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Hassan et al.

(54) COMPOSITIONS COMPRISING SEL1-DERIVED PEPTIDES AND METHODS OF TREATMENT/PREVENTION OF EXCESS OXALATE LEVELS AND ASSOCIATED CONDITIONS/DISEASES THEREWITH

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	A61P 13/04	(2006.01)
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(57) **ABSTRACT**

Provided herein are compositions comprising Sel1-derived peptides thereof, and method of use thereof for the treatment/prevention excess oxalate levels and conditions and diseases related thereto. In particular, peptides comprise Sel-like repeat (SLR) domains and/or tetratricopeptide (TPR) domains and may be linked together or with other peptides or polypeptides to treat/prevent diseases/conditions related to excess oxalate levels, such as hyperoxaluria and/or hyperoxalemia.

13 Claims, 15 Drawing Sheets

Specification includes a Sequence Listing.

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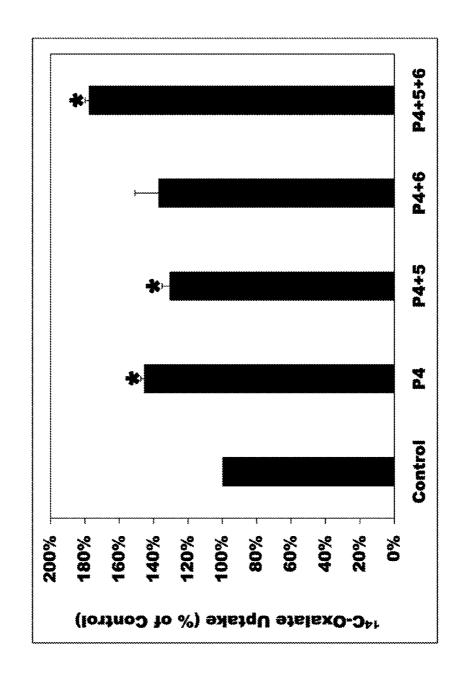
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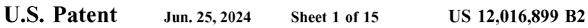
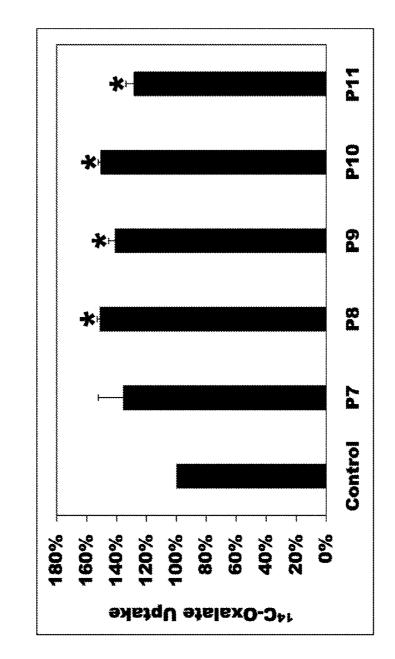
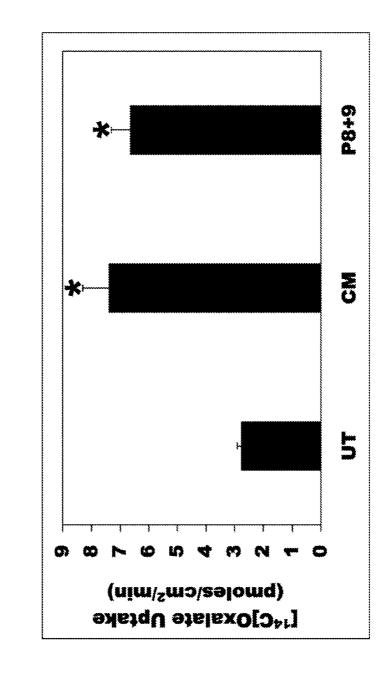


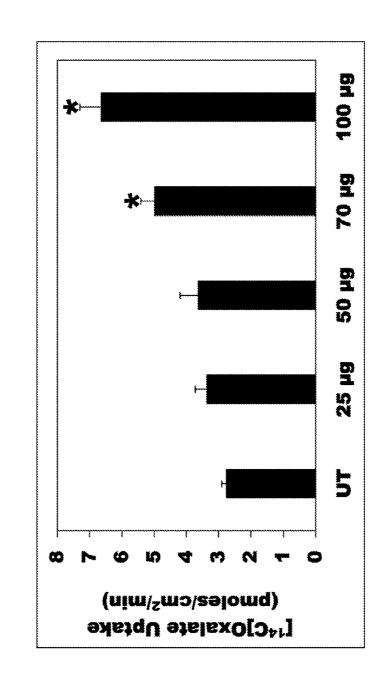
FIG. 1



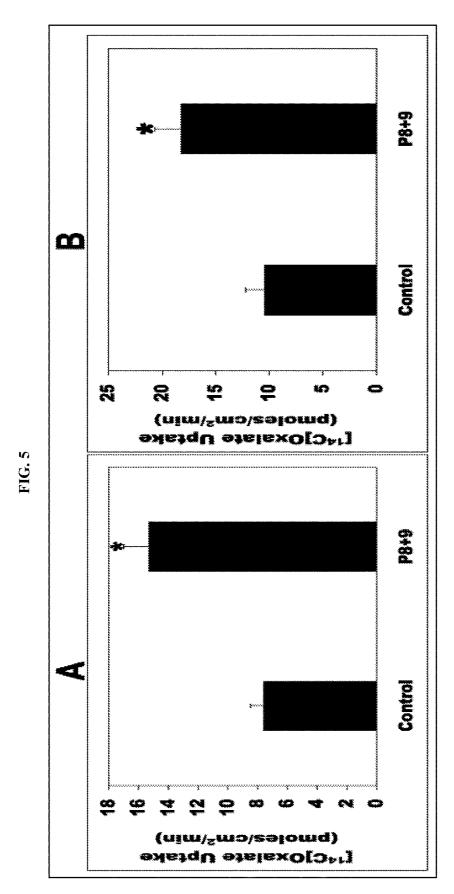


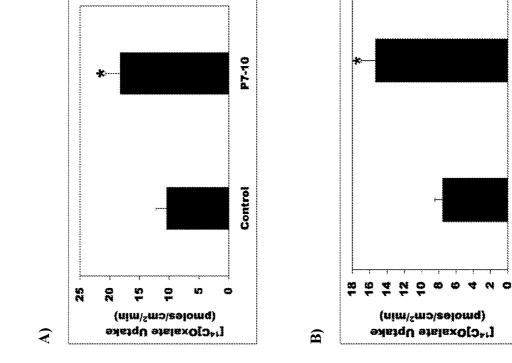








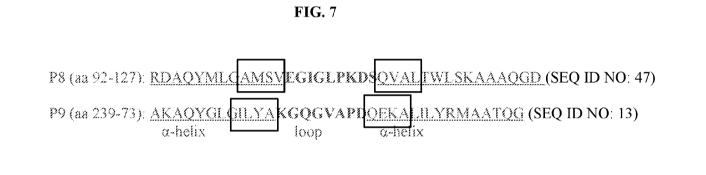




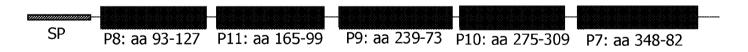


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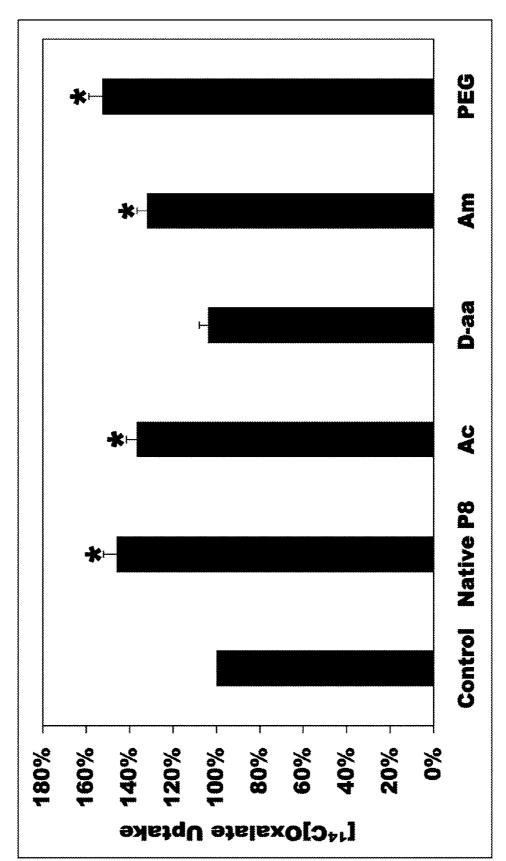
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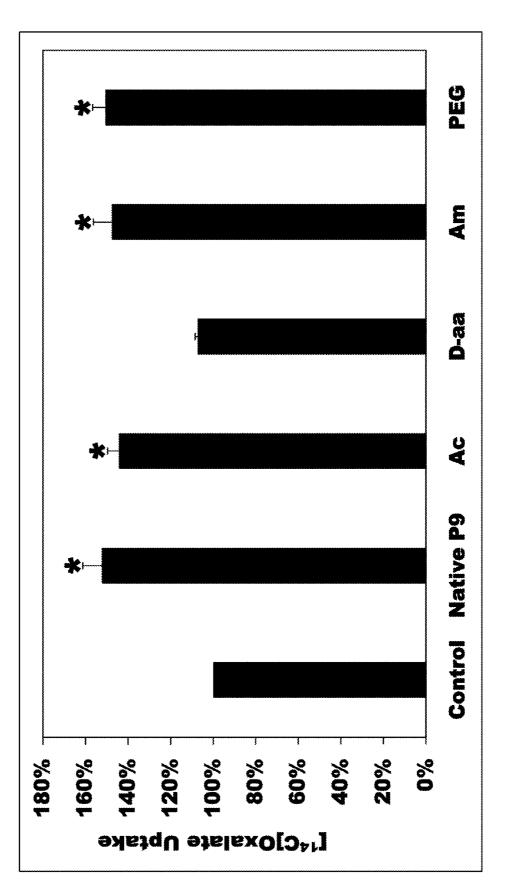
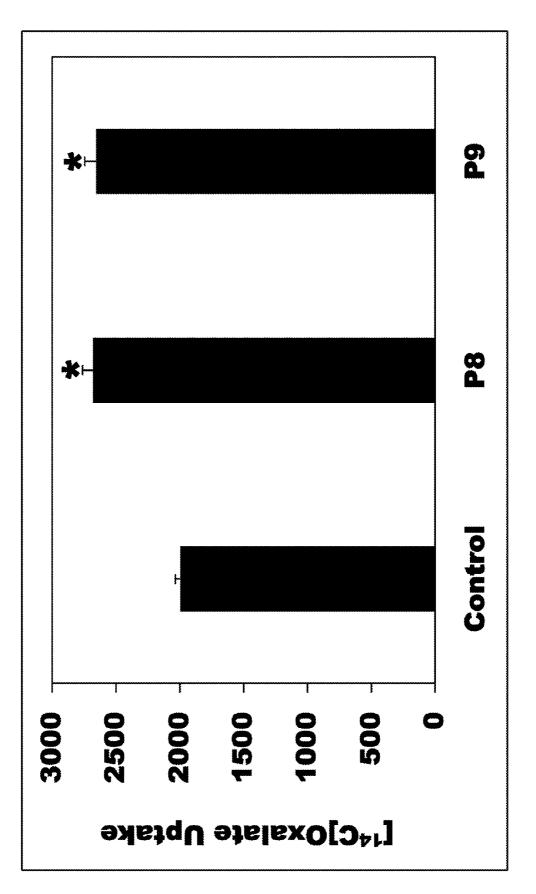
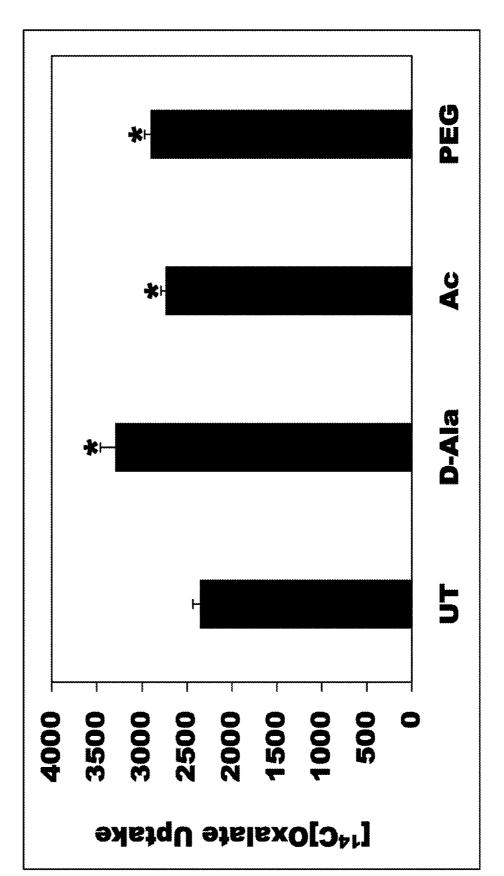
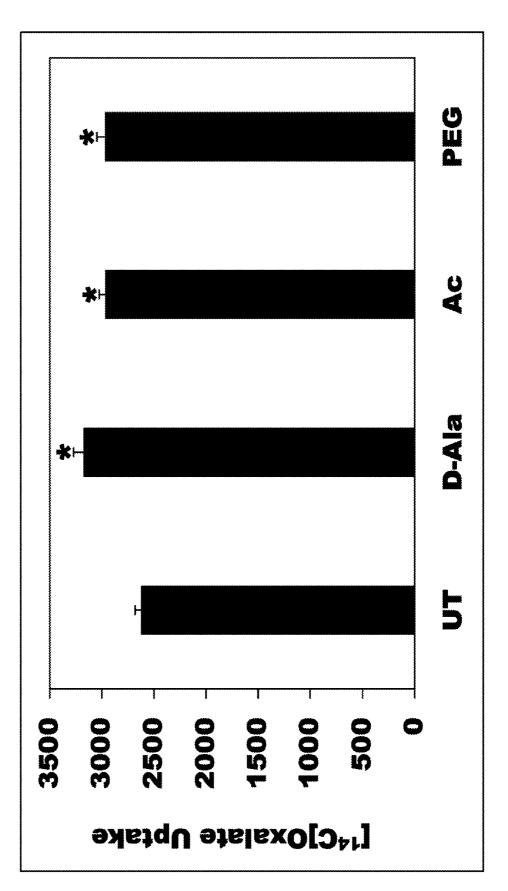


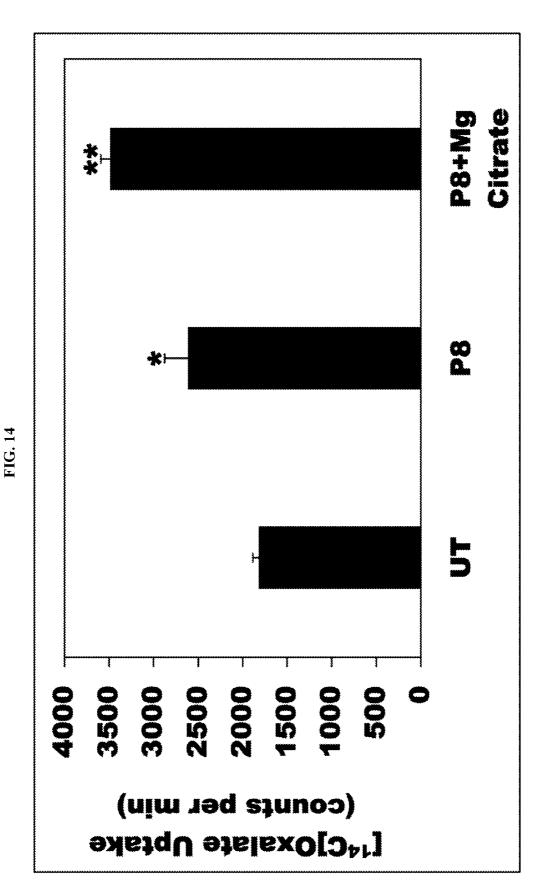
FIG. 10



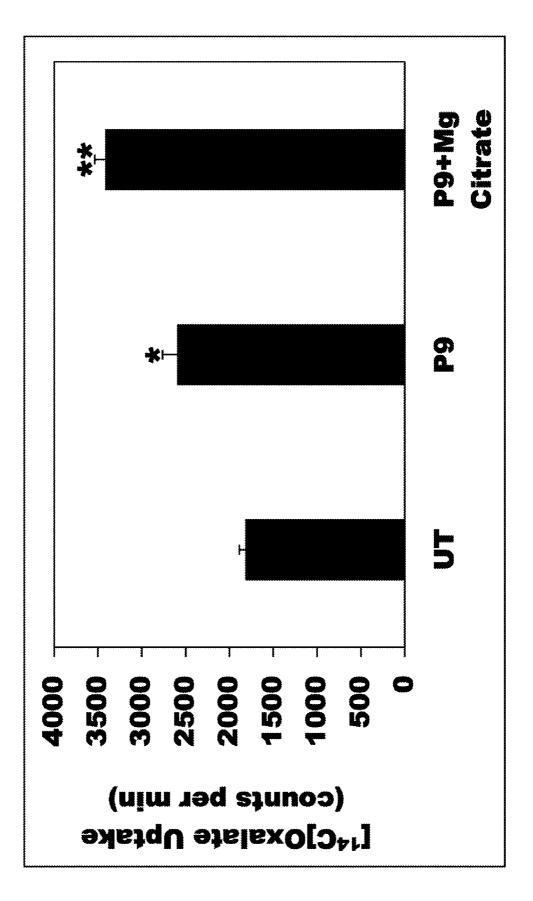




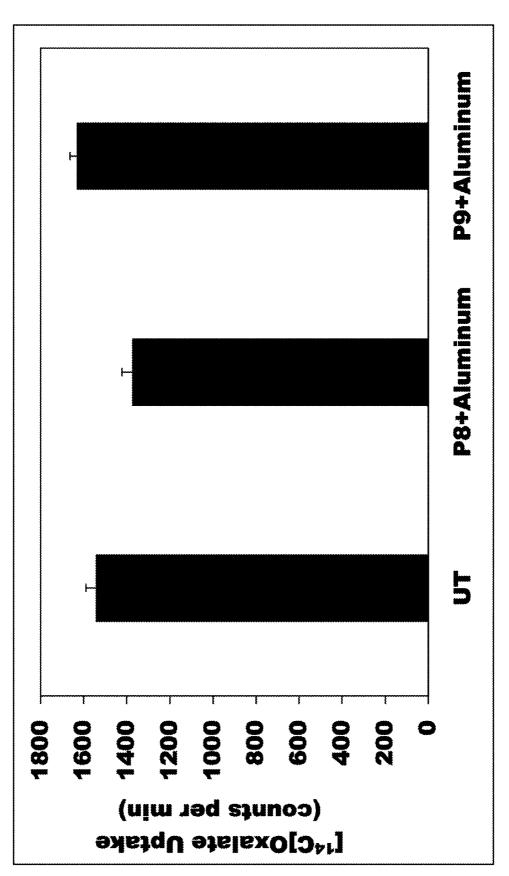




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COMPOSITIONS COMPRISING SEL1-DERIVED PEPTIDES AND METHODS OF TREATMENT/PREVENTION OF EXCESS OXALATE LEVELS AND ASSOCIATED CONDITIONS/DISEASES THEREWITH

This invention was made with government support under grant numbers DK067245, DK042086 and DK101643 awarded by The National Institutes of Health. The government has certain rights in the invention.

SEQUENCE LISTING

The text of the computer readable sequence listing filed herewith, titled "36476-¹⁵ 252_SEQUENCE_LISTING_ST25.txt", created Nov. 2, 2023, having a file size of 32.391 bytes, is hereby incorporated by reference in its entirety.

FIELD

Provided herein are compositions comprising Sel1-derived peptides, and method of use thereof for the treatment/ prevention of excess oxalate levels and conditions and diseases related thereto. In particular, peptides comprise ²⁵ Sel-like repeat (SLR) domains and/or tetratricopeptide (TPR) domains and may be linked together or administered with other peptides or polypeptides to treat/prevent diseases/ conditions related to excess oxalate levels, such as hyperoxaluria and/or hyperoxalemia. ³⁰

BACKGROUND

Nephrolithiasis, or the formation of mineral deposit blockages in the kidney (kidney stones (KS)), is the second 35 most prevalent kidney disease in USA after hypertension, with a rising prevalence and complications including advanced chronic kidney disease (CKD) and end stage renal disease (ESRD). It remains a major source of patient discomfort and disability, lost working days, and health-care 40 expenditure, with an annual economic cost approaching \$10 billion. Hyperoxaluria (HO) is a major risk factor for KS, and 70-80% of KS are composed of calcium oxalate. Urinary oxalate is an important determinant of supersaturation, and the risk for stone formation is affected by small 45 increases in urine oxalate. Oxalate is a metabolic end product that cannot be further metabolized and is highly toxic. The mammalian intestine plays a crucial role in oxalate homeostasis, by regulating the amount of absorbed dietary oxalate and providing an avenue for enteric oxalate 50 excretion. Anion exchanger SLC26A6 (A6)-mediated intestinal oxalate secretion plays a critical role in preventing hyperoxaluria and calcium oxalate kidney stones (COKS). Inflammatory bowel disease patients have a significantly increased risk of KS due to the associated enteric hyperox- 55 aluria. Obesity is a risk factor for KS and obese stone formers often have mild to moderate hyperoxaluria. Hyperoxaluria is also emerging as a major complication (developing in >50% of patients) of bariatric surgery for obesity. With the rising prevalence of obesity and increased utiliza- 60 tion of bariatric surgery, it is expected that the incidence of hyperoxaluria and related COKS (including the associated cost burden) will continue to increase at a significant rate. Primary hyperoxaluria (PH) is an inherited disease in which there is endogenous oxalate overproduction, which leads to 65 recurrent KS and/or progressive nephrocalcinosis, ESRD, as well as significant hyperoxalemia, systemic oxalosis and

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premature death. Systemic deposition of calcium oxalate (oxalosis) leads to bone disease, cardiac arrhythmias, cardiomyopathy, skin ulcers, erythropoietin refractory anemia, and digital gangrene. The only treatment known to fully correct the underlying metabolic defect is liver transplantation or combined kidney-liver transplantation once ESRD develops. In addition, significant hyperoxalemia is also seen in ESRD. Cardiovascular diseases are the leading cause of morbidity and mortality in ESRD patients, and a recent report suggest that the ESRD-associated hyperoxalemia may contribute to this increased risk.

Kidney stones (KS) affect ~1 in 5 men and ~1 in 11 women, are costly (>\$10B annually), and are associated with CKD and ESRD. High recurrence rates (50% in 5 years and up to 80% in 10 years), indicate that current interventions are inadequate and alternative therapies are needed. Most KS are composed of calcium oxalate and very small increases in urine oxalate concentration enhance the risk for stone formation. Lower urinary calcium oxalate (CaOx) 20 supersaturation definitively reduces KS formation. Currently no 1-DA approved drugs reduce urinary oxalate excretion. The gut bacterium Oxalobacter formigens (Of) induces colonic oxalate secretion and reduces urinary oxalate excretion via a secretagogue. Given the difficulties with recolonization, Of alone is not therapeutically feasible and underscores the need to utilize the secretagogue that induces colonic oxalate secretion.

SUMMARY

Provided herein are compositions comprising Sel1-derived peptides, and method of use thereof for the treatment/ prevention of excess oxalate levels and conditions and diseases related thereto. In particular, peptides comprise Sel-like repeat (SLR) domains and/or tetratricopeptide (TPR) domains and may be linked together or administered with other peptides or polypeptides to treat/prevent diseases/ conditions related to excess oxalate levels, such as hyperoxaluria and/or hyperoxalemia.

In some embodiments, provided herein are compositions comprising a Sel1-derived peptide sequence at least 60% (e.g., 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 99%, 100%, or ranges therebetween) sequence identity to one of SEQ ID NOs: 5-46, wherein the composition is not a product of nature. In some embodiments, the Sel1-derived peptide sequence has less than 100% sequence identity to SEQ ID NOs: 5-46. In some embodiments, the Sel1-derived peptide sequence is fused to a second peptide or polypeptide sequence. In some embodiments, the second peptide or polypeptide sequence is a carrier moiety, therapeutic moiety, detectable moiety, or Sell-derived peptide sequence. In some embodiments, the composition comprises 2 or more Sel1-derived peptide sequences with at least 60% sequence identity to one of SEQ ID NOs: 5-46. In some embodiments, the Sel1-derived peptide sequences are fused directly or indirectly to each other. In some embodiments, the Sel1derived peptide sequences are not fused to each other. In some embodiments, (i) one or more of the amino acid residues in the peptide sequence(s) are D-enantiomers, (ii) the peptide sequence(s) comprise a one or more unnatural amino acids, (iii) the peptide sequence(s) comprise a one or more amino acid analogs. and/or (iv) the peptide sequence(s) comprise a one or more peptoid amino acids. In some embodiments, the peptide sequence(s) or an amino acid therein comprises a modification selected from the group consisting of phosphorylation, glycosylation, ubiquitination, S-nitrosylation, methylation, N-acetylation, lipidation,

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lipovlation, deimination, eliminylation, disulfide bridging, isoaspartate formation, racemization, glycation; carbamylation, carbonylation, isopeptide bond formation, sulfation, succinvlation, S-sulfonylation, S-sulfinylation, S-sulfenylation, S-glutathionylation, pyroglutamate formation, pro-5 pionvlation, adenvlvlation, nucleotide addition, iodination, hydroxylation, malonylation, butyrylation, amidation, alkylation, acylation, biotinylation, carbamylation, oxidation, and pegylation. In some embodiments, the peptide sequence(s) exhibit enhanced stability relative to one of SEQ ID NOs: 5-46. In some embodiments, the peptide sequence(s) exhibit enhanced oxalate-transport stimulation relative to one of SEQ ID NOs: 5-46.

In some embodiments, provided herein are pharmaceuti-15 cal compositions comprising a Sel1-derived peptide composition described herein and a pharmaceutically-acceptable carrier. In some embodiments, the pharmaceutical composition further comprises one or more additional therapeutic agents. In some embodiments, the additional therapeutic 20 agent is an oxalate degrading enzyme (e.g., oxalate oxidase, oxalate decarboxylase, oxalyl-CoA decarboxylase, formyl-CoA transferase, etc.). In some embodiments, the additional therapeutic agent is an oxalate degrading organism (e.g., bacteria, etc.).

In some embodiments, provided herein are methods of stimulating oxalate transport comprising administering to a subject a composition described herein. In some embodiments, administration comprises rectal administration, oral administration, and/or injection. In some embodiments, the 30 subject suffers from or is at risk for hyperoxaluria and/or hyperoxalemia. In some embodiments, the subject's risk of the risk of calcium oxalate kidney stones, nephrocalcinosis, oxalate nephropathy, end stage renal disease, chronic kidney disease, and/or systemic oxalosis is lowered by the method. 35

In some embodiments, provided herein are methods of treating or preventing hyperoxaluria and/or hyperoxalemia comprising administering a pharmaceutical composition (e.g., comprising a Sel1-derived peptide) described herein to a subject. In some embodiments, treating or preventing 40 hyperoxaluria and/or hyperoxalemia lowers the subject's risk of the risk of calcium oxalate kidney stones, nephrocalcinosis, oxalate nephropathy, end stage renal disease, and/or systemic oxalosis.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1. 301-derived peptides (individually or in combination) significantly stimulate ¹⁴C-oxalate influx into human Caco2-BBE (C2) cells.

FIG. 2. Several 318-derived peptides significantly stimulate ¹⁴C-oxalate influx into C2 cells.

FIG. 3. P8+9 peptides significantly stimulate (>2.4-fold) oxalate transport by C2 cells and similar to the conditioned medium (CM).

FIG. 4. P8+9 peptides significantly stimulate oxalate transport by C2 cells in a dose-dependent manner.

FIG. 5. P8+9 peptides significantly stimulated oxalate transport by an ileum (A) and sigmoid (B) Organoids.

FIG. 6. P7-10 peptides significantly stimulated oxalate 60 transport by an ileum (A) and sigmoid (B) Organoids.

FIG. 7. P8 and P9 peptides sequences.

FIG. 8. 318 Sel1 protein. SP=signal peptide. Boxes=SLR motifs and they represent P7-11 peptides.

FIG. 9. Compared with native P8 peptide, P8 with N-ter- 65 minal acetylation (Ac), C-terminal amidation (Am), all glycines replaced with PEG6 significantly stimulate 14C-

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oxalate influx into Caco2-BBE cells. Replacing natural amino acids with D-amino acids (D-aa) produces a nonfunctional P8 peptide.

FIG. 10. Compared with native P9 peptide, P9 with N-terminal acetvlation (Ac), C-terminal amidation (Am), all glycines replaced with PEG6 significantly stimulate 14Coxalate influx into Caco2-BBE cells. Replacing natural amino acids with D-amino acids (D-aa) produces a nonfunctional P9 peptide.

FIG. 11. P8 with G15 replaced with D-alanine and P9 with G16 replaced with D-alanine significantly stimulate ¹⁴Coxalate influx into Caco2-BBE cells.

FIG. 12. P9 with N-terminal acetylation (Ac), C-terminal amidation (Am), and G16 replaced with D-alanine (D-Ala) significantly stimulate oxalate transport by a human distal colon organoid.

FIG. 13. P9 with N-terminal acetylation (Ac), C-terminal amidation (Am), and G16 replaced with D-alanine (D-Ala) significantly stimulate oxalate transport by a human sigmoid colon organoid.

FIG. 14. P8 and magnesium citrate produced enhancement of oxalate uptake over P8 peptide alone.

FIG. 15. P9 and magnesium citrate produced enhancement of oxalate uptake over P8 peptide alone.

FIG. 16. The addition of aluminum to P8 or P9 did not result in increased oxalate transport and oxalate transport induction was similar to untreated

DEFINITIONS

Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of embodiments described herein, some preferred methods, compositions, devices, and materials are described herein. However, before the present materials and methods are described, it is to be understood that this invention is not limited to the particular molecules, compositions, methodologies or protocols herein described, as these may vary in accordance with routine experimentation and optimization. It is also to be understood that the terminology used in the description is for the purpose of describing the particular versions or embodiments only, and is not intended to limit the scope of the embodiments described herein.

Unless otherwise defined, all technical and scientific 45 terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. However, in case of conflict, the present specification, including definitions, will control. Accordingly, in the context of the embodiments described herein, the following definitions apply.

As used herein and in the appended claims, the singular forms "a", "an" and "the" include plural reference unless the context clearly dictates otherwise. Thus, for example, reference to "a Sel1-derived peptide" is a reference to one or more Sel1-derived peptides and equivalents thereof known to those skilled in the art, and so forth.

As used herein, the term "comprise" and linguistic variations thereof denote the presence of recited feature(s), element(s), method step(s), etc. without the exclusion of the presence of additional feature(s), element(s), method step(s), etc. Conversely, the term "consisting of" and linguistic variations thereof, denotes the presence of recited feature(s), element(s), method step(s), etc. and excludes any unrecited feature(s), element(s), method step(s), etc., except for ordinarily-associated impurities. The phrase "consisting essentially of" denotes the recited feature(s), element(s), method step(s), etc. and any additional feature(s), element(s),

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method step(s), etc. that do not materially affect the basic nature of the composition, system, or method. Many embodiments herein are described using open "comprising" language. Such embodiments encompass multiple closed "consisting of" and/or "consisting essentially of" embodi-5 ments, which may alternatively be claimed or described using such language.

As used herein, the term "hyperoxaluria" refers to the excessive urinary excretion of oxalate by a subject (e.g., >25 mg/day).

As used herein, the term "hyperoxalemia" refers to excessive plasma levels of oxalate in a subject. Various studies report a normal range of oxalate in the plasma of 1 to 3 μ mol per liter. Subjects with levels exceeding that range are considered to suffer from hyperoxalemia.

The term "amino acid" refers to natural amino acids, unnatural amino acids, and amino acid analogs, all in their D and L stereoisomers, unless otherwise indicated, if their structures allow such stereoisomeric forms.

Natural amino acids include alanine (Ala or A), arginine 20 (Arg or R), asparagine (Asn or N), aspartic acid (Asp or D), cysteine (Cys or C), glutamine (Gln or Q), glutamic acid (Glu or E), glycine (Gly or G), histidine (His or H), isoleucine (Ile or I), leucine (Leu or L), Lysine (Lys or K), methionine (Met or M), phenylalanine (Phe or F), proline 25 (Pro or P), serine (Ser or S), threonine (Thr or T), tryptophan (Trp or W), tyrosine (Tyr or Y) and valine (Val or V).

Unnatural amino acids include, but are not limited to, azetidinecarboxylic acid, 2-aminoadipic acid, 3-aminoadipic acid, beta-alanine, naphthylalanine ("naph"), aminopro- 30 pionic acid, 2-aminobutyric acid, 4-aminobutyric acid, 6-aminocaproic acid, 2-aminoheptanoic acid, 2-aminoisobutyric acid, 3-aminoisbutyric acid, 2-aminopimelic acid. tertiary-butylglycine ("tBuG"), 2,4-diaminoisobutyric acid, desmosine, 2,2'-diaminopimelic acid, 2,3-diaminopropionic 35 acid, N-ethylglycine, N-ethylasparagine, homoproline ("hPro" or "homoP"), hydroxylysine, allo-hydroxylysine, 3-hydroxyproline ("3Hyp"), 4-hydroxyproline ("4Hyp"), isodesmosine, allo-isoleucine, N-methylalanine ("MeAla" or "Nime"), N-alkylglycine ("NAG") including N-methyl- 40 glycine. N-methylisoleucine, N-alkylpentylglycine ("NAPG") including N-methylpentylglycine. N-methylvaline, naphthylalanine, norvaline ("Norval"), norleucine ("Norleu"), octylglycine ("OctG"), ornithine ("Orn"), pentylglycine ("pG" or "PGly"), pipecolic acid, thioproline 45 ("ThioP" or "tPro"), homoLysine ("hLys"), and homoArginine ("hArg").

The term "amino acid analog" refers to a natural or unnatural amino acid where one or more of the C-terminal carboxy group, the N-terminal amino group and side-chain 50 functional group has been chemically blocked, reversibly or irreversibly, or otherwise modified to another functional group. For example, aspartic acid-(beta-methyl ester) is an amino acid analog of aspartic acid; N-ethylglycine is an amino acid analog of glycine; or alanine carboxamide is an 55 amino acid analog of alanine. Other amino acid analogs include methionine sulfoxide, methionine sulfone, S-(carboxymethyl)-cysteine, S-(carboxymethyl)-cysteine sulfoxide and S-(carboxymethyl)-cysteine sulfone.

As used herein, the term "peptide" refers a short polymer 60 of amino acids linked together by peptide bonds. In contrast to other amino acid polymers (e.g., proteins, polypeptides, etc.), peptides are of about 50 amino acids or less in length. A peptide may comprise natural amino acids, non-natural amino acids, amino acid analogs, and/or modified amino 65 acids. A peptide may be a subsequence of naturally occurring protein or a non-natural (synthetic) sequence.

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As used herein, the term "mutant peptide" or "variant peptide" refers to a peptide having a distinct amino acid sequence from the most common variant occurring in nature, referred to as the "wild-type" sequence. A mutant peptide may be a subsequence of a mutant protein or polypeptide (e.g., a subsequence of a naturally-occurring protein that is not the most common sequence in nature) or may be a peptide that is not a subsequence of a naturally occurring protein or polypeptide. For example, a "mutant SLR peptide" (e.g., a "mutant Sel1 protein") may be a subsequence of a mutant version of SLR protein (e.g., Sel1 protein) or may be distinct sequence not found in naturallyoccurring SLR proteins (e.g., Sel1 proteins).

As used herein, the term "mutant polypeptide" or "variant polypeptide" refers to a polypeptide having a distinct amino acid sequence from the "wild-type" sequence. A mutant polypeptide may be a naturally-occurring protein that is not the most common sequence in nature (or a polypeptide fragment thereof) or may be a polypeptide that is not a subsequence of a naturally occurring protein or polypeptide. For example, a "mutant SLR polypeptide" may be a naturally occurring SLR protein (e.g., Sel1 protein), a polypeptide fragment of a SLR protein (e.g., Sel1 protein), or may be distinct sequence not found in naturally-occurring SLR proteins (e.g., Sel1 proteins).

As used herein, the term "artificial peptide" or "artificial polypeptide" refers to a peptide or polypeptide having a distinct amino acid sequence from those found in natural peptides and/or proteins. An artificial protein is not a subsequence of a naturally occurring protein, either the wildtype (i.e., most abundant) or mutant versions thereof. For example, an artificial SLR peptide or polypeptide is not a subsequence of naturally occurring SLR protein (e.g., Sel1 protein). An artificial peptide or polypeptide may be produced or synthesized by any suitable method (e.g., recombinant expression, chemical synthesis, enzymatic synthesis, etc.).

The terms "peptide mimetic" or "peptidomimetic" refer to a peptide-like molecule that emulates a sequence derived from a protein or peptide. A peptide mimetic or peptidomimetic may contain amino acids and/or non-amino acid components. Examples of peptidomimtecs include chemically modified peptides, peptoids (side chains are appended to the nitrogen atom of the peptide backbone, rather than to the α -carbons), (β -peptides (amino group bonded to the β carbon rather than the α carbon), etc.

As used herein, a "conservative" amino acid substitution refers to the substitution of an amino acid in a peptide or polypeptide with another amino acid having similar chemical properties, such as size or charge. For purposes of the present disclosure, each of the following eight groups contains amino acids that are conservative substitutions for one another:

- 1) Alanine (A) and Glycine (G);
- 2) Aspartic acid (D) and Glutamic acid (E);
- 3) Asparagine (N) and Glutamine (Q);
- 4) Arginine (R) and Lysine (K);
- 5) Isoleucine (I), Leucine (L), Methionine (M), and Valine
- (V);6) Phenylalanine (F), Tyrosine (Y), and Tryptophan (W);
- 7) Serine (S) and Threonine (T); and
- 8) Cysteine (C) and Methionine (M).

Naturally occurring residues may be divided into classes based on common side chain properties, for example: polar positive (histidine (H), lysine (K), and arginine (R)); polar negative (aspartic acid (D), glutamic acid (E)); polar neutral (serine (S), threonine (T), asparagine (N). glutamine (Q)); non-polar aliphatic (alanine (A), valine (V), leucine (L), isoleucine (I), methionine (M)); non-polar aromatic (phenylalanine (F), tyrosine (Y), tryptophan (W)); proline and glycine; and cysteine. As used herein, a "semi-conservative" amino acid substitution refers to the substitution of an amino acid in a peptide or polypeptide with another amino acid within the same class.

In some embodiments, unless otherwise specified, a conservative or semi-conservative amino acid substitution may also encompass non-naturally occurring amino acid residues ¹⁰ that have similar chemical properties to the natural residue. These non-natural residues are typically incorporated by chemical peptide synthesis rather than by synthesis in biological systems. These include, but are not limited to, ¹⁵ peptidomimetics and other reversed or inverted forms of amino acid moieties. Embodiments herein may, in some embodiments, be limited to natural amino acids, non-natural amino acids, and/or amino acid analogs. Non-conservative substitutions may involve the exchange of a member of one ²⁰ class for a member from another class.

As used herein, the term "sequence identity" refers to the degree to which two polymer sequences (e.g., peptide, polypeptide, nucleic acid, etc.) have the same sequential composition of monomer subunits. The term "sequence 25 similarity" refers to the degree with which two polymer sequences (e.g., peptide, polypeptide, nucleic acid, etc.) differ only by conservative and/or semi-conservative amino acid substitutions. The "percent sequence identity" (or "percent sequence similarity") is calculated by: (1) comparing 30 two optimally aligned sequences over a window of comparison (e.g., the length of the longer sequence, the length of the shorter sequence, a specified window, etc.), (2) determining the number of positions containing identical (or similar) monomers (e.g., same amino acids occurs in both 35 sequences, similar amino acid occurs in both sequences) to yield the number of matched positions. (3) dividing the number of matched positions by the total number of positions in the comparison window (e.g., the length of the longer sequence, the length of the shorter sequence, a 40 specified window), and (4) multiplying the result by 100 to yield the percent sequence identity or percent sequence similarity. For example, if peptides A and B are both 20 amino acids in length and have identical amino acids at all but 1 position, then peptide A and peptide B have 95% 45 sequence identity. If the amino acids at the non-identical position shared the same biophysical characteristics (e.g., both were acidic), then peptide A and peptide B would have 100% sequence similarity. As another example, if peptide C is 20 amino acids in length and peptide D is 15 amino acids 50 in length, and 14 out of 15 amino acids in peptide D are identical to those of a portion of peptide C, then peptides C and D have 70% sequence identity, but peptide D has 93.3% sequence identity to an optimal comparison window of peptide C. For the purpose of calculating "percent sequence 55 identity" (or "percent sequence similarity") herein, any gaps in aligned sequences are treated as mismatches at that position.

As used herein, the term "subject" broadly refers to any animal, including but not limited to, human and non-human 60 animals (e.g., dogs, cats, cows, horses, sheep, poultry, fish, crustaceans, etc.). As used herein, the term "patient" typically refers to a human subject that is being treated for a disease or condition.

As used herein, the term "effective amount" refers to the 65 amount of a sufficient to effect beneficial or desired results. An effective amount can be administered in one or more

administrations, applications or dosages and is not intended to be limited to a particular formulation or administration route.

As used herein, the terms "administration" and "administering" refer to the act of giving a drug, prodrug, or other agent, or therapeutic treatment to a subject or in vivo, in vitro, or ex vivo cells, tissues, and organs. Exemplary routes of administration to the human body can be through space under the arachnoid membrane of the brain or spinal cord (intrathecal), the eyes (ophthalmic), mouth (oral), skin (topical or transdermal), nose (nasal), lungs (inhalant), oral mucosa (buccal), ear, rectal, vaginal, by injection (e.g., intravenously, subcutaneously, intratumorally, intraperitoneally, etc.) and the like.

As used herein, the terms "co-administration" and "coadministering" refer to the administration of at least two agent(s) or therapies to a subject. In some embodiments, the co-administration of two or more agents or therapies is concurrent. In other embodiments, a first agent/therapy is administered prior to a second agent/therapy. Those of skill in the art understand that the formulations and/or routes of administration of the various agents or therapies used may vary. The appropriate dosage for co-administration can be readily determined by one skilled in the art. In some embodiments, when agents or therapies are co-administered, the respective agents or therapies are administered at lower dosages than appropriate for their administration alone. Thus, co-administration is especially desirable in embodiments where the co-administration of the agents or therapies lowers the requisite dosage of a potentially harmful (e.g., toxic) agent(s), and/or when co-administration of two or more agents results in sensitization of a subject to beneficial effects of one of the agents via co-administration of the other agent.

As used herein, the term "treatment" means an approach to obtaining a beneficial or intended clinical result. The beneficial or intended clinical result may include alleviation of symptoms, a reduction in the severity of the disease, inhibiting an underlying cause of a disease or condition, steadying diseases in a non-advanced state, delaying the progress of a disease, and/or improvement or alleviation of disease conditions.

As used herein, the term "pharmaceutical composition" refers to the combination of an active agent (e.g., Sellderived peptide) with a carrier, inert or active, making the composition especially suitable for diagnostic or therapeutic use in vitro, in vivo or ex vivo. The terms "pharmaceutically acceptable" or "pharmacologically acceptable," as used herein, refer to compositions that do not substantially produce adverse reactions, e.g., toxic, allergic, or immunological reactions, when administered to a subject.

As used herein, the term "pharmaceutically acceptable carrier" refers to any of the standard pharmaceutical carriers including, but not limited to, phosphate buffered saline solution, water, emulsions (e.g., such as an oil/water or water/oil emulsions), and various types of wetting agents, any and all solvents, dispersion media, coatings, sodium lauryl sulfate, isotonic and absorption delaying agents, disintegrants (e.g., potato starch or sodium starch glycolate), and the like. The compositions also can include stabilizers and preservatives. For examples of carriers, stabilizers and adjuvants, see, e.g., Martin, Remington's Pharmaceutical Sciences, 15th Ed., Mack Publ. Co., Easton, Pa. (1975), incorporated herein by reference in its entirety.

DETAILED DESCRIPTION

Provided herein are compositions comprising Sell-derived peptides thereof, and method of use thereof for the treatment/prevention of excess oxalate levels and conditions and diseases related thereto. In particular, peptides comprise Sel-like repeat (SLR) domains and/or tetratricopeptide (TPR) domains and may be linked together or administered with other peptides or polypeptides to treat/prevent diseases/ 5 conditions related to excess oxalate levels, such as hyperoxaluria and/or hyperoxalemia.

Most kidney stones (KS) are composed of calcium oxalate, and small increases in urine oxalate affect the stone risk. The mammalian intestine plays a crucial role in oxalate 10 homeostasis. Intestinal oxalate secretion mediated by anion exchanger SLC26A6 (A6) plays a major role in limiting net intestinal absorption of ingested oxalate; thereby preventing hyperoxaluria and calcium oxalate kidney stones (COKS). Hyperoxaluria and a high incidence of KS are commonly 15 seen in IBD patients. Hyperoxaluria is also emerging as a major complication of bariatric surgery for obesity. Primary hyperoxaluria (PH) is an inherited disease in which there is endogenous oxalate overproduction. Enhancing intestinal oxalate secretion is expected to lead to reduced urine and 20 plasma oxalate levels. In addition to degrading intraluminal dietary oxalate, the probiotic bacterium Oxalobacter formigenes (Of) also interacts with colonic epithelium by inducing colonic oxalate secretion, leading to reduced urinary excretion. Significant difficulties exist in sustaining Of colo- 25 nization in animals and humans in the absence of high exogenous oxalate.

Sel1-like repeat (SLR) proteins (e.g. Sel1, Hrd3, Chs4, Nif1, PodJ, ExoR, AlgK, HcpA, Hsp12, EnhC, LpnE, MotX, and MerG) are involved in signal transduction pathways. 30 SLR proteins (e.g., Sel1 proteins) have repeat units (e.g., repeat peptides). Most repeats are 5 to 40 amino acids (e.g., 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, or ranges therebetween), but longer repeat peptides 35 (e.g., 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 100, or longer or ranges therebetween) are within the scope of the SLR proteins (e.g., Sel1 proteins) herein. In some embodiments, repeat units fold into two to four secondary structural elements. In some embodiments, SLR proteins (e.g., Sel1 40 proteins) serve as adaptor proteins for the assembly of membrane-bound macromolecular complexes. Several bacterial and eukaryotic SLR proteins (e.g. Sel1 & Hrd3) are activated upon cellular stress. In some embodiments, Of Sel1 proteins are activated when oxalate is low in the culture 45 medium (e.g., as evidenced by the observation of a CM of higher (>2-fold) bioactivity under this condition). Bacterial LpnE, EnhC, HcpA, ExoR, and AlgK proteins mediate the interactions between bacterial and eukaryotic host cells. In some embodiments, the SLR motif establishes a link 50 between signal transduction pathways from eukaryotes and bacteria. In some embodiments, an SLR protein (e.g., Sel1 proteins) comprises leader sequences. In some embodiments, SLR proteins (e.g., Sel1 proteins) without leader sequences, such as PodJ or leaderless analogs of natural SLR 55 proteins (e.g., Sel1 proteins), are active in the periplasmic space. In some embodiments, bacterial SLR proteins (e.g., Sel1 proteins), such as HcpA, ExoR, EnhC and LpnE are responsible for the adaptation of bacteria to different eukaryotic hosts. 60

Provided herein are compositions (e.g., Sel1-derived peptides, fusions of multiple Sel1-derived peptides, combinations of Sel1-derived peptides, fusions of Sel1-derived peptides with other peptides/polypeptides, etc.) which stimulate the clearance of oxalate (e.g., activate oxalate transport) 65 from a biological environment (e.g., blood, urine, etc.). In some embodiments, compositions significantly reduce

oxalate concentrations (e.g., urine oxalate, blood oxalate, etc.) for example, by at least 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, or ranges therebetween. In some embodiments, compositions herein stimulate oxalate transport, thereby reducing in vivo oxalate levels in the blood (e.g., plasma oxalate levels), urine, etc., through mechanisms such as, for example, PKA activation and increased activity of SLC26 family members (e.g., SLC26A6) or other transporter(s); although embodiments herein are not limited to any particular mechanism of action and an understanding of the mechanism of action is not necessary to practice such embodiments.

In some embodiments. provided herein are compositions, kits, systems, and/or methods to treat, prevent, reduce the likelihood, treat/prevent a side effect of one or more of: hyperoxalemia, hyperoxaluria, nephrolithiasis, chronic kidney disease, end stage renal disease, calcium oxalate kidney stones, nephrocalcinosis, oxalate nephropathy, primary hyperoxaluria (PH), enteric hyperoxaluria (seen for example in IBD, following small bowel surgery or bariatric surgery, obesity, and celiac disease) and systemic oxalosis. In some embodiments, the reduction in oxalate levels and/or activation of oxalate transport is activated by compositions and methods described herein. In some embodiments, oxalate transport pathways are activated by the compositions and methods described herein. In some embodiments, compositions and methods are utilized in the treatment and/or prevention of hyperoxalemia, hyperoxaluria, and/or related diseases and conditions. In some embodiments, compositions and methods are utilized in screening for peptides and polypeptides useful in the treatment and/or prevention of hyperoxalemia, hyperoxaluria, and/or related diseases and conditions.

In some embodiments, provided herein are pharmaceutical compositions, SLR peptides (e.g., Sel1 peptides), TPR peptides (and fusions thereof), fusions of two or more Sel1-derived peptides (e.g., Sel1 peptides, Sel1 peptide analogs, etc.), fusions of Sel1-derived peptides (e.g., Sel1 peptides, Sell peptide analogs, etc.) with other peptides/ polypeptides, nucleic acids encoding the peptides, proteins and polypeptides herein, molecular complexes of the foregoing, etc. for the treatment or prevention of hyperoxalemia, hyperoxaluria, and/or related diseases and conditions. In some embodiments, provided herein are peptides derived from (e.g., a fragment of, derived from a fragment of, etc.) SEQ ID NO: 2 or SEQ ID NO: 4. In some embodiments, peptides comprise at least 50% (e.g., 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, less than 100%, 100%, or ranges therebetween) sequence identity to a portion of SEQ ID NO: 2 or SEQ ID NO: 4. In some embodiments, peptides comprise at least 50% (e.g., 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, less than 100%, 100%, or ranges therebetween) sequence similarity to a portion of SEQ ID NO: 2 or SEQ ID NO: 4. In some embodiments, provided herein are peptides derived from (e.g., a fragment of, derived from a fragment of, etc.) SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, SEQ ID NO: 30, SEQ ID NO: 31, SEQ ID NO: 32, SEQ ID NO: 33, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 36, SEQ ID NO: 37, SEQ ID NO: 38, SEQ ID NO: 39, SEQ ID NO: 40, SEQ ID NO:

41, SEQ ID NO: 42, SEQ ID NO: 43, SEQ ID NO: 44, SEQ ID NO: 45, or SEQ ID NO: 46. In some embodiments, peptides comprise at least 50% (e.g., 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, less than 100%, 100%, or ranges therebetween) sequence identity to all or a portion of 5 SEO ID NO: 5, SEO ID NO: 6, SEO ID NO: 7, SEO ID NO: 8, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ 10 ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, SEQ ID NO: 30, SEQ ID NO: 31, SEQ ID NO: 32, SEQ ID NO: 33, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 15 36, SEQ ID NO: 37, SEQ ID NO: 38, SEQ ID NO: 39, SEQ ID NO: 40, SEQ ID NO: 41, SEQ ID NO: 42, SEQ ID NO: 43, SEQ ID NO: 44, SEQ ID NO: 45, or SEQ ID NO: 46. In some embodiments, peptides comprise at least 50% (e.g., 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 20 less than 100%, 100%, or ranges therebetween) sequence similarity to all or a portion of SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9. SEQ ID NO: 10, SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID 25 NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, SEQ ID NO: 30, SEQ ID NO: 31, SEQ ID NO: 32, SEQ ID NO: 33, SEQ ID NO: 34, 30 SEQ ID NO: 35, SEQ ID NO: 36, SEQ ID NO: 37, SEQ ID NO: 38, SEQ ID NO: 39, SEQ ID NO: 40, SEQ ID NO: 41, SEQ ID NO: 42, SEQ ID NO: 43, SEQ ID NO: 44, SEQ ID NO: 45, or SEQ ID NO: 46.

In some embodiments, provided herein are fusions of two 35 or more Scl1-derived peptides. In some embodiments, a fusion comprises a first peptide sequence derived from SEQ ID NOs: 1-46 linked directly or indirectly (via a linker or connector) to a second peptide sequence derived from SEQ ID NOs: 1-46. A fusion may additionally comprise one or 40 more (e.g., 2, 3, 4, 5, 6, etc.) peptide sequence derived from SEQ ID NOs: 1-46 linked directly or indirectly. Embodiments are not limited by the combinations of various Sel1-derived peptides or the order in which they are fused.

In some embodiments, a peptide/polypeptide/fusion pro- 45 vided herein is an artificial, not naturally-occurring, sequence. In some embodiments, a peptide/polypeptide/ fusion described herein is prepared by methods known to those of ordinary skill in the art. For example, the peptide/ polypeptide/fusion can be synthesized using solid phase 50 polypeptide synthesis techniques (e.g. Fmoc or Boc chemistry). Alternatively, the peptide/polypeptide/fusion can be produced using recombinant DNA technology (e.g., using bacterial or eukaryotic expression systems). Further, a peptide/polypeptide/fusion may be expressed within a subject 55 (e.g., following administration of an appropriate vector). Accordingly, to facilitate such methods, provided herein are genetic vectors (e.g., plasmids, viral vectors (e.g. AAV), etc.) comprising a sequence encoding the peptide/polypeptide/fusion, as well as host cells comprising such vectors. 60 Furthermore, provided herein are the peptide/polypeptide/ fusion produced via such methods.

In some embodiments, the administration of compositions described herein (e.g. Sel1 peptides, fusions of Sel1 peptides, variants and mimetics of Sel1-derived peptides, 65 nucleic acids encoding Sel1-derived peptides, etc.) is provided. In some embodiments, provided herein is the admin-

istration of bioactive agents which reduce oxalate levels in vivo, or are otherwise described herein.

In some embodiments, a peptide is provided comprising or consisting of all or a portion of SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, SEQ ID NO: 30, SEQ ID NO: 31, SEQ ID NO: 32, SEQ ID NO: 33, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 36, SEQ ID NO: 37, SEQ ID NO: 38, SEQ ID NO: 39, SEQ ID NO: 40, SEQ ID NO: 41, SEQ ID NO: 42, SEQ ID NO: 43, SEQ ID NO: 44, SEQ ID NO: 45, or SEQ ID NO: 46. In some embodiments, a peptide is provided comprising at least 50% sequence identity to one of SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, SEQ ID NO: 30, SEQ ID NO: 31, SEQ ID NO: 32, SEQ ID NO: 33, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 36, SEQ ID NO: 37, SEQ ID NO: 38, SEQ ID NO: 39, SEQ ID NO: 40, SEQ ID NO: 41, SEQ ID NO: 42, SEQ ID NO: 43, SEQ ID NO: 44, SEQ ID NO: 45, or SEQ ID NO: 46. (e.g. at least 60% sequence identity, at least 70% sequence identity, at least 80% sequence identity, at least 90% sequence identity, at least 95% sequence identity, etc.). In some embodiments, a peptide comprises at least one substitution (e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, or ranges therebetween (e.g., 1-10 substitutions)) from a Sel1 peptide sequence described herein (e.g., SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, SEQ ID NO: 30, SEQ ID NO: 31, SEQ ID NO: 32, SEQ ID NO: 33, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 36, SEQ ID NO: 37, SEQ ID NO: 38, SEQ ID NO: 39, SEQ ID NO: 40, SEQ ID NO: 41, SEQ ID NO: 42, SEQ ID NO: 43, SEQ ID NO: 44, SEQ ID NO: 45, or SEQ ID NO: 46.).

In some embodiments, a peptide/polypeptide is provided that is a fusion of two or more of SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, SEQ ID NO: 20. SEO ID NO: 21. SEO ID NO: 22. SEO ID NO: 23. SEO ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, SEQ ID NO: 30, SEQ ID NO: 31, SEQ ID NO: 32, SEQ ID NO: 33, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 36, SEQ ID NO: 37, SEQ ID NO: 38, SEQ ID NO: 39, SEQ ID NO: 40, SEQ ID NO: 41, SEQ ID NO: 42, SEQ ID NO: 43, SEQ ID NO: 44, SEQ ID NO: 45, or SEQ ID NO: 46. In some embodiments, a peptide/polypeptide is provided that is a fusion of one or more peptides derived from (e.g., fragments of, comprising substitutions relative to, etc.) SEQ ID NO: 5. SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9, SEQ ID

NO: 10, SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, SEQ ID NO: 30, SEQ ID NO: 31, SEQ ID NO: 32, SEQ ID NO: 33, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 36, SEQ ID NO: 37, SEQ ID NO: 38, SEQ ID NO: 39, SEQ ID NO: 40, SEQ ID NO: 41, SEQ ID NO: 42, SEQ ID NO: 43, SEQ ID NO: 44, SEQ ID 10 NO: 45, or SEQ ID NO: 46. In some embodiments, a fusion comprises a portion (or portions) having one or more substitutions (e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, or more) compared to SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID 15 NO: 9, SEQ ID NO: 10, SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, 20 SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, SEQ ID NO: 30, SEQ ID NO: 31, SEQ ID NO: 32, SEQ ID NO: 33, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 36, SEQ ID NO: 37, SEQ ID NO: 38, SEQ ID NO: 39, SEQ ID NO: 40, SEQ ID NO: 41, SEQ ID NO: 42, SEQ ID NO: 43, SEQ ID 25 NO: 44, SEQ ID NO: 45, or SEQ ID NO: 46.

In some embodiments, fusions are provided comprising two or more (e.g., 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) of the peptides herein or variants thereof. In some embodiments, a fusion may comprise multiple (e.g., 2, 3, 4, 5, 6, 7, 8, 9, 10, 30 or more) copies of a single Sel1-derived peptide or variant thereof. In some embodiments, fusions comprise one or more (e.g., 2, 3, 4, 5, 6, 7, 8, 9, 10, or more) of the peptides herein or variants thereof and one or more non-Sel1-derived peptides or polypeptides. In some embodiments, a fusion 35 may comprise a functional peptide or polypeptide segment. In some embodiments, the functional peptide or polypeptide segment comprises a signaling moiety, therapeutic moiety, localization moiety (e.g., cellular import signal, nuclear localization signal, etc.), detectable moiety (e.g., fluorescent 40 moiety, contrast agent), or isolation/purification moiety (e.g., streptavidin, His₆, etc.). Such fusions may be expressed from a recombinant DNA which encodes the Sel1-derived peptide(s) and the additional peptide/polypeptide or may be formed by chemical synthesis. For instance, 45 the fusion may comprise a Sel1-derived peptide(s) and an enzyme of interest, a luciferase, RNasin or RNase, and/or a channel protein (e.g., ion channel protein), a receptor, a membrane protein, a cytosolic protein, a nuclear protein, a structural protein, a phosphoprotein, a kinase, a signaling 50 protein, a metabolic protein, a mitochondrial protein, a receptor associated protein, a fluorescent protein, an enzyme substrate, a transcription factor, selectable marker protein, nucleic acid binding protein, extracellular matrix protein, secreted protein, receptor ligand, serum protein, a protein 55 with reactive cysteines, a transporter protein, a targeting sequence (e.g., a myristylation sequence), a mitochondrial localization sequence, or a nuclear localization sequence. The additional peptide/polypeptide may be fused to the N-terminus and/or the C-terminus of the Sel1-derived peptide(s). In one embodiment, the fusion protein comprises a first peptide/polypeptide at the N-terminus and another (different) peptide/polypeptide at the C-terminus of the Sel1-derived peptide(s). Optionally, the elements in the fusion are separated by a connector sequence, e.g., prefer- 65 ably one having at least 2 amino acid residues, such as one having 13 and up to 40 or 50 amino acid residues. In some

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embodiments, the presence of a connector sequence in a fusion protein of the invention does not substantially alter the function of either element (e.g., Sel1-derived peptide(s)) in the fusion relative to the function of each individual element, likely due to the connector sequence providing flexibility (autonomy) for each element in the fusion. In certain embodiments, the connector sequence is a sequence recognized by an enzyme or is photocleavable. For example, the connector sequence may include a protease recognition site.

In some embodiments, compositions are provided containing two or more unlinked peptides/polypeptides/fusions comprising SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25. SEO ID NO: 26. SEO ID NO: 27. SEO ID NO: 28. SEQ ID NO: 29, SEQ ID NO: 30, SEQ ID NO: 31, SEQ ID NO: 32, SEQ ID NO: 33, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 36, SEQ ID NO: 37, SEQ ID NO: 38, SEQ ID NO: 39, SEQ ID NO: 40, SEQ ID NO: 41, SEQ ID NO: 42, SEQ ID NO: 43, SEQ ID NO: 44, SEQ ID NO: 45, and SEQ ID NO: 46. In some embodiments, a composition is provided two or more peptides derived from (e.g., fragments of, comprising substitutions relative to, etc.) SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEO ID NO: 14, SEO ID NO: 15, SEO ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, SEQ ID NO: 30, SEQ ID NO: 31, SEQ ID NO: 32, SEQ ID NO: 33, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 36, SEQ ID NO: 37, SEQ ID NO: 38, SEQ ID NO: 39, SEQ ID NO: 40, SEQ ID NO: 41, SEQ ID NO: 42, SEQ ID NO: 43, SEQ ID NO: 44, SEQ ID NO: 45, and SEQ ID NO: 46. In some embodiments, a composition comprises two or more peptides comprising one or more substitutions (e.g., 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, or more) compared to SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, SEQ ID NO: 30, SEQ ID NO: 31, SEQ ID NO: 32, SEQ ID NO: 33, SEQ ID NO: 34, SEQ ID NO: 35, SEQ ID NO: 36, SEQ ID NO: 37, SEQ ID NO: 38, SEO ID NO: 39, SEQ ID NO: 40, SEQ ID NO: 41, SEQ ID NO: 42, SEQ ID NO: 43, SEQ ID NO: 44, SEQ ID NO: 45, and SEQ ID NO: 46.

Embodiments are not limited to the specific sequences listed herein. In some embodiments, peptides/polypeptides/ fusions meeting limitations described herein and having substitutions not explicitly described are within the scope of embodiments here. In some embodiments, the peptides/ polypeptides/fusions described herein are further modified (e.g., substitution, deletion, or addition of standard amino acids; chemical modification; etc.). Modifications that are understood in the field include N-terminal modification, C-terminal modification (which protects the peptide from proteolytic degradation), alkylation of amide groups, hydrocarbon "stapling" (e.g., to stabilize conformations). In some 10

embodiments, the peptides/polypeptides described herein may be modified by conservative residue substitutions, for example, of the charged residues (K to R, R to K. D to E and E to D). Modifications of the terminal carboxy group include, without limitation, the amide, lower alkyl amide, constrained alkyls (e.g. branched, cyclic, fused, adamantyl) alkyl, dialkyl amide, and lower alkyl ester modifications. Lower alkyl is C1-C4 alkyl. Furthermore, one or more side groups, or terminal groups, may be protected by protective groups known to the ordinarily-skilled peptide chemist. The α -carbon of an amino acid may be mono- or dimethylated.

In some embodiments, peptides (or fusions thereof) are provided comprising: (i) one or more of the amino acid residues in the peptide are D-enantiomers, (ii) an N-terminally acetyl group, (iii) a deamidated C-terminal group, (iv) one or more unnatural amino acids, (v) one or more amino acid analogs, and/or (vi) one or more peptoid amino acids. In some embodiments, the peptide (or fusions thereof) or an amino acid therein comprises a modification selected from 20 the group consisting of phosphorylation, glycosylation, ubiquitination, S-nitrosylation, methylation, N-acetylation, lipidation, lipoylation, deimination, eliminylation, disulfide bridging, isoaspartate formation, racemization, glycation; carbamylation, carbonylation, isopeptide bond formation, 25 sulfation, succinylation, S-sulfonylation, S-sulfinylation, S-sulfenylation, S-glutathionylation, pyroglutamate formation, propionylation, adenylylation, nucleotide addition, iodination, hydroxylation, malonylation, butyrylation, amidation, C-terminal amidation, de-amidation, alkylation, acy- 30 lation, biotinylation, carbamylation, oxidation, and pegylation. In some embodiments, the peptide exhibits enhanced stability relative to one of SEQ ID NOs: 5-46. In some embodiments, the peptide exhibits enhanced oxalate transport stimulation activity relative to one of SEQ ID NOs: 35 5-46

In some embodiments, any embodiments described herein may comprise mimetics corresponding to Sel1-derived peptide and/or variants thereof, with various modifications that are understood in the field. In some embodiments, residues 40 in the peptide sequences described herein may be substituted with amino acids having similar characteristics (e.g., hydrophobic to hydrophobic, neutral to neutral, etc.) or having other desired characteristics (e.g., more acidic, more hydrophobic, less bulky, more bulky, etc.). In some embodiments, 45 non-natural amino acids (or naturally-occurring amino acids other than the standard 20 amino acids) are substituted in order to achieve desired properties.

In some embodiments, residues having a side chain that is positively charged under physiological conditions, or resi- 50 dues where a positively-charged side chain is desired, are substituted with a residue including, but not limited to: lysine, homolysine, 6-hydroxylysine, homoarginine, 2,4diaminobutyric acid, 3-homoarginine, D-arginine, arginal (-COOH in arginine is replaced by -CHO), 2-amino-3- 55 guanidinopropionic acid, nitroarginine (N(G)-nitroarginine), nitrosoarginine (N(G)-nitrosoarginine), methylarginine (N-methyl-arginine), c-N-methyllysine, allo-hydroxylysine, 2,3-diaminopropionic acid, 2,2'-diaminopimelic acid, ornithine, sym-dimethylarginine, asym-dimethylarginine, 2,6diaminohexinic acid, p-aminobenzoic acid and 3-aminotyrosine and, histidine, 1-methylhistidine, and 3-methylhistidine.

A neutral residue is a residue having a side chain that is uncharged under physiological conditions. A polar residue 65 preferably has at least one polar group in the side chain. In some embodiments, polar groups are selected from

hydroxyl, sulfhydryl, amine, amide and ester groups or other groups which permit the formation of hydrogen bridges.

In some embodiments, residues having a side chain that is neutral/polar under physiological conditions, or residues where a neutral side chain is desired, are substituted with a residue including, but not limited to: asparagine, cysteine, glutamine, serine, threonine, tyrosine, citrulline, N-methylserine, homoserine, allo-threonine and 3,5-dinitro-tyrosine, and β -homoserine.

Residues having a non-polar, hydrophobic side chain are residues that are uncharged under physiological conditions, preferably with a hydropathy index above 0, particularly above 3. In some embodiments, non-polar, hydrophobic side chains are selected from alkyl, alkylene, alkoxy, alkenoxy, alkylsulfanyl and alkenylsulfanyl residues having from 1 to 10, preferably from 2 to 6, carbon atoms, or aryl residues having from 5 to 12 carbon atoms. In some embodiments, residues having a non-polar, hydrophobic side chain are, or residues where a non-polar, hydrophobic side chain is desired, are substituted with a residue including, but not limited to: leucine, isoleucine, valine, methionine, alanine, phenylalanine, N-methylleucine, tert-butylglycine, octylglycine, cyclohexylalanine, β-alanine, 1-aminocyclohexylcarboxylic acid, N-methylisoleucine, norleucine, norvaline, and N-methylvaline

In some embodiments, peptide and polypeptides are isolated and/or purified (or substantially isolated and/or substantially purified). Accordingly, in such embodiments, peptides and/or polypeptides are provided in substantially isolated form. In some embodiments, peptides and/or polypeptides are isolated from other peptides and/or polypeptides as a result of solid phase peptide synthesis, for example. Alternatively, peptides and/or polypeptides can be substantially isolated from other proteins after cell lysis from recombinant production. Standard methods of protein purification (e.g., HPLC) can be employed to substantially purify peptides and/or polypeptides. In some embodiments, the present invention provides a preparation of peptides and/or polypeptides in a number of formulations, depending on the desired use. For example, where the polypeptide is substantially isolated (or even nearly completely isolated from other proteins), it can be formulated in a suitable medium solution for storage (e.g., under refrigerated conditions or under frozen conditions). Such preparations may contain protective agents, such as buffers, preservatives, cryprotectants (e.g., sugars such as trehalose), etc. The form of such preparations can be solutions, gels, etc. In some embodiments, peptides and/or polypeptides are prepared in lyophilized form. Moreover, such preparations can include other desired agents, such as small molecules or other peptides, polypeptides or proteins. Indeed, such a preparation comprising a mixture of different embodiments of the peptides and/or polypeptides described here may be provided.

In some embodiments, provided herein are peptidomimetic versions of the peptide sequences described herein or variants thereof. In some embodiments, a peptidomimetic is characterized by an entity that retains the polarity (or non-polarity, hydrophobicity, etc.), three-dimensional size, and functionality (bioactivity) of its peptide equivalent but wherein all or a portion of the peptide bonds have been replaced (e.g., by more stable linkages). In some embodiments, 'stable' refers to being more resistant to chemical degradation or enzymatic degradation by hydrolytic enzymes. In some embodiments, the bond which replaces the amide bond (e.g., amide bond surrogate) conserves some properties of the amide bond (e.g., conformation, steric bulk, electrostatic character, capacity for hydrogen bonding, etc.). Cyclization (head-to-tail, head/tail-to-side-chain, and/or side-chain-to-side-chain) enhances peptide stability and permeability by introducing conformation constraint, thereby reducing peptide flexibility, and a cyclic enkephalin analog is highly resistant to enzymatic degradation. Chapter 14 of "Drug Design and Development", Krogsgaard, Larsen, Liljefors and Madsen (Eds) 1996, Horwood Acad. Publishers provides a general discussion of techniques for the design and synthesis of peptidomimetics and is herein 10 incorporated by reference in its entirety. Suitable amide bond surrogates include, but are not limited to: N-alkylation (Schmidt, R. et al., Int. J. Peptide Protein Res., 1995, 46,47; herein incorporated by reference in its entirety), retroinverse amide (Chorev, M. and Goodman, M., Acc. Chem. 15 Res, 1993, 26, 266; herein incorporated by reference in its entirety), thioamide (Sherman D. B. and Spatola, A. F. J. Am. Chem. Soc., 1990, 112, 433; herein incorporated by reference in its entirety), thioester, phosphonate, ketomethylene (Hoffman, R. V. and Kim, H. O. J. Org. Chem., 1995, 20 60, 5107; herein incorporated by reference in its entirety), hydroxymethylene, fluorovinyl (Allmendinger, T. et al., Tetrahydron Lett., 1990, 31, 7297; herein incorporated by reference in its entirety), vinyl, methyleneamino (Sasaki, Y and Abe, J. Chem. Pharm. Bull. 1997 45, 13; herein incor- 25 porated by reference in its entirety), methylenethio (Spatola, A. F., Methods Neurosci, 1993, 13, 19; herein incorporated by reference in its entirety), alkane (Lavielle, S. et. al., Int. J. Peptide Protein Res., 1993, 42, 270; herein incorporated by reference in its entirety) and sulfonamido (Luisi, G. et al. 30 Tetrahedron Lett. 1993, 34, 2391; herein incorporated by reference in its entirety).

As well as replacement of amide bonds, peptidomimetics may involve the replacement of larger structural moieties with di- or tripeptidomimetic structures and in this case, 35 mimetic moieties involving the peptide bond, such as azolederived mimetics may be used as dipeptide replacements. Suitable peptidomimetics include reduced peptides where the amide bond has been reduced to a methylene amine by treatment with a reducing agent (e.g. borane or a hydride 40 reagent such as lithium aluminum-hydride); such a reduction has the added advantage of increasing the overall cationicity of the molecule.

Other peptidomimetics include peptoids formed, for example, by the stepwise synthesis of amide-functionalised 45 polyglycines. Some peptidomimetic backbones will be readily available from their peptide precursors, such as peptides which have been permethylated, suitable methods are described by Ostresh, J. M. et al. in Proc. Natl. Acad. Sci. USA (1994) 91, 11138-11142; herein incorporated by ref- 50 erence in its entirety.

In some embodiments, provided herein are pharmaceutical compositions comprising of one or more Sel1-derived peptides or fusions or variants thereof and a pharmaceutically acceptable carrier. Any carrier which can supply an 55 active peptide or polypeptide (e.g., without destroying the peptide or polypeptide within the carrier) is a suitable carrier, and such carriers are well known in the art. In some embodiments, compositions are formulated for administration by any suitable route, including but not limited to, orally (e.g., such as in the form of tablets, capsules, granules or powders), sublingually, bucally, parenterally (such as by subcutaneous, intravenous, intramuscular, intradermal, or intracisternal injection or infusion (e.g., as sterile injectable aqueous or non-aqueous solutions or suspensions, etc.)), 65 nasally (including administration to the nasal membranes, such as by inhalation spray), topically (such as in the form

of a cream or ointment), transdermally (such as by transdermal patch), rectally (such as in the form of suppositories), etc.

In some embodiments, provided herein are methods for treating patients suffering from (or at risk of) hyperoxaluria, hyperoxalemia, and/or in need of treatment (or preventative therapy). In some embodiments, a pharmaceutical composition comprising at least one Sel1-derived peptide described herein (or fusions or variants thereof) is delivered to such a patient in an amount and at a location sufficient to treat the condition. In some embodiments, peptides and/or polypeptides (or pharmaceutical composition comprising such) can be delivered to the patient systemically or locally, and it will be within the ordinary skill of the medical professional treating such patient to ascertain the most appropriate delivery route, time course, and dosage for treatment. It will be appreciated that application methods of treating a patient most preferably substantially alleviates or even eliminates such symptoms; however, as with many medical treatments, application of the inventive method is deemed successful if, during, following, or otherwise as a result of the inventive method, the symptoms of the disease or disorder in the patient subside to an ascertainable degree.

A pharmaceutical composition may be administered in the form which is formulated with a pharmaceutically acceptable carrier and optional excipients, adjuvants, etc. in accordance with good pharmaceutical practice. The Sel1-derived peptide (or fusions or variants thereof) pharmaceutical composition may be in the form of a solid, semi-solid or liquid dosage form: such as powder, solution, elixir, syrup, suspension, cream, drops, paste and spray. As those skilled in the art would recognize, depending on the chosen route of administration (e.g. pill. injection, etc.), the composition form is determined. In general, it is preferred to use a unit dosage form in order to achieve an easy and accurate administration of the active pharmaceutical peptide or polypeptide. In general, the therapeutically effective pharmaceutical compound is present in such a dosage form at a concentration level ranging from about 0.5% to about 99% by weight of the total composition, e.g., in an amount sufficient to provide the desired unit dose. In some embodiments, the pharmaceutical composition may be administered in single or multiple doses. The particular route of administration and the dosage regimen will be determined by one of skill in keeping with the condition of the individual to be treated and said individual's response to the treatment. In some embodiments, Sel1-derived peptide described herein (or fusions or variants thereof) pharmaceutical composition is provided in a unit dosage form for administration to a subject, comprising one or more nontoxic pharmaceutically acceptable carriers, adjuvants or vehicles. The amount of the active ingredient that may be combined with such materials to produce a single dosage form will vary depending upon various factors, as indicated above. A variety of materials can be used as carriers, adjuvants and vehicles in the composition of the invention, as available in the pharmaceutical art. Injectable preparations, such as oleaginous solutions, suspensions or emulsions, may be formulated as known in the art, using suitable dispersing or wetting agents and suspending agents, as needed. The sterile injectable preparation may employ a nontoxic parenterally acceptable diluent or solvent such as sterile nonpyrogenic water or 1,3-butanediol. Among the other acceptable vehicles and solvents that may be employed are 5% dextrose injection, Ringer's injection and isotonic sodium chloride injection (as described in the USP/NF). In addition, sterile, fixed oils may be conventionally employed as solvents or suspending

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media. For this purpose, any bland fixed oil may be used, including synthetic mono-, di- or triglycerides. Fatty acids such as oleic acid can also be used in the preparation of injectable compositions.

In various embodiments, the peptides disclosed herein are 5 derivatized by conjugation to one or more polymers or small molecule substituents.

In certain of these embodiments, the Sel1-derived peptides described herein (or fusions or variants thereof) are derivatized by coupling to polyethylene glycol (PEG). Cou- 10 pling may be performed using known processes. See, Int. J. Hematology, 68:1 (1998); Bioconjugate Chem., 6:150 (1995); and Crit. Rev. Therap. Drug Carrier Sys., 9:249 (1992) all of which are incorporated herein by reference in their entirety. Those skilled in the art, therefore, will be able 15 to utilize such well-known techniques for linking one or more polyethylene glycol polymers to the peptides and polypeptides described herein. Suitable polyethylene glycol polymers typically are commercially available or may be made by techniques well known to those skilled in the art. 20 The polyethylene glycol polymers preferably have molecular weights between 500 and 20,000 and may be branched or straight chain polymers.

The attachment of a PEG to a peptide or polypeptide described herein can be accomplished by coupling to amino, 25 carboxyl or thiol groups. These groups will typically be the N- and C-termini and on the side chains of such naturally occurring amino acids as lysine, aspartic acid, glutamic acid and cysteine. Since the peptides and polypeptides of the present disclosure can be prepared by solid phase peptide 30 chemistry techniques, a variety of moieties containing diamino and dicarboxylic groups with orthogonal protecting groups can be introduced for conjugation to PEG.

The present disclosure also provides for conjugation of the Sel1-derived peptides described herein (or fusions or 35 variants thereof) to one or more polymers other than polyethylene glycol.

In some embodiments, the Sel1-derived peptides described herein (or fusions or variants thereof) are derivatized by conjugation or linkage to, or attachment of, 40 polyamino acids (e.g., poly-his, poly-arg, poly-lys, etc.) and/or fatty acid chains of various lengths to the N- or C-terminus or amino acid residue side chains. In certain embodiments, the peptides and polypeptides described herein are derivatized by the addition of polyamide chains, 45 particularly polyamide chains of precise lengths, as described in U.S. Pat. No. 6,552,167, which is incorporated by reference in its entirety. In yet other embodiments, the peptides and polypeptides are modified by the addition of alkyIPEG moieties as described in U.S. Pat. Nos. 5,359,030 50 and 5,681,811, which are incorporated by reference in their entireties.

In select embodiments, the Sel1-derived peptides described herein (or fusions or variants thereof) are derivatized by conjugation to polymers that include albumin and 55 gelatin. See, Gombotz and Pettit, Bioconjugate Chem., 6:332-351, 1995, which is incorporated herein by reference in its entirety.

In further embodiments, the Sel1-derived peptides described herein (or fusions or variants thereof) are conjugated or fused to immunoglobulins or immunoglobulin fragments, such as antibody Fc regions.

In some embodiments, the pharmaceutical compositions described herein (e.g., comprising the Sel1-derived peptides described herein (or fusions or variants thereof) find use in 65 the treatment and/or prevention of hyperoxaluria, hyperoxalemia, and related conditions. In some embodiments, the

compositions are administered to a subject. In certain embodiments, the patient is an adult. In other embodiments, the patient is a child.

In various embodiments, the Sel1-derived peptides described herein (or fusions or variants thereof) are administered in an amount, on a schedule, and for a duration sufficient to decrease triglyceride levels by at least 5%, 10%, 15%, 20% or 25% or more as compared to levels just prior to initiation of treatment. In some embodiments, the Sel1derived peptides described herein (or fusions or variants thereof) are administered in an amount, on a dosage schedule, and for a duration sufficient to decrease oxalate levels (e.g., in urine, in plasma) by at least 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45% or 50%. In particular embodiments, the Sel1-derived peptides described herein (or fusions or variants thereof) are administered in an amount, on a schedule, and for a time sufficient to decrease oxalate levels (e.g., in urine, in plasma) by at least 55%, 60%. 65%, even at least about 70% or more.

In certain embodiments, the Sel1-derived peptides described herein (or fusions or variants thereof) are administered in an amount, expressed as a daily equivalent dose regardless of dosing frequency, of 50 micrograms ("mcg") per day, 60 mcg per day, 70 mcg per day, 75 mcg per day, 100 mcg per day, 150 mcg per day, 200 mcg per day, or 250 mcg per day. In some embodiments, the Sel1-derived peptides described herein (or fusions or variants thereof) are administered in an amount of 500 mcg per day, 750 mcg per day, or 1 milligram ("mg") per day. In yet further embodiments, the Sel1-derived peptides described herein (or fusions or variants thereof) are administered in an amount. expressed as a daily equivalent dose regardless of dosing frequency, of 1-10 mg per day, including 1 mg per day, 1.5 mg per day, 1.75 mg per day, 2 mg per day, 2.5 mg per day, 3 mg per day, 3.5 mg per day, 4 mg per day, 4.5 mg per day, 5 mg per day, 5.5 mg per day, 6 mg per day, 6.5 mg per day, 7 mg per day, 7.5 mg per day, 8 mg per day, 8.5 mg per day, 9 mg per day, 9.5 mg per day, or 10 mg per day.

In various embodiments, the Sel1-derived peptides described herein (or fusions or variants thereof) are administered on a monthly, biweekly, weekly, daily ("QD"), or twice a day ("BID") dosage schedule. In other embodiments, the peptide/polypeptide is administered. In typical embodiments, the peptide/polypeptide is administered for at least 3 months, at least 6 months, at least 12 months, or more. In some embodiments, Sel1-derived peptides described herein (or fusions or variants thereof) are administered for at least 18 months, 2 years, 3 years, or more.

EXPERIMENTAL

Of-derived factors are secreted in its culture conditioned medium (CM) that significantly stimulate (>2.8-fold) oxalate transport by human intestinal Caco2-BBE (C2) cells through PKA activation and stimulation of the oxalate transporters SLC26A6 (A6) and SLC26A2 (A2). In vivo, rectal administration of Of CM reduced urinary oxalate excretion >32.5% in hyperoxaluric mice, and stimulated colonic oxalate secretion >42%, reflecting the therapeutic impact of these factors. Multiple Sel1 proteins are involved in protein-protein interactions and signal transduction pathways as the major Of-derived factors. These Sel1 proteins closely recapitulate the effects of the Of-derived factors and similarly stimulate (1.4-2.4-fold) oxalate transport by C2 cells via PKA and the A2/A6 transporters. 35-amino acid peptides (P7-10) within a Sel1 protein are identified which significantly stimulate (1.5-fold individually and >2.4-fold

by P8+9) oxalate transport by C2 cells. P8+9 peptides also significantly stimulated oxalate transport by human sigmoid colon (1.8-fold), distal colon (1.7-fold), and ileum (2-fold) organoids (ex vivo intestinal epithelia models fully mimicking the in vivo physiological responses), indicating that P8+9 peptides will stimulate oxalate transport in human colonic and ileal epithelia in vivo. Therefore, the identification and characterization of the active motifs responsible for colonic oxalate transport provides an effective drug for reducing plasma and urine oxalate levels.

Sel1 (sel: suppressor-enhancer of lin) proteins are identified as the major Of-derived secreted factors. Sel1 proteins are in the family of solenoid proteins, distinguishable from general globular proteins by the presence of intra-molecular sequence similarities (repeats) that lead to their modular architectures (Ref. 25; incorporated by reference in its entirety). A cross-genome analysis of repeats revealed an a/a-repeat consisting of 36 to 44 amino acids called Sel1-like repeat (SLR). SLR proteins (e.g. Sel1, Hrd3, etc.) are involved in protein-protein interactions and signal transduc- 20 tion pathways (Ref. 25; incorporated by reference in its entirety) and the latter is important since CM stimulates oxalate transport through PKA activation. SLR proteins serve as adaptor proteins for the assembly of membranebound macromolecular complexes (Ref. 25; incorporated by 25 reference in its entirety), which might include oxalate transporters (e.g. A6) and other regulatory proteins. Several bacterial and eukaryotic SLR proteins (e.g. Sel1) are activated upon cellular stress (Ref. 25; incorporated by refer-

ence in its entirety), which of significant interest since Of Sel1 proteins might be activated when oxalate is limited given the observation of >2-fold higher stimulatory effects from the CM under this condition (Ref. 4; incorporated by reference in its entirety). The Of genome has 44 Sel1 genes, which are significantly over-represented and reflects the importance of Sel1 proteins for Of. The dependence of Of on oxalate for viability might explain why it has 44 Sel1 proteins, and it is possible that Of utilizes Sel1 proteins to derive oxalate from blood by inducing colonic secretion, as observed with CM, to sustain its survival when dietary oxalate is limited. Supporting this notion, SLR proteins mediate the interactions between bacterial and eukaryotic host cells and establishes a link between signal transduction pathways from eukaryotes and bacteria. Most of the 44 Sel1 proteins have signal peptides and therefore are secreted proteins. SLR proteins are involved in the ER-associated protein degradation (ERAD) and may also be responsible for the adaptation of bacteria to different eukaryotic hosts.

Experiments conducted during development of embodiments herein demonstrated that certain Sel1 proteins (e.g., 301, 310, 318, and 321) are capable of significantly stimulating oxalate transport by cells (See, PCT/US18/14494; herein incorporated by reference in its entirety). Sel1 proteins 301 and 318 have been demonstrated to be the most potent stimulators of oxalate transport in human Caco2-BBE (C2) cells, and are predicted to have six (P1-P6; Table 1) and five (P7-P11; Table 2) SLR domains (each 30-35 amino acids in length and called peptides here), respectively.

TABLE 1

	Predicted 301 SLR domains				
P1	ADAQNRLGIAYRYGTGVRKNPALSVKWLEKAAKQG	SEQ	ID	NO :	5
Ρ2	QYNLGVAYYYGRGI KKDFSEAVSWYKKSAE	SEQ	ID	NO :	6
РЗ	AYHNLGTAYYDGIGVDKNPHEAVRWWKKAAELG	SEQ	ID	NO :	7
P4	QSQYNLGIAYEEGWGAEKNPENAVFWYRKAAEQGH	SEQ	ID	NO :	8
Р5	ARAQFNLGKTFYIGAGINKNTDKAVYWFIKAANQG	SEQ	ID	NO :	9
P6	ESQNNLGALYNDGNGVDRDYQEAVFWYRKSALQGD	SEQ	ID	N O :	10

TABLE 2

	Predicted 318 SLR domain:	S			
P7	EAQYNMGYHYAEGKGVPRDQGKAVFWYEKAAAAGD	SEQ	ID	NO :	11
P8	DAQYMLGAMSVEGIGLPKDSQVALTWLSKAAAQGD	SEQ	ID	NO :	12
P9	AKAQYGLGILYAKGQGVAPDQEKALILYRMAATQG	SEQ	ID	NO:	13
P10	ATAEYAVGLAYAYGRGTAQNDVKAADWFEAAAQQG	SEQ	ID	NO:	14
P11	EAQRRWALMLASGRGVAKNEGEALKWFKKAAVAGD	SEQ	ID	NO:	15

50

Sel1 proteins 310, 301 (from the human strain OXCC13), 317, 319, 321, 304, and 322 also contain predicted SLR domains (Tables 3-9).

	TABLE 3			
	Predicted 310 SLR domains			
P12	ALAQSNLGVLYASGRGVESSPKRALEWYKKAAVQGN	SEQ I	ID NO:	16
P13	SQAQFSLGNMYEDGSGVEKNLAVAAAWYQKSAEQGN	SEQ I	ID NO:	17
P14	AEAQTNLGVLYSYGLGVDKDLSKAFYWY	SEQ I	ID NO:	18
P15	ESQDRLGLMLTNGVGVKQDYKQAYSWFRKAARQG	SEQ I	ID NO:	19
P16	AESQNNLGVLYARGLGVEKDYKQAVAWYRKA	SEQ I	ID NO:	20
P17	QAQFNLGTMYLQGHGVKQDVKQARHWFTKAAAQ	SEQ I	ID NO:	21

	TABLE 4				
	Predicted 301 (human strain OXCC13)	SLR	domains		
P1H	AQAQHNLGVTYYEGEGIKKDYAKAVYWWKKAAEQG	SEQ	ID NO: 22		
P2H	PQSQYNLGIAYEEGWGAEKNPENAVFWYRKAAEQGH	SEQ	ID NO: 23		
РЗН	EAQAYIGMIYFKGKYVAKNEKKGFYWLKKAAEKDS	SEQ	ID NO: 24		

TABLE 5			
Predicted 317 SLR domains			
ALYGLGVMATNGLGMPRNDEKALVWFREGAAKG SEQ ID NO: 25			
EAQFGLGAMYDLSRGVRQDMTLAIDWYEKSARAG SEQ ID NO: 26	35		

TABLE	6
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Predicted 319 SLR domains	3				40
QLYLGLMYGHGKGVPRDLNKSLFWVEKAADRG	SEQ	ID	NO :	27	
AQYLMGMAYLEGKSVPQDLPVAAAWFYKAAMQGN	SEQ	ID	NO :	28	
ADAQLRLGYMYARGIGVPVDKPKAVAWLEKAASAGN	SEQ	ID	NO:	29	45

TABLE 7					
Predicted 321 SLR domai	ns				
GSMLSQGKGVEKDPKKGLEWFVQAGQDGD	SEQ	ID	NO :	30	
SEAQQMMGFLYGEGWGAKRDPVKAEYWFDKAAASGD	SEQ	ID	NO :	31	

TABLE 8	
Predicted 304 SLR domai	ins
QAEHEMGSLYLMGIGVAQSNVMAVAWYRKAAIQG	SEQ ID NO: 32
APSQTAMGYAYEEGAGV PQDADLARYWFDKAAAQGN	SEQ ID NO: 33
TABLE 9	
Predicted 322 SLR domai	ins
AQAGLGWMYAAGRGVNKDETLSFSWYERAAVAG	SEQ ID NO: 34
AQYMLGRYYEKGIGVAKDRVLAKEWYEKAAAQGN	SEQ ID NO: 35

Also identified herein are tetratricopeptide repeat (TPR) sequences. A TPR is a structural motif comprising a degenerate 34 amino acid sequence. The TPR sequences identified herein (Table 10) and variants and mimetics thereof find use in the embodiments described herein (e.g., in place of SLR peptides, with SLR peptides, etc.).

TABLE	10
	T O

	Predicted TPR sequences				
TRP1	KAAKSGNAEAQYLFGMLVYDGRGVQQDNCVAMLW WMKAAEQNHAKALVMLGNLHRKGQCIAENYPKAIA YWKRAAVQNNV	SEQ	ID	NO :	36
TRP2	LGTAYYDGIGVDKNPHEAVRWWKKAAELGFPESQN NLGALYNDGNGVDRDYQEAVFWYRKSALQEDELG QYNLGVAYYGRGIKKDFSEAVSWYKKSAE	SEQ	ID	NO:	37

30

60

	Predicted TPR sequences				
TRP3	YRKAAEQGHADAQNRLGIAYRYGTGVRKNPALSVK WLEKAAKQGLARAQFNLGKTFYIGAGINKNTDKAV	SEQ	ID	NO:	38
	YWFIKAANOGFTEAOAY				

Experiments were conducted during development of 10embodiments herein to determine whether individual Sel1 301 SLR peptides and/or combinations of multiple Sel1 301 SLR peptides are capable of stimulating oxalate influx into human Caco2-BBE (C2) cells. Significant stimulation of ¹⁴C-oxalate influx into human C2 cells was observed (FIG. 1).

Sel1 318 is the only Sel1 protein identified by mass spectroscopic analysis of the human Oxalobacter formigenes (Of) strain OXCC13 genome using a stringent search (Maxquant: 20 ppm, with additional filtering at 1% FDR); 20 several Sel1 proteins are identified by a less stringent search. Sel1 318 is separated by 3 genes from 5 other Sel1 genes located together in a potential operon (304/319-322); Sel1 318 is also a few genes away from Sell 317 and Sell 310, which are located near each other. Collectively, these obser- 25 vations indicate that Sell 318 a key Sell protein for Of. Several Sel1 318-derived peptides individually stimulate 14C-oxalate influx into Caco2-BBE cells (FIG. 2).

301 (54 kDa) and 318 (52 kDa) Sel1 proteins have several SLR motifs (30-35 amino acids [aa] in length; called here peptides 1-6 [P1-6 for 301] and 7-11 [P7-11 for 318]; FIG. 8); it is contemplated that one or more of these domains mediate the observed stimulation, given their importance in protein-protein interactions and signal transduction pathways. To test this hypothesis, P1-6 and P7-11 peptides from 35 the human Of strain OXCC13 were chemically synthetized by Genscript (NJ, USA). P4 (1.5-fold) and P4-6 (1.8-fold) significantly stimulated oxalate transport by C2 cells (FIG. 1), while P1-3, 5 & 6 had no effects. Individually, P7-10 significantly stimulated (~1.5-fold) oxalate transport by C2 40 cells (FIG. 2). The combination of P8+9 stimulated oxalate transport >2.4-fold by C2 cells, similar to the CM (FIG. 3), in a dose-dependent manner (FIG. 4) and more than any other combination. Of interest is that 318 (P7-11) is the only Sel1 protein identified when the Mass-Spec data was ana- 45 lyzed against OXCC13 genome when a highly stringent search was used. 318 gene is near 7 other Sel1 genes (319-323, located together in a potential operon, and 310/ 317), collectively making 318 a key Sel1 protein for Of. P8+9-induced stimulation is better than the 318-induced 50 stimulation potentially due to the batch to batch variability observed with recombinant 318 generated in E. coli. In addition to SLR motifs, 318 is also predicted to have 2 TPR (tetratricopeptide repeat; 34 aa) motifs, and studies are in progress to evaluate their effects on oxalate transport given 55 their important roles in protein-protein interactions and signaling pathways. Having identified these small peptides, the potential for therapeutic development increases since from a drug development prospective it is highly desirable to work with small peptides rather than large proteins.

Two of the most potent stimulators of oxalate influx were tested together, and it was demonstrated that P8+9 peptides significantly stimulate (>2.4-fold) oxalate transport by C2 cells to a level about similar to the Oxalobacter-derived culture conditioned medium (CM: >2.6-fold) (FIG. 3). The 65 P8+9 peptides significantly stimulate oxalate transport by C2 cells in a dose-dependent manner, indicating a receptor-

ligand-type interaction (FIG. 4). Several other P7-P11 peptide combinations (including all of them together) stimulate oxalate transport by C2 cells to a level less than observed with P8+9 in preliminary studies.

Although the transformed C2 cells resemble the native epithelium, they don't fully recapitulate the in vivo physiological responses. Primary stem-cell derived intestinal organoids (Orgs) were recently established as an ex vivo model of the intestinal epithelium, and they mimic the full spectrum of physiological responses in vivo. Therefore, experiments were conducted during development of embodiments herein to confirm the C2 findings in colonic organoids. P7-10 peptides significantly stimulate (1.8-fold) oxalate transport by human sigmoid colon organoid and human ileum organoid (FIG. 6); thereby confirming the results determined in C2 cells. Additionally, experiments conducted during development of embodiments herein demonstrate that P8+9 peptides significantly stimulated oxalate transport by an ileum (FIG. 5A; ~2-fold), sigmoid colon (FIG. 5B; ~1.8-fold), and distal colon (~1.7-fold; not shown) organoids.

The intestine plays a major role in oxalate homeostasis, responsible for dietary oxalate absorption and gastrointestinal oxalate secretion/excretion. Animals and humans studies show that Of metabolizes dietary oxalate leading to reduced intestinal absorption and urinary oxalate excretion (Refs. 8, 30; incorporated by reference in their entireties). Of is an anaerobic colonic bacterium which utilizes oxalate as its exclusive energy source (Ref. 17; incorporated by reference in its entirety). In addition to degrading dietary oxalate, Of, for its own potential survival benefit, also interacts with the colonic epithelium to induce colonic oxalate secretion via a secretagogue, leading to reduced urinary oxalate excretion (Ref. 15; incorporated by reference in its entirety). Of colonization of a mouse model for PH type I was associated with 50% reduction in serum and urinary oxalate levels due to induction of colonic oxalate secretion (Ref. 18; incorporated by reference in its entirety). However, PH1 mice lost colonization within 18 days when switched from a high oxalate/low calcium diet (1.5% oxalate/0.5% calcium) to regular mouse chow (0.25% oxalate/1% calcium) (Ref. 18; incorporated by reference in its entirety). In addition, sustaining Of colonization in rats required a 3% dietary oxalate and colonized rats lost Of colonization within 5 days of dietary oxalate removal. Of colonized rats on high oxalate diets demonstrated greater urinary oxalate levels than that of non-colonized rats on a low oxalate diet and colonization was not maintained without reducing dietary calcium intake. Furthermore, studies in PH patients & PH1 mice raise the possibility that the intraluminal environment in PH is not supportive of sustained Of colonization (Refs. 18-19; incorporated by reference in their entireties). These limitations have resulted in the failure of several phase I-III trials using Of to lower urine or plasma oxalate levels in PH patients. Therefore, an appropriate strategy is to use Of-derived factors that can be directly administered to stimulate intestinal oxalate secretion to lower urine and plasma oxalate levels.

Toward this end, Of culture conditioned medium (CM) was prepared as recently reported (Ref. 4; herein incorporated by reference in its entirety) and human intestinal Caco2-BBE (C2) cells were used to evaluate the effects of Of CM on intestinal oxalate transport. Apical oxalate uptake by C2 cells was measured by imposing an outward Cl gradient by removing extracellular Cl [Cli>Clo] (Refs. 3, 4, 14; incorporated by reference in their entireties) and measuring DIDS (anion exchange inhibitor)-sensitive influx of 14C-oxalate in exchange for intracellular Cl [i.e. apical 10 Cl-oxalate exchange activity, ≥49% of which is mediated by the oxalate transporter SLC26A6 (A6) in C2 cells (4, 12))]. A6 is the critical transport mechanism for exchanging intracellular oxalate for mucosal CI during the process of transepithelial intestinal oxalate secretion, but its activity was 15 measured by the more convenient assay of cellular oxalate uptake since it can transport oxalate in either direction (Ref. 22; incorporated by reference in its entirety) Of note is that A6 null mice have a critical defect in intestinal oxalate secretion resulting in enhanced net absorption of dietary 20 oxalate, hyperoxalemia, hyperoxaluria, and a high incidence of COKS (Refs. 13, 21; incorporated by reference in their entireties).

Of-derived factors, such as the peptides described herein, stimulate oxalate transport in vitro: Preincubation of C2 25 cells with Of CM (1:50 dilution×24 h) stimulated oxalate transport >2.8-fold compared to untreated cells (UT) or cells treated with Of growth medium without bacteria included (OM) (UT=3.1±0.2; OM=2.9±0.3; CM=9.1±0.9 pmoles/ cm2/min) (Ref. 4: herein incorporated by reference in its 30 entirety). CM from Lactobacillus acidophilus did not impact oxalate transport, indicating specificity (Ref. 4: herein incorporated by reference in its entirety. Pretreatment of the CM with heat, pepsin, or trypsin completely abolished its stimulatory effects, indicating that the secreted factor(s) is/are 35 likely to be proteins and/or peptides (Ref. 4: herein incorporated by reference in its entirety). Selective ultrafiltration revealed that the secreted factors have molecular weights mostly between 10-30 kDa (Ref. 4: herein incorporated by reference in its entirety). Pretreatment with the PKA inhibi- 40 tor H89 completely blocked CM-induced oxalate transport, indicating that it is mediated by PKA activation (4). CMinduced stimulation is DIDS-sensitive and knockdown studies showed that stimulation is largely mediated by A6 (Ref. 4: herein incorporated by reference in its entirety), as well as 45 A2. Lowering oxalate concentration in the culture medium from 5 g/L to 1 g/L led to a CM with >2-fold higher stimulatory effect (Ref. 4: herein incorporated by reference in its entirety), while increasing oxalate to 25 g/L led to a CM with 50% reduced activity, indicating that the secretion 50 of these factors is inducible. CM-induced transport is due to >2-fold increase in Vmax (i.e. enhanced transport capacity) and >3.4-fold reduction in Km [i.e. increased affinity for oxalate] of the involved transporter(s) (Ref. 4: herein incorporated by reference in its entirety). CM did not affect A6 55 surface protein expression (Ref. 4: herein incorporated by reference in its entirety), and given the reduced Km (greater A6 affinity for oxalate), the observed stimulation is due to CM-induced increase in the intrinsic activity of the preexisting A6 membrane transporters. 60

To recapitulate the findings in vivo, primary stem-cell derived intestinal organoids (Org) were used which have been established as an ex vivo model of the intestinal epithelium with a full spectrum of physiological responses found in vivo (Ref. 35; incorporated by reference in its 65 entirety). A human sigmoid colonic Org (from a healthy individual) was grown and maintained as previously

reported (Ref. 34; incorporated by reference in its entirety). Org cells were plated onto collagen-coated transwell filters and formed monolayers. Monolayer confluency and differentiation was confirmed by transepithelial electrical resistance measurements (Ref. 34; incorporated by reference in its entirety). Org cells demonstrated transport of oxalate measured as influx of ¹⁴C-oxalate in exchange for intracellular Cl. The influx of ¹⁴C-oxalate is DIDS-sensitive (inhibited by \sim 70% with 100 μ M DIDS), indicating that it is anion exchange mediated oxalate transport. An ileum Org was developed which demonstrates DIDS-sensitive (inhibited by ~80% with 500 µM DIDS) oxalate transport. Importantly, P8+9 (100 µg/ml×24 h) significantly stimulated oxalate transport by the ileum Org (2-fold; FIG. 5A) and the sigmoid Org (1.8-fold; FIG. 5B). These results show that P8+9 stimulates oxalate transport by human Org as observed in C2 cells, suggesting that these peptides are very likely to work in humans in vivo.

Peptides Modifications to Enhance their In Vivo Stabilities

Given P4-10 peptides susceptibilities to proteolytic degradation, the lead peptides are optimized to improve peptide druggability through structural modification approaches that reduce proteolysis and thus enhance stability. Proteolytic enzymes like aminopeptidases and carboxypeptidases break down peptide sequences from the N- and C-termini, and therefore N-(acetylation) and/or C-(amidation) terminal modifications can improve peptide stability (Ref. 36; herein incorporated by reference in its entirety). Of note is that the N-terminal acetylated forms of GLP1 7-34 and somatostatin analogs were shown to be much more stable than the native peptides (Ref. 36; herein incorporated by reference in its entirety). In addition, substitution of natural L-amino acids with non-natural D-amino is known to reduce the substrate recognition and binding affinity of proteolytic enzymes and enhances peptide stability (Ref. 36; herein incorporated by reference in its entirety). An example is the drug-octreotide, in which replacing L-amino acids with D-amino acids prolonged the human in vivo half-life from a few minutes to 1.5 h (Ref. 36; herein incorporated by reference in its entirety). Moreover, cyclization (head-to-tail, head/tail-toside-chain, and/or side-chain-to-side-chain) enhances peptide stability and permeability by introducing conformation constraint, thereby reducing peptide flexibility, and a cyclic enkephalin analog is highly resistant to enzymatic degradation (Ref. 36; herein incorporated by reference in its entirety). Furthermore, modification of natural amino acids improves peptide stability by introducing steric hindrance or disrupting enzyme recognition. An example is buserelin, in which one Gly is replaced with a t-butyl-D-Ser and another Gly is replaced with ethylamide, leading to prolonged halflife in humans compared to a very short half-life (minutes) in the unmodified GRH (Ref. 36; herein incorporated by reference in its entirety). P4-11 peptides are subjected to all of the above described modifications. The Gly residues in each peptide are substituted with PEG6, which is much better than PEG4 for peptide stabilization. In addition, both N-acetylation and C-amidation in the same peptide as well as applying N-acetylation or C-amidation in conjunction with amino acid substitution are made. Moreover, peptides that are cationic show fast degradation due to their arginine (R) and lysine (K) contents and therefore, the R residues in the peptides of this invention are replaced with the arginine homologue Agp to enhance their stabilities (Ref. 36: herein incorporated by reference in its entirety). Replacing all R

with Agp in peptide Sub3 led to a dramatic increase in its stability (Ref. 37; herein incorporated by reference in its entirety).

Identification of P8 and P9 Functional Subdomains

The 318 homology-based structure has a long helical 5 structure composed of many alpha helix hairpins. 318 is predicted to have 5 SLR motifs (P7-11; FIG. 8), and each SLR motif is composed of 2 alpha helices (underlined aa for P8 and P9; FIG. 7), separated by a loop (bolded aa; FIG. 7). G14 & 16 in the loop are conserved in all P1-11 peptides and 10 they are likely important for the structure. Y96, A122, Q125, and G126 in P8 and Y243, A269, O272, and G273 in P9 are close to each other and potentially form the binding sites. Similar sequences are also found in P4 (from 301), P7, and P10. Of remarkable interest is that only P4 and P7-10 15 individually stimulated oxalate transport (~1.5-fold) by C2 cells. P1-3, 5, 6, and 11 don't have similar sequences, especially Y, and they don't stimulate oxalate transport by C2 cells. Conserved P8 & P9 aa are the same in the two sequences (FIG. 7). Detailed analysis of the P8 and 9 20 structures indicates that the only regions that can be potentially deleted without affecting their functions will be the 4 aa (boxed) from each helix. Deleting >4 aa would likely disrupt the helix function. Therefore, AMSV (SEQ ID NO: 48)/QVAL (SEQ ID NO: 49) and ILYA (SEQ ID NO: 25 50)/QEKA (SEQ ID NO: 51) will be deleted from P8 and P9, respectively, to identify the smallest P8 and P9 subdomains that stimulate oxalate transport similar to the full-length peptides. The truncated peptides (truncated P8: RDAQYMLGEGIGLPKDSTWLSKAAAQGD; truncated P9: AKAQYGLGKGQGVAPDLILYRMAATQG) will be chemically synthesized by Genscript, 1-4 mg which is sufficient for the studies, and >95% purity. If the truncated P8 and truncated P9 similarly stimulate oxalate transport by C2 cells as the full-length peptides, this will confirm suc- 35 cessful shortening of each peptide by 8 aa. Additional modifications include deleting the first and last aa in each loop (E/S in P8 and K/D in P9) and the effects of the truncated P8 and truncated 9 will be similarly assessed. If the truncated P8 and truncated P9 similarly stimulate oxalate 40 transport by C2 cells as the full-length peptides, this will confirm successful shortening of each peptide by 10 aa. Additional modifications include deleting the first and last aa in each loop (E/S in P8 and K/D in P9) and the effects of the truncated P8 and 9 will be similarly assessed. If the trun- 45 cated P8 and P9 similarly stimulate oxalate transport by C2 cells as the full-length peptides, this will confirm successful shortening of each peptide by 10 aa.

The SLR consensus sequence highlights conserved alanine (A) and glycine (G) residues at positions 3, 8, 14, 24, 50 32, 39, 40 and 43. The G residues at positions 14, 24, and 43 facilitate sharp turns in the loop regions and small residues at positions 3, 8, 32, 39, and 40 allow tight packing of α -helices. Both P8 and P9 have A at positions 3, 24, & 32 (A94/241, A115/262, and A123/270, respectively) and G at 55 positions 8 & 14 (G99/246 and G105/252). In addition, Y96, A122, Q125, and G126 in P8 and Y243, A269, Q272, and G273 in P9 form the potential binding sites. It is contemplated that one or more of these conserved aa (individually or in combination), especially those form the potential 60 binding site, is/are crucial for the observed stimulation by facilitating the binding to a receptor and/or other interacting proteins. To define the critical aa in the identified subdomains, the above described conserved aa is substituted as follows. Y96/243 will be substituted with phenylalanine (F), 65 while the specified A, Q, & G will be substituted with tyrosine (Y). The individual substitutions are Y96/243,

A122/269, Q125/272, G126/G273, A94/241, A115/262, A123/270G8, G99/246, and G105/252 or combinations thereof. Further deletion combinations include 3 aa (AMS/QVA or MSV/VAL and ILY/QEK or LYA/EKA), 2 aa (AM/QV, MS/VA, or SV/AL and IL/QE, LY/EK, or YA/KA), or single aa deletions (e.g. A/Q or V/L). Complete abolishment or reduction of subdomain-mediated stimulation of oxalate transport by C2 cells denote critical residues needed to activity.

P8+9 are designed to have N-terminal or C-terminal biotin tags used to pull down complexes. Biotinylated P4-6 peptides with N-terminal biotin tags stimulated oxalate transport by C2 cells similar to the untagged P4-6 peptides, indicating that the biotin tags are unlikely to affect P8+9 activities.

In vivo efficacy testing of peptides.

Having effectively optimized the characterized P8+9 subdomains, the optimized subdomains are tested to determine whether the peptides retain their biological activities in vivo and significantly reduce urine and plasma oxalate levels in hyperoxaluric and hyperoxalemic mice. The CM significantly reduced (>32.5%) urinary oxalate excretion in PH1 mice (Ref. 4; herein incorporated by reference in its entirety), and therefore these mice are used to evaluate the therapeutic potential of the optimized peptides and peptides subdomains. PH1 mice using one sex (males; 3 doses; n-5/dose; Control mice receive a vehicle) receive optimized subdomains SD8 and SD9 in pilot dose ranging (3 different doses for each of SD8, SD9, and SD8+9 for up to 4 weeks). The doses found to work well (leading to the highest reduction in urine and plasma oxalate levels) are further tested in both sexes. To this end, PH1 mice (12-20 weeks old; n=14/sex/treatment group) are placed individually in metabolic cages and urine & feces are collected on days 0, 7, 14, 21, & 28. Blood samples are collected on days 0, 14, and 28. Following baseline urine, blood, and fecal samples collection, the mice receive SD8+9 (if found to give the highest reduction in urine and plasma oxalate levels in the pilot studies described above) rectally as enemas (100 µl twice daily×28 days) using the optimal doses developed in the pilot studies. Control mice receive vehicle (the buffer in which the subdomains are dissolved). The urine and blood samples are acidified with 4N HCL and stored at -80° C. Urine, plasma, and fecal oxalate levels are measured enzymatically using oxalate oxidase, employing sodium nitrite (Ref. 32; herein incorporated by reference in its entirety) to prevent interference by ascorbic acid.

Observing normalization or significantly reduced urine and plasma oxalate levels in SD8+9-treated PH1 mice compared with vehicle-treated ones reflect in vivo retention of SD8+9 biological activities and the significant therapeutic potential of these peptides. As a result of enhanced colonic oxalate secretion, fecal oxalate levels are higher in the SD8+9-treated mice. In addition to reducing urinary oxalate excretion in PH and without being bound by theory, SD8+9 effectively lowers urinary oxalate excretion in EH, which is seen in association with many conditions, including IBD, obesity, and post-bariatric surgery. This is very important since showing that SD8+9 can significantly reduce urine oxalate levels in both PH and EH prove that SD8+9 are effective in all forms of hyperoxalurias and thus have a wider clinical applicability. To this end, SD8+9 is administered rectally to SAPM1/YitFc (SAM) and ob/ob (ob) mice (8-12 weeks old; n=14/sex/treatment group). SAM is a mouse model with remarkable similarities to human Crohn's disease that the inventors developed as a good model for the IBD-associated hyperoxaluria. SAM mice have 2-fold

hyperoxaluria compared to controls, as well as significantly reduced A6 ileal mRNA (>87%) and total protein (>60%) levels. ob is an obesity model developed as a good model for the obesity-associated hyperoxaluria (>3.2-fold hyperoxaluria compared to controls) (Ref. 32; incorporated by 5 reference in its entirety). Of note is that the doses found to work in PH1 mice are expected to work in SAM mice (similar body weights), but unlikely to work in the obese ob mice (having at least twice the body weights of PH1 mice). Therefore, a dose ranging study will be done in ob mice. 10 SAM and ob mice are similarly treated with SD8+9 and the data are analyzed as described above with the PH1 mice.

Having shown that SD8+9 significantly reduce plasma and urine oxalate levels in PH1 as well as urine oxalate levels in the ob and SAM mice, experiments are conducted 15 to demonstrate that the observed reduction in urine and plasma oxalate levels is due to SD8+9-induced enhanced colonic oxalate secretion. Toward this end, cecal, proximal, and distal colonic tissues from SD8+9- and vehicle-treated PH1, ob, and SAM mice are isolated and mounted in Ussing 20 chambers at the end of treatment. Two unidirectional [mucosa to serosa=JMS (absorptive flux), and serosa to mucosa=JSM (secretory flux)] are assessed and the magnitude and direction of the net flux (Jnet) across conductancematched tissues are determined by calculating the difference 2 between the two measured unidirectional fluxes as previously reported (Ref. 4; herein incorporated by reference in its entirety). Compared to vehicle-treated mice, if net basal cecal oxalate flux is converted from absorption to secretion, or significantly higher net basal secretory flux is observed in

the proximal and/or the distal colon of SD8+9-treated mice [due to significantly increased secretory flux and/or reduced absorptive flux(es)], then such findings strongly support the idea that SD8+9 reduce serum and urine oxalate levels in PH1 mice and urine oxalate levels in ob and SAM mice by enhancing colonic oxalate secretion. Collectively, such findings provide a molecular basis for therapeutic application of SD8+9 for prevention and/or treatment of hyperoxaluria, hyperoxalemia, and related COKS.

Experiments were conducted during development of embodiments herein to determine the effects of various peptide modifications (Table 11) including N-terminal acetylation, C-terminal amidation, replacing glycines with PEG, and replacing L-amino acids with D-amino acids on stimulation of oxalate transport. Results are depicted in FIG. **9-13**.

The results in FIGS. **9-13** show that P8 and P9 peptides with N-terminal acetylation and C-terminal amidation significantly stimulated oxalate transport by C2 cells to a level similar to native P8 and P9 peptides. In addition, P8 and P9 peptides with all Glycines replaced with PEG6 also similarly stimulated oxalate transport by C2 cells compared with native P8 and P9 peptides. Moreover, replacing G15 in P8 and G16 in P9 with D-alanine also similarly stimulated oxalate transport by C2 cells compared with native P8 and P9 peptides. Collectively, these results indicate that all of the above described structural modifications did not impact the functional activities of P8 and P9 peptides. P8 and P9 peptides having all natural L-amino acids replaced by D-amino acids were nonfunctional.

TABLE 11

	Modified 318 SLR domain peptides		
P8-Ac	Ac-DAQYMLGAMSVEGIGLPKDSQVALTWLSKAAAQGD	SEQ	ID
		NO :	39
P8-Daa	DAQYMLGAMSVEGI (d-alanine)LPKDSQVALTWLSKAAAQGD	SEQ	ID
		NO :	40
P8-Am	DAQYMLGAMSVEGIGLPKDSQVALTWLSKAAAQGD-Am	SEQ	ID
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P8-Peg	DAQYMLA (Peg) MSVE (Peg) I (Peg) LPKDSQVALTWLSKAAAQ	SEQ	ID
	(Peg)D	NO :	42
P9-Ac	Ac-AKAQYGLGILYAKGQGVAPDQEKALILYRMAATQG	SEQ	ID
		NO :	43
P9-Daa	AKAQYGLGILYAKGQ(d-alanine)VAPDQEKALILYRMAATQG	SEQ	ID
		NO:	44
P9-Am	AKAOYGLGILYAKGOGVAPDOEKALILYRMAATOG-Am	SEO	ID
		NO :	45
P9-Peq	AKAQY (Peq) L (Peq) ILYAK (Peq) Q (Peq) VAPDQEKALI LYRMAA	SEO	ID
5	TQ(Peg)	NO :	

Oxalate Transport Enhanced by Addition of Select Trace Elements

The inventors sought to identify trace elements to better stimulate oxalate transport by facilitating the binding of P8 5 and/or P9 to a cell surface receptor. The inventors analyzed combinations of P8 and P9 alone or with aluminum, zinc, or magnesium by measuring C14 oxalate transport in C2 cells. Peptides P8 and P9 were premixed with magnesium citrate prior to administration to human Caco2-BBE cells. The addition of magnesium citrate induced a 1.9-fold increase in oxalate transport (FIGS. 14 and 15) compared to peptide alone. Zinc citrate was observed to have similar stimulatory effects but not from zinc chloride or zinc sulfate. The addition of aluminum to P8 or P9 did not result in increased oxalate transport and oxalate transport induction was similar to untreated (FIG. 16).

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The invention claimed is:

1. A method of stimulating oxalate transport comprising administering to a subject a pharmaceutical composition ₃₀ comprising a suppressor-enhancer of lin 1 (Sel1)-derived peptide, wherein the Sel1-derived peptide comprises (i) the amino acid sequence of SEQ ID NO: 52 or 53 or (ii) the amino acid sequence that is at least 90% identical to SEQ ID NO: 12 or 13, and wherein the Sel1-derived peptide is no 35 more than about 50 amino acids in length.

2. The method of claim **1**, wherein the administration is rectal administration, oral administration. or injection.

3. The method of claim **1**, wherein the subject suffers from or is at risk for hyperoxaluria and/or hyperoxalemia.

4. The method of claim 1, wherein the subject's risk of calcium oxalate kidney stones, nephrocalcinosis, oxalate nephropathy, end stage renal disease, chronic kidney disease, and/or systemic oxalosis is lowered by the administration.

5. The method of claim **1**, wherein the Sel1-derived peptide comprises the amino acid sequence that is at least 90% identical to SEQ ID NO: 12 or 13.

6. The method of claim **1**, wherein the Sel1-derived peptide comprises the amino acid sequence of SEQ ID NO: 50 52 or 53.

7. The method of claim 1, wherein the composition comprises 2 or more of the Sell-derived peptides.

8. The method of claim **7**, wherein the two or more Sell-derived peptides are fused directly or indirectly to each 55 other.

9. The method of claim 7, wherein the two or more Sell-derived peptides are not fused to each other.

10. The method of claim 1, wherein:

- (i) the Sel1-derived peptide comprises one or more D enantiomers,
- (ii) the Sel1-derived peptide comprises one or more unnatural amino acids,
- (iii) the Sel1-derived peptide comprises one or more amino acid analogs, and/or
- (iv) the Sel1-derived peptide comprises one or more peptoid amino acids.

11. The method of claim 1, wherein the Sel1-derived peptide comprises a modification selected from the group consisting of phosphorylation, glycosylation, ubiquitination, S-nitrosylation, methylation, N-acetylation, C-terminal amidation, cyclization, substitution of natural L-amino acids with non-natural D-amino acids, lipidation, lipoylation, deimination, climinylation, disulfide bridging, isoaspartate formation, racemization, glycation, carbamylation, carbonylation, S-sulfonylation, S-sulfinylation, S-sulfenylation, S-glutathionylation, pyroglutamate formation, propio-nylation, adenylylation, nucleotide addition, iodination, hydroxylation, biotinylation, carbamylation, axidation, and pegylation.

12. The method of claim 1, wherein the pharmaceutical composition further comprises a pharmaceutically acceptable carrier.

13. The method of claim 12, wherein the pharmaceutical composition further comprises one or more additional therapeutic agents.

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