser Lys Glu Asn Lys Ser Ser His
  100     105     110

Thr Glu Met Asn Gly Gln Ser Asp Ile Asp Ser Lys Leu Asp Gln Leu
  115     120     125

Leu Lys Asp Leu Asn Glu Val Ser Ser Asn Val Asp Arg Gly Gln Gln
  130     135     140

Ser Gly Glu Asp Asp Leu Asn Ala Met Lys Asn Asp Met Ser Gln Thr
  145     150     155     160

Ala Thr Thr Lys Tyr Gly Glu Lys Asp Asp Lys Asn Asp Glu Ala Met
  165     170     175

Val Asn Lys Ala Leu Glu Asp Leu Asp His Leu Asn Gln Gln Ile His
  180     185     190

Lys Ser Lys Asp Ala Leu Lys Asp Ala Ser Lys Asp Pro Ala Val Ser
  195     200     205

Thr Thr Asp Ser Asn His Glu Val Ala Lys Thr Pro Asn Asn Asp Gly
  210     215     220

Ser Gly His Val Val Leu Asn Lys Phe Leu Ser Asn Glu Glu Asn Gln
  225     230     235     240

Ser His Ser Asn Gln Leu Thr Asp Lys Leu Gln Gly Ser Asp Lys Ile
  245     250     255

Asn His Ala Met Ile Glu Lys Leu Ala Lys Ser Asn Ala Ser Thr Gln
  260     265     270

His Tyr Thr Tyr His Lys Leu Asn Thr Leu Gln Ser Leu Asp Gln Arg
  275     280     285

Ile Ala Asn Thr Gln Leu Pro Lys Asn Gln Lys Ser Asp Leu Met Ser
  290     295     300

Glu Val Asn Lys Thr Lys Glu Arg Ile Lys Ser Gln Arg Asn Ile Ile
  305     310     315     320

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Leu Glu Glu Leu Ala Arg Thr Asp Asp Lys Lys Tyr Ala Thr Gln Ser
 325 330 335

Ile Leu Glu Ser Ile Phe Asn Lys Asp Glu Ala Asp Lys Ile Leu Lys
 340 345 350

Asp Ile Arg Val Asp Gly Lys Thr Asp Gln Gln Ile Ala Asp Gln Ile
 355 360 365

Thr Arg His Ile Asp Gln Leu Ser Leu Thr Thr Ser Asp Asp Leu Leu
 370 375 380

Thr Ser Leu Ile Asp Gln Ser Gln Asp Lys Ser Leu Leu Ile Ser Gln
 385 390 395 400

Ile Leu Gln Thr Lys Leu Gly Lys Ala Glu Ala Asp Lys Leu Ala Lys
 405 410 415

Asp Trp Thr Asn Lys Gly Leu Ser Asn Arg Gln Ile Val Asp Gln Leu
 420 425 430

Lys Lys His Phe Ala Ser Thr Gly Asp Thr Ser Ser Asp Asp Ile Leu
 435 440 445

Lys Ala Ile Leu Asn Asn Ala Lys Asp Lys Lys Gln Ala Ile Glu Thr
 450 455 460

Ile Leu Ala Thr Arg Ile Glu Arg Gln Lys Ala Lys Leu Leu Ala Asp
 465 470 475 480

Leu Ile Thr Lys Ile Glu Thr Asp Gln Asn Lys Ile Phe Asn Leu Val
 485 490 495

Lys Ser Ala Leu Asn Gly Lys Ala Asp Asp Leu Leu Asn Leu Gln Lys
 500 505 510

Arg Leu Asn Gln Thr Lys Lys Asp Ile Asp Tyr Ile Leu Ser Pro Ile
 515 520 525

Val Asn Arg Pro Ser Leu Leu Asp Arg Leu Asn Lys Asn Gly Lys Thr
 530 535 540

Thr Asp Leu Asn Lys Leu Ala Asn Leu Met Asn Gln Gly Ser Asn Leu
 545 550 555 560

Leu Asp Ser Ile Pro Asp Ile Pro Thr Pro Lys Pro Glu Lys Thr Leu
 565 570 575

Thr Leu Gly Lys Gly Asn Gly Leu Leu Ser Gly Leu Leu Asn Ala Asp
 580 585 590

Gly Asn Val Ser Leu Pro Lys Ala Gly Glu Thr Ile Lys Glu His Trp
 595 600 605

Leu Pro Ile Ser Val Ile Val Gly Ala Met Gly Val Leu Met Ile Trp
 610 615 620

Leu Ser Arg Arg Asn Lys Leu Lys Asn Lys Ala
 625 630 635

<210> SEQ ID NO 21
 <211> LENGTH: 953
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 21

Met Asn Asn Lys Lys Thr Ala Thr Asn Arg Lys Gly Met Ile Pro Asn
 1 5 10 15

Arg Leu Asn Lys Phe Ser Ile Arg Lys Tyr Ser Val Gly Thr Ala Ser
 20 25 30

Ile Leu Val Gly Thr Thr Leu Ile Phe Gly Leu Ser Gly His Glu Ala
 35 40 45

Lys Ala Ala Glu His Thr Asn Gly Glu Leu Asn Gln Ser Lys Asn Glu
 50 55 60

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Thr Thr Ala Pro Ser Glu Asn Lys Thr Thr Glu Lys Val Asp Ser Arg
 65 70 75 80
 Gln Leu Lys Asp Asn Thr Gln Thr Ala Thr Ala Asp Gln Pro Lys Val
 85 90 95
 Thr Met Ser Asp Ser Ala Thr Val Lys Glu Thr Ser Ser Asn Met Gln
 100 105 110
 Ser Pro Gln Asn Ala Thr Ala Ser Gln Ser Thr Thr Gln Thr Ser Asn
 115 120 125
 Val Thr Thr Asn Asp Lys Ser Ser Thr Thr Tyr Ser Asn Glu Thr Asp
 130 135 140
 Lys Ser Asn Leu Thr Gln Ala Lys Asn Val Ser Thr Thr Pro Lys Thr
 145 150 155 160
 Thr Thr Ile Lys Gln Arg Ala Leu Asn Arg Met Ala Val Asn Thr Val
 165 170 175
 Ala Ala Pro Gln Gln Gly Thr Asn Val Asn Asp Lys Val His Phe Thr
 180 185 190
 Asn Ile Asp Ile Ala Ile Asp Lys Gly His Val Asn Lys Thr Thr Gly
 195 200 205
 Asn Thr Glu Phe Trp Ala Thr Ser Ser Asp Val Leu Lys Leu Lys Ala
 210 215 220
 Asn Tyr Thr Ile Asp Asp Ser Val Lys Glu Gly Asp Thr Phe Thr Phe
 225 230 235 240
 Lys Tyr Gly Gln Tyr Phe Arg Pro Gly Ser Val Arg Leu Pro Ser Gln
 245 250 255
 Thr Gln Asn Leu Tyr Asn Ala Gln Gly Asn Ile Ile Ala Lys Gly Ile
 260 265 270
 Tyr Asp Ser Lys Thr Asn Thr Thr Thr Tyr Thr Phe Thr Asn Tyr Val
 275 280 285
 Asp Gln Tyr Thr Asn Val Ser Gly Ser Phe Glu Gln Val Ala Phe Ala
 290 295 300
 Lys Arg Glu Asn Ala Thr Thr Asp Lys Thr Ala Tyr Lys Met Glu Val
 305 310 315 320
 Thr Leu Gly Asn Asp Thr Tyr Ser Lys Asp Val Ile Val Asp Tyr Gly
 325 330 335
 Asn Gln Lys Gly Gln Gln Leu Ile Ser Ser Thr Asn Tyr Ile Asn Asn
 340 345 350
 Glu Asp Leu Ser Arg Asn Met Thr Val Tyr Val Asn Gln Pro Lys Lys
 355 360 365
 Thr Tyr Thr Lys Glu Thr Phe Val Thr Asn Leu Thr Gly Tyr Lys Phe
 370 375 380
 Asn Pro Asp Ala Lys Asn Phe Lys Ile Tyr Glu Val Thr Asp Gln Asn
 385 390 395 400
 Gln Phe Val Asp Ser Phe Thr Pro Asp Thr Ser Lys Leu Lys Asp Val
 405 410 415
 Thr Gly Gln Phe Asp Val Ile Tyr Ser Asn Asp Asn Lys Thr Ala Thr
 420 425 430
 Val Asp Leu Leu Asn Gly Gln Ser Ser Ser Asp Lys Gln Tyr Ile Ile
 435 440 445
 Gln Gln Val Ala Tyr Pro Asp Asn Ser Ser Thr Asp Asn Gly Lys Ile
 450 455 460
 Asp Tyr Thr Leu Glu Thr Gln Asn Gly Lys Ser Ser Trp Ser Asn Ser
 465 470 475 480

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Ala Asp Leu Ile Asn Thr Ser Glu Ile Lys Ala Ile Asn Ile Asn Val
 130 135 140

Asp Thr Lys Lys Gln Val Glu Asp Lys Lys Lys Asp Lys Ala Asn Tyr
 145 150 155 160

Gln Val Pro Tyr Thr Ile Thr Val Asn Gly Thr Ser Gln Asn Ile Leu
 165 170 175

Ser Asn Leu Thr Phe Asn Lys Asn Gln Asn Ile Ser Tyr Lys Asp Leu
 180 185 190

Glu Asp Lys Val Lys Ser Val Leu Glu Ser Asn Arg Gly Ile Thr Asp
 195 200 205

Val Asp Leu Arg Leu Ser Lys Gln Ala Lys Tyr Thr Val Asn Phe Lys
 210 215 220

Asn Gly Thr Lys Lys Val Ile Asp Leu Lys Ser Gly Ile Tyr Thr Ala
 225 230 235 240

Asn Leu Ile Asn Ser Ser Asp Ile Lys Ser Ile Asn Ile Asn Val Asp
 245 250 255

Thr Lys Lys His Ile Glu Asn Lys Ala Lys Arg Asn Tyr Gln Val Pro
 260 265 270

Tyr Ser Ile Asn Leu Asn Gly Thr Ser Thr Asn Ile Leu Ser Asn Leu
 275 280 285

Ser Phe Ser Asn Lys Pro Trp Thr Asn Tyr Lys Asn Leu Thr Ser Gln
 290 295 300

Ile Lys Ser Val Leu Lys His Asp Arg Gly Ile Ser Glu Gln Asp Leu
 305 310 315 320

Lys Tyr Ala Lys Lys Ala Tyr Tyr Thr Val Tyr Phe Lys Asn Gly Gly
 325 330 335

Lys Arg Ile Leu Gln Leu Asn Ser Lys Asn Tyr Thr Ala Asn Leu Val
 340 345 350

His Ala Lys Asp Val Lys Arg Ile Glu Ile Thr Val Lys Thr Gly Thr
 355 360 365

Lys Ala Lys Ala Asp Arg Tyr Val Pro Tyr Thr Ile Ala Val Asn Gly
 370 375 380

Thr Ser Thr Pro Ile Leu Ser Asp Leu Lys Phe Thr Gly Asp Pro Arg
 385 390 395 400

Val Gly Tyr Lys Asp Ile Ser Lys Lys Val Lys Ser Val Leu Lys His
 405 410 415

Asp Arg Gly Ile Gly Glu Arg Glu Leu Lys Tyr Ala Lys Lys Ala Thr
 420 425 430

Tyr Thr Val His Phe Lys Asn Gly Thr Lys Lys Val Ile Asn Ile Asn
 435 440 445

Ser Asn Ile Ser Gln Leu Asn Leu Leu Tyr Val Gln Asp Ile Lys Lys
 450 455 460

Ile Asp Ile Asp Val Lys Thr Gly Thr Lys Ala Lys Ala Asp Ser Tyr
 465 470 475 480

Val Pro Tyr Thr Ile Ala Val Asn Gly Thr Ser Thr Pro Ile Leu Ser
 485 490 495

Lys Leu Lys Ile Ser Asn Lys Gln Leu Ile Ser Tyr Lys Tyr Leu Asn
 500 505 510

Asp Lys Val Lys Ser Val Leu Lys Ser Glu Arg Gly Ile Ser Asp Leu
 515 520 525

Asp Leu Lys Phe Ala Lys Gln Ala Lys Tyr Thr Val Tyr Phe Lys Asn
 530 535 540

Gly Lys Lys Gln Val Val Asn Leu Lys Ser Asp Ile Phe Thr Pro Asn

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Lys Gln Thr Gly Asp Gly Tyr Gln Trp Val Ile Lys Phe Asn Lys Gly
 340 345 350
 His Ala Pro His Gln Asn Met Ile Phe Trp Phe Ala Leu Pro Ala Asp
 355 360 365
 Gln Val Pro Val Gly Arg Thr Asp Phe Val Thr Val Asn Ser Asp Gly
 370 375 380
 Thr Asn Val Gln Trp Ser His Gly Ala Gly Ala Gly Ala Asn Lys Pro
 385 390 395 400
 Leu Gln Gln Met Trp Glu Tyr Gly Val Asn Asp Pro His Arg Ser His
 405 410 415
 Asp Phe Lys Ile Arg Asn Arg Ser Gly Gln Val Ile Tyr Asp Trp Pro
 420 425 430
 Thr Val His Ile Tyr Ser Leu Glu Asp Leu Ser Arg Ala Ser Asp Tyr
 435 440 445
 Phe Ser Glu Ala Gly Ala Thr Pro Ala Thr Lys Ala Phe Gly Arg Gln
 450 455 460
 Asn Phe Glu Tyr Ile Asn Gly Gln Lys Pro Ala Glu Ser Pro Gly Val
 465 470 475 480
 Pro Lys Val Tyr Thr Phe Ile Gly Gln Gly Asp Ala Ser Tyr Thr Ile
 485 490 495
 Ser Phe Lys Thr Gln Gly Pro Thr Val Asn Lys Leu Tyr Tyr Ala Ala
 500 505 510
 Gly Gly Arg Ala Leu Glu Tyr Asn Gln Leu Phe Met Tyr Ser Gln Leu
 515 520 525
 Tyr Val Glu Ser Thr Gln Asp His Gln Gln Arg Leu Asn Gly Leu Arg
 530 535 540
 Gln Val Val Asn Arg Thr Tyr Arg Ile Gly Thr Thr Lys Arg Val Glu
 545 550 555 560
 Val Ser Gln Gly Asn Val Gln Thr Lys Lys Val Leu Glu Ser Thr Asn
 565 570 575
 Leu Asn Ile Asp Asp Phe Val Asp Asp Pro Leu Ser Tyr Val Lys Thr
 580 585 590
 Pro Ser Asn Lys Val Leu Gly Phe Tyr Ser Asn Asn Ala Asn Thr Asn
 595 600 605
 Ala Phe Arg Pro Gly Gly Ala Gln Gln Leu Asn Glu Tyr Gln Leu Ser
 610 615 620
 Gln Leu Phe Thr Asp Gln Lys Leu Gln Glu Ala Ala Arg Thr Arg Asn
 625 630 635 640
 Pro Ile Arg Leu Met Ile Gly Phe Asp Tyr Pro Asp Ala Tyr Gly Asn
 645 650 655
 Ser Glu Thr Leu Val Pro Val Asn Leu Thr Val Leu Pro Glu Ile Gln
 660 665 670
 His Asn Ile Lys Phe Phe Lys Asn Asp Asp Thr Gln Asn Ile Ala Glu
 675 680 685
 Lys Pro Phe Ser Lys Gln Ala Gly His Pro Val Phe Tyr Val Tyr Ala
 690 695 700
 Gly Asn Gln Gly Asn Ala Ser Val Asn Leu Gly Gly Ser Val Thr Ser
 705 710 715 720
 Ile Gln Pro Leu Arg Ile Asn Leu Thr Ser Asn Glu Asn Phe Thr Asp
 725 730 735
 Lys Asp Trp Gln Ile Thr Gly Ile Pro Arg Thr Leu His Ile Glu Asn
 740 745 750
 Ser Thr Asn Arg Pro Asn Asn Ala Arg Glu Arg Asn Ile Glu Leu Val

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Thr	Asn	Ala	Tyr	His	Lys	Asp	Ser	Glu	Asp	Phe	Tyr	Ile	Ser	Leu
1175						1180					1185			
Lys	Ile	Ile	Asp	Val	Lys	Gln	Pro	Glu	Gly	Asp	Gln	Arg	Val	Tyr
1190						1195					1200			
Arg	Thr	Ser	Thr	Tyr	Asp	Leu	Thr	Thr	Asp	Glu	Ile	Ser	Lys	Val
1205						1210					1215			
Lys	Gln	Ala	Phe	Ile	Asn	Ala	Asn	Arg	Asp	Val	Ile	Thr	Leu	Ala
1220						1225					1230			
Glu	Gly	Asp	Ile	Ser	Val	Thr	Asn	Thr	Pro	Asn	Gly	Ala	Asn	Val
1235						1240					1245			
Ser	Thr	Ile	Thr	Val	Asn	Ile	Asn	Lys	Gly	Arg	Leu	Thr	Lys	Ser
1250						1255					1260			
Phe	Ala	Ser	Asn	Leu	Ala	Asn	Met	Asn	Phe	Leu	Arg	Trp	Val	Asn
1265						1270					1275			
Phe	Pro	Gln	Asp	Tyr	Thr	Val	Thr	Trp	Thr	Asn	Ala	Lys	Ile	Ala
1280						1285					1290			
Asn	Arg	Pro	Thr	Asp	Gly	Gly	Leu	Ser	Trp	Ser	Asp	Asp	His	Lys
1295						1300					1305			
Ser	Leu	Ile	Tyr	Arg	Tyr	Asp	Ala	Thr	Leu	Gly	Thr	Gln	Ile	Thr
1310						1315					1320			
Thr	Asn	Asp	Ile	Leu	Thr	Met	Leu	Lys	Ala	Thr	Thr	Thr	Val	Pro
1325						1330					1335			
Gly	Leu	Arg	Asn	Asn	Ile	Thr	Gly	Asn	Glu	Lys	Ser	Gln	Ala	Glu
1340						1345					1350			
Ala	Gly	Gly	Arg	Pro	Asn	Phe	Arg	Thr	Thr	Gly	Tyr	Ser	Gln	Ser
1355						1360					1365			
Asn	Ala	Thr	Thr	Asp	Gly	Gln	Arg	Gln	Phe	Thr	Leu	Asn	Gly	Gln
1370						1375					1380			
Val	Ile	Gln	Val	Leu	Asp	Ile	Ile	Asn	Pro	Ser	Asn	Gly	Tyr	Gly
1385						1390					1395			
Gly	Gln	Pro	Val	Thr	Asn	Ser	Asn	Thr	Arg	Ala	Asn	His	Ser	Asn
1400						1405					1410			
Ser	Thr	Val	Val	Asn	Val	Asn	Glu	Pro	Ala	Ala	Asn	Gly	Ala	Gly
1415						1420					1425			
Ala	Phe	Thr	Ile	Asp	His	Val	Val	Lys	Ser	Asn	Ser	Thr	His	Asn
1430						1435					1440			
Ala	Ser	Asp	Ala	Val	Tyr	Lys	Ala	Gln	Leu	Tyr	Leu	Thr	Pro	Tyr
1445						1450					1455			
Gly	Pro	Lys	Gln	Tyr	Val	Glu	His	Leu	Asn	Gln	Asn	Thr	Gly	Asn
1460						1465					1470			
Thr	Thr	Asp	Ala	Ile	Asn	Ile	Tyr	Phe	Val	Pro	Ser	Asp	Leu	Val
1475						1480					1485			
Asn	Pro	Thr	Ile	Ser	Val	Gly	Asn	Tyr	Thr	Asn	His	Gln	Val	Phe
1490						1495					1500			
Ser	Gly	Glu	Thr	Phe	Thr	Asn	Thr	Ile	Thr	Ala	Asn	Asp	Asn	Phe
1505						1510					1515			
Gly	Val	Gln	Ser	Val	Thr	Val	Pro	Asn	Thr	Ser	Gln	Ile	Thr	Gly
1520						1525					1530			
Thr	Val	Asp	Asn	Asn	His	Gln	His	Val	Ser	Ala	Thr	Ala	Pro	Asn
1535						1540					1545			
Val	Thr	Ser	Ala	Thr	Asn	Lys	Thr	Ile	Asn	Leu	Leu	Ala	Thr	Asp
1550						1555					1560			

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Thr	Ser	Gly	Asn	Thr	Ala	Thr	Thr	Ser	Phe	Asn	Val	Thr	Val	Lys
1565						1570					1575			
Pro	Leu	Arg	Asp	Lys	Tyr	Arg	Val	Gly	Thr	Ser	Ser	Thr	Ala	Ala
1580						1585					1590			
Asn	Pro	Val	Arg	Ile	Ala	Asn	Ile	Ser	Asn	Asn	Ala	Thr	Val	Ser
1595						1600					1605			
Gln	Ala	Asp	Gln	Thr	Thr	Ile	Ile	Asn	Ser	Leu	Thr	Phe	Thr	Glu
1610						1615					1620			
Thr	Val	Pro	Asn	Arg	Ser	Tyr	Ala	Arg	Ala	Ser	Ala	Asn	Glu	Ile
1625						1630					1635			
Thr	Ser	Lys	Thr	Val	Ser	Asn	Val	Ser	Arg	Thr	Gly	Asn	Asn	Ala
1640						1645					1650			
Asn	Val	Thr	Val	Thr	Val	Thr	Tyr	Gln	Asp	Gly	Thr	Thr	Ser	Thr
1655						1660					1665			
Val	Thr	Val	Pro	Val	Lys	His	Val	Ile	Pro	Glu	Ile	Val	Ala	His
1670						1675					1680			
Ser	His	Tyr	Thr	Val	Gln	Gly	Gln	Asp	Phe	Pro	Ala	Gly	Asn	Gly
1685						1690					1695			
Ser	Ser	Ala	Ser	Asp	Tyr	Phe	Lys	Leu	Ser	Asn	Gly	Ser	Asp	Ile
1700						1705					1710			
Ala	Asp	Ala	Thr	Ile	Thr	Trp	Val	Ser	Gly	Gln	Ala	Pro	Asn	Lys
1715						1720					1725			
Asp	Asn	Thr	Arg	Ile	Gly	Glu	Asp	Ile	Thr	Val	Thr	Ala	His	Ile
1730						1735					1740			
Leu	Ile	Asp	Gly	Glu	Thr	Thr	Pro	Ile	Thr	Lys	Thr	Ala	Thr	Tyr
1745						1750					1755			
Lys	Val	Val	Arg	Thr	Val	Pro	Lys	His	Val	Phe	Glu	Thr	Ala	Arg
1760						1765					1770			
Gly	Val	Leu	Tyr	Pro	Gly	Val	Ser	Asp	Met	Tyr	Asp	Ala	Lys	Gln
1775						1780					1785			
Tyr	Val	Lys	Pro	Val	Asn	Asn	Ser	Trp	Ser	Thr	Asn	Ala	Gln	His
1790						1795					1800			
Met	Asn	Phe	Gln	Phe	Val	Gly	Thr	Tyr	Gly	Pro	Asn	Lys	Asp	Val
1805						1810					1815			
Val	Gly	Ile	Ser	Thr	Arg	Leu	Ile	Arg	Val	Thr	Tyr	Asp	Asn	Arg
1820						1825					1830			
Gln	Thr	Glu	Asp	Leu	Thr	Ile	Leu	Ser	Lys	Val	Lys	Pro	Asp	Pro
1835						1840					1845			
Pro	Arg	Ile	Asp	Ala	Asn	Ser	Val	Thr	Tyr	Lys	Ala	Gly	Leu	Thr
1850						1855					1860			
Asn	Gln	Glu	Ile	Lys	Val	Asn	Asn	Val	Leu	Asn	Asn	Ser	Ser	Val
1865						1870					1875			
Lys	Leu	Phe	Lys	Ala	Asp	Asn	Thr	Pro	Leu	Asn	Val	Thr	Asn	Ile
1880						1885					1890			
Thr	His	Gly	Ser	Gly	Phe	Ser	Ser	Val	Val	Thr	Val	Ser	Asp	Ala
1895						1900					1905			
Leu	Pro	Asn	Gly	Gly	Ile	Lys	Ala	Lys	Ser	Ser	Ile	Ser	Met	Asn
1910						1915					1920			
Asn	Val	Thr	Tyr	Thr	Thr	Gln	Asp	Glu	His	Gly	Gln	Val	Val	Thr
1925						1930					1935			
Val	Thr	Arg	Asn	Glu	Ser	Val	Asp	Ser	Asn	Asp	Ser	Ala	Thr	Val
1940						1945					1950			
Thr	Val	Thr	Pro	Gln	Leu	Gln	Ala	Thr	Thr	Glu	Gly	Ala	Val	Phe

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1955	1960	1965
Ile Lys Gly Gly Asp Gly 1970	Phe Asp Phe Gly His 1975	Val Glu Arg Phe 1980
Ile Gln Asn Pro Pro His 1985	Gly Ala Thr Val Ala 1990	Trp His Asp Ser 1995
Pro Asp Thr Trp Lys Asn 2000	Thr Val Gly Asn Thr 2005	His Lys Thr Ala 2010
Val Val Thr Leu Pro Asn 2015	Gly Gln Gly Thr Arg 2020	Asn Val Glu Val 2025
Pro Val Lys Val Tyr Pro 2030	Val Ala Asn Ala Lys 2035	Ala Pro Ser Arg 2040
Asp Val Lys Gly Gln Asn 2045	Leu Thr Asn Gly Thr 2050	Asp Ala Met Asn 2055
Tyr Ile Thr Phe Asp Pro 2060	Asn Thr Asn Thr Asn 2065	Gly Ile Thr Ala 2070
Ala Trp Ala Asn Arg Gln 2075	Gln Pro Asn Asn Gln 2080	Gln Ala Gly Val 2085
Gln His Leu Asn Val Asp 2090	Val Thr Tyr Pro Gly 2095	Ile Ser Ala Ala 2100
Lys Arg Val Pro Val Thr 2105	Val Asn Val Tyr Gln 2110	Phe Glu Phe Pro 2115
Gln Thr Thr Tyr Thr Thr 2120	Thr Val Gly Gly Thr 2125	Leu Ala Ser Gly 2130
Thr Gln Ala Ser Gly Tyr 2135	Ala His Met Gln Asn 2140	Ala Thr Gly Leu 2145
Pro Thr Asp Gly Phe Thr 2150	Tyr Lys Trp Asn Arg 2155	Asp Thr Thr Gly 2160
Thr Asn Asp Ala Asn Trp 2165	Ser Ala Met Asn Lys 2170	Pro Asn Val Ala 2175
Lys Val Val Asn Ala Lys 2180	Tyr Asp Val Ile Tyr 2185	Asn Gly His Thr 2190
Phe Ala Thr Ser Leu Pro 2195	Ala Lys Phe Val Val 2200	Lys Asp Val Gln 2205
Pro Ala Lys Pro Thr Val 2210	Thr Glu Thr Ala Ala 2215	Gly Ala Ile Thr 2220
Ile Ala Pro Gly Ala Asn 2225	Gln Thr Val Asn Thr 2230	His Ala Gly Asn 2235
Val Thr Thr Tyr Ala Asp 2240	Lys Leu Val Ile Lys 2245	Arg Asn Gly Asn 2250
Val Val Thr Thr Phe Thr 2255	Arg Arg Asn Asn Thr 2260	Ser Pro Trp Val 2265
Lys Glu Ala Ser Ala Ala 2270	Thr Val Ala Gly Ile 2275	Ala Gly Thr Asn 2280
Asn Gly Ile Thr Val Ala 2285	Ala Gly Thr Phe Asn 2290	Pro Ala Asp Thr 2295
Ile Gln Val Val Ala Thr 2300	Gln Gly Ser Gly Glu 2305	Thr Val Ser Asp 2310
Glu Gln Arg Ser Asp Asp 2315	Phe Thr Val Val Ala 2320	Pro Gln Pro Asn 2325
Gln Ala Thr Thr Lys Ile 2330	Trp Gln Asn Gly His 2335	Ile Asp Ile Thr 2340
Pro Asn Asn Pro Ser Gly 2345	His Leu Ile Asn Pro 2350	Thr Gln Ala Met 2355

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Asp	Ile	Ala	Tyr	Thr	Glu	Lys	Val	Gly	Asn	Gly	Ala	Glu	His	Ser
2360						2365					2370			
Lys	Thr	Ile	Asn	Val	Val	Arg	Gly	Gln	Asn	Asn	Gln	Trp	Thr	Ile
2375						2380					2385			
Ala	Asn	Lys	Pro	Asp	Tyr	Val	Thr	Leu	Asp	Ala	Gln	Thr	Gly	Lys
2390						2395					2400			
Val	Thr	Phe	Asn	Ala	Asn	Thr	Ile	Lys	Pro	Asn	Ser	Ser	Ile	Thr
2405						2410					2415			
Ile	Thr	Pro	Lys	Ala	Gly	Thr	Gly	His	Ser	Val	Ser	Ser	Asn	Pro
2420						2425					2430			
Ser	Thr	Leu	Thr	Ala	Pro	Ala	Ala	His	Thr	Val	Asn	Thr	Thr	Glu
2435						2440					2445			
Ile	Val	Lys	Asp	Tyr	Gly	Ser	Asn	Val	Thr	Ala	Ala	Glu	Ile	Asn
2450						2455					2460			
Asn	Ala	Val	Gln	Val	Ala	Asn	Lys	Arg	Thr	Ala	Thr	Ile	Lys	Asn
2465						2470					2475			
Gly	Thr	Ala	Met	Pro	Thr	Asn	Leu	Ala	Gly	Gly	Ser	Thr	Thr	Thr
2480						2485					2490			
Ile	Pro	Val	Thr	Val	Thr	Tyr	Asn	Asp	Gly	Ser	Thr	Glu	Glu	Val
2495						2500					2505			
Gln	Glu	Ser	Ile	Phe	Thr	Lys	Ala	Asp	Lys	Arg	Glu	Leu	Ile	Thr
2510						2515					2520			
Ala	Lys	Asn	His	Leu	Asp	Asp	Pro	Val	Ser	Thr	Glu	Gly	Lys	Lys
2525						2530					2535			
Pro	Gly	Thr	Ile	Thr	Gln	Tyr	Asn	Asn	Ala	Met	His	Asn	Ala	Gln
2540						2545					2550			
Gln	Gln	Ile	Asn	Thr	Ala	Lys	Thr	Glu	Ala	Gln	Gln	Val	Ile	Asn
2555						2560					2565			
Asn	Glu	Arg	Ala	Thr	Pro	Gln	Gln	Val	Ser	Asp	Ala	Leu	Thr	Lys
2570						2575					2580			
Val	Arg	Ala	Ala	Gln	Thr	Lys	Ile	Asp	Gln	Ala	Lys	Ala	Leu	Leu
2585						2590					2595			
Gln	Asn	Lys	Glu	Asp	Asn	Ser	Gln	Leu	Val	Thr	Ser	Lys	Asn	Asn
2600						2605					2610			
Leu	Gln	Ser	Ser	Val	Asn	Gln	Val	Pro	Ser	Thr	Ala	Gly	Met	Thr
2615						2620					2625			
Gln	Gln	Ser	Ile	Asp	Asn	Tyr	Asn	Ala	Lys	Lys	Arg	Glu	Ala	Glu
2630						2635					2640			
Thr	Glu	Ile	Thr	Ala	Ala	Gln	Arg	Val	Ile	Asp	Asn	Gly	Asp	Ala
2645						2650					2655			
Thr	Ala	Gln	Gln	Ile	Ser	Asp	Glu	Lys	His	Arg	Val	Asp	Asn	Ala
2660						2665					2670			
Leu	Thr	Ala	Leu	Asn	Gln	Ala	Lys	His	Asp	Leu	Thr	Ala	Asp	Thr
2675						2680					2685			
His	Ala	Leu	Glu	Gln	Ala	Val	Gln	Gln	Leu	Asn	Arg	Thr	Gly	Thr
2690						2695					2700			
Thr	Thr	Gly	Lys	Lys	Pro	Ala	Ser	Ile	Thr	Ala	Tyr	Asn	Asn	Ser
2705						2710					2715			
Ile	Arg	Ala	Leu	Gln	Ser	Asp	Leu	Thr	Ser	Ala	Lys	Asn	Ser	Ala
2720						2725					2730			
Asn	Ala	Ile	Ile	Gln	Lys	Pro	Ile	Arg	Thr	Val	Gln	Glu	Val	Gln
2735						2740					2745			

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Ser 2750	Ala	Leu	Thr	Asn	Val	Asn 2755	Arg	Val	Asn	Glu	Arg 2760	Leu	Thr	Gln
Ala 2765	Ile	Asn	Gln	Leu	Val	Pro 2770	Leu	Ala	Asp	Asn	Ser 2775	Ala	Leu	Lys
Thr 2780	Ala	Lys	Thr	Lys	Leu	Asp 2785	Glu	Glu	Ile	Asn	Lys 2790	Ser	Val	Thr
Thr 2795	Asp	Gly	Met	Thr	Gln	Ser 2800	Ser	Ile	Gln	Ala	Tyr 2805	Glu	Asn	Ala
Lys 2810	Arg	Ala	Gly	Gln	Thr	Glu 2815	Ser	Thr	Asn	Ala	Gln 2820	Asn	Val	Ile
Asn 2825	Asn	Gly	Asp	Ala	Thr	Asp 2830	Gln	Gln	Ile	Ala	Ala 2835	Glu	Lys	Thr
Lys 2840	Val	Glu	Glu	Lys	Tyr	Asn 2845	Ser	Leu	Lys	Gln	Ala 2850	Ile	Ala	Gly
Leu 2855	Thr	Pro	Asp	Leu	Ala	Pro 2860	Leu	Gln	Thr	Ala	Lys 2865	Thr	Gln	Leu
Gln 2870	Asn	Asp	Ile	Asp	Gln	Pro 2875	Thr	Ser	Thr	Thr	Gly 2880	Met	Thr	Ser
Ala 2885	Ser	Ile	Ala	Ala	Phe	Asn 2890	Glu	Lys	Leu	Ser	Ala 2895	Ala	Arg	Thr
Lys 2900	Ile	Gln	Glu	Ile	Asp	Arg 2905	Val	Leu	Ala	Ser	His 2910	Pro	Asp	Val
Ala 2915	Thr	Ile	Arg	Gln	Asn	Val 2920	Thr	Ala	Ala	Asn	Ala 2925	Ala	Lys	Ser
Ala 2930	Leu	Asp	Gln	Ala	Arg	Asn 2935	Gly	Leu	Thr	Val	Asp 2940	Lys	Ala	Pro
Leu 2945	Glu	Asn	Ala	Lys	Asn	Gln 2950	Leu	Gln	His	Ser	Ile 2955	Asp	Thr	Gln
Thr 2960	Ser	Thr	Thr	Gly	Met	Thr 2965	Gln	Asp	Ser	Ile	Asn 2970	Ala	Tyr	Asn
Ala 2975	Lys	Leu	Thr	Ala	Ala	Arg 2980	Asn	Lys	Ile	Gln	Gln 2985	Ile	Asn	Gln
Val 2990	Leu	Ala	Gly	Ser	Pro	Thr 2995	Val	Glu	Gln	Ile	Asn 3000	Thr	Asn	Thr
Ser 3005	Thr	Ala	Asn	Gln	Ala	Lys 3010	Ser	Asp	Leu	Asp	His 3015	Ala	Arg	Gln
Ala 3020	Leu	Thr	Pro	Asp	Lys	Ala 3025	Pro	Leu	Gln	Thr	Ala 3030	Lys	Thr	Gln
Leu 3035	Glu	Gln	Ser	Ile	Asn	Gln 3040	Pro	Thr	Asp	Thr	Thr 3045	Gly	Met	Thr
Thr 3050	Ala	Ser	Leu	Asn	Ala	Tyr 3055	Asn	Gln	Lys	Leu	Gln 3060	Ala	Ala	Arg
Gln 3065	Lys	Leu	Thr	Glu	Ile	Asn 3070	Gln	Val	Leu	Asn	Gly 3075	Asn	Pro	Thr
Val 3080	Gln	Asn	Ile	Asn	Asp	Lys 3085	Val	Thr	Glu	Ala	Asn 3090	Gln	Ala	Lys
Asp 3095	Gln	Leu	Asn	Thr	Ala	Arg 3100	Gln	Gly	Leu	Thr	Leu 3105	Asp	Arg	Gln
Pro 3110	Ala	Leu	Thr	Thr	Leu	His 3115	Gly	Ala	Ser	Asn	Leu 3120	Asn	Gln	Ala
Gln 3125	Gln	Asn	Asn	Phe	Thr	Gln 3130	Gln	Ile	Asn	Ala	Ala 3135	Gln	Asn	His
Ala 3140	Ala	Leu	Glu	Thr	Ile	Lys 3145	Ser	Asn	Ile	Thr	Ala 3150	Leu	Asn	Thr

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3140	3145	3150
Ala Met Thr Lys Leu Lys Asp Ser Val Ala Asp Asn Asn Thr Ile 3155 3160 3165		
Lys Ser Asp Gln Asn Tyr Thr Asp Ala Thr Pro Ala Asn Lys Gln 3170 3175 3180		
Ala Tyr Asp Asn Ala Val Asn Ala Ala Lys Gly Val Ile Gly Glu 3185 3190 3195		
Thr Thr Asn Pro Thr Met Asp Val Asn Thr Val Asn Gln Lys Ala 3200 3205 3210		
Ala Ser Val Lys Ser Thr Lys Asp Ala Leu Asp Gly Gln Gln Asn 3215 3220 3225		
Leu Gln Arg Ala Lys Thr Glu Ala Thr Asn Ala Ile Thr His Ala 3230 3235 3240		
Ser Asp Leu Asn Gln Ala Gln Lys Asn Ala Leu Thr Gln Gln Val 3245 3250 3255		
Asn Ser Ala Gln Asn Val Gln Ala Val Asn Asp Ile Lys Gln Thr 3260 3265 3270		
Thr Gln Ser Leu Asn Thr Ala Met Thr Gly Leu Lys Arg Gly Val 3275 3280 3285		
Ala Asn His Asn Gln Val Val Gln Ser Asp Asn Tyr Val Asn Ala 3290 3295 3300		
Asp Thr Asn Lys Lys Asn Asp Tyr Asn Asn Ala Tyr Asn His Ala 3305 3310 3315		
Asn Asp Ile Ile Asn Gly Asn Ala Gln His Pro Val Ile Thr Pro 3320 3325 3330		
Ser Asp Val Asn Asn Ala Leu Ser Asn Val Thr Ser Lys Glu His 3335 3340 3345		
Ala Leu Asn Gly Glu Ala Lys Leu Asn Ala Ala Lys Gln Glu Ala 3350 3355 3360		
Asn Thr Ala Leu Gly His Leu Asn Asn Leu Asn Asn Ala Gln Arg 3365 3370 3375		
Gln Asn Leu Gln Ser Gln Ile Asn Gly Ala His Gln Ile Asp Ala 3380 3385 3390		
Val Asn Thr Ile Lys Gln Asn Ala Thr Asn Leu Asn Ser Ala Met 3395 3400 3405		
Gly Asn Leu Arg Gln Ala Val Ala Asp Lys Asp Gln Val Lys Arg 3410 3415 3420		
Thr Glu Asp Tyr Ala Asp Ala Asp Thr Ala Lys Gln Asn Ala Tyr 3425 3430 3435		
Asn Ser Ala Val Ser Ser Ala Glu Thr Ile Ile Asn Gln Thr Thr 3440 3445 3450		
Asn Pro Thr Met Ser Val Asp Asp Val Asn Arg Ala Thr Ser Ala 3455 3460 3465		
Val Thr Ser Asn Lys Asn Ala Leu Asn Gly Tyr Glu Lys Leu Ala 3470 3475 3480		
Gln Ser Lys Thr Asp Ala Ala Arg Ala Ile Asp Ala Leu Pro His 3485 3490 3495		
Leu Asn Asn Ala Gln Lys Ala Asp Val Lys Ser Lys Ile Asn Ala 3500 3505 3510		
Ala Ser Asn Ile Ala Gly Val Asn Thr Val Lys Gln Gln Gly Thr 3515 3520 3525		
Asp Leu Asn Thr Ala Met Gly Asn Leu Gln Gly Ala Ile Asn Asp 3530 3535 3540		

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Glu	Gln	Thr	Thr	Leu	Asn	Ser	Gln	Asn	Tyr	Gln	Asp	Ala	Thr	Pro
3545						3550					3555			
Ser	Lys	Lys	Thr	Ala	Tyr	Thr	Asn	Ala	Val	Gln	Ala	Ala	Lys	Asp
3560						3565					3570			
Ile	Leu	Asn	Lys	Ser	Asn	Gly	Gln	Asn	Lys	Thr	Lys	Asp	Gln	Val
3575						3580					3585			
Thr	Glu	Ala	Met	Asn	Gln	Val	Asn	Ser	Ala	Lys	Asn	Asn	Leu	Asp
3590						3595					3600			
Gly	Thr	Arg	Leu	Leu	Asp	Gln	Ala	Lys	Gln	Thr	Ala	Lys	Gln	Gln
3605						3610					3615			
Leu	Asn	Asn	Met	Thr	His	Leu	Thr	Thr	Ala	Gln	Lys	Thr	Asn	Leu
3620						3625					3630			
Thr	Asn	Gln	Ile	Asn	Ser	Gly	Thr	Thr	Val	Ala	Gly	Val	Gln	Thr
3635						3640					3645			
Val	Gln	Ser	Asn	Ala	Asn	Thr	Leu	Asp	Gln	Ala	Met	Asn	Thr	Leu
3650						3655					3660			
Arg	Gln	Ser	Ile	Ala	Asn	Lys	Asp	Ala	Thr	Lys	Ala	Ser	Glu	Asp
3665						3670					3675			
Tyr	Val	Asp	Ala	Asn	Asn	Asp	Lys	Gln	Thr	Ala	Tyr	Asn	Asn	Ala
3680						3685					3690			
Val	Ala	Ala	Ala	Glu	Thr	Ile	Ile	Asn	Ala	Asn	Ser	Asn	Pro	Glu
3695						3700					3705			
Met	Asn	Pro	Ser	Thr	Ile	Thr	Gln	Lys	Ala	Glu	Gln	Val	Asn	Ser
3710						3715					3720			
Ser	Lys	Thr	Ala	Leu	Asn	Gly	Asp	Glu	Asn	Leu	Ala	Ala	Ala	Lys
3725						3730					3735			
Gln	Asn	Ala	Lys	Thr	Tyr	Leu	Asn	Thr	Leu	Thr	Ser	Ile	Thr	Asp
3740						3745					3750			
Ala	Gln	Lys	Asn	Asn	Leu	Ile	Ser	Gln	Ile	Thr	Ser	Ala	Thr	Arg
3755						3760					3765			
Val	Ser	Gly	Val	Asp	Thr	Val	Lys	Gln	Asn	Ala	Gln	His	Leu	Asp
3770						3775					3780			
Gln	Ala	Met	Ala	Ser	Leu	Gln	Asn	Gly	Ile	Asn	Asn	Glu	Ser	Gln
3785						3790					3795			
Val	Lys	Ser	Ser	Glu	Lys	Tyr	Arg	Asp	Ala	Asp	Thr	Asn	Lys	Gln
3800						3805					3810			
Gln	Glu	Tyr	Asp	Asn	Ala	Ile	Thr	Ala	Ala	Lys	Ala	Ile	Leu	Asn
3815						3820					3825			
Lys	Ser	Thr	Gly	Pro	Asn	Thr	Ala	Gln	Asn	Ala	Val	Glu	Ala	Ala
3830						3835					3840			
Leu	Gln	Arg	Val	Asn	Asn	Ala	Lys	Asp	Ala	Leu	Asn	Gly	Asp	Ala
3845						3850					3855			
Lys	Leu	Ile	Ala	Ala	Gln	Asn	Ala	Ala	Lys	Gln	His	Leu	Gly	Thr
3860						3865					3870			
Leu	Thr	His	Ile	Thr	Thr	Ala	Gln	Arg	Asn	Asp	Leu	Thr	Asn	Gln
3875						3880					3885			
Ile	Ser	Gln	Ala	Thr	Asn	Leu	Ala	Gly	Val	Glu	Ser	Val	Lys	Gln
3890						3895					3900			
Asn	Ala	Asn	Ser	Leu	Asp	Gly	Ala	Met	Gly	Asn	Leu	Gln	Thr	Ala
3905						3910					3915			
Ile	Asn	Asp	Lys	Ser	Gly	Thr	Leu	Ala	Ser	Gln	Asn	Phe	Leu	Asp
3920						3925					3930			

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Ala Asp 3935	Glu Gln Lys Arg	Asn 3940	Ala Tyr Asn Gln Ala 3945	Val Ser Ala
Ala Glu 3950	Thr Ile Leu Asn Lys 3955	Gln Thr Gly Pro	Asn Thr Ala Lys 3960	
Thr Ala 3965	Val Glu Gln Ala Leu 3970	Asn Asn Val Asn Asn	Ala Lys His 3975	
Ala Leu 3980	Asn Gly Thr Gln Asn 3985	Leu Asn Asn Ala Lys 3990	Gln Ala Ala	
Ile Thr 3995	Ala Ile Asn Gly Ala 4000	Ser Asp Leu Asn Gln 4005	Lys Gln Lys	
Asp Ala 4010	Leu Lys Ala Gln Ala 4015	Asn Gly Ala Gln Arg 4020	Val Ser Asn	
Ala Gln 4025	Asp Val Gln His Asn 4030	Ala Thr Glu Leu Asn 4035	Thr Ala Met	
Gly Thr 4040	Leu Lys His Ala Ile 4045	Ala Asp Lys Thr Asn 4050	Thr Leu Ala	
Ser Ser 4055	Lys Tyr Val Asn Ala 4060	Asp Ser Thr Lys Gln 4065	Asn Ala Tyr	
Thr Thr 4070	Lys Val Thr Asn Ala 4075	Glu His Ile Ile Ser 4080	Gly Thr Pro	
Thr Val 4085	Val Thr Thr Pro Ser 4090	Glu Val Thr Ala Ala 4095	Ala Asn Gln	
Val Asn 4100	Ser Ala Lys Gln Glu 4105	Leu Asn Gly Asp Glu 4110	Arg Leu Arg	
Glu Ala 4115	Lys Gln Asn Ala Asn 4120	Thr Ala Ile Asp Ala 4125	Leu Thr Gln	
Leu Asn 4130	Thr Pro Gln Lys Ala 4135	Lys Leu Lys Glu Gln 4140	Val Gly Gln	
Ala Asn 4145	Arg Leu Glu Asp Val 4150	Gln Thr Val Gln Thr 4155	Asn Gly Gln	
Ala Leu 4160	Asn Asn Ala Met Lys 4165	Gly Leu Arg Asp Ser 4170	Ile Ala Asn	
Glu Thr 4175	Thr Val Lys Thr Ser 4180	Gln Asn Tyr Thr Asp 4185	Ala Ser Pro	
Asn Asn 4190	Gln Ser Thr Tyr Asn 4195	Ser Ala Val Ser Asn 4200	Ala Lys Gly	
Ile Ile 4205	Asn Gln Thr Asn Asn 4210	Pro Thr Met Asp Thr 4215	Ser Ala Ile	
Thr Gln 4220	Ala Thr Thr Gln Val 4225	Asn Asn Ala Lys Asn 4230	Gly Leu Asn	
Gly Ala 4235	Glu Asn Leu Arg Asn 4240	Ala Gln Asn Thr Ala 4245	Lys Gln Asn	
Leu Asn 4250	Thr Leu Ser His Leu 4255	Thr Asn Asn Gln Lys 4260	Ser Ala Ile	
Ser Ser 4265	Gln Ile Asp Arg Ala 4270	Gly His Val Ser Glu 4275	Val Thr Ala	
Thr Lys 4280	Asn Ala Ala Thr Glu 4285	Leu Asn Thr Gln Met 4290	Gly Asn Leu	
Glu Gln 4295	Ala Ile His Asp Gln 4300	Asn Thr Val Lys Gln 4305	Ser Val Lys	
Phe Thr 4310	Asp Ala Asp Lys Ala 4315	Lys Arg Asp Ala Tyr 4320	Thr Asn Ala	
Val Ser	Arg Ala Glu Ala Ile	Leu Asn Lys Thr Gln	Gly Ala Asn	

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4325	4330	4335
Thr Ser Lys Gln Asp Val	Glu Ala Ala Ile Gln Asn	Val Ser Ser
4340	4345	4350
Ala Lys Asn Ala Leu Asn	Gly Asp Gln Asn Val Thr	Asn Ala Lys
4355	4360	4365
Asn Ala Ala Lys Asn Ala	Leu Asn Asn Leu Thr Ser	Ile Asn Asn
4370	4375	4380
Ala Gln Lys Arg Asp Leu	Thr Thr Lys Ile Asp Gln	Ala Thr Thr
4385	4390	4395
Val Ala Gly Val Glu Ala	Val Ser Asn Thr Ser Thr	Gln Leu Asn
4400	4405	4410
Thr Ala Met Ala Asn Leu	Gln Asn Gly Ile Asn Asp	Lys Thr Asn
4415	4420	4425
Thr Leu Ala Ser Glu Asn	Tyr His Asp Ala Asp Ser	Asp Lys Lys
4430	4435	4440
Thr Ala Tyr Thr Gln Ala	Val Thr Asn Ala Glu Asn	Ile Leu Asn
4445	4450	4455
Lys Asn Ser Gly Ser Asn	Leu Asp Lys Thr Ala Val	Glu Asn Ala
4460	4465	4470
Leu Ser Gln Val Ala Asn	Ala Lys Gly Ala Leu Asn	Gly Asn His
4475	4480	4485
Asn Leu Glu Gln Ala Lys	Ser Asn Ala Asn Thr Thr	Ile Asn Gly
4490	4495	4500
Leu Gln His Leu Thr Thr	Ala Gln Lys Asp Lys Leu	Lys Gln Gln
4505	4510	4515
Val Gln Gln Ala Gln Asn	Val Ala Gly Val Asp Thr	Val Lys Ser
4520	4525	4530
Ser Ala Asn Thr Leu Asn	Gly Ala Met Gly Thr Leu	Arg Asn Ser
4535	4540	4545
Ile Gln Asp Asn Thr Ala	Thr Lys Asn Gly Gln Asn	Tyr Leu Asp
4550	4555	4560
Ala Thr Glu Arg Asn Lys	Thr Asn Tyr Asn Asn Ala	Val Asp Ser
4565	4570	4575
Ala Asn Gly Val Ile Asn	Ala Thr Ser Asn Pro Asn	Met Asp Ala
4580	4585	4590
Asn Ala Ile Asn Gln Ile	Ala Thr Gln Val Thr Ser	Thr Lys Asn
4595	4600	4605
Ala Leu Asp Gly Thr His	Asn Leu Thr Gln Ala Lys	Gln Thr Ala
4610	4615	4620
Thr Asn Ala Ile Asp Gly	Ala Thr Asn Leu Asn Lys	Ala Gln Lys
4625	4630	4635
Asp Ala Leu Lys Ala Gln	Val Thr Ser Ala Gln Arg	Val Ala Asn
4640	4645	4650
Val Thr Ser Ile Gln Gln	Thr Ala Asn Glu Leu Asn	Thr Ala Met
4655	4660	4665
Gly Gln Leu Gln His Gly	Ile Asp Asp Glu Asn Ala	Thr Lys Gln
4670	4675	4680
Thr Gln Lys Tyr Arg Asp	Ala Glu Gln Ser Lys Lys	Thr Ala Tyr
4685	4690	4695
Asp Gln Ala Val Ala Ala	Ala Lys Ala Ile Leu Asn	Lys Gln Thr
4700	4705	4710
Gly Ser Asn Ser Asp Lys	Ala Ala Val Asp Arg Ala	Leu Gln Gln
4715	4720	4725

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Val Thr Ser Thr Lys Asp Ala Leu Asn Gly Asp Ala Lys Leu Ala 4730 4735 4740
Glu Ala Lys Ala Ala Ala Lys Gln Asn Leu Gly Thr Leu Asn His 4745 4750 4755
Ile Thr Asn Ala Gln Arg Thr Asp Leu Glu Gly Gln Ile Asn Gln 4760 4765 4770
Ala Thr Thr Val Asp Gly Val Asn Thr Val Lys Thr Asn Ala Asn 4775 4780 4785
Thr Leu Asp Gly Ala Met Asn Ser Leu Gln Gly Ser Ile Asn Asp 4790 4795 4800
Lys Asp Ala Thr Leu Arg Asn Gln Asn Tyr Leu Asp Ala Asp Glu 4805 4810 4815
Ser Lys Arg Asn Ala Tyr Thr Gln Ala Val Thr Ala Ala Glu Gly 4820 4825 4830
Ile Leu Asn Lys Gln Thr Gly Gly Asn Thr Ser Lys Ala Asp Val 4835 4840 4845
Asp Asn Ala Leu Asn Ala Val Thr Arg Ala Lys Ala Ala Leu Asn 4850 4855 4860
Gly Ala Asp Asn Leu Arg Asn Ala Lys Thr Ser Ala Thr Asn Thr 4865 4870 4875
Ile Asp Gly Leu Pro Asn Leu Thr Gln Leu Gln Lys Asp Asn Leu 4880 4885 4890
Lys His Gln Val Glu Gln Ala Gln Asn Val Ala Gly Val Asn Gly 4895 4900 4905
Val Lys Asp Lys Gly Asn Thr Leu Asn Thr Ala Met Gly Ala Leu 4910 4915 4920
Arg Thr Ser Ile Gln Asn Asp Asn Thr Thr Lys Thr Ser Gln Asn 4925 4930 4935
Tyr Leu Asp Ala Ser Asp Ser Asn Lys Asn Asn Tyr Asn Thr Ala 4940 4945 4950
Val Asn Asn Ala Asn Gly Val Ile Asn Ala Thr Asn Asn Pro Asn 4955 4960 4965
Met Asp Ala Asn Ala Ile Asn Gly Met Ala Asn Gln Val Asn Thr 4970 4975 4980
Thr Lys Ala Ala Leu Asn Gly Ala Gln Asn Leu Ala Gln Ala Lys 4985 4990 4995
Thr Asn Ala Thr Asn Thr Ile Asn Asn Ala His Asp Leu Asn Gln 5000 5005 5010
Lys Gln Lys Asp Ala Leu Lys Thr Gln Val Asn Asn Ala Gln Arg 5015 5020 5025
Val Ser Asp Ala Asn Asn Val Gln His Thr Ala Thr Glu Leu Asn 5030 5035 5040
Ser Ala Met Thr Ala Leu Lys Ala Ala Ile Ala Asp Lys Glu Arg 5045 5050 5055
Thr Lys Ala Ser Gly Asn Tyr Val Asn Ala Asp Gln Glu Lys Arg 5060 5065 5070
Gln Ala Tyr Asp Ser Lys Val Thr Asn Ala Glu Asn Ile Ile Ser 5075 5080 5085
Gly Thr Pro Asn Ala Thr Leu Thr Val Asn Asp Val Asn Ser Ala 5090 5095 5100
Ala Ser Gln Val Asn Ala Ala Lys Thr Ala Leu Asn Gly Asp Asn 5105 5110 5115

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Asn 5120	Leu	Arg	Val	Ala	Lys	Glu 5125	His	Ala	Asn	Asn	Thr 5130	Ile	Asp	Gly
Leu 5135	Ala	Gln	Leu	Asn	Asn	Ala 5140	Gln	Lys	Ala	Lys	Leu 5145	Lys	Glu	Gln
Val 5150	Gln	Ser	Ala	Thr	Thr	Leu 5155	Asp	Gly	Val	Gln	Thr 5160	Val	Lys	Asn
Ser 5165	Ser	Gln	Thr	Leu	Asn	Thr 5170	Ala	Met	Lys	Gly	Leu 5175	Arg	Asp	Ser
Ile 5180	Ala	Asn	Glu	Ala	Thr	Ile 5185	Lys	Ala	Gly	Gln	Asn 5190	Tyr	Thr	Asp
Ala 5195	Ser	Pro	Asn	Asn	Arg	Asn 5200	Glu	Tyr	Asp	Ser	Ala 5205	Val	Thr	Ala
Ala 5210	Lys	Ala	Ile	Ile	Asn	Gln 5215	Thr	Ser	Asn	Pro	Thr 5220	Met	Glu	Pro
Asn 5225	Thr	Ile	Thr	Gln	Val	Thr 5230	Ser	Gln	Val	Thr	Thr 5235	Lys	Glu	Gln
Ala 5240	Leu	Asn	Gly	Ala	Arg	Asn 5245	Leu	Ala	Gln	Ala	Lys 5250	Thr	Thr	Ala
Lys 5255	Asn	Asn	Leu	Asn	Asn	Leu 5260	Thr	Ser	Ile	Asn	Asn 5265	Ala	Gln	Lys
Asp 5270	Ala	Leu	Thr	Arg	Ser	Ile 5275	Asp	Gly	Ala	Thr	Thr 5280	Val	Ala	Gly
Val 5285	Asn	Gln	Glu	Thr	Ala	Lys 5290	Ala	Thr	Glu	Leu	Asn 5295	Asn	Ala	Met
His 5300	Ser	Leu	Gln	Asn	Gly	Ile 5305	Asn	Asp	Glu	Thr	Gln 5310	Thr	Lys	Gln
Thr 5315	Gln	Lys	Tyr	Leu	Asp	Ala 5320	Glu	Pro	Ser	Lys	Lys 5325	Ser	Ala	Tyr
Asp 5330	Gln	Ala	Val	Asn	Ala	Ala 5335	Lys	Ala	Ile	Leu	Thr 5340	Lys	Ala	Ser
Gly 5345	Gln	Asn	Val	Asp	Lys	Ala 5350	Ala	Val	Glu	Gln	Ala 5355	Leu	Gln	Asn
Val 5360	Asn	Ser	Thr	Lys	Thr	Ala 5365	Leu	Asn	Gly	Asp	Ala 5370	Lys	Leu	Asn
Glu 5375	Ala	Lys	Ala	Ala	Ala	Lys 5380	Gln	Thr	Leu	Gly	Thr 5385	Leu	Thr	His
Ile 5390	Asn	Asn	Ala	Gln	Arg	Thr 5395	Ala	Leu	Asp	Asn	Glu 5400	Ile	Thr	Gln
Ala 5405	Thr	Asn	Val	Glu	Gly	Val 5410	Asn	Thr	Val	Lys	Ala 5415	Lys	Ala	Gln
Gln 5420	Leu	Asp	Gly	Ala	Met	Gly 5425	Gln	Leu	Glu	Thr	Ser 5430	Ile	Arg	Asp
Lys 5435	Asp	Thr	Thr	Leu	Gln	Ser 5440	Gln	Asn	Tyr	Gln	Asp 5445	Ala	Asp	Asp
Ala 5450	Lys	Arg	Thr	Ala	Tyr	Ser 5455	Gln	Ala	Val	Asn	Ala 5460	Ala	Ala	Thr
Ile 5465	Leu	Asn	Lys	Thr	Ala	Gly 5470	Gly	Asn	Thr	Pro	Lys 5475	Ala	Asp	Val
Glu 5480	Arg	Ala	Met	Gln	Ala	Val 5485	Thr	Gln	Ala	Asn	Thr 5490	Ala	Leu	Asn
Gly 5495	Ile	Gln	Asn	Leu	Asp	Arg 5500	Ala	Lys	Gln	Ala	Ala 5505	Asn	Thr	Ala
Ile 5510	Thr	Asn	Ala	Ser	Asp	Leu 5515	Asn	Thr	Lys	Gln	Lys 5520	Glu	Ala	Leu

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5510	5515	5520
Lys Ala Gln Val Thr Ser 5525	Ala Gly Arg Val Ser 5530	Ala Ala Asn Gly 5535
Val Glu His Thr Ala Thr 5540	Glu Leu Asn Thr Ala 5545	Met Thr Ala Leu 5550
Lys Arg Ala Ile Ala Asp 5555	Lys Ala Glu Thr Lys 5560	Ala Ser Gly Asn 5565
Tyr Val Asn Ala Asp Ala 5570	Asn Lys Arg Gln Ala 5575	Tyr Asp Glu Lys 5580
Val Thr Ala Ala Glu Asn 5585	Ile Val Ser Gly Thr 5590	Pro Thr Pro Thr 5595
Leu Thr Pro Ala Asp Val 5600	Thr Asn Ala Ala Thr 5605	Gln Val Thr Asn 5610
Ala Lys Thr Gln Leu Asn 5615	Gly Asn His Asn Leu 5620	Glu Val Ala Lys 5625
Gln Asn Ala Asn Thr Ala 5630	Ile Asp Gly Leu Thr 5635	Ser Leu Asn Gly 5640
Pro Gln Lys Ala Lys Leu 5645	Lys Glu Gln Val Gly 5650	Gln Ala Thr Thr 5655
Leu Pro Asn Val Gln Thr 5660	Val Arg Asp Asn Ala 5665	Gln Thr Leu Asn 5670
Thr Ala Met Lys Gly Leu 5675	Arg Asp Ser Ile Ala 5680	Asn Glu Ala Thr 5685
Ile Lys Ala Gly Gln Asn 5690	Tyr Thr Asp Ala Ser 5695	Gln Asn Lys Gln 5700
Thr Asp Tyr Asn Ser Ala 5705	Val Thr Ala Ala Lys 5710	Ala Ile Ile Gly 5715
Gln Thr Thr Ser Pro Ser 5720	Met Asn Ala Gln Glu 5725	Ile Asn Gln Ala 5730
Lys Asp Gln Val Thr Ala 5735	Lys Gln Gln Ala Leu 5740	Asn Gly Gln Glu 5745
Asn Leu Arg Thr Ala Gln 5750	Thr Asn Ala Lys Gln 5755	His Leu Asn Gly 5760
Leu Ser Asp Leu Thr Asp 5765	Ala Gln Lys Asp Ala 5770	Val Lys Arg Gln 5775
Ile Glu Gly Ala Thr His 5780	Val Asn Glu Val Thr 5785	Gln Ala Gln Asn 5790
Asn Ala Asp Ala Leu Asn 5795	Thr Ala Met Thr Asn 5800	Leu Lys Asn Gly 5805
Ile Gln Asp Gln Asn Thr 5810	Ile Lys Gln Gly Val 5815	Asn Phe Thr Asp 5820
Ala Asp Glu Ala Lys Arg 5825	Asn Ala Tyr Thr Asn 5830	Ala Val Thr Gln 5835
Ala Glu Gln Ile Leu Asn 5840	Lys Ala Gln Gly Pro 5845	Asn Thr Ser Lys 5850
Asp Gly Val Glu Thr Ala 5855	Leu Glu Asn Val Gln 5860	Arg Ala Lys Asn 5865
Glu Leu Asn Gly Asn Gln 5870	Asn Val Ala Asn Ala 5875	Lys Thr Thr Ala 5880
Lys Asn Ala Leu Asn Asn 5885	Leu Thr Ser Ile Asn 5890	Asn Ala Gln Lys 5895
Glu Ala Leu Lys Ser Gln 5900	Ile Glu Gly Ala Thr 5905	Thr Val Ala Gly 5910

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Val	Asn	Gln	Val	Ser	Thr	Thr	Ala	Ser	Glu	Leu	Asn	Thr	Ala	Met
5915						5920					5925			
Ser	Asn	Leu	Gln	Asn	Gly	Ile	Asn	Asp	Glu	Ala	Ala	Thr	Lys	Ala
5930						5935					5940			
Ala	Gln	Lys	Tyr	Thr	Asp	Ala	Asp	Arg	Glu	Lys	Gln	Thr	Ala	Tyr
5945						5950					5955			
Asn	Asp	Ala	Val	Thr	Ala	Ala	Lys	Thr	Leu	Leu	Asp	Lys	Thr	Ala
5960						5965					5970			
Gly	Ser	Asn	Asp	Asn	Lys	Ala	Ala	Val	Glu	Gln	Ala	Leu	Gln	Arg
5975						5980					5985			
Val	Asn	Thr	Ala	Lys	Thr	Ala	Leu	Asn	Gly	Asp	Glu	Arg	Leu	Asn
5990						5995					6000			
Glu	Ala	Lys	Asn	Thr	Ala	Lys	Gln	Gln	Val	Ala	Thr	Met	Ser	His
6005						6010					6015			
Leu	Thr	Asp	Ala	Gln	Lys	Ala	Asn	Leu	Thr	Ser	Gln	Ile	Glu	Ser
6020						6025					6030			
Gly	Thr	Thr	Val	Ala	Gly	Val	Gln	Gly	Ile	Gln	Ala	Asn	Ala	Gly
6035						6040					6045			
Thr	Leu	Asp	Gln	Ala	Met	Asn	Gln	Leu	Arg	Gln	Ser	Ile	Ala	Ser
6050						6055					6060			
Lys	Asp	Ala	Thr	Lys	Ser	Ser	Glu	Asp	Tyr	Gln	Asp	Ala	Asn	Ala
6065						6070					6075			
Asp	Leu	Gln	Asn	Ala	Tyr	Asn	Asp	Ala	Val	Thr	Asn	Ala	Glu	Gly
6080						6085					6090			
Ile	Ile	Ser	Ala	Thr	Asn	Asn	Pro	Glu	Met	Asn	Pro	Asp	Thr	Ile
6095						6100					6105			
Asn	Gln	Lys	Ala	Ser	Gln	Val	Asn	Ser	Ala	Lys	Ser	Ala	Leu	Asn
6110						6115					6120			
Gly	Asp	Glu	Lys	Leu	Ala	Ala	Ala	Lys	Gln	Thr	Ala	Lys	Ser	Asp
6125						6130					6135			
Ile	Gly	Arg	Leu	Thr	Asp	Leu	Asn	Asn	Ala	Gln	Arg	Thr	Ala	Ala
6140						6145					6150			
Asn	Ala	Glu	Val	Asp	Gln	Ala	Pro	Asn	Leu	Ala	Ala	Val	Thr	Ala
6155						6160					6165			
Ala	Lys	Asn	Lys	Ala	Thr	Ser	Leu	Asn	Thr	Ala	Met	Gly	Asn	Leu
6170						6175					6180			
Lys	His	Ala	Leu	Ala	Glu	Lys	Asp	Asn	Thr	Lys	Arg	Ser	Val	Asn
6185						6190					6195			
Tyr	Thr	Asp	Ala	Asp	Gln	Pro	Lys	Gln	Gln	Ala	Tyr	Asp	Thr	Ala
6200						6205					6210			
Val	Thr	Gln	Ala	Glu	Ala	Ile	Thr	Asn	Ala	Asn	Gly	Ser	Asn	Ala
6215						6220					6225			
Asn	Glu	Thr	Gln	Val	Gln	Ala	Ala	Leu	Asn	Gln	Leu	Asn	Gln	Ala
6230						6235					6240			
Lys	Asn	Asp	Leu	Asn	Gly	Asp	Asn	Lys	Val	Ala	Gln	Ala	Lys	Glu
6245						6250					6255			
Ser	Ala	Lys	Arg	Ala	Leu	Ala	Ser	Tyr	Ser	Asn	Leu	Asn	Asn	Ala
6260						6265					6270			
Gln	Ser	Thr	Ala	Ala	Ile	Ser	Gln	Ile	Asp	Asn	Ala	Thr	Thr	Val
6275						6280					6285			
Ala	Gly	Val	Thr	Ala	Ala	Gln	Asn	Thr	Ala	Asn	Glu	Leu	Asn	Thr
6290						6295					6300			

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Ala Met 6305	Gly Gln Leu Gln 6310	Asn Gly Ile Asn Asp 6315	Gln Asn Thr Val 6315
Lys Gln 6320	Gln Val Asn Phe Thr 6325	Asp Ala Asp Gln Gly 6330	Lys Lys Asp 6330
Ala Tyr 6335	Thr Asn Ala Val Thr 6340	Asn Ala Gln Gly Ile 6345	Leu Asp Lys 6345
Ala His 6350	Gly Gln Asn Met Thr 6355	Lys Ala Gln Val Glu 6360	Ala Ala Leu 6360
Asn Gln 6365	Val Thr Thr Ala Lys 6370	Asn Ala Leu Asn Gly 6375	Asp Ala Asn 6375
Val Arg 6380	Gln Ala Lys Ser Asp 6385	Ala Lys Ala Asn Leu 6390	Gly Thr Leu 6390
Thr His 6395	Leu Asn Asn Ala Gln 6400	Lys Gln Asp Leu Thr 6405	Ser Gln Ile 6405
Glu Gly 6410	Ala Thr Thr Val Asn 6415	Gly Val Asn Gly Val 6420	Lys Thr Lys 6420
Ala Gln 6425	Asp Leu Asp Gly Ala 6430	Met Gln Arg Leu Gln 6435	Ser Ala Ile 6435
Ala Asn 6440	Lys Asp Gln Thr Lys 6445	Ala Ser Glu Asn Tyr 6450	Ile Asp Ala 6450
Asp Pro 6455	Thr Lys Lys Thr Ala 6460	Phe Asp Asn Ala Ile 6465	Thr Gln Ala 6465
Glu Ser 6470	Tyr Leu Asn Lys Asp 6475	His Gly Ala Asn Lys 6480	Asp Lys Gln 6480
Ala Val 6485	Glu Gln Ala Ile Gln 6490	Ser Val Thr Ser Thr 6495	Glu Asn Ala 6495
Leu Asn 6500	Gly Asp Ala Asn Leu 6505	Gln Arg Ala Lys Thr 6510	Glu Ala Ile 6510
Gln Ala 6515	Ile Asp Asn Leu Thr 6520	His Leu Asn Thr Pro 6525	Gln Lys Thr 6525
Ala Leu 6530	Lys Gln Gln Val Asn 6535	Ala Ala Gln Arg Val 6540	Ser Gly Val 6540
Thr Asp 6545	Leu Lys Asn Ser Ala 6550	Thr Ser Leu Asn Asn 6555	Ala Met Asp 6555
Gln Leu 6560	Lys Gln Ala Ile Ala 6565	Asp His Asp Thr Ile 6570	Val Ala Ser 6570
Gly Asn 6575	Tyr Thr Asn Ala Ser 6580	Pro Asp Lys Gln Gly 6585	Ala Tyr Thr 6585
Asp Ala 6590	Tyr Asn Ala Ala Lys 6595	Asn Ile Val Asn Gly 6600	Ser Pro Asn 6600
Val Ile 6605	Thr Asn Ala Ala Asp 6610	Val Thr Ala Ala Thr 6615	Gln Arg Val 6615
Asn Asn 6620	Ala Glu Thr Gly Leu 6625	Asn Gly Asp Thr Asn 6630	Leu Ala Thr 6630
Ala Lys 6635	Gln Gln Ala Lys Asp 6640	Ala Leu Arg Gln Met 6645	Thr His Leu 6645
Ser Asp 6650	Ala Gln Lys Gln Ser 6655	Ile Thr Gly Gln Ile 6660	Asp Ser Ala 6660
Thr Gln 6665	Val Thr Gly Val Gln 6670	Ser Val Lys Asp Asn 6675	Ala Thr Asn 6675
Leu Asp 6680	Asn Ala Met Asn Gln 6685	Leu Arg Asn Ser Ile 6690	Ala Asn Lys 6690
Asp Asp 6695	Val Lys Ala Ser Gln 6700	Pro Tyr Val Asp Ala 6705	Asp Arg Asp 6705

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6695	6700	6705
Lys Gln Asn Ala Tyr Asn Thr 6710	Ala Val Thr Asn Ala 6715	Glu Asn Ile 6720
Ile Asn Ala Thr Ser Gln Pro 6725	Thr Leu Asp Pro Ser 6730	Ala Val Thr 6735
Gln Ala Ala Asn Gln Val Ser 6740	Thr Asn Lys Thr Ala 6745	Leu Asn Gly 6750
Ala Gln Asn Leu Ala Asn Lys 6755	Lys Gln Glu Thr Thr 6760	Ala Asn Ile 6765
Asn Gln Leu Ser His Leu Asn 6770	Asn Ala Gln Lys Gln 6775	Asp Leu Asn 6780
Thr Gln Val Thr Asn Ala Pro 6785	Asn Ile Ser Thr Val 6790	Asn Gln Val 6795
Lys Thr Lys Ala Glu Gln Leu 6800	Asp Gln Ala Met Glu 6805	Arg Leu Ile 6810
Asn Gly Ile Gln Asp Lys Asp 6815	Gln Val Lys Gln Ser 6820	Val Asn Phe 6825
Thr Asp Ala Asp Pro Glu Lys 6830	Gln Thr Ala Tyr Asn 6835	Asn Ala Val 6840
Thr Ala Ala Glu Asn Ile Ile 6845	Asn Gln Ala Asn Gly 6850	Thr Asn Ala 6855
Asn Gln Ser Gln Val Glu Ala 6860	Ala Leu Ser Thr Val 6865	Thr Thr Thr 6870
Lys Gln Ala Leu Asn Gly Asp 6875	Arg Lys Val Thr Asp 6880	Ala Lys Asn 6885
Asn Ala Asn Gln Thr Leu Ser 6890	Thr Leu Asp Asn Leu 6895	Asn Asn Ala 6900
Gln Lys Gly Ala Val Thr Gly 6905	Asn Ile Asn Gln Ala 6910	His Thr Val 6915
Ala Glu Val Thr Gln Ala Ile 6920	Gln Thr Ala Gln Glu 6925	Leu Asn Thr 6930
Ala Met Gly Asn Leu Lys Asn 6935	Ser Leu Asn Asp Lys 6940	Asp Thr Thr 6945
Leu Gly Ser Gln Asn Phe Ala 6950	Asp Ala Asp Pro Glu 6955	Lys Lys Asn 6960
Ala Tyr Asn Glu Ala Val His 6965	Asn Ala Glu Asn Ile 6970	Leu Asn Lys 6975
Ser Thr Gly Thr Asn Val Pro 6980	Lys Asp Gln Val Glu 6985	Ala Ala Met 6990
Asn Gln Val Asn Ala Thr Lys 6995	Ala Ala Leu Asn Gly 7000	Thr Gln Asn 7005
Leu Glu Lys Ala Lys Gln His 7010	Ala Asn Thr Ala Ile 7015	Asp Gly Leu 7020
Ser His Leu Thr Asn Ala Gln 7025	Lys Glu Ala Leu Lys 7030	Gln Leu Val 7035
Gln Gln Ser Thr Thr Val Ala 7040	Glu Ala Gln Gly Asn 7045	Glu Gln Lys 7050
Ala Asn Asn Val Asp Ala Ala 7055	Met Asp Lys Leu Arg 7060	Gln Ser Ile 7065
Ala Asp Asn Ala Thr Thr Lys 7070	Gln Asn Gln Asn Tyr 7075	Thr Asp Ala 7080
Ser Gln Asn Lys Lys Asp Ala 7085	Tyr Asn Asn Ala Val 7090	Thr Thr Ala 7095

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Gln Gly	Ile Ile Asp	Gln Thr	Thr Ser Pro	Thr Leu	Asp Pro Thr	7100	7105	7110
Val Ile	Asn Gln Ala Ala	Gly	Gln Val Ser Thr	Thr	Lys Asn Ala	7115	7120	7125
Leu Asn	Gly Asn Glu Asn	Leu	Glu Ala Ala Lys	Gln	Gln Ala Ser	7130	7135	7140
Gln Ser	Leu Gly Ser Leu	Asp	Asn Leu Asn Asn	Ala	Gln Lys Gln	7145	7150	7155
Thr Val	Thr Asp Gln Ile	Asn	Gly Ala His Thr	Val	Asp Glu Ala	7160	7165	7170
Asn Gln	Ile Lys Gln Asn	Ala	Gln Asn Leu Asn	Thr	Ala Met Gly	7175	7180	7185
Asn Leu	Lys Gln Ala Ile	Ala	Asp Lys Asp Ala	Thr	Lys Ala Thr	7190	7195	7200
Val Asn	Phe Thr Asp Ala	Asp	Gln Ala Lys Gln	Gln	Ala Tyr Asn	7205	7210	7215
Thr Ala	Val Thr Asn Ala	Glu	Asn Ile Ser Lys	Ala	Asn Gly Asn	7220	7225	7230
Ala Thr	Gln Ala Glu Val	Glu	Gln Ala Ile Lys	Gln	Val Asn Ala	7235	7240	7245
Ala Lys	Gln Ala Leu Asn	Gly	Asn Ala Asn Val	Gln	His Ala Lys	7250	7255	7260
Asp Glu	Ala Thr Ala Leu	Ile	Asn Ser Ser Asn	Asp	Leu Asn Gln	7265	7270	7275
Ala Gln	Lys Asp Ala Leu	Lys	Gln Gln Val Gln	Asn	Ala Thr Thr	7280	7285	7290
Val Ala	Gly Val Asn Asn	Val	Lys Gln Thr Ala	Gln	Glu Leu Asn	7295	7300	7305
Asn Ala	Met Thr Gln Leu	Lys	Gln Gly Ile Ala	Asp	Lys Glu Gln	7310	7315	7320
Thr Lys	Ala Asp Gly Asn	Phe	Val Asn Ala Asp	Pro	Asp Lys Gln	7325	7330	7335
Asn Ala	Tyr Asn Gln Ala	Val	Ala Lys Ala Glu	Ala	Leu Ile Ser	7340	7345	7350
Ala Thr	Pro Asp Val Val	Val	Thr Pro Ser Glu	Ile	Thr Ala Ala	7355	7360	7365
Leu Asn	Lys Val Thr Gln	Ala	Lys Asn Asp Leu	Asn	Gly Asn Thr	7370	7375	7380
Asn Leu	Ala Thr Ala Lys	Gln	Asn Val Gln His	Ala	Ile Asp Gln	7385	7390	7395
Leu Pro	Asn Leu Asn Gln	Ala	Gln Arg Asp Glu	Tyr	Ser Lys Gln	7400	7405	7410
Ile Thr	Gln Ala Thr Leu	Val	Pro Asn Val Asn	Ala	Ile Gln Gln	7415	7420	7425
Ala Ala	Thr Thr Leu Asn	Asp	Ala Met Thr Gln	Leu	Lys Gln Gly	7430	7435	7440
Ile Ala	Asn Lys Ala Gln	Ile	Lys Gly Ser Glu	Asn	Tyr His Asp	7445	7450	7455
Ala Asp	Thr Asp Lys Gln	Thr	Ala Tyr Asp Asn	Ala	Val Thr Lys	7460	7465	7470
Ala Glu	Glu Leu Leu Lys	Gln	Thr Thr Asn Pro	Thr	Met Asp Pro	7475	7480	7485

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Asn Thr 7490	Ile Gln Gln Ala 7495	Leu Thr Lys Val 7495	Asn Asp 7500	Thr Asn Gln
Ala Leu 7505	Asn Gly Asn Gln Lys 7510	Leu Ala Asp Ala Lys 7515	Gln Asp Ala	
Lys Thr 7520	Thr Leu Gly Thr 7525	Leu Asp His Leu Asn Asp 7530	Ala Gln Lys	
Gln Ala 7535	Leu Thr Thr Gln Val 7540	Glu Gln Ala Pro Asp 7545	Ile Ala Thr	
Val Asn 7550	Asn Val Lys Gln Asn 7555	Ala Gln Asn Leu Asn 7560	Asn Ala Met	
Thr Asn 7565	Leu Asn Asn Ala 7570	Gln Asp Lys Thr Glu 7575	Thr Leu Asn	
Ser Ile 7580	Asn Phe Thr Asp Ala 7585	Asp Gln Ala Lys Lys 7590	Asp Ala Tyr	
Thr Asn 7595	Ala Val Ser His Ala 7600	Glu Gly Ile Leu Ser 7605	Lys Ala Asn	
Gly Ser 7610	Asn Ala Ser Gln Thr 7615	Glu Val Glu Gln Ala 7620	Met Gln Arg	
Val Asn 7625	Glu Ala Lys Gln Ala 7630	Leu Asn Gly Asn Asp 7635	Asn Val Gln	
Arg Ala 7640	Lys Asp Ala Ala Lys 7645	Gln Val Ile Thr Asn 7650	Ala Asn Asp	
Leu Asn 7655	Gln Ala Gln Lys Asp 7660	Ala Leu Lys Gln Gln 7665	Val Asp Ala	
Ala Gln 7670	Thr Val Ala Asn Val 7675	Asn Thr Ile Lys Gln 7680	Thr Ala Gln	
Asp Leu 7685	Asn Gln Ala Met Thr 7690	Gln Leu Lys Gln Gly 7695	Ile Ala Asp	
Lys Asp 7700	Gln Thr Lys Ala Asn 7705	Gly Asn Phe Val Asn 7710	Ala Asp Thr	
Asp Lys 7715	Gln Asn Ala Tyr Asn 7720	Asn Ala Val Ala His 7725	Ala Glu Gln	
Ile Ile 7730	Ser Gly Thr Pro Asn 7735	Ala Asn Val Asp Pro 7740	Gln Gln Val	
Ala Gln 7745	Ala Leu Gln Gln Val 7750	Asn Gln Ala Lys Gly 7755	Asp Leu Asn	
Gly Asn 7760	His Asn Leu Gln Val 7765	Ala Lys Asp Asn Ala 7770	Asn Thr Ala	
Ile Asp 7775	Gln Leu Pro Asn Leu 7780	Asn Gln Pro Gln Lys 7785	Thr Ala Leu	
Lys Asp 7790	Gln Val Ser His Ala 7795	Glu Leu Val Thr Gly 7800	Val Asn Ala	
Ile Lys 7805	Gln Asn Ala Asp Ala 7810	Leu Asn Asn Ala Met 7815	Gly Thr Leu	
Lys Gln 7820	Gln Ile Gln Ala Asn 7825	Ser Gln Val Pro Gln 7830	Ser Val Asp	
Phe Thr 7835	Gln Ala Asp Gln Asp 7840	Lys Gln Gln Ala Tyr 7845	Asn Asn Ala	
Ala Asn 7850	Gln Ala Gln Gln Ile 7855	Ala Asn Gly Ile Pro 7860	Thr Pro Val	
Leu Thr 7865	Pro Asp Thr Val Thr 7870	Gln Ala Val Thr Thr 7875	Met Asn Gln	
Ala Lys	Asp Ala Leu Asn Gly	Asp Glu Lys Leu Ala	Gln Ala Lys	

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7880	7885	7890
Gln Glu Ala Leu Ala Asn 7895	Leu Asp Thr Leu Arg Asp 7900	Leu Asn Gln 7905
Pro Gln Arg Asp Ala Leu 7910	Arg Asn Gln Ile Asn 7915	Gln Ala Gln Ala 7920
Leu Ala Thr Val Glu Gln 7925	Thr Lys Gln Asn Ala 7930	Gln Asn Val Asn 7935
Thr Ala Met Ser Asn Leu 7940	Lys Gln Gly Ile Ala 7945	Asn Lys Asp Thr 7950
Val Lys Ala Ser Glu Asn 7955	Tyr His Asp Ala Asp 7960	Ala Asp Lys Gln 7965
Thr Ala Tyr Thr Asn Ala 7970	Val Ser Gln Ala Glu 7975	Gly Ile Ile Asn 7980
Gln Thr Thr Asn Pro Thr 7985	Leu Asn Pro Asp Glu 7990	Ile Thr Arg Ala 7995
Leu Thr Gln Val Thr Asp 8000	Ala Lys Asn Gly Leu 8005	Asn Gly Glu Ala 8010
Lys Leu Ala Thr Glu Lys 8015	Gln Asn Ala Lys Asp 8020	Ala Val Ser Gly 8025
Met Thr His Leu Asn Asp 8030	Ala Gln Lys Gln Ala 8035	Leu Lys Gly Gln 8040
Ile Asp Gln Ser Pro Glu 8045	Ile Ala Thr Val Asn 8050	Gln Val Lys Gln 8055
Thr Ala Thr Ser Leu Asp 8060	Gln Ala Met Asp Gln 8065	Leu Ser Gln Ala 8070
Ile Asn Asp Lys Ala Gln 8075	Thr Leu Ala Asp Gly 8080	Asn Tyr Leu Asn 8085
Ala Asp Pro Asp Lys Gln 8090	Asn Ala Tyr Lys Gln 8095	Ala Val Ala Lys 8100
Ala Glu Ala Leu Leu Asn 8105	Lys Gln Ser Gly Thr 8110	Asn Glu Val Gln 8115
Ala Gln Val Glu Ser Ile 8120	Thr Asn Glu Val Asn 8125	Ala Ala Lys Gln 8130
Ala Leu Asn Gly Asn Asp 8135	Asn Leu Ala Asn Ala 8140	Lys Gln Gln Ala 8145
Lys Gln Gln Leu Ala Asn 8150	Leu Thr His Leu Asn 8155	Asp Ala Gln Lys 8160
Gln Ser Phe Glu Ser Gln 8165	Ile Thr Gln Ala Pro 8170	Leu Val Thr Asp 8175
Val Thr Thr Ile Asn Gln 8180	Lys Ala Gln Thr Leu 8185	Asp His Ala Met 8190
Glu Leu Leu Arg Asn Ser 8195	Val Ala Asp Asn Gln 8200	Thr Thr Leu Ala 8205
Ser Glu Asp Tyr His Asp 8210	Ala Thr Ala Gln Arg 8215	Gln Asn Asp Tyr 8220
Asn Gln Ala Val Thr Ala 8225	Ala Asn Asn Ile Ile 8230	Asn Gln Thr Thr 8235
Ser Pro Thr Met Asn Pro 8240	Asp Asp Val Asn Gly 8245	Ala Thr Thr Gln 8250
Val Asn Asn Thr Lys Val 8255	Ala Leu Asp Gly Asp 8260	Glu Asn Leu Ala 8265
Ala Ala Lys Gln Gln Ala 8270	Asn Asn Arg Leu Asp 8275	Gln Leu Asp His 8280

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Leu	Asn	Asn	Ala	Gln	Lys	Gln	Gln	Leu	Gln	Ser	Gln	Ile	Thr	Gln
8285						8290					8295			
Ser	Ser	Asp	Ile	Ala	Ala	Val	Asn	Gly	His	Lys	Gln	Thr	Ala	Glu
8300						8305					8310			
Ser	Leu	Asn	Thr	Ala	Met	Gly	Asn	Leu	Ile	Asn	Ala	Ile	Ala	Asp
8315						8320					8325			
His	Gln	Ala	Val	Glu	Gln	Arg	Gly	Asn	Phe	Ile	Asn	Ala	Asp	Thr
8330						8335					8340			
Asp	Lys	Gln	Thr	Ala	Tyr	Asn	Thr	Ala	Val	Asn	Glu	Ala	Ala	Ala
8345						8350					8355			
Met	Ile	Asn	Lys	Gln	Thr	Gly	Gln	Asn	Ala	Asn	Gln	Thr	Glu	Val
8360						8365					8370			
Glu	Gln	Ala	Ile	Thr	Lys	Val	Gln	Thr	Thr	Leu	Gln	Ala	Leu	Asn
8375						8380					8385			
Gly	Asp	His	Asn	Leu	Gln	Val	Ala	Lys	Thr	Asn	Ala	Thr	Gln	Ala
8390						8395					8400			
Ile	Asp	Ala	Leu	Thr	Ser	Leu	Asn	Asp	Pro	Gln	Lys	Thr	Ala	Leu
8405						8410					8415			
Lys	Asp	Gln	Val	Thr	Ala	Ala	Thr	Leu	Val	Thr	Ala	Val	His	Gln
8420						8425					8430			
Ile	Glu	Gln	Asn	Ala	Asn	Thr	Leu	Asn	Gln	Ala	Met	His	Gly	Leu
8435						8440					8445			
Arg	Gln	Ser	Ile	Gln	Asp	Asn	Ala	Ala	Thr	Lys	Ala	Asn	Ser	Lys
8450						8455					8460			
Tyr	Ile	Asn	Glu	Asp	Gln	Pro	Glu	Gln	Gln	Asn	Tyr	Asp	Gln	Ala
8465						8470					8475			
Val	Gln	Ala	Ala	Asn	Asn	Ile	Ile	Asn	Glu	Gln	Thr	Ala	Thr	Leu
8480						8485					8490			
Asp	Asn	Asn	Ala	Ile	Asn	Gln	Ala	Ala	Thr	Thr	Val	Asn	Thr	Thr
8495						8500					8505			
Lys	Ala	Ala	Leu	His	Gly	Asp	Val	Lys	Leu	Gln	Asn	Asp	Lys	Asp
8510						8515					8520			
His	Ala	Lys	Gln	Thr	Val	Ser	Gln	Leu	Ala	His	Leu	Asn	Asn	Ala
8525						8530					8535			
Gln	Lys	His	Met	Glu	Asp	Thr	Leu	Ile	Asp	Ser	Glu	Thr	Thr	Arg
8540						8545					8550			
Thr	Ala	Val	Lys	Gln	Asp	Leu	Thr	Glu	Ala	Gln	Ala	Leu	Asp	Gln
8555						8560					8565			
Leu	Met	Asp	Ala	Leu	Gln	Gln	Ser	Ile	Ala	Asp	Lys	Asp	Ala	Thr
8570						8575					8580			
Arg	Ala	Ser	Ser	Ala	Tyr	Val	Asn	Ala	Glu	Pro	Asn	Lys	Lys	Gln
8585						8590					8595			
Ser	Tyr	Asp	Glu	Ala	Val	Gln	Asn	Ala	Glu	Ser	Ile	Ile	Ala	Gly
8600						8605					8610			
Leu	Asn	Asn	Pro	Thr	Ile	Asn	Lys	Gly	Asn	Val	Ser	Ser	Ala	Thr
8615						8620					8625			
Gln	Ala	Val	Ile	Ser	Ser	Lys	Asn	Ala	Leu	Asp	Gly	Val	Glu	Arg
8630						8635					8640			
Leu	Ala	Gln	Asp	Lys	Gln	Thr	Ala	Gly	Asn	Ser	Leu	Asn	His	Leu
8645						8650					8655			
Asp	Gln	Leu	Thr	Pro	Ala	Gln	Gln	Gln	Ala	Leu	Glu	Asn	Gln	Ile
8660						8665					8670			

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Asn 8675	Asn	Ala	Thr	Thr	Arg	Gly 8680	Glu	Val	Ala	Gln	Lys 8685	Leu	Thr	Glu
Ala 8690	Gln	Ala	Leu	Asn	Gln	Ala 8695	Met	Glu	Ala	Leu	Arg 8700	Asn	Ser	Ile
Gln 8705	Asp	Gln	Gln	Gln	Thr	Glu 8710	Ala	Gly	Ser	Lys	Phe 8715	Ile	Asn	Glu
Asp 8720	Lys	Pro	Gln	Lys	Asp	Ala 8725	Tyr	Gln	Ala	Ala	Val 8730	Gln	Asn	Ala
Lys 8735	Asp	Leu	Ile	Asn	Gln	Thr 8740	Asn	Asn	Pro	Thr	Leu 8745	Asp	Lys	Ala
Gln 8750	Val	Glu	Gln	Leu	Thr	Gln 8755	Ala	Val	Asn	Gln	Ala 8760	Lys	Asp	Asn
Leu 8765	His	Gly	Asp	Gln	Lys	Leu 8770	Ala	Asp	Asp	Lys	Gln 8775	His	Ala	Val
Thr 8780	Asp	Leu	Asn	Gln	Leu	Asn 8785	Gly	Leu	Asn	Asn	Pro 8790	Gln	Arg	Gln
Ala 8795	Leu	Glu	Ser	Gln	Ile	Asn 8800	Asn	Ala	Ala	Thr	Arg 8805	Gly	Glu	Val
Ala 8810	Gln	Lys	Leu	Ala	Glu	Ala 8815	Lys	Ala	Leu	Asp	Gln 8820	Ala	Met	Gln
Ala 8825	Leu	Arg	Asn	Ser	Ile	Gln 8830	Asp	Gln	Gln	Gln	Thr 8835	Glu	Ser	Gly
Ser 8840	Lys	Phe	Ile	Asn	Glu	Asp 8845	Lys	Pro	Gln	Lys	Asp 8850	Ala	Tyr	Gln
Ala 8855	Ala	Val	Gln	Asn	Ala	Lys 8860	Asp	Leu	Ile	Asn	Gln 8865	Thr	Gly	Asn
Pro 8870	Thr	Leu	Asp	Lys	Ser	Gln 8875	Val	Glu	Gln	Leu	Thr 8880	Gln	Ala	Val
Thr 8885	Thr	Ala	Lys	Asp	Asn	Leu 8890	His	Gly	Asp	Gln	Lys 8895	Leu	Ala	Arg
Asp 8900	Gln	Gln	Gln	Ala	Val	Thr 8905	Thr	Val	Asn	Ala	Leu 8910	Pro	Asn	Leu
Asn 8915	His	Ala	Gln	Gln	Gln	Ala 8920	Leu	Thr	Asp	Ala	Ile 8925	Asn	Ala	Ala
Pro 8930	Thr	Arg	Thr	Glu	Val	Ala 8935	Gln	His	Val	Gln	Thr 8940	Ala	Thr	Glu
Leu 8945	Asp	His	Ala	Met	Glu	Thr 8950	Leu	Lys	Asn	Lys	Val 8955	Asp	Gln	Val
Asn 8960	Thr	Asp	Lys	Ala	Gln	Pro 8965	Asn	Tyr	Thr	Glu	Ala 8970	Ser	Thr	Asp
Lys 8975	Lys	Glu	Ala	Val	Asp	Gln 8980	Ala	Leu	Gln	Ala	Ala 8985	Glu	Ser	Ile
Thr 8990	Asp	Pro	Thr	Asn	Gly	Ser 8995	Asn	Ala	Asn	Lys	Asp 9000	Ala	Val	Asp
Gln 9005	Val	Leu	Thr	Lys	Leu	Gln 9010	Glu	Lys	Glu	Asn	Glu 9015	Leu	Asn	Gly
Asn 9020	Glu	Arg	Val	Ala	Glu	Ala 9025	Lys	Thr	Gln	Ala	Lys 9030	Gln	Thr	Ile
Asp 9035	Gln	Leu	Thr	His	Leu	Asn 9040	Ala	Asp	Gln	Ile	Ala 9045	Thr	Ala	Lys
Gln 9050	Asn	Ile	Asp	Gln	Ala	Thr 9055	Lys	Leu	Gln	Pro	Ile 9060	Ala	Glu	Leu
Val 9065	Asp	Gln	Ala	Thr	Gln	Leu 9070	Asn	Gln	Ser	Met	Asp 9075	Gln	Leu	Gln

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9065	9070	9075
Gln Ala Val Asn Glu His 9080	Ala Asn Val Glu Gln Thr 9085	Val Asp Tyr 9090
Thr Gln Ala Asp Ser Asp 9095	Lys Gln Asn Ala Tyr Lys 9100	Gln Ala Ile 9105
Ala Asp Ala Glu Asn Val 9110	Leu Lys Gln Asn Ala 9115	Lys Gln Gln 9120
Val Asp Gln Ala Leu Gln 9125	Asn Ile Leu Asn Ala Lys 9130	Gln Ala Leu 9135
Asn Gly Asp Glu Arg Val 9140	Ala Leu Ala Lys Thr Asn 9145	Gly Lys His 9150
Asp Ile Asp Gln Leu Asn 9155	Ala Leu Asn Asn Ala Gln 9160	Gln Asp Gly 9165
Phe Lys Gly Arg Ile Asp 9170	Gln Ser Asn Asp Leu Asn 9175	Gln Ile Gln 9180
Gln Ile Val Asp Glu Ala 9185	Lys Ala Leu Asn Arg Ala 9190	Met Asp Gln 9195
Leu Ser Gln Glu Ile Thr 9200	Asp Asn Glu Gly Arg Thr 9205	Lys Gly Ser 9210
Thr Asn Tyr Val Asn Ala 9215	Asp Thr Gln Val Lys Gln 9220	Val Tyr Asp 9225
Glu Thr Val Asp Lys Ala 9230	Lys Gln Ala Leu Asp Lys 9235	Ser Thr Gly 9240
Gln Asn Leu Thr Ala Lys 9245	Gln Val Ile Lys Leu Asn 9250	Asp Ala Val 9255
Thr Ala Ala Lys Lys Ala 9260	Leu Asn Gly Glu Glu Arg 9265	Leu Asn Asn 9270
Arg Lys Ala Glu Ala Leu 9275	Gln Arg Leu Asp Gln Leu 9280	Thr His Leu 9285
Asn Asn Ala Gln Arg Gln 9290	Leu Ala Ile Gln Gln Ile 9295	Asn Asn Ala 9300
Glu Thr Leu Asn Lys Ala 9305	Ser Arg Ala Ile Asn Arg 9310	Ala Thr Lys 9315
Leu Asp Asn Ala Met Gly 9320	Ala Val Gln Gln Tyr Ile 9325	Asp Glu Gln 9330
His Leu Gly Val Ile Ser 9335	Ser Thr Asn Tyr Ile Asn 9340	Ala Asp Asp 9345
Asn Leu Lys Ala Asn Tyr 9350	Asp Asn Ala Ile Ala Asn 9355	Ala Ala His 9360
Glu Leu Asp Lys Val Gln 9365	Gly Asn Ala Ile Ala Lys 9370	Ala Glu Ala 9375
Glu Gln Leu Lys Gln Asn 9380	Ile Ile Asp Ala Gln Asn 9385	Ala Leu Asn 9390
Gly Asp Gln Asn Leu Ala 9395	Asn Ala Lys Asp Lys Ala 9400	Asn Ala Phe 9405
Val Asn Ser Leu Asn Gly 9410	Leu Asn Gln Gln Gln Gln 9415	Asp Leu Ala 9420
His Lys Ala Ile Asn Asn 9425	Ala Asp Thr Val Ser Asp 9430	Val Thr Asp 9435
Ile Val Asn Asn Gln Ile 9440	Asp Leu Asn Asp Ala Met 9445	Glu Thr Leu 9450
Lys His Leu Val Asp Asn 9455	Glu Ile Pro Asn Ala Glu 9460	Gln Thr Val 9465

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Asn Tyr	Gln Asn Ala Asp	Asp	Asn Ala Lys Thr	Asn	Phe Asp Asp
9470		9475		9480	
Ala Lys	Arg Leu Ala Asn Thr	Leu Leu Asn Ser	Asp	Asn Thr Asn	
9485		9490		9495	
Val Asn	Asp Ile Asn Gly Ala	Ile Gln Ala Val	Asn	Asp Ala Ile	
9500		9505		9510	
His Asn	Leu Asn Gly Asp Gln	Arg Leu Gln Asp	Ala	Lys Asp Lys	
9515		9520		9525	
Ala Ile	Gln Ser Ile Asn Gln	Ala Leu Ala Asn Lys	Leu Lys Glu		
9530		9535	9540		
Ile Glu	Ala Ser Asn Ala Thr	Asp Gln Asp Lys	Leu	Ile Ala Lys	
9545		9550	9555		
Asn Lys	Ala Glu Glu Leu Ala	Asn Ser Ile Ile	Asn	Asn Ile Asn	
9560		9565	9570		
Lys Ala	Thr Ser Asn Gln Ala	Val Ser Gln Val	Gln	Thr Ala Gly	
9575		9580	9585		
Asn His	Ala Ile Glu Gln Val	His Ala Asn Glu	Ile	Pro Lys Ala	
9590		9595	9600		
Lys Ile	Asp Ala Asn Lys Asp	Val Asp Lys Gln	Val	Gln Ala Leu	
9605		9610	9615		
Ile Asp	Glu Ile Asp Arg Asn	Pro Asn Leu Thr	Asp	Lys Glu Lys	
9620		9625	9630		
Gln Ala	Leu Lys Asp Arg Ile	Asn Gln Ile Leu	Gln	Gln Gly His	
9635		9640	9645		
Asn Gly	Ile Asn Asn Ala Met	Thr Lys Glu Glu	Ile	Glu Gln Ala	
9650		9655	9660		
Lys Ala	Gln Leu Ala Gln Ala	Leu Gln Asp Ile	Lys	Asp Leu Val	
9665		9670	9675		
Lys Ala	Lys Glu Asp Ala Lys	Gln Asp Val Asp	Lys	Gln Val Gln	
9680		9685	9690		
Ala Leu	Ile Asp Glu Ile Asp	Gln Asn Pro Asn	Leu	Thr Asp Lys	
9695		9700	9705		
Glu Lys	Gln Ala Leu Lys Tyr	Arg Ile Asn Gln	Ile	Leu Gln Gln	
9710		9715	9720		
Gly His	Asn Asp Ile Asn Asn	Ala Leu Thr Lys	Glu	Glu Ile Glu	
9725		9730	9735		
Gln Ala	Lys Ala Gln Leu Ala	Gln Ala Leu Gln	Asp	Ile Lys Asp	
9740		9745	9750		
Leu Val	Lys Ala Lys Glu Asp	Ala Lys Asn Ala	Ile	Lys Ala Leu	
9755		9760	9765		
Ala Asn	Ala Lys Arg Asp Gln	Ile Asn Ser Asn	Pro	Asp Leu Thr	
9770		9775	9780		
Pro Glu	Gln Lys Ala Lys Ala	Leu Lys Glu Ile	Asp	Glu Ala Glu	
9785		9790	9795		
Lys Arg	Ala Leu Gln Asn Val	Glu Asn Ala Gln	Thr	Ile Asp Gln	
9800		9805	9810		
Leu Asn	Arg Gly Leu Asn Leu	Gly Leu Asp Asp	Ile	Arg Asn Thr	
9815		9820	9825		
His Val	Trp Glu Val Asp Glu	Gln Pro Ala Val	Asn	Glu Ile Phe	
9830		9835	9840		
Glu Ala	Thr Pro Glu Gln Ile	Leu Val Asn Gly	Glu	Leu Ile Val	
9845		9850	9855		

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10250	10255	10260
Asn Lys Asp Ser Ile Lys Glu Thr Leu Asp Asp Thr Lys His Leu		
10265	10270	10275
Pro Leu Leu Phe Ala Lys Arg Arg Arg Lys Glu Asp Glu Glu Asp		
10280	10285	10290
Val Thr Val Glu Glu Lys Asp Ser Leu Asn Asn Gly Glu Ser Leu		
10295	10300	10305
Asp Lys Val Lys His Thr Pro Phe Phe Leu Pro Lys Arg Arg Arg		
10310	10315	10320
Lys Glu Asp Glu Glu Asp Val Glu Val Thr Asn Glu Asn Thr Asp		
10325	10330	10335
Glu Lys Val Leu Lys Asp Asn Glu His Ser Pro Leu Leu Phe Ala		
10340	10345	10350
Lys Arg Arg Lys Asp Lys Glu Glu Asp Val Glu Thr Thr Thr Ser		
10355	10360	10365
Ile Glu Ser Lys Asp Glu Asp Val Pro Leu Leu Leu Ala Lys Lys		
10370	10375	10380
Lys Asn Gln Lys Asp Asn Gln Ser Lys Asp Lys Lys Ser Ala Ser		
10385	10390	10395
Lys Asn Thr Ser Lys Lys Val Ala Ala Lys Lys Lys Lys Lys Lys		
10400	10405	10410
Ala Lys Lys Asn Lys Lys		
10415		

<210> SEQ ID NO 25
 <211> LENGTH: 340
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 25

Met Lys Lys Lys Leu Leu Val Leu Thr Met Ser Thr Leu Phe Ala Thr			
1	5	10	15
Gln Ile Met Asn Ser Asn His Ala Lys Ala Ser Val Thr Glu Ser Val			
	20	25	30
Asp Lys Lys Phe Val Val Pro Glu Ser Gly Ile Asn Lys Ile Ile Pro			
	35	40	45
Ala Tyr Asp Glu Phe Lys Asn Ser Pro Lys Val Asn Val Ser Asn Leu			
	50	55	60
Thr Asp Asn Lys Asn Phe Val Ala Ser Glu Asp Lys Leu Asn Lys Ile			
	65	70	75
Ala Asp Ser Ser Ala Ala Ser Lys Ile Val Asp Lys Asn Phe Val Val			
	85	90	95
Pro Glu Ser Lys Leu Gly Asn Ile Val Pro Glu Tyr Lys Glu Ile Asn			
	100	105	110
Asn Arg Val Asn Val Ala Thr Asn Asn Pro Ala Ser Gln Gln Val Asp			
	115	120	125
Lys His Phe Val Ala Lys Gly Pro Glu Val Asn Arg Phe Ile Thr Gln			
	130	135	140
Asn Lys Val Asn His His Phe Ile Thr Thr Gln Thr His Tyr Lys Lys			
	145	150	155
Val Ile Thr Ser Tyr Lys Ser Thr His Val His Lys His Val Asn His			
	165	170	175
Ala Lys Asp Ser Ile Asn Lys His Phe Ile Val Lys Pro Ser Glu Ser			
	180	185	190

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Pro Arg Tyr Thr His Pro Ser Gln Ser Leu Ile Ile Lys His His Phe
 195 200 205

Ala Val Pro Gly Tyr His Ala His Lys Phe Val Thr Pro Gly His Ala
 210 215 220

Ser Ile Lys Ile Asn His Phe Cys Val Val Pro Gln Ile Asn Ser Phe
 225 230 235 240

Lys Val Ile Pro Pro Tyr Gly His Asn Ser His Arg Met His Val Pro
 245 250 255

Ser Phe Gln Asn Asn Thr Thr Ala Thr His Gln Asn Ala Lys Val Asn
 260 265 270

Lys Ala Tyr Asp Tyr Lys Tyr Phe Tyr Ser Tyr Lys Val Val Lys Gly
 275 280 285

Val Lys Lys Tyr Phe Ser Phe Ser Gln Ser Asn Gly Tyr Lys Ile Gly
 290 295 300

Lys Pro Ser Leu Asn Ile Lys Asn Val Asn Tyr Gln Tyr Ala Val Pro
 305 310 315 320

Ser Tyr Ser Pro Thr His Tyr Val Pro Glu Phe Lys Gly Ser Leu Pro
 325 330 335

Ala Pro Arg Val
 340

<210> SEQ ID NO 26
 <211> LENGTH: 130
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 26

Met Asn Phe Asn Asp Ile Glu Thr Met Val Lys Ser Lys Phe Lys Asp
 1 5 10 15

Ile Lys Lys His Ala Glu Glu Ile Ala His Glu Ile Glu Val Arg Ser
 20 25 30

Gly Tyr Leu Arg Lys Ala Glu Gln Tyr Lys Arg Leu Glu Phe Asn Leu
 35 40 45

Ser Phe Ala Leu Asp Asp Ile Glu Ser Thr Ala Lys Asp Val Gln Thr
 50 55 60

Ala Lys Ser Ser Ala Asn Lys Asp Ser Val Thr Val Lys Gly Lys Ala
 65 70 75 80

Pro Asn Thr Leu Tyr Ile Glu Lys Arg Asn Leu Met Lys Gln Lys Leu
 85 90 95

Glu Met Leu Gly Glu Asp Ile Asp Lys Asn Lys Glu Ser Leu Gln Lys
 100 105 110

Ala Lys Glu Ile Ala Gly Glu Lys Ala Ser Glu Tyr Phe Asn Lys Ala
 115 120 125

Met Asn
 130

<210> SEQ ID NO 27
 <211> LENGTH: 636
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 27

Met Lys Lys Gln Ile Ile Ser Leu Gly Ala Leu Ala Val Ala Ser Ser
 1 5 10 15

Leu Phe Thr Trp Asp Asn Lys Ala Asp Ala Ile Val Thr Lys Asp Tyr
 20 25 30

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Ser Gly Lys Ser Gln Val Asn Ala Gly Ser Lys Asn Gly Thr Leu Ile
 35 40 45

Asp Ser Arg Tyr Leu Asn Ser Ala Leu Tyr Tyr Leu Glu Asp Tyr Ile
 50 55 60

Ile Tyr Ala Ile Gly Leu Thr Asn Lys Tyr Glu Tyr Gly Asp Asn Ile
 65 70 75 80

Tyr Lys Glu Ala Lys Asp Arg Leu Leu Glu Lys Val Leu Arg Glu Asp
 85 90 95

Gln Tyr Leu Leu Glu Arg Lys Lys Ser Gln Tyr Glu Asp Tyr Lys Gln
 100 105 110

Trp Tyr Ala Asn Tyr Lys Lys Glu Asn Pro Arg Thr Asp Leu Lys Met
 115 120 125

Ala Asn Phe His Lys Tyr Asn Leu Glu Glu Leu Ser Met Lys Glu Tyr
 130 135 140

Asn Glu Leu Gln Asp Ala Leu Lys Arg Ala Leu Asp Asp Phe His Arg
 145 150 155 160

Glu Val Lys Asp Ile Lys Asp Lys Asn Ser Asp Leu Lys Thr Phe Asn
 165 170 175

Ala Ala Glu Glu Asp Lys Ala Thr Lys Glu Val Tyr Asp Leu Val Ser
 180 185 190

Glu Ile Asp Thr Leu Val Val Ser Tyr Tyr Gly Asp Lys Asp Tyr Gly
 195 200 205

Glu His Ala Lys Glu Leu Arg Ala Lys Leu Asp Leu Ile Leu Gly Asp
 210 215 220

Thr Asp Asn Pro His Lys Ile Thr Asn Glu Arg Ile Lys Lys Glu Met
 225 230 235 240

Ile Asp Asp Leu Asn Ser Ile Ile Asp Asp Phe Phe Met Glu Thr Lys
 245 250 255

Gln Asn Arg Pro Lys Ser Ile Thr Lys Tyr Asn Pro Thr Thr His Asn
 260 265 270

Tyr Lys Thr Asn Ser Asp Asn Lys Pro Asn Phe Asp Lys Leu Val Glu
 275 280 285

Glu Thr Lys Lys Ala Val Lys Glu Ala Asp Asp Ser Trp Lys Lys Lys
 290 295 300

Thr Val Lys Lys Tyr Gly Glu Thr Glu Thr Lys Ser Pro Val Val Lys
 305 310 315 320

Glu Glu Lys Lys Val Glu Glu Pro Gln Ala Pro Lys Val Asp Asn Gln
 325 330 335

Gln Glu Val Lys Thr Thr Ala Gly Lys Ala Glu Glu Thr Thr Gln Pro
 340 345 350

Val Ala Gln Pro Leu Val Lys Ile Pro Gln Gly Thr Ile Thr Gly Glu
 355 360 365

Ile Val Lys Gly Pro Glu Tyr Pro Thr Met Glu Asn Lys Thr Val Gln
 370 375 380

Gly Glu Ile Val Gln Gly Pro Asp Phe Leu Thr Met Glu Gln Ser Gly
 385 390 395 400

Pro Ser Leu Ser Asn Asn Tyr Thr Asn Pro Pro Leu Thr Asn Pro Ile
 405 410 415

Leu Glu Gly Leu Glu Gly Ser Ser Ser Lys Leu Glu Ile Lys Pro Gln
 420 425 430

Gly Thr Glu Ser Thr Leu Lys Gly Thr Gln Gly Glu Ser Ser Asp Ile
 435 440 445

Glu Val Lys Pro Gln Ala Thr Glu Thr Thr Glu Ala Ser Gln Tyr Gly

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Ala Val Gly Asn His Glu Phe Asp Phe Gly Tyr Asp Gln Leu Lys Lys
 195 200 205

Leu Glu Gly Met Leu Asp Phe Pro Met Leu Ser Thr Asn Val Tyr Lys
 210 215 220

Asp Gly Lys Arg Ala Phe Lys Pro Ser Thr Ile Val Thr Lys Asn Gly
 225 230 235 240

Ile Arg Tyr Gly Ile Ile Gly Val Thr Thr Pro Glu Thr Lys Thr Lys
 245 250 255

Thr Arg Pro Glu Gly Ile Lys Gly Val Glu Phe Arg Asp Pro Leu Gln
 260 265 270

Ser Val Thr Ala Glu Met Met Arg Ile Tyr Lys Asp Val Asp Thr Phe
 275 280 285

Val Val Ile Ser His Leu Gly Ile Asp Pro Ser Thr Gln Glu Thr Trp
 290 295 300

Arg Gly Asp Tyr Leu Val Lys Gln Leu Ser Gln Asn Pro Gln Leu Lys
 305 310 315 320

Lys Arg Ile Thr Val Ile Asp Gly His Ser His Thr Val Leu Gln Asn
 325 330 335

Gly Gln Ile Tyr Asn Asn Asp Ala Leu Ala Gln Thr Gly Thr Ala Leu
 340 345 350

Ala Asn Ile Gly Lys Ile Thr Phe Asn Tyr Arg Asn Gly Glu Val Ser
 355 360 365

Asn Ile Lys Pro Ser Leu Ile Asn Val Lys Asp Val Glu Asn Val Thr
 370 375 380

Pro Asn Lys Ala Leu Ala Glu Gln Ile Asn Gln Ala Asp Gln Thr Phe
 385 390 395 400

Arg Ala Gln Thr Ala Glu Val Ile Ile Pro Asn Asn Thr Ile Asp Phe
 405 410 415

Lys Gly Glu Arg Asp Asp Val Arg Thr Arg Glu Thr Asn Leu Gly Asn
 420 425 430

Ala Ile Ala Asp Ala Met Glu Ala Tyr Gly Val Lys Asn Phe Ser Lys
 435 440 445

Lys Thr Asp Phe Ala Val Thr Asn Gly Gly Gly Ile Arg Ala Ser Ile
 450 455 460

Ala Lys Gly Lys Val Thr Arg Tyr Asp Leu Ile Ser Val Leu Pro Phe
 465 470 475 480

Gly Asn Thr Ile Ala Gln Ile Asp Val Lys Gly Ser Asp Val Trp Thr
 485 490 495

Ala Phe Glu His Ser Leu Gly Ala Pro Thr Thr Gln Lys Asp Gly Lys
 500 505 510

Thr Val Leu Thr Ala Asn Gly Gly Leu Leu His Ile Ser Asp Ser Ile
 515 520 525

Arg Val Tyr Tyr Asp Ile Asn Lys Pro Ser Gly Lys Arg Ile Asn Ala
 530 535 540

Ile Gln Ile Leu Asn Lys Glu Thr Gly Lys Phe Glu Asn Ile Asp Leu
 545 550 555 560

Lys Arg Val Tyr His Val Thr Met Asn Asp Phe Thr Ala Ser Gly Gly
 565 570 575

Asp Gly Tyr Ser Met Phe Gly Gly Pro Arg Glu Glu Gly Ile Ser Leu
 580 585 590

Asp Gln Val Leu Ala Ser Tyr Leu Lys Thr Ala Asn Leu Ala Lys Tyr
 595 600 605

Asp Thr Thr Glu Pro Gln Arg Met Leu Leu Gly Lys Pro Ala Val Ser

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Pro Glu Phe Asp Asp Leu Lys Arg Thr Asp Ala Ser Gln Val Asp Gly
 245 250 255

Gln Asp Asp Thr Ser Glu Asp Asp Asp Asn Glu Ser Glu Lys Ala Asp
 260 265 270

Ser Lys Ser Ala Asp Ser Glu Ser Lys Gly Gly Ala Tyr Leu Glu Met
 275 280 285

Glu Leu His Glu Gly Gln Asn Ser Glu Thr Leu Gly Asn Asp Glu Ala
 290 295 300

Arg Glu Gly Asp Ala Thr Asp Asp Met Thr Asp Met Met Thr Lys Lys
 305 310 315 320

Gly Lys Gly Ser Asn Asp Thr Leu Asn Arg Glu Glu Gly Asp Ala Val
 325 330 335

Gly Gln Ser Gln Ala Phe Gln Leu Asp Gly Val Asn Lys Asn Val Glu
 340 345 350

Ile Lys Trp Gln Ile Pro Glu Ile Glu Pro Gln Tyr Val Leu Glu Tyr
 355 360 365

Gln Glu Ser Lys Gln Asp Val Gln Tyr Glu Ile Lys Asp Leu Ile Gln
 370 375 380

Ile Ile Lys Lys Thr Ile Glu Arg Glu Gln Arg Asp Ala Arg Phe Asn
 385 390 395 400

Leu Thr Lys Gly Arg Leu Gln Lys Asp Leu Ile Asn Trp Phe Ile Asp
 405 410 415

Asp Gln Tyr Lys Leu Phe Tyr Lys Lys Gln Asp Leu Ser Lys Ser Phe
 420 425 430

Asp Ala Thr Phe Thr Leu Leu Ile Asp Ala Ser Ala Ser Met His Asp
 435 440 445

Lys Met Ala Glu Thr Lys Lys Gly Val Val Leu Phe His Glu Thr Leu
 450 455 460

Lys Ala Leu Asn Ile Lys His Glu Ile Leu Ser Phe Ser Glu Asp Ala
 465 470 475 480

Phe Asp Ser Asp Glu His Ala Gln Pro Asn Ile Ile Asn Glu Ile Ile
 485 490 495

Asn Tyr Asp Tyr Ser Thr Phe Glu Lys Asp Gly Pro Arg Ile Met Ala
 500 505 510

Leu Glu Pro Gln Asp Asp Asn Arg Asp Gly Val Ala Ile Arg Val Ala
 515 520 525

Ser Glu Arg Leu Met Arg Arg Asn Gln His Gln Arg Phe Leu Ile Val
 530 535 540

Phe Ser Asp Gly Glu Pro Ser Ala Phe Asn Tyr Ser Gln Asp Gly Ile
 545 550 555 560

Ile Asp Thr Tyr Glu Ala Val Glu Met Ser Arg Lys Phe Gly Ile Glu
 565 570 575

Val Phe Asn Val Phe Leu Ser Gln Asp Pro Ile Thr Glu Asp Val Glu
 580 585 590

Gln Thr Ile His Asn Ile Tyr Gly Gln Tyr Ala Ile Phe Val Glu Gly
 595 600 605

Val Ala His Leu Pro Gly His Leu Ser Pro Leu Leu Lys Lys Leu Leu
 610 615 620

Leu Lys Ser Leu
 625

<210> SEQ ID NO 30
 <211> LENGTH: 154
 <212> TYPE: PRT

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<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 30

Ala Glu Ile Asn Lys Gln Thr Thr Ser Gln Gly Val Thr Thr Glu Lys
 1 5 10 15
 Asn Asn Gly Ile Ala Val Leu Glu Gln Asp Val Ile Thr Pro Thr Val
 20 25 30
 Lys Pro Gln Ala Lys Gln Asp Ile Ile Gln Ala Val Thr Thr Arg Lys
 35 40 45
 Gln Gln Ile Lys Lys Ser Asn Ala Ser Leu Gln Asp Glu Lys Asp Val
 50 55 60
 Ala Asn Asp Lys Ile Gly Lys Ile Glu Thr Lys Ala Ile Lys Asp Ile
 65 70 75 80
 Asp Ala Ala Thr Thr Asn Ala Gln Val Glu Ala Ile Lys Thr Lys Ala
 85 90 95
 Ile Asn Asp Ile Asn Gln Thr Thr Pro Ala Thr Thr Ala Lys Ala Ala
 100 105 110
 Ala Leu Glu Glu Phe Asp Glu Val Val Gln Ala Gln Ile Asp Gln Ala
 115 120 125
 Pro Leu Asn Pro Asp Thr Thr Asn Glu Glu Val Ala Glu Ala Ile Glu
 130 135 140
 Arg Ile Asn Ala Ala Lys Val Ser Gly Val
 145 150

<210> SEQ ID NO 31

<211> LENGTH: 584

<212> TYPE: PRT

<213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 31

Met Lys Phe Lys Ser Leu Ile Thr Thr Thr Leu Ala Leu Gly Val Leu
 1 5 10 15
 Ala Ser Thr Gly Ala Asn Phe Asn Asn Asn Glu Ala Ser Ala Ala Ala
 20 25 30
 Lys Pro Leu Asp Lys Ser Ser Ser Ser Leu His His Gly Tyr Ser Lys
 35 40 45
 Val His Val Pro Tyr Ala Ile Thr Val Asn Gly Thr Ser Gln Asn Ile
 50 55 60
 Leu Ser Ser Leu Thr Phe Asn Lys Asn Gln Asn Ile Ser Tyr Lys Asp
 65 70 75 80
 Leu Glu Asp Arg Val Lys Ser Val Leu Lys Ser Asp Arg Gly Ile Ser
 85 90 95
 Asp Ile Asp Leu Arg Leu Ser Lys Gln Ala Lys Tyr Thr Val Tyr Phe
 100 105 110
 Lys Asn Gly Thr Lys Lys Val Ile Asp Leu Lys Ala Gly Ile Tyr Thr
 115 120 125
 Ala Asp Leu Ile Asn Thr Ser Glu Ile Lys Ala Ile Asn Ile Asn Val
 130 135 140
 Asp Thr Lys Lys Gln Val Glu Asp Lys Lys Lys Asp Lys Ala Asn Tyr
 145 150 155 160
 Gln Val Pro Tyr Thr Ile Thr Val Asn Gly Thr Ser Gln Asn Ile Leu
 165 170 175
 Ser Asn Leu Thr Phe Asn Lys Asn Gln Asn Ile Ser Tyr Lys Asp Leu
 180 185 190
 Glu Asp Lys Val Lys Ser Val Leu Glu Ser Asn Arg Gly Ile Thr Asp

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195				200				205							
Val	Asp	Leu	Arg	Leu	Ser	Lys	Gln	Ala	Lys	Tyr	Thr	Val	Asn	Phe	Lys
210						215						220			
Asn	Gly	Thr	Lys	Lys	Val	Ile	Asp	Leu	Lys	Ser	Gly	Ile	Tyr	Thr	Ala
225				230						235					240
Asn	Leu	Ile	Asn	Ser	Ser	Asp	Ile	Lys	Ser	Ile	Asn	Ile	Asn	Val	Asp
				245					250					255	
Thr	Lys	Lys	His	Ile	Glu	Asn	Lys	Ala	Lys	Arg	Asn	Tyr	Gln	Val	Pro
			260					265					270		
Tyr	Ser	Ile	Asn	Leu	Asn	Gly	Thr	Ser	Thr	Asn	Ile	Leu	Ser	Asn	Leu
		275					280					285			
Ser	Phe	Ser	Asn	Lys	Pro	Trp	Thr	Asn	Tyr	Lys	Asn	Leu	Thr	Ser	Gln
	290					295					300				
Ile	Lys	Ser	Val	Leu	Lys	His	Asp	Arg	Gly	Ile	Ser	Glu	Gln	Asp	Leu
305				310						315					320
Lys	Tyr	Ala	Lys	Lys	Ala	Tyr	Tyr	Thr	Val	Tyr	Phe	Lys	Asn	Gly	Gly
			325						330					335	
Lys	Arg	Ile	Leu	Gln	Leu	Asn	Ser	Lys	Asn	Tyr	Thr	Ala	Asn	Leu	Val
			340					345					350		
His	Ala	Lys	Asp	Val	Lys	Arg	Ile	Glu	Ile	Thr	Val	Lys	Thr	Gly	Thr
		355					360					365			
Lys	Ala	Lys	Ala	Asp	Arg	Tyr	Val	Pro	Tyr	Thr	Ile	Ala	Val	Asn	Gly
	370				375						380				
Thr	Ser	Thr	Pro	Ile	Leu	Ser	Asp	Leu	Lys	Phe	Thr	Gly	Asp	Pro	Arg
385				390						395					400
Val	Gly	Tyr	Lys	Asp	Ile	Ser	Lys	Lys	Val	Lys	Ser	Val	Leu	Lys	His
			405					410						415	
Asp	Arg	Gly	Ile	Gly	Glu	Arg	Glu	Leu	Lys	Tyr	Ala	Lys	Lys	Ala	Thr
			420					425				430			
Tyr	Thr	Val	His	Phe	Lys	Asn	Gly	Thr	Lys	Lys	Val	Ile	Asn	Ile	Asn
		435					440					445			
Ser	Asn	Ile	Ser	Gln	Leu	Asn	Leu	Leu	Tyr	Val	Gln	Asp	Ile	Lys	Lys
	450				455						460				
Ile	Asp	Ile	Asp	Val	Lys	Thr	Gly	Thr	Lys	Ala	Lys	Ala	Asp	Ser	Tyr
465				470						475					480
Val	Pro	Tyr	Thr	Ile	Ala	Val	Asn	Gly	Thr	Ser	Thr	Pro	Ile	Leu	Ser
				485					490					495	
Lys	Leu	Lys	Ile	Ser	Asn	Lys	Gln	Leu	Ile	Ser	Tyr	Lys	Tyr	Leu	Asn
			500					505					510		
Asp	Lys	Val	Lys	Ser	Val	Leu	Lys	Ser	Glu	Arg	Gly	Ile	Ser	Asp	Leu
		515					520					525			
Asp	Leu	Lys	Phe	Ala	Lys	Gln	Ala	Lys	Tyr	Thr	Val	Tyr	Phe	Lys	Asn
	530					535					540				
Gly	Lys	Lys	Gln	Val	Val	Asn	Leu	Lys	Ser	Asp	Ile	Phe	Thr	Pro	Asn
545				550						555					560
Leu	Phe	Ser	Ala	Lys	Asp	Ile	Lys	Lys	Ile	Asp	Ile	Asp	Val	Lys	Gln
			565						570					575	
Tyr	Thr	Lys	Ser	Lys	Lys	Asn	Lys								
			580												

<210> SEQ ID NO 32
 <211> LENGTH: 508
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus sp.

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<400> SEQUENCE: 32

Met Lys Asn Lys Leu Leu Val Leu Ser Leu Gly Ala Leu Cys Val Ser
 1 5 10 15
 Gln Ile Trp Glu Ser Asn Arg Ala Ser Ala Val Val Ser Gly Glu Lys
 20 25 30
 Asn Pro Tyr Val Ser Glu Ser Leu Lys Leu Thr Asn Asn Lys Asn Lys
 35 40 45
 Ser Arg Thr Val Glu Glu Tyr Lys Lys Ser Leu Asp Asp Leu Ile Trp
 50 55 60
 Ser Phe Pro Asn Leu Asp Asn Glu Arg Phe Asp Asn Pro Glu Tyr Lys
 65 70 75 80
 Glu Ala Met Lys Lys Tyr Gln Gln Arg Phe Met Ala Glu Asp Glu Ala
 85 90 95
 Leu Lys Lys Phe Phe Ser Glu Glu Lys Lys Ile Lys Asn Gly Asn Thr
 100 105 110
 Asp Asn Leu Asp Tyr Leu Gly Leu Ser His Glu Arg Tyr Glu Ser Val
 115 120 125
 Phe Asn Thr Leu Lys Lys Gln Ser Glu Glu Phe Leu Lys Glu Ile Glu
 130 135 140
 Asp Ile Lys Lys Asp Asn Pro Glu Leu Lys Asp Phe Asn Glu Glu Glu
 145 150 155 160
 Gln Leu Lys Cys Asp Leu Glu Leu Asn Lys Leu Glu Asn Gln Ile Leu
 165 170 175
 Met Leu Gly Lys Thr Phe Tyr Gln Asn Tyr Arg Asp Asp Val Glu Ser
 180 185 190
 Leu Tyr Ser Lys Leu Asp Leu Ile Met Gly Tyr Lys Asp Glu Glu Arg
 195 200 205
 Ala Asn Lys Lys Ala Val Asn Lys Arg Met Leu Glu Asn Lys Lys Glu
 210 215 220
 Asp Leu Glu Thr Ile Ile Asp Glu Phe Phe Ser Asp Ile Asp Lys Thr
 225 230 235 240
 Arg Pro Asn Asn Ile Pro Val Leu Glu Asp Glu Lys Gln Glu Glu Lys
 245 250 255
 Asn His Lys Asn Met Ala Gln Leu Lys Ser Asp Thr Glu Ala Ala Lys
 260 265 270
 Ser Asp Glu Ser Lys Arg Ser Lys Arg Ser Lys Arg Ser Leu Asn Thr
 275 280 285
 Gln Asn His Lys Pro Ala Ser Gln Glu Val Ser Glu Gln Gln Lys Ala
 290 295 300
 Glu Tyr Asp Lys Arg Ala Glu Glu Arg Lys Ala Arg Phe Leu Asp Asn
 305 310 315 320
 Gln Lys Ile Lys Lys Thr Pro Val Val Ser Leu Glu Tyr Asp Phe Glu
 325 330 335
 His Lys Gln Arg Ile Asp Asn Glu Asn Asp Lys Lys Leu Val Val Ser
 340 345 350
 Ala Pro Thr Lys Lys Pro Thr Ser Pro Thr Thr Tyr Thr Glu Thr Thr
 355 360 365
 Thr Gln Val Pro Met Pro Thr Val Glu Arg Gln Thr Gln Gln Gln Ile
 370 375 380
 Ile Tyr Asn Ala Pro Lys Gln Leu Ala Gly Leu Asn Gly Glu Ser His
 385 390 395 400
 Asp Phe Thr Thr Thr His Gln Ser Pro Thr Thr Ser Asn His Thr His

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Lys Leu Asn Asp Ala Gln Ala Pro Lys Ala Asp Asn Lys Phe Asn Lys
 275 280 285

Glu Gln Gln Asn Ala Phe Tyr Glu Ile Leu His Leu Pro Asn Leu Thr
 290 295 300

Glu Glu Gln Arg Asn Gly Phe Ile Gln Ser Leu Lys Asp Asp Pro Ser
 305 310 315 320

Val Ser Lys Glu Ile Leu Ala Glu Ala Lys Lys Leu Asn Asp Ala Gln
 325 330 335

Ala Pro Lys Glu Glu Asp Asn Asn Lys Pro Gly Lys Glu Asp Gly Asn
 340 345 350

Lys Pro Gly Lys Glu Asp Asn Asn Lys Pro Gly Lys Glu Asp Asn Lys
 355 360 365

Lys Pro Gly Lys Glu Asp Asn Asn Lys Pro Gly Lys Glu Asp Asn Asn
 370 375 380

Lys Pro Gly Lys Glu Asp Gly Asn Lys Pro Gly Lys Glu Asp Asn Lys
 385 390 395 400

Lys Pro Gly Lys Glu Asp Asn Asn Lys Pro Gly Lys Glu Asp Gly Asn
 405 410 415

Lys Pro Gly Lys Glu Asp Gly Asn Gly Val His Val Val Lys Pro Gly
 420 425 430

Asp Thr Val Asn Asp Ile Ala Lys Ala Asn Gly Thr Thr Ala Asp Lys
 435 440 445

Ile Ala Ala Asp Asn Lys Leu Ala Asp Lys Asn Met Ile Lys Pro Gly
 450 455 460

Gln Glu Leu Val Val Asp Lys Lys Gln Pro Ala Asn His Ala Asp Ala
 465 470 475 480

Asn Lys Ala Gln Ala Leu Pro Glu Thr Gly Glu Glu Asn Pro Phe Ile
 485 490 495

Gly Thr Thr Val Phe Gly Gly Leu Ser Leu Ala Leu Gly Ala Ala Leu
 500 505 510

Leu Ala Gly Arg Arg Arg Glu Leu
 515 520

<210> SEQ ID NO 34
 <211> LENGTH: 291
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 34

Ala Gln His Asp Glu Ala Lys Lys Asn Ala Phe Tyr Gln Val Leu Asn
 1 5 10 15

Met Pro Asn Leu Asn Ala Asp Gln Arg Asn Gly Phe Ile Gln Ser Leu
 20 25 30

Lys Ala Ala Pro Ser Gln Ser Ala Asn Val Leu Gly Glu Ala Gln Lys
 35 40 45

Leu Asn Asp Ser Gln Ala Pro Lys Ala Asp Ala Gln Gln Asn Asn Phe
 50 55 60

Asn Lys Asp Lys Lys Ser Ala Phe Tyr Glu Ile Leu Asn Met Pro Asn
 65 70 75 80

Leu Asn Glu Ala Gln Arg Asn Gly Phe Ile Gln Ser Leu Lys Ala Ala
 85 90 95

Pro Ser Gln Ser Thr Asn Val Leu Gly Glu Ala Lys Lys Leu Asn Glu
 100 105 110

Ser Gln Ala Pro Lys Ala Asp Asn Asn Phe Asn Lys Glu Lys Lys Asn
 115 120 125

-continued

Ala Phe Tyr Glu Ile Leu Asn Met Pro Asn Leu Asn Glu Glu Gln Arg
 130 135 140
 Asn Gly Phe Ile Gln Ser Leu Lys Ala Ala Pro Ser Gln Ser Ala Asn
 145 150 155 160
 Leu Leu Ser Glu Ala Lys Lys Leu Asn Glu Ser Gln Ala Pro Lys Ala
 165 170 175
 Asp Asn Lys Phe Asn Lys Glu Lys Lys Asn Ala Phe Tyr Glu Ile Leu
 180 185 190
 His Leu Pro Asn Leu Asn Glu Glu Gln Arg Asn Gly Phe Ile Gln Ser
 195 200 205
 Leu Lys Ala Ala Pro Ser Gln Ser Ala Asn Leu Leu Ala Glu Ala Lys
 210 215 220
 Lys Leu Asn Asp Ala Gln Ala Pro Lys Ala Asp Asn Lys Phe Asn Lys
 225 230 235 240
 Glu Lys Lys Asn Ala Phe Tyr Glu Ile Leu His Leu Pro Asn Leu Thr
 245 250 255
 Glu Glu Gln Arg Asn Gly Phe Ile Gln Ser Leu Lys Ala Ala Pro Ser
 260 265 270
 Val Ser Lys Glu Ile Leu Ala Glu Ala Lys Lys Leu Asn Asp Ala Gln
 275 280 285
 Ala Pro Lys
 290

<210> SEQ ID NO 35
 <211> LENGTH: 34
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 35

gctgcacata tggcgcaaca cgatgaagct caac

34

<210> SEQ ID NO 36
 <211> LENGTH: 30
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 36

agtggatcct tatgctttgt tagcatctgc

30

<210> SEQ ID NO 37
 <211> LENGTH: 19
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 37

Met Gly Ser Ser His His His His His His Ser Ser Gly Leu Val Pro
 1 5 10 15

Arg Gly Ser

<210> SEQ ID NO 38
 <211> LENGTH: 29
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 38

aacatattgt caacaaagat caacaaagc

29

<210> SEQ ID NO 39

-continued

<211> LENGTH: 29
 <212> TYPE: DNA
 <213> ORGANISM: *Staphylococcus sp.*
 <400> SEQUENCE: 39
 aaggatccag attcgtttaa ttttttagc 29

<210> SEQ ID NO 40
 <211> LENGTH: 43
 <212> TYPE: DNA
 <213> ORGANISM: *Staphylococcus sp.*
 <400> SEQUENCE: 40
 cttcattcaa agtcttaaag cgcocccaag ccaaagcact aac 43

<210> SEQ ID NO 41
 <211> LENGTH: 43
 <212> TYPE: DNA
 <213> ORGANISM: *Staphylococcus sp.*
 <400> SEQUENCE: 41
 gttagtgtt tggcttgggg cggctttaag actttgaatg aag 43

<210> SEQ ID NO 42
 <211> LENGTH: 42
 <212> TYPE: DNA
 <213> ORGANISM: *Staphylococcus sp.*
 <400> SEQUENCE: 42
 catatgttca acaaagataa aaaaagcgcc ttctatgaaa tc 42

<210> SEQ ID NO 43
 <211> LENGTH: 42
 <212> TYPE: DNA
 <213> ORGANISM: *Staphylococcus sp.*
 <400> SEQUENCE: 43
 gatttcatag aaggcgcttt ttttatcttt gttgaacata tg 42

<210> SEQ ID NO 44
 <211> LENGTH: 42
 <212> TYPE: DNA
 <213> ORGANISM: *Staphylococcus sp.*
 <400> SEQUENCE: 44
 catatgttca acaaagatgg aggaagcgcc ttctatgaaa tc 42

<210> SEQ ID NO 45
 <211> LENGTH: 42
 <212> TYPE: DNA
 <213> ORGANISM: *Staphylococcus sp.*
 <400> SEQUENCE: 45
 gatttcatag aaggcgcttc ctccatcttt gttgaacata tg 42

<210> SEQ ID NO 46
 <211> LENGTH: 52
 <212> TYPE: DNA
 <213> ORGANISM: *Staphylococcus sp.*
 <400> SEQUENCE: 46
 ggggacaagt ttgtacaaaa aagcaggctg atgactaagt tgaaaaaaga ag 52

-continued

<210> SEQ ID NO 47
 <211> LENGTH: 28
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 47
 aaggatcccc tccaaaatgt aattgccc 28

<210> SEQ ID NO 48
 <211> LENGTH: 30
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 48
 aaggatccgt ttgtaactct atccaaagac 30

<210> SEQ ID NO 49
 <211> LENGTH: 49
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 49
 ggggaccact ttgtacaaga aagctgggtg acacctattg cagcattcg 49

<210> SEQ ID NO 50
 <211> LENGTH: 50
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 50
 ggggacaagt ttgtacaaaa aagcaggctc agatagcgat tcagattcag 50

<210> SEQ ID NO 51
 <211> LENGTH: 31
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 51
 aaggatccct gtattttctc ctttaatttc c 31

<210> SEQ ID NO 52
 <211> LENGTH: 30
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 52
 aaggatccca tggctgcaaa gcaataatg 30

<210> SEQ ID NO 53
 <211> LENGTH: 51
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 53
 ggggaccact ttgtacaaga aagctgggtg ccctgggtga acaaatttat g 51

<210> SEQ ID NO 54
 <211> LENGTH: 37
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.
 <400> SEQUENCE: 54
 gaaggatccg tttattctag ttaatatata gttaatg 37

-continued

<210> SEQ ID NO 55
 <211> LENGTH: 33
 <212> TYPE: DNA
 <213> ORGANISM: *Staphylococcus* sp.
 <400> SEQUENCE: 55
 gaactgcagc tgtatgtctt tggatagagt tac 33

<210> SEQ ID NO 56
 <211> LENGTH: 33
 <212> TYPE: DNA
 <213> ORGANISM: *Staphylococcus* sp.
 <400> SEQUENCE: 56
 gaaggatocg gtggcttttt tacttggatt ttc 33

<210> SEQ ID NO 57
 <211> LENGTH: 33
 <212> TYPE: DNA
 <213> ORGANISM: *Staphylococcus* sp.
 <400> SEQUENCE: 57
 gaactgcagc gacaaactca ttatttgctt tgc 33

<210> SEQ ID NO 58
 <211> LENGTH: 27
 <212> TYPE: DNA
 <213> ORGANISM: *Staphylococcus* sp.
 <400> SEQUENCE: 58
 gaactcgagt ctagcttatt tacatgg 27

<210> SEQ ID NO 59
 <211> LENGTH: 45
 <212> TYPE: DNA
 <213> ORGANISM: *Staphylococcus* sp.
 <400> SEQUENCE: 59
 gaactcgaga tagaaggcag aatagtaaca aaggattata gtggg 45

<210> SEQ ID NO 60
 <211> LENGTH: 27
 <212> TYPE: DNA
 <213> ORGANISM: *Staphylococcus* sp.
 <400> SEQUENCE: 60
 gtaggatcct gggatagagt tacaaac 27

<210> SEQ ID NO 61
 <211> LENGTH: 34
 <212> TYPE: DNA
 <213> ORGANISM: *Staphylococcus* sp.
 <400> SEQUENCE: 61
 gaactcgagg cattatgtgt atcacaatt tggg 34

<210> SEQ ID NO 62
 <211> LENGTH: 43
 <212> TYPE: DNA
 <213> ORGANISM: *Staphylococcus* sp.
 <400> SEQUENCE: 62
 gaactcgaga tagaaggcag agtggtttct ggggagaaga atc 43

-continued

<210> SEQ ID NO 63
 <211> LENGTH: 33
 <212> TYPE: DNA
 <213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 63

gaactcgagg cagccatgca ttaattattt gcc

33

<210> SEQ ID NO 64
 <211> LENGTH: 677
 <212> TYPE: PRT
 <213> ORGANISM: Staphylococcus sp.

<400> SEQUENCE: 64

Met Lys Ser Asn Leu Arg Tyr Gly Ile Arg Lys His Lys Leu Gly Ala
 1 5 10 15
 Ala Ser Val Phe Leu Gly Thr Met Ile Val Val Gly Met Gly Gln Glu
 20 25 30
 Lys Glu Ala Ala Ala Ser Glu Gln Asn Asn Thr Thr Val Glu Glu Ser
 35 40 45
 Gly Ser Ser Ala Thr Glu Ser Lys Ala Ser Glu Thr Gln Thr Thr Thr
 50 55 60
 Asn Asn Val Asn Thr Ile Asp Glu Thr Gln Ser Tyr Ser Ala Thr Ser
 65 70 75 80
 Thr Glu Gln Pro Ser Gln Ser Thr Gln Val Thr Thr Glu Glu Ala Pro
 85 90 95
 Lys Thr Val Gln Ala Pro Lys Val Glu Thr Ser Arg Val Asp Leu Pro
 100 105 110
 Ser Glu Lys Val Ala Asp Lys Glu Thr Thr Gly Thr Gln Val Asp Ile
 115 120 125
 Ala Gln Pro Ser Asn Val Ser Glu Ile Lys Pro Arg Met Lys Arg Ser
 130 135 140
 Thr Asp Val Thr Ala Val Ala Glu Lys Glu Val Val Glu Glu Thr Lys
 145 150 155 160
 Ala Thr Gly Thr Asp Val Thr Asn Lys Val Glu Val Glu Glu Gly Ser
 165 170 175
 Glu Ile Val Gly His Lys Gln Asp Thr Asn Val Val Asn Pro His Asn
 180 185 190
 Ala Glu Arg Val Thr Leu Lys Tyr Lys Trp Lys Phe Gly Glu Gly Ile
 195 200 205
 Lys Ala Gly Asp Tyr Phe Asp Phe Thr Leu Ser Asp Asn Val Glu Thr
 210 215 220
 His Gly Ile Ser Thr Leu Arg Lys Val Pro Glu Ile Lys Ser Thr Asp
 225 230 235 240
 Gly Gln Val Met Ala Thr Gly Glu Ile Ile Gly Glu Arg Lys Val Arg
 245 250 255
 Tyr Thr Phe Lys Glu Tyr Val Gln Glu Lys Lys Asp Leu Thr Ala Glu
 260 265 270
 Leu Ser Leu Asn Leu Phe Ile Asp Pro Thr Thr Val Thr Gln Lys Gly
 275 280 285
 Asn Gln Asn Val Glu Val Lys Leu Gly Glu Thr Thr Val Ser Lys Ile
 290 295 300
 Phe Asn Ile Gln Tyr Leu Gly Gly Val Arg Asp Asn Trp Gly Val Thr
 305 310 315 320

-continued

Ala Asn Gly Arg Ile Asp Thr Leu Asn Lys Val Asp Gly Lys Phe Ser
 325 330 335

His Phe Ala Tyr Met Lys Pro Asn Asn Gln Ser Leu Ser Ser Val Thr
 340 345 350

Val Thr Gly Gln Val Thr Lys Gly Asn Lys Pro Gly Val Asn Asn Pro
 355 360 365

Thr Val Lys Val Tyr Lys His Ile Gly Ser Asp Asp Leu Ala Glu Ser
 370 375 380

Val Tyr Ala Lys Leu Asp Asp Val Ser Lys Phe Glu Asp Val Thr Asp
 385 390 395 400

Asn Met Ser Leu Asp Phe Asp Thr Asn Gly Gly Tyr Ser Leu Asn Phe
 405 410 415

Asn Asn Leu Asp Gln Ser Lys Asn Tyr Val Ile Lys Tyr Glu Gly Tyr
 420 425 430

Tyr Asp Ser Asn Ala Ser Asn Leu Glu Phe Gln Thr His Leu Phe Gly
 435 440 445

Tyr Tyr Asn Tyr Tyr Tyr Thr Ser Asn Leu Thr Trp Lys Asn Gly Val
 450 455 460

Ala Phe Tyr Ser Asn Asn Ala Gln Gly Asp Gly Lys Asp Lys Leu Lys
 465 470 475 480

Glu Pro Ile Ile Glu His Ser Thr Pro Ile Glu Leu Glu Phe Lys Ser
 485 490 495

Glu Pro Pro Val Glu Lys His Glu Leu Thr Gly Thr Ile Glu Glu Ser
 500 505 510

Asn Asp Ser Lys Pro Ile Asp Phe Glu Tyr His Thr Ala Val Glu Gly
 515 520 525

Ala Glu Gly His Ala Glu Gly Thr Ile Glu Thr Glu Glu Asp Ser Ile
 530 535 540

His Val Asp Phe Glu Glu Ser Thr His Glu Asn Ser Lys His His Ala
 545 550 555 560

Asp Val Val Glu Tyr Glu Glu Asp Thr Asn Pro Gly Gly Gly Gln Val
 565 570 575

Thr Thr Glu Ser Asn Leu Val Glu Phe Asp Glu Asp Ser Thr Lys Gly
 580 585 590

Ile Val Thr Gly Ala Val Ser Asp His Thr Thr Ile Glu Asp Thr Lys
 595 600 605

Glu Tyr Thr Thr Glu Ser Asn Leu Ile Glu Leu Val Asp Glu Leu Pro
 610 615 620

Glu Glu His Gly Gln Ala Gln Gly Pro Ile Glu Glu Ile Thr Glu Asn
 625 630 635 640

Asn His His Ile Ser His Ser Gly Leu Gly Thr Glu Asn Gly His Gly
 645 650 655

Asn Tyr Gly Val Ile Glu Glu Ile Glu Glu Asn Ser His Val Asp Ile
 660 665 670

Lys Ser Glu Leu Gly
 675

What is claimed is:

1. A polypeptide comprising a variant Staphylococcal Protein A (SpA) having an amino acid substitution at amino acid positions 9 and 10 of SEQ ID NO:2 and an amino acid sequence that is at least 70% identical to the amino acid sequence of SEQ ID NO:2.

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2. The polypeptide of claim 1, wherein the variant SpA comprises a variant domain D segment.

65 3. The isolated polypeptide of claim 1, wherein the SpA variant has an amino acid sequence that is at least 80% identical to the amino acid sequence of SEQ ID NO:2.

4. The polypeptide of claim 3, wherein the amino acid substitution is a lysine residue for a glutamine residue.

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5. The polypeptide of claim 3, further comprising one or more amino acid substitutions at amino acid residue Q26, G29, F30, S33, D36, D37, Q40, N43, and/or E47 of SEQ ID NO:2.

6. The polypeptide of claim 5, wherein the amino acid sequence of the domain D comprises an amino acid substitution at amino acid position 33 of SEQ ID NO:2.

7. The polypeptide of claim 2, further comprising one or more variants of an SpA E domain, A domain, B domain, or C domain.

8. A composition comprising, in a pharmaceutically acceptable composition, the polypeptide of claim 1.

9. The composition of claim 8, further comprising at least a second staphylococcal antigen.

10. The composition of claim 8, wherein the composition further comprises an adjuvant.

11. An immunogenic composition comprising an isolated peptide comprising a Protein A (SpA) variant having an amino acid substitution at amino acid positions 9, 10, and 33 of SEQ ID NO:2.

12. A method for eliciting an immune response against a *staphylococcus* bacterium in a subject comprising providing to the subject an effective amount of a composition comprising a Protein A (SpA) variant having an amino acid substitution at amino acid positions 9 and 10 of SEQ ID NO:2.

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13. The method of claim 12, wherein the subject is provided with an effective amount of an SpA variant by administering to the subject a composition comprising:

- i) an isolated SpA variant having an amino acid substitution at amino acid positions 9 and 10 of SEQ ID NO:2, or
- ii) at least one isolated recombinant nucleic acid molecule encoding a SpA variant having an amino acid substitution at amino acid positions 9 and 10 of SEQ ID NO:2.

14. The method of claim 12, wherein the composition comprises an adjuvant.

15. The method of claim 12, wherein the SpA variant is at least 70% identical to SEQ ID NO:2 and having an amino acid substitution at amino acid position 9 and 10 of SEQ ID NO:2.

16. The method of claim 12, wherein the *staphylococcus* bacterium is a *S. aureus* bacterium.

17. The method of claim 16, wherein the bacterium is methicillin resistant.

18. The method of claim 12, further comprising administering to the subject a composition comprising a second staphylococcal antigen.

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