# PNAS www.pnas.org 

## Supplementary Information for

DNA methylation signatures in airway cells from adult children of asthmatic mothers reflect subtypes of severe asthma

Kevin M. Magnaye, Selene M. Clay, Jessie Nicodemus-Johnson, Katherine A. Naughton, Janel Huffman, Matthew C. Altman, Daniel J. Jackson, James E. Gern, Douglas K. Hogarth, Edward T. Naureckas, Steven R. White, Carole Ober.

Corresponding authors: Carole Ober and Kevin Magnaye
Email: c-ober@genetics.uchicago.edu and kevin.magnaye@ucsf.edu

## This PDF file includes:

Supplementary Methods
Figures S1 to S11
Tables S1 to S8
Supplementary References

## Supplementary Methods

## Inclusion and Exclusion Criteria

Subjects with asthma met the following criteria: (1) a physician's diagnosis of asthma; (2) either (a) a fall in baseline $F E V_{1} \geq 20 \%$ at $\leq 25 \mathrm{mg} / \mathrm{ml}$ methacholine in subjects whose $\mathrm{FEV}_{1}$ predicted was $\geq 70 \%$ or (b) a $\geq 15 \%$ increase in baseline $\mathrm{FEV}_{1}$ after inhalation of a bronchodilator (albuterol) or over time with treatment in subjects whose $\mathrm{FEV}_{1}$ predicted was $<70 \%$; (3) at least two symptoms (cough, wheeze, and dyspnea); (4) fewer than three pack years of cigarette exposure; and (5) no conflicting pulmonary diagnoses. Controls had no current or previous diagnosis of asthma, normal spirometry, and no evidence of bronchial hyperresponsiveness by methacholine challenge tests. For both cases and controls, exclusion criteria included: (1) a smoking history of $\geq 10$ pack years; (2) born premature ( $\geq 37$ weeks gestation); (3) a history of conflicting pulmonary diagnosis, such as chronic obstructive pulmonary disease, allergic bronchopulmonary aspergillosis, cystic fibrosis, or Churg-Strauss syndrome; or (4) any medical contraindication to bronchoscopy.

## Genotyping and QC

SNPs were genotyped using the Illumina Omni2.5-8v1A, Omni1MDuo, or HumanCore+Exome arrays. SNPs on each array were excluded with HWE P<0.0001 within each ethnic group (European American, African American), MAF < 0.05, SNP call rate < 0.95, and subject call rates $<0.95$. Ancestry principal component analysis (PCA) was performed in R (prcomp function) using the 676 ancestry informative markers (1) that were available in our sample and overlapped with three representative populations from the HapMap release 3 (YRI, Yoruba in Ibadan, Nigeria; CEU, Utah residents with Northern and Western European ancestry; CHB, Han Chinese in Beijing, China).

## RNA extraction, sequencing, and QC

RNAs were extracted from bronchial epithelial epithelial cells and purified using the QIAGEN AllPrep DNA/RNA/miRNA Universal Kit (Hilden, Germany), following manufacterer's instructions. Quality and quantity assessment of RNA were measured at the University of Chicago Functional Genomics Core using an Agilent RNA 6000 Pico assay and the Agilent 2100 Bioanalyzer. cDNA libraries were constructed using the Illumina TruSeq RNA Library Prep Kit v2. RNA sequencing was performed at the University of Chicago Genomics Core on either the Illumina HiSeq 2500 or 4000 platforms.

For the RNA-seq data, potential sample contamination and sample swaps were assessed using VerifyBamID (2). No cross-contamination was detected. Two sample swaps between individuals were identified and corrected. Quality control checks were performed using FastQC (3). RNA sequences were then aligned and annotated to known RNA sequences using Spliced Transcripts Alignment to a Reference using STAR (4). Genes with low count data (<1 CPM in at least $25 \%$ of the sample) and those on the $\mathrm{X}, \mathrm{Y}$, and mitochondrial chromosomes were removed. Samples containing >8M mapped RNA reads were kept, retaining 124 subjects for downstream analyses.

Raw counts were normalized using the trimmed mean of M-values (TMM) method (5). Mean-variance trend was adjusted using variance modeling in voom (6)). Technical sources of variation were identified using PCA for the RNA expression profiles. All potential confounders are described in the Methods section of the main paper.

## Sample Composition and 'Omics

- 88 adults with asthma
- 56 without an asthmatic mother
(NMA)
- 32 with an asthmatic mother (MA) - 42 adult controls without asthma
- Collect cryopreserved epithelial cells from bronchial brushings
- Thaw and purify DNA and RNA
- Measure global DNA methylation and gene expression

Analytical Pipeline


Figure S1. Overview of study design.


Figure S2. Flow chart of sample selection. Analyses included the 130 subjects shown in the shaded box, as described in Table 1.


Figure S3. Scatterplot of beta coefficients for the 2,326 CpGs that were differentially methylated in all asthma cases compared to controls (shared DMCs). The beta coefficients are plotted in the NMA cases (x-axis) and MA cases (y-axis). The 2,147 (92.30\%) DMCs with larger effects in MA cases compared to NMA cases are shown as blue points; the 179 DMCs ( $33.15 \%$ ) with larger effects in NMA cases are shown as orange points. Paired signed-rank test; $\mathrm{P}<2.2 \times 10^{-16}$.

## Cluster Dendrogram



Figure S4. Cluster dendrogram of 69 non-maternal asthma-associated differentially methylated CpGs (NMA-DMCs). Of the 69 NMA-DMCs, 66 ( $95.65 \%$ ) formed two co-methylation modules (green and orange). After merging closely related modules using the MergeCloseModules function, as implemented in WGCNA (7), the two co-methylation modules remained distinct (31 and 35 CpGs in each module). The average pairwise Spearman correlation for the CpGs within the orange and green modules was $0.28,0.35$, respectively. There were $3(4.35 \%)$ remaining CpGs assigned to the gray module. The average pairwise Spearman correlation of the three remaining CpGs was 0.50 . We only considered modules that contained at least 15 CpGs in our analyses (see Materials and Methods).

## Cluster Dendrogram



Figure S5. Cluster dendrogram of 554 maternal asthma-associated differentially methylated CpGs (MA-DMCs). Of the 554 MA-DMCs, 203 (36.64\%) formed seven co-methylation modules (black, turquoise, green, blue, brown, red, and yellow). After merging closely related modules using the MergeCloseModules function, as implemented in WGCNA (7), five co-methylation modules remained (black, turquoise, blue, red, and yellow; 15 to 74 CpGs in each module). The average pairwise Spearman correlation for the CpGs within the black, turquoise, blue, red, and yellow modules was $0.61,0.57,0.52,0.60$, and 0.56 , respectively. There were 351 ( $63.40 \%$ ) uncorrelated CpGs assigned to the gray module (average pairwise Spearman correlation = 0.04 ). We only considered the five correlated modules in our analysis (see Materials and Methods).


Figure S6. Distribution of NMA-DMCs in each co-methylation module across chromosomes and functional annotations. The $y$-axes show the $-\log _{10}(p$-values) corresponding to methylation differences between NMA cases and non-asthma controls. Blue horizontal lines show the q value threshold (FDR 0.10). Orange (upper) and green (lower) points are the CpGs assigned to the orange or green modules, respectively. The pie charts on the right show the distribution of the module-specific NMA-DMCs by functional annotations.


Figure S7. Distribution of MA-DMCs in each co-methylation module across chromosomes and functional annotations. The y-axes show the - $\log _{10}(\mathrm{p}$-values) corresponding to methylation differences between MA cases and non-asthma controls. Blue horizontal lines show the $q$ value threshold (FDR 0.10). Purple (upper left), blue (upper right), red (middle left), turquoise (middle right) and yellow (lower left) points are the CpGs assigned to the black, blue, red, turquoise, and yellow modules, respectively. The pie charts show the distribution of the module-specific MA-DMCs by functional annotations.


Figure S8. No difference in median and covariate-adjusted gene expression of the 231 genes of the black module and the 675 genes of the yellow module between NMA and MA cases. See list of genes in Table S4.


Figure S9. Selection of hub DMCs for each module (module membership $>0.80$ ). The number of hub DMCs of the total DMCs are shown below each module. For each hub DMC, detailed functional information is in Table S6.


Figure S10. QQ plots before and after controlling for genomic inflation factors (lambda) in each of the three DNA methylation analyses.


Figure S11. Selection of soft thresholding power for the NMA (left) and MA-focused (right) WGCNA analyses.

Table S1. Characteristics of 130 subjects at the time of bronchoscopy by risk group. Severity was defined by STEP classification (8) of mild, moderate and severe asthma. MA, Maternal Asthma; Af Am, African American; Eur Am, European American; ICS, Inhaled Corticosteroid; OCS, Oral Corticosteroid; FVC, Forced Vital Capacity. Pairwise comparisons were made between controls, asthma cases without an asthmatic mother (Case NMA) and asthma cases with an asthmatic mother (Case MA). Continuous variables were tested with a Wilcoxon rank-sum test and categorical variables were tested using a Fisher's Exact Test. ${ }^{\text {A }} 52$ Case NMA subjects and 30 Case MA subjects had FeNO measurements. ${ }^{B} 41$ controls and 55 Case NMA had BAL eosinophil measurements. 41 controls and 55 Case NMA subjects had BAL neutrophil measurements. ${ }^{\circ} 41$ controls had blood eosinophil measurements. Significant pvalues after Bonferroni-correcting for 15 tests (variables; $p<3.33 \times 10^{-3}$ ) are bolded; ns, not significant ( $p>0.05$ ).

|  | Maternal Asthma Risk Groups |  |  | Comparisons |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Control $(\mathrm{N}=42)$ | $\begin{gathered} \text { Case NMA } \\ (\mathrm{N}=56) \\ \hline \end{gathered}$ | $\begin{gathered} \text { Case MA } \\ (\mathrm{N}=32) \end{gathered}$ | Control vs. Case NMA | Control vs. Case MA | Case NMA vs. Case MA |
| Covariates |  |  |  |  |  |  |
| Age (mean yr $\pm$ SD) | $37.45 \pm 11.75$ | $41.41 \pm 12.24$ | $37.56 \pm 13.39$ | ns | ns | ns |
| Gender (\% female) | 62 | 79 | 66 | ns | ns | ns |
| Ethnicity (\%) (Af Am/Eur Am/ Other) | 64/29/7 | 55/45/0 | 66/34/0 | $2.0 \times 10^{-3}$ | 0.02 | ns |
| \% smoker at bronchoscopy | 17 | 4 | 6 | 0.04 | ns | ns |
| Clinical Measures |  |  |  |  |  |  |
| ICS use (\%) | - | 75 | 75 | - | - | ns |
| OCS use (\%) | - | 45 | 25 | - | - | ns |
| STEP Category (\% mild/\% moderate/\% severe) | - | 22/23/55 | 28/25/47 | ${ }^{-}$ | ${ }^{-}$ | ns |
| Mean FEV ${ }_{1}$ \% Predicted ( $\pm$ SD) | $95.36 \pm 11.63$ | $75.45 \pm 19.73$ | $70.41 \pm 18.15$ | $6.6 \times 10^{-8}$ | $9.7 \times 10^{-8}$ | ns |
| Mean FEV $1 / \mathrm{FVC}$ ( $\pm$ SD) | $0.82 \pm 0.05$ | $0.73 \pm 0.10$ | $0.83 \pm 0.48$ | $9.3 \times 10^{-6}$ | $2.7 \times 10^{-5}$ | ns |
| Median total serum IgE (IU/mL) (lower, upper quartile) | $\begin{gathered} 56.50 \\ (22.00,169.00) \end{gathered}$ | $\begin{gathered} 117.50 \\ (22.00,305.25) \end{gathered}$ | $\begin{gathered} 156.50 \\ (70.25,626.75) \end{gathered}$ | ns | $6.3 \times 10^{-3}$ | ns |
| Median FeNO (ppb) (lower, upper quartile) ${ }^{\text {A }}$ | $\begin{gathered} 14.00 \\ (10.50,17.75) \\ \hline \end{gathered}$ | $\begin{gathered} 23.50 \\ (13.00,45.75) \\ \hline \end{gathered}$ | $\begin{gathered} 27.00 \\ (13.25,54.75) \\ \hline \end{gathered}$ | $2.5 \times 10^{-3}$ | $2.9 \times 10^{-3}$ | ns |
| Median BAL eosinophilia (\%) (lower, upper quartile) ${ }^{\mathrm{B}}$ | $\begin{gathered} 0.0 \\ (0.0,0.4) \\ \hline \end{gathered}$ | $\begin{gathered} 2.9 \\ (1.5,5.6) \end{gathered}$ | $\begin{gathered} 3.6 \\ (1.5,7.1) \\ \hline \end{gathered}$ | $7.0 \times 10^{-13}$ | $5.2 \times 10^{-11}$ | ns |
| Median BAL neutrophilia (\%) (lower, upper quartile) ${ }^{\text {C }}$ | $\begin{gathered} 4.9 \\ (3.0,5.9) \\ \hline \end{gathered}$ | $\begin{gathered} 5.2 \\ (4.2,6.8) \\ \hline \end{gathered}$ | $\begin{gathered} 3.5 \\ (2.2,6.4) \\ \hline \end{gathered}$ | ns | ns | 0.04 |
| Median blood eosinophilia (cells/ $\mu \mathrm{l}$ ) (lower, upper quartile) ${ }^{\text {D }}$ | $\begin{gathered} 70 \\ (100,170) \end{gathered}$ | $\begin{gathered} 170 \\ (70,283) \end{gathered}$ | $\begin{gathered} 225 \\ (108,323) \end{gathered}$ | ns | $8.4 \times 10^{-4}$ | ns |
| Mean Body Mass Index ( $\pm$ SD) | $28.52 \pm 5.71$ | $34.64 \pm 8.83$ | $33.68 \pm 10.48$ | $4.5 \times 10^{-4}$ | 0.04 | ns |

Table S2. Genes that were uniquely correlated (FDR < 0.10) with the two NMA-associated and five MA-associated co-methylation module eigenvectors.

| Module (\# of genes) | Unique genes |
| :---: | :---: |
| Orange ( $\mathrm{N}=115$ ) | GRIK2, WNT16, C10orf143, SERPINE2, TSHZ3, RTEL1, MARCHF4, RPS6KA5, NAB2, RGS1, HDC, RETREG1, SPECC1L, GRASP, SHC2, NOB1, PIP4K2B, ZNF329, GSR, ZNF740, HS3ST1, PMP22, ZMAT4, ERLIN1, TMEM79, SOX30, ST8SIA4, WSB2, RASAL2, E2F7, TCP11L2, BCO2, H3C8, TAB2, NOS2, MCAM, IFRD2, WDR61, POLD3, ARMC10, SMAP1, ING5, KNOP1, MYO3A, DLX4, STK40, MGA, ALG10B, PAQR3, WDR82, ZNF555, DPH2, PTPRJ, CDC16, SERPINF2, NBL1, HDHD5, KRT19, ABCA1, ARL13B, TMEM52, CPEB4, DHX9, PLIN5, OAZ2, KCNE4, GALK1, BCAS1, LAMC1, PRXL2B, NDUFA4L2, ATP8B2, RALGAPA1, ZNF620, AATF, PLD6, POSTN, ZNF846, TTLL6, STOX2, VASN, C1orf74, PLA1A, NFU1, GTF3C4, FAM184A, ZNF613, ZNF766, BAHCC1, SLC2A8, ITGA6, MTHFSD, GTSE1, MTNR1A, NBPF10, FBL, OTUD4, UBXN2A, ZNF155, TJP1, C14orf93, C15orf40, WIPF2, ZDHHC16, GTF2IRD1, DTL, FEN1, JCHAIN, ATP6V0D1, MYCL, ELL3, RAB33B, IQGAP1, ELOVL5, HHLA2 |
| Green ( $\mathrm{N}=110$ ) | CAPN13, MYLIP, LRRC17, KRT10, RAPGEFL1, RNASEL, CTSV, NRCAM, SOX21, ADCY2, SF3B3, CANX, TNRC6C, BCAT1, SNX25, CXXC4, PPIF, COL17A1, TMEM243, EML1, C6orf141, NDUFAF6, NAA60, MED29, CHKB, SYT7, DHTKD1, ARHGEF37, SNRNP200, CACNG8, TBCD, TXNIP, ZBTB46, DENND2C, H2BC11, IP6K3, MFSD6, GLG1, THBS1, LSM2, DPF2, ZBTB5, ZNF117, ITFG1, SNX9, AGPAT5, RNF43, PCDHB14, PHF2, TP53BP1, RPL18A, HNRNPAB, TSTD2, CSRP2, MSMB, LAMA4, <br> WDR4, TFCP2, ZNF280B, OCLN, OLFML1, MEX3D, PROM1, SPON2, CPM, LMLN, TMEM141, PPP1R13L, FHL2, TRNP1, TMEM64, NEO1, SEMA4F, LRMDA, DCAF5, THNSL2, TXN, TMEM116, COX7A2, TFAP4, HSPB7, ETFB, NR4A1, AGPS, ARHGAP20, HSF2BP, SLC44A2, SORCS2, TMPPE, SARS2, ARHGEF4, RGCC, SMO, SMCO4, SDR39U1, UQCRFS1, APBA1, OSBP, CHCHD10, GRHL2, SLC6A16, EMC1, ZNF345, CEACAM19, CAVIN2, FIGN, BCAR3, C1orf53, ATG4B, DHRS1 |
| Black ( $\mathrm{N}=794$ ) | PSMG3, NAA30, FBXL13, SLC4A11, C12orf57, GSS, U2AF1L4, PARD6B, STAP2, SLC35E3, YIPF4, SGSM3, DPY19L4, GPN3, TMEM128, CPEB2, IFITM3, SPIN1, FAM237B, PHIP, THOC6, BCKDHB, AEBP2, LRRC57, N4BP2, CCDC126, WDR17, TFAM, ZYG11B, TGOLN2, SRSF8, CRKL, RIN1, AGAP2, DPH7, HSD17B12, SEC22C, PAXX, ALKBH8, KIAA0825, BORCS7, WWTR1, ZAP70, HACE1, ITGAV, CREBRF, SPAST, METTL15, ZMYND19, ZC3H12A, ZMIZ2, RO60, EXOC6B, PPP1R12C, PLPBP, BMPR1B, AIMP2, GET1, ZBTB41, NRXN3, NBEAL2, MAD2L2, ZNF529, SEPTIN1, PGLS, SLC35F5, C9orf64, NELL2, AKAP10, GYS1, HEXD, HOXC4, CORO1A, TRMT5, SCAMP1, SMAD4, SLC25A28, CXXC1, ANKRD26, SCRIB, PSMD13, FAM189B, SIT1, PI4K2B, SAMD10, TMEM134, MBD3, PLEKHF1, EID1, ADH5, CEP97, FBXL17, OGFR, SDR42E1, ZNF221, WRN, CYP20A1, ZFAND2B, RNF111, |

RBBP5, PDLIM5, CD3E, CARF, PRPF31, DEGS1, EIF2B4, ZFR, C1orf226, TRIB2, CEP44, MICALL2, ZFAND4, C5orf51, HDAC10, MARCHF6, HYOU1, TMEM214, CYTH2, DPH6, CSNK1G2, TCEA1,
ZNF264, PEF1, SLC25A27, SH3BP1, ZNF776, FAM126B, ZNF721, SKP2, SEC24B, ETFBKMT, SBF1, SPTY2D1, LMBRD2, UHRF1BP1L, SPOCK2, MFSD4B, SMIM14, BTN3A3, UBALD1, LNX2, AP2A2,
RB1CC1, CST3, AATK, TMEM41B, DCP1A, TMED8, FNIP1, CD3D, ARID4B, TRIM2, CALR, OSBPL11, FAM227B, DAAM1, NFAT5, DDX11, AP1B1, ARFRP1, EIF3B, TRIM33, OTUD7B, WDR11, PIH1D1, UBQLN4, GPR85, RAB21, IMPAD1, PUS10, CARM1, SQOR, DPP8, ABTB2, EVI5, RFX3, NPRL2, CLPP, FBXL3, S100A2, UGDH, CXCR6, RAB34, MPP3, SNRNP48, SMAD2, NAA16, E4F1, BCKDK, CEP112, AUP1, GDPGP1, C1D, CISD3, TUBA4A, DDX56, TMEM51, ZNF383, NMRAL1, NEK7, HGH1, MTMR10, SMAD9, AGGF1, PRR22, B3GNT10, LYRM7, CLCN3, KIFC1, ZNF843, HARS2, FXYD3, ARHGAP5,
BTBD6, GCC2, METTL1, PRKCI, MOGS, PSMB10, PCBD2, ZNF579, GPALPP1, FBXL4, GP9, TAF1C,
ALG3, CHMP3, PTPN4, CDC40, EXOC8, MIB2, FZR1, BEND7, ZFYVE16, WDR41, MANEA, ZDHHC20, OLFML3, LRRC29, RPRD1B, SIAH1, UTP23, CD8A, SEPTIN9, MYO9B, NR2F6, PELP1, JMY, PTGR2, DENND6A, PSMD4, AP1M1, TMEM184C, ZNF619, CTDSPL2, ATF2, TBCB, ARMH3, TSC2, DIXDC1, OSBPL8, ANKRD36, WDR7, TLCD4, RCOR3, CSTB, TBX21, TTC33, SIRT6, PAFAH1B3, PLEKHA3,
BTF3L4, ZNF654, NDOR1, FRA10AC1, JADE1, MICAL3, ZNF708, ROPN1B, ADRM1, ATF7IP, PRRX2, USP8, DTWD2, PCYT2, CTBP1, TSC22D4, SATB1, JAKMIP1, MTAP, XRCC3, SPATA13, ZNF277, RFC3, CEBPZOS, ELF1, ATP2C1, LIMD2, PGGT1B, CLUH, DNAJC30, PCNP, GFI1, CDC42EP1,
RHOG, PDLIM1, ZNF566, CERT1, MED13, PRDM5, ARIH1, RBM48, ULK1, NOVA1, ZNF780B, CCDC110, CACNA1A, ARL2, MICU2, AGL, TMEM30A, NUP43, DBR1, CCL5, ORMDL2, ZNF720, SGMS2, NARS2, MINDY1, TIMM50, CHD1, DNAJB4, GIMAP1, TPP2, CNIH3, ABHD13, NRBP1,
TMEM106B, ANO5, UQCRC1, CAMTA1, RC3H1, FNDC3A, ZBTB26, MARK2, DNAJC14, EDF1, CCSAP, SAMD13, PIK3CA, EXOSC4, MED17, CD247, RASIP1, KLHL20, C19orf73, GIMAP7, CAT, PLXNB1, KDM7A, EVA1C, ANKAR, TIMM23, MOAP1, PKN2, PSMB3, RPRM, QPCTL, CBFB, FBXO16, HBS1L, CCDC152, SMG1, ANKUB1, TSPAN31, CD2, ZNF829, SDF4, B4GALT7, ODF3B, ALG10, TAF1B, SHQ1, CHCHD5, PDCD1, GTF2I, YIPF5, TMEM68, FBXW9, PGRMC2, KIF2A, EHMT1, CACTIN, ZNF606, SF3A2, RECQL5, NOC2L, PRR14, CAPN7, PDIA4, METTL26, FAM168B, ABI3, MAPK8IP3, ZDHHC17, HNRNPH3, PNKD, MCM8, HBP1, PPM1K, GRSF1, MRPL55, URI1, ROR2, GPR137, STK11, VPS41, MRM1, R3HDM2, VPS28, POLR2E, GCFC2, ICAM3, ARCN1, KCNAB1, PYCARD, DMXL1, VAV3, ACBD5, ANAPC16, CLDN4, PCNX3, SEPTIN8, CC2D1B, ZNF800, OTUB1, SELENOW, DGKZ, TNNC1, SKIL, FAM207A, LENG8, ERO1B, SPATA5, HRAS, ETV4, CARNMT1, MNT, THEM6, SH2D2A, ABCC1, RAPGEF6, CDC73, MAP3K2, NSD2, PDE4C, PYGB, TRIM11, PTPN11, PHF10, MLST8, WIZ, MRPL28, TRH, DRC7, RFESD, GPR171, CSNK2B, STMP1, RPN2, WDPCP, ATE1, IL32, MICOS13, GPSM1, FLOT1, SOX6, CCDC28A, DICER1, DIS3L, KLHDC1, PPP4C, SINHCAF, SHTN1, IDH3B, TRIP11, SH2B1, ABCB8, PUM1, COX11, HSPA4L, DGKH, TOM1L2, G3BP2, BDP1, FAM160B2, KIN, GLMP, UBA7, CRCP, ATXN3, PLEKHF2, SOS2, TASP1, FDXR, PPP2R5D, SRGAP2B, YARS2, BLVRB, SCML4,

IMP4, SPAG17, CLDN3, YTHDC1, PDE5A, PPP3CA, NAGPA, DPP9, CDK10, GRWD1, AP1G2, ELP4, MCHR1, GRINA, SCAF11, ZBTB25, MRPL10, BRWD1, SORT1, LCORL, RPN1, MOB3A, SCP2, ATP6V1A, WDR92, NHLRC2, TMOD3, TMEM179B, CDC34, FIGNL1, ZBTB18, GIPC2, TMEM54, ECHS1, ZNF664, NFKBIB, DHX36, TRAPPC12, TONSL, RNF11, SLC35A3, CACNG4, COPG1, BAIAP2, TTC21B, CTDNEP1, VCPIP1, FANCC, NCKAP1, MAST3, FZD3, TASOR, LRP8, SLC38A9, AKAP8L, PHOSPHO2, TRMT1, CDKAL1, ZDBF2, PPIP5K2, DENND4C, IKZF2, SELENOH, GCAT, ADAM17, DHX16, ZNF24, CCDC186, PDE6D, ASB4, SMTNL1, MCM5, ITSN2, SLMAP, MED16, MINK1, ZNF286A, CCDC97, ACTN4, COX6B1, TICAM1, DIPK2A, VPS4B, POLDIP2, RCSD1, GFPT1, PITPNM1, PRMT1, RNF138, ADPRM, NEK4, RNF170, CUL3, SF3B5, SMG5, SAMD8, GALK2, ZNF431, CTNNA1, MED7, NT5DC1, KIAA1586, MIER3, STX19, SZRD1, TAOK1, GTF3C5, HSD11B1L, PLIN3, DDHD2, KDF1, SMIM31, SCAND1, DCLRE1B, CDK17, LMNTD2, DNASE1L2, ATP13A2, GARS1, HAUS3, TSNAX, YPEL3, STOML2, PCMTD1, ANKRD52, ADSS2, SLC30A9, PEX16, TIMM13, RAB2A, TCTEX1D4, IFI27L1, KIF5B, CD200R1, CAPZB, RNH1, TMEM102, ARHGDIA, SNAPC4, RGS14, AK9, DNAJB14, ZNF580, SNX13, DDX17, ROGDI, FAM216B, AZIN1, KIAA1328, GP2, USF1, AVL9, AGO1, ASXL3, FAM8A1, ATXN2L, H3-3A, ZBTB7B, PM20D2, LRRC45, DIS3, EIF3A, CDK18, PSMC3, DHX29, IPMK, PES1, SLF2, IGSF9B, SHOC2, NCF1, LY6G5C, PIGN, GGA3, HMCES, CCDC12, PRIMPOL, CCDC30, TTC6, ARID3A, ZMPSTE24, PEX1, PSPH, C1orf87, ATP6V1F, PIDD1, CASC3, TBC1D10B, AURKAIP1, AHI1, VPS72, GPLD1, TRAF2, RING1, EML5, SWAP70, ANKMY2, MKLN1, MXRA8, PHAX, TMEM212, MGAT4B, BCLAF1, CCDC69, FAAP100, ZNF880, SSBP4, TTC23L, MRPL37, PCDHB1, NCLN, CRELD1, MST1, PDRG1, SLC22A18, TMEM190, HP1BP3, C8orf82, RAD18, P3H4, PLAAT4, PPP3CB, VWA1, CMTM6, SEPTIN3, TEPP, AGBL3, PPP1R11, MRC2, IMPACT, ST3GAL6, SLC25A17, POLE3, GNS, WDR60, LAMTOR4, PRDX2, CXCR1, CYC1, FNTB, PPIC, BTBD9, CLASRP, ABCC5, TMEM160, UHMK1, NANP, ATG2A, LLGL2, ABHD3, COPS2, PFN2, ITGB7, C1orf35, NDUFS7, KPNA5, TUBGCP4, SNRNP70, FRMPD2, UBE3D, ZNF747, FAM98C, TELO2, PYGL, GRM5, BCDIN3D, GRPEL2, FCHO1, MRPS6, DOLK, PPIB, MRPS17, NFKBIA, CPSF2, TMEM250, MIF4GD, DNAJC24, ZNF260, ZMYM1,

MED25, IGFBP7, PIN1, CDKN2AIP, SDCCAG8
B4GALT1, RAB7B, NOLC1, SLC35F6, HSPA2, MINPP1, MSLNL, SNAI2, ADAMTS17, TYW3, FOXA1, OVGP1, RIBC2, KITLG, FOXN1, DLK2, RNF212, PDGFC, ITGB1, TINCR, SULT1E1, SMU1, RGMB, DIRAS3, ZFP64, ANKRD29, VIPAS39, PROB1, FAM114A2, RERG, SYTL2, RHEBL1, EIF5, SNF8, ADAMTSL5, RAB20, FAM171B, HYKK, ZFYVE27, HHATL, SUPT16H, SEPTIN2, CDK5RAP2, TET1, ZNF227, PSCA, BMP8A, S100A1, EPHB6, ELAC1, TMPRSS2, CAMLG, AFAP1L2, SULT1C2, TMEM9B, DST, RARB, KRT12, ATAD1, CYP2U1, MMP7, FOXA2, ARF3, AK6, SENP8, CYBRD1, BIK, SV2A, DSEL, EMID1, SELENOS, PNRC2, SNX5, ORC1, XRCC5, LRPPRC, MLIP, KLF5, PERP, RAB3B, TTC32,
TMED3, CDK20, BLOC1S5, L3MBTL2, FIBIN, ICA1, KLHL9, BSCL2, ZZEF1, ZNF821, ZNF624, CD164L2, SYNGR1, TCF25, MAST4, STIMATE-MUSTN1, ESF1, HMGB1, FUT4, DERL3, ZNF473, TPST2, OCIAD2, GNPDA1, EPB41L5, PCDHGB4, DACT2, PIGQ, JAK1, ROM1, KCTD17, SEMA3D,

|  | INO80B, CNOT7, ABHD2, FJX1, FERMT2, ASPRV1, TAF5, CNOT11, SHH, VPS26A, ZNF595, FAM83F, STAMBPL1, SPNS1, THUMPD1, FHIT, LARGE1, USP1, INTS10, SPATA5L1, KIRREL1, FPR1, MMP25, PTPRG, C22orf15, TNS1, MED4, SLIT3, APPL2, RPS6KC1, LGALS9B, PPP1R1C, ADARB2, PRUNE1, SDHD, ACER3, TRMU, ABCB6, B3GALT2, FUT10, FANCE, ST20, EXOC2, LGALS9C, MARS2, CWC22, SLC16A8, ZNF420, POLR3D, ZFP2, CCDC200, VWA2, NEU4, MZF1, CDH5, LRRC23, MEGF6, DIO2, QRICH2, NUP54, SUPT5H, GSE1, ABCB4, FAM131A, NPIPB5, PDE7A, ZC3HC1, MEIS1, MYDGF, WDR20, RAB36, ISL1, NDUFB6, ELAVL1, VLDLR, NUP107, ZFP36L2, B3GLCT, FAM153B, ZNF512, ZNF552, CDC23, DRICH1, ZNF146, ZNF436, RILP, STK10, CCL3L1, TESK2, CAPN8, C11orf95, STK3, CR1L, DENND6B, FOXRED2, GJC2, DDX54, ELP3, LONP1, CSE1L, PUS7, MISP3, MAP2K3, LRRC31, TENT2, NOA1, LGALS8, PSMG4, TSPAN5, BRIX1, PTGR1, ITLN1, PCSK5, ZNF571, DIP2A, TGFB111, ABHD16A, ZNF181, CD36, VMAC, ZNF559 |
| :---: | :---: |
| Red ( $\mathrm{N}=46$ ) | TTC7B, DNHD1, ZNF839, PIAS3, EIF4EBP2, HNRNPUL2, CCNG2, CD83, OGDHL, SMUG1, PATL2, LPP, TDRD9, NPM2, CDHR2, ST6GALNAC5, SAR1A, SNIP1, PLXDC2, TIMP2, TACC2, COLCA2, MKS1, APOC1, LHFPL6, MEX3C, DEPDC5, CLASP1, AGAP9, SLC12A2, LANCL2, RBM27, TMEM130, DNAAF1, POLRMT, FNDC3B, KCNK15, EPC2, ERG28, FN1, DDAH2, ANAPC5, PLEKHA2, WASHC1, CBFA2T3, ELP6 |
| Turquoise ( $\mathrm{N}=85$ ) | IFIT2, MILR1, CCR2, HLA-DRB5, FCGR2B, LTB, CSF1R, IL21R, CXCL10, ADGRG5, PLXND1, SYT11, CAPN2, PRKG2, PALD1, SFMBT2, ASB2, CSF1, ATP10A, LAG3, COL3A1, AADAC, NPW, APBA2, BBS5, SALL4, CALCOCO2, C1S, LMNB1, AOAH, FNBP1, TLR10, TNFSF14, CARD9, ADSS1, FKBP15, A2M, HLA-DOA, ZNF609, RASSF5, DPYD, FYB1, HLA-DPB1, PTPN7, LRRN1, CNTLN, ADAR, LCP2, GPR18, CGREF1, TTYH2, ST8SIA1, CRTAM, PFDN6, SIRPG, ARHGAP25, TNFRSF11A, PCED1B, PPM1M, TMEM86A, QPRT, CD209, ANG, SLC26A4, CD74, STK33, OPTN, EPB41, DOCK10, IFI44L, AKR1C3, JAK2, CALHM6, RGS7BP, ANKRD39, LAX1, CCR5, HLA-DRA, HLA-DMA, CSF3, IGSF11, CXCL9, SYNPO2, IL4I1, CLEC10A |
| Yellow ( $\mathrm{N}=391$ ) | NAAA, C2orf50, TNC, TNFRSF10B, SERPINB4, NOP53, LSM14B, NEURL1B, AK2, CMTM8, SLC16A11, ATXN1, IFI16, FAR2, IRX3, RPS3, RPL7A, STOM, ANAPC15, DNAJC17, RPL3, ANKRD45, MTFR2, SMIM35, APBB3, GBP6, AKAP6, BIRC5, MYH15, DMAC2L, RPL12, MTRF1, LAMC2, SLC5A8, EMC4, DHDDS, TRMT9B, DSTN, TAF11, EIF3F, AUH, FCGR3B, RPS20, CD55, ISCA2, FAM111B, RNF183, <br> AURKB, EGLN2, GABRP, WDR62, DSP, RNF5, TIGD5, BORA, AKAP9, ST7, CEP85, CTF1, <br> MPHOSPH8, NFIC, TRIM38, LDHA, NABP1, ALDH1A3, LNP1, RUFY4, FAM174A, ACAD8, RPL19, HMGN2, PLEK2, RPL6, RPL29, DPY19L1, SCRN1, SLC8B1, TTC12, CYP51A1, KHDC1, CPT1B, RPL10A, PMM2, PPIP5K1, UBXN10, GLRB, XAF1, SOGA1, ENO4, ZNF133, CLEC2D, TOB2, ZNF32, TGM3, CEP55, TPK1, FBXO27, NFE2L3, ZNF562, SLC23A1, DTD1, FKBP14, STAT2, LMO1, RPL13A, ARHGAP44, EFCAB11, ADRB1, POC1B, CASS4, ARG2, NBAS, KRTCAP3, CPLANE2, RPLPO, PTGS2, TIMM22, MRFAP1, UTP20, PLPPR3, MUL1, CXCR2, LSMEM1, CD58, HEATR3, MOB3C, ATP6V1FNB, DDX47, PLEKHG4B, ST6GALNAC2, AK8, LRRIQ4, ATP4B, FILIP1, RPL32, DNM1L, MED30, VEPH1, |

BBS4, GALNT14, NFKBID, KIF20A, RAB4B, PSTPIP2, AURKA, SARAF, FAM118B, SUPT6H, TRAPPC9, JDP2, EPHX2, VEGFB, EEF1B2, RPS14, LPCAT3, SHOC1, RGS16, KLK11, P4HA2, RPS13, EFCAB6, SLC16A14, SETD7, MRPL9, HNRNPA0, PDGFA, HTRA1, TTC39A, PDK2, KHDC4, EGLN3, IVNS1ABP, RPL13, ANLN, RNF19A, SPRYD4, ECSIT, SH3PXD2B, ANKRD66, FARS2, LRRC69, KLF15, ZSCAN18, SP140L, PPA1, DSCAML1, CMC2, SAP30L, CLEC2B, CASP6, MBTPS1, ITPRIPL2, BACE2, FAM135A, FUCA2, PJVK, TAF6, SYTL3, RALBP1, TNFRSF10C, TRIM16, LDLRAD1, TMEM8B, GALNT5, IGF1R, RABIF, FBXO36, USP15, CPNE5, MCU, BUB1B, TOMM7, KDM1B, FYCO1, FBLN7, LNX1, ALPK3, C1RL, ZDHHC4, GNL3, YWHAQ, MCCC2, RND3, CCNI2, GAB2, MAP2K5, ERCC5, DMTN, RACK1, FPR2, ASPM, CKMT1A, SERGEF, ITPRID2, PTPRN2, RELB, FAM222B, DNAH6, FAM98A, ZC2HC1C, GNAZ, VWA5A, PARP14, TGIF1, APCDD1, SPAG7, MRPS18B, TMEM181, PRDM1, RPL30, SLC20A1, COL16A1, GRAMD1A, SHCBP1, C5orf49, COBL, NFKB2, NUPR2, NEK11, FKBP4, RPL37A, PANX2, PROM2, MRPL45, STIP1, C11orf21, WBP2, DMRTA1, KAT5, KBTBD8, ACSL3, BMP8B, GPR39, KCTD14, CDC42EP2, PRR18, AZU1, INPP5F, MAP3K8, ZNF200, MYL12A, AKIP1, SON, PLEKHD1, OAS1, AMPH, PLEKHB2, SAMD9, ZBTB7C, SPATA18, SKA3, WASF1, MGAT5, SEC31B, C1orf21, SOWAHA, TMEM14C, ZFYVE9, LOX, GCNT1, TIMM23B, KCNB1, TIGD3, ECT2, NBPF1, ITGB6, FAM86B1, VPS45, PDK1, TMEM203, MAB21L4, REM2, NUSAP1, SAMD12, PCSK9, CRY2, RPA3, TSTD1, GJB2, LARGE2, RNF10, DAAM2, SLC7A1, KIAA1671, ZNF841, TXNL4B, C6orf223, UNC119B, MPZL1, RHBDL3, TRIM31, MRPS23, LACTB2, RPSA, SDHC, PLEKHS1, CMAS, DYRK3, RBMS2, CDC42SE1, STMND1, COPS5, SRPK1, ADPRHL2, ATP5MC2, SYS1, REXO4, RPS5, C11orf97, GNAS, AK3, CEP152, TMEM117, DHX57, CDC6, HAS3, ARPC3, CFAP57, CSF3R, ITPKB, S100A5, RPS12, TIMM17A, DEPDC1, OXA1L, DDX28, GAPT, JHY, RASSF2, TAF7, PCLAF, NCAPD3, RARS1, PLAAT3, FBXW2, CCNE1, RDH10, RPS11, RAC2, RPLP1, ACAT2, CYP4F3, HACD4, GPSM2, TMEM106C

Table S3. Significant pathways for the genes that were correlated with the MA-associated black, turquoise and yellow co-methylation module eigenvectors. The uniquely correlated genes of each module ( $F D R<0.10$ ) were included as input for pathway analysis (TopFunn (9)). Eleven, 36, and 33 pathways were significantly enriched for the uniquely correlated genes of the black, turquoise and yellow modules, respectively, at an FDR-adjusted $p$-value of $<0.10$. No pathways were enriched for the uniquely correlated genes of the NMA-associated orange or green modules or the MA-associated blue and red modules. The 3, 16, and 18 pathways of the black, turquoise, and yellow modules that discriminated between NMA and MA cases using Random Forests are bolded and listed in Table S4.

| Black module (794 uniquely correlated genes) |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ID | Name | Source | P -value | FDRadjusted Pvalue | \# of input genes / \# of total genes in annotation | Genes from input |
| 1269649 | Gene Expression | BioSystems: REACTOME | 7.33E-07 | 1.91E-03 | 111 / 1844 | NR2F6, RPN1, RPN2, MRM1, PIDD1, FANCC, METTL1, CDC40, C1D, ZNF606, TRMT5, <br> DHX16, CARM1, CASC3, TRMT1, SKIL, DIS3, ZNF529, MBD3, TRIM33, SNRNP48, LAMTOR4, PELP1, ZNF747, EIF3A, EIF3B, SNAPC4, SNRNP70, DCP1A, BDP1, SMG1, HBS1L, ARID4B, GARS1, ZNF619, PHAX, JMY, STK11, CRCP, IMP4, EIF2B4, AEBP2, CPSF2, MLST8, TCEA1, GCFC2, TAF1C, TAF1B, NUP43, SMG5, PRDX2, TFAM, DICER1, HARS2, PES1, ZNF566, H3-3A, ZNF431, ZNF721, ZNF664, U2AF1L4, TSC2, TSNAX, GTF3C5, RPRD1B, PIN1, PRMT1, ZNF264, MED17, MED7, EXOSC4, NRBP1, ZNF383, POLR2E, COX6B1, WRN, COX11, WWTR1, ATF2, THOC6, ZNF708, CSNK2B, ZNF221, SF3B5, RNF111, ZNF720, PRPF31, NOC2L, PSMB3, PSMB10, PSMC3, PSMD4, PSMD13, CDKAL1, ZNF776, ALKBH8, CDC73, MED13, ARID3A, AIMP2, MED16, RFC3, NARS2, YARS2, AGO1, MED25, EHMT1, SF3A2, SMAD2, ZNF286A, SMAD4 |
| M9904 | T cell receptor signaling pathway | MSigDB C2 BIOCARTA (v7.3) | 1.65E-05 | 1.53E-02 | 15 / 108 | VAV3, NFAT5, SOS2, NFKBIA, NFKBIB, CD3D, CD3E, CD247, CD8A, PDCD1, PIK3CA, HRAS, ZAP70, PPP3CA, PPP3CB |
| M39843 | Cancer immunotherapy by PD-1 blockade | MSigDB C2 BIOCARTA (v7.3) | 1.76E-05 | 1.53E-02 | 7 / 23 | NFAT5, CD3D, CD3E, CD8A, PDCD1, ZAP70, PTPN11 |
| M19784 | T Cell Receptor Signaling Pathway | MSigDB C2 BIOCARTA (v7.3) | 3.77E-05 | 2.09E-02 | 9 / 44 | NFKBIA, CD3D, CD3E, CD247, PIK3CA, HRAS, ZAP70, PPP3CA, PPP3CB |
| 83080 | T cell receptor signaling pathway | BioSystems: KEGG | 4.01E-05 | 2.09E-02 | 14 / 103 | VAV3, SOS2, NFKBIA, NFKBIB, CD3D, CD3E, CD247, CD8A, PDCD1, PIK3CA, HRAS, ZAP70, PPP3CA, PPP3CB |


| M13247 | T Cytotoxic Cell Surface Molecules | MSigDB C2 BIOCARTA (v7.3) | 5.32E-05 | $2.31 \mathrm{E}-02$ | 5 / 12 | CD2, CD3D, CD3E, CD247, CD8A |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| M16966 | Stathmin and breast cancer resistance to antimicrotubule agents | MSigDB C2 <br> BIOCARTA <br> (v7.3) | 7.85E-05 | 2.92E-02 | 6 / 20 | CD2, CD3D, CD3E, CD247, UHMK1, RB1CC1 |
| 1269171 | Adaptive Immune System | BioSystems: REACTOME | 9.62E-05 | 3.13E-02 | 54 / 826 | BTBD6, NCF1, FZR1, BTN3A3, AP2A2, AP1B1, HACE1, SEC24B, RNF138, CUL3, FBXW9, SIAH1, SKP2, CD200R1, ASB4, KLHL20, NFKBIA, NFKBIB, AP1M1, MLST8, CALR, CD3D, CD3E, CD247, CD8A, CDC34, TPP2, PDCD1, FBXL13, TSC2, UBA7, PIK3CA, HRAS, ICAM3, MIB2, ZAP70, PPP2R5D, RNF111, PSMB3, PSMB10, PSMC3, PSMD4, PSMD13, ITGAV, ITGB7, FBXL3, FBXL4, PTPN11, TRIM11, MRC2, KIF2A, KIF5B, AGO1, UBE3D |
| 1269192 | Class I MHC mediated antigen processing \& presentation | BioSystems: REACTOME | 1.39E-04 | 3.92E-02 | $30 / 376$ | BTBD6, NCF1, FZR1, HACE1, SEC24B, RNF138, CUL3, FBXW9, SIAH1, SKP2, ASB4, KLHL20, CALR, CDC34, TPP2, FBXL13, UBA7, MIB2, RNF111, PSMB3, PSMB10, PSMC3, PSMD4, PSMD13, ITGAV, FBXL3, FBXL4, TRIM11, MRC2, UBE3D |
| P00053 | T cell activation | PantherDB | 1.51E-04 | 3.92E-02 | 11 / 76 | VAV3, SOS2, NFKBIA, CD3D, CD3E, CD247, PIK3CA, HRAS, ZAP70, PPP3CA, PPP3CB |
| M19422 | IL 17 Signaling Pathway | MSigDB C2 BIOCARTA (v7.3) | 1.83E-04 | 4.34E-02 | 5 / 15 | CD2, CD3D, CD3E, CD247, CD8A |
| 83059 | mTOR signaling pathway | BioSystems: KEGG | $2.41 \mathrm{E}-04$ | 4.87E-02 | 16 / 151 | FNIP1, ULK1, SKP2, NPRL2, LAMTOR4, SOS2, ATP6V1A, STK11, MLST8, ATP6V1F, TSC2, PIK3CA, HRAS, SLC38A9, TELO2, FZD3 |
| M39602 | Neurodegeneration with brain iron accumulation (NBIA) subtypes pathway | MSigDB C2 <br> BIOCARTA <br> (v7.3) | 2.43E-04 | 4.87E-02 | $8 / 44$ | SCP2, ULK1, ATG2A, STK11, MLST8, ATP13A2, TSC2, RB1CC1 |
| Turquoise module (85 uniquely correlated genes) |  |  |  |  |  |  |


| ID | Name | Source | P-value | FDRadjusted $P$ value | \# of input genes / \# of total genes in annotation | Genes from input |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 172846 | Staphylococcus aureus infection | $\begin{gathered} \hline \text { BioSystems: } \\ \text { KEGG } \end{gathered}$ | 8.68E-09 | 3.24E-06 | 7 / 56 | HLA-DMA, FCGR2B, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5, C1S |
| 200309 | Rheumatoid arthritis | BioSystems: KEGG | 1.11E-08 | 3.24E-06 | 8 / 90 | CSF1, HLA-DMA, HLA-DOA, HLA-DPB1, HLADRA, HLA-DRB5, LTB, TNFRSF11A |
| 213780 | Tuberculosis | BioSystems: KEGG | 1.33E-08 | 3.24E-06 | 10 / 179 | JAK2, HLA-DMA, FCGR2B, HLA-DOA, CARD9, HLA-DPB1, HLA-DRA, HLA-DRB5, CD74, CD209 |
| 83078 | Hematopoietic cell lineage | $\begin{gathered} \text { BioSystems: } \\ \text { KEGG } \end{gathered}$ | 2.01E-08 | 3.68E-06 | 8 / 97 | CSF1, CSF1R, CSF3, HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5 |
| M9809 | Cytokine-cytokine receptor interaction | MSigDB C2 BIOCARTA (v7.3) | 5.10E-08 | 7.47E-06 | 11 / 265 | CCR2, CSF1, CSF1R, CSF3, TNFSF14, CXCL10, IL21R, CXCL9, LTB, CCR5, TNFRSF11A |
| 83051 | Cytokine-cytokine receptor interaction | BioSystems: KEGG | 6.18E-08 | 7.53E-06 | 11 / 270 | CCR2, CSF1, CSF1R, CSF3, TNFSF14, CXCL10, IL21R, CXCL9, LTB, CCR5, TNFRSF11A |
| M13950 | Asthma | MSigDB C2 BIOCARTA (v7.3) | 2.97E-07 | 3.10E-05 | 5 / 30 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5 |
| 83120 | Asthma | BioSystems: KEGG | 3.52E-07 | 3.22E-05 | 5 / 31 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5 |
| 1269200 | MHC class II antigen presentation | BioSystems: REACTOME | 6.20E-07 | 4.90E-05 | 7 / 103 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5, LAG3, CD74 |
| 842771 | Inflammatory bowel disease (IBD) | BioSystems: KEGG | 6.76E-07 | 4.90E-05 | 6 / 65 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5, IL21R |
| 1269175 | Generation of second messenger molecules | BioSystems: REACTOME | 7.67E-07 | 4.90E-05 | 5 / 36 | HLA-DPB1, HLA-DRA, HLA-DRB5, LCP2, FYB1 |
| 1469482 | Th17 cell differentiation | BioSystems: KEGG | 8.04E-07 | 4.90E-05 | 7 / 107 | JAK2, HLA-DMA, HLA-DOA, HLA-DPB1, HLADRA, HLA-DRB5, IL21R |
| M18615 | Allograft rejection | MSigDB C2 <br> BIOCARTA <br> (v7.3) | 8.84E-07 | 4.98E-05 | 5 / 37 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5 |


| 83123 | Allograft rejection | BioSystems: KEGG | 1.01E-06 | 5.30E-05 | 5 / 38 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 169642 | Toxoplasmosis | BioSystems: KEGG | 1.16E-06 | 5.68E-05 | 7 / 113 | JAK2, HLA-DMA, HLA-DOA, HLA-DPB1, HLADRA, HLA-DRB5, CCR5 |
| M3126 | Leishmania infection | MSigDB C2 BIOCARTA (v7.3) | 1.25E-06 | 5.70E-05 | 6 / 72 | JAK2, HLA-DMA, HLA-DOA, HLA-DPB1, HLADRA, HLA-DRB5 |
| 144181 | Leishmaniasis | BioSystems: KEGG | 1.35E-06 | 5.77E-05 | 6 / 73 | JAK2, HLA-DMA, HLA-DOA, HLA-DPB1, HLADRA, HLA-DRB5 |
| M13519 | Graft-versus-host disease | MSigDB C2 <br> BIOCARTA <br> (v7.3) | 1.50E-06 | 5.77E-05 | 5 / 41 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5 |
| 83124 | Graft-versus-host disease | BioSystems: KEGG | 1.50E-06 | 5.77E-05 | 5 / 41 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5 |
| 217173 | Influenza A | BioSystems: KEGG | 1.76E-06 | 6.07E-05 | $8 / 173$ | JAK2, HLA-DMA, HLA-DOA, CXCL10, HLADPB1, HLA-DRA, HLA-DRB5, ADAR |
| 83074 | Antigen processing and presentation | BioSystems: KEGG | 1.86E-06 | 6.07E-05 | 6 / 77 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5, CD74 |
| M12617 | Type I diabetes mellitus | MSigDB C2 BIOCARTA (v7.3) | 1.91E-06 | $6.07 \mathrm{E}-05$ | 5 / 43 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5 |
| 83095 | Type I diabetes mellitus | BioSystems: KEGG | 1.91E-06 | 6.07E-05 | 5 / 43 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5 |
| 1269310 | Cytokine Signaling in Immune system | BioSystems: REACTOME | 2.68E-06 | 8.17E-05 | 15 / 763 | CCR2, JAK2, CSF1, CSF1R, CSF3, TNFSF14, CXCL10, HLA-DPB1, HLA-DRA, HLA-DRB5, LTB, CCR5, TNFRSF11A, ADAR, IFIT2 |
| 377873 | Herpes simplex infection | BioSystems: KEGG | 2.91E-06 | 8.52E-05 | $8 / 185$ | JAK2, TNFSF14, HLA-DMA, HLA-DOA, HLADPB1, HLA-DRA, HLA-DRB5, CD74 |
| M39693 | Ebola Virus Pathway on Host | MSigDB C2 <br> BIOCARTA <br> (v7.3) | 3.14E-06 | 8.84E-05 | 7 / 131 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5, CLEC10A, CD209 |
| M615 | Intestinal immune network for $\lg \mathrm{A}$ production | MSigDB C2 BIOCARTA (v7.3) | 3.33E-06 | 9.03E-05 | 5 / 48 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5 |
| 1470924 | Interleukin-10 signaling | BioSystems: REACTOME | 3.70E-06 | 9.33E-05 | 5 / 49 | CCR2, CSF1, CSF3, CXCL10, CCR5 |


| 128760 | Intestinal immune network for $\lg A$ production | BioSystems: KEGG | 3.70E-06 | 9.33E-05 | 5 / 49 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| M16004 | Antigen processing and presentation | MSigDB C2 <br> BIOCARTA <br> (v7.3) | 4.07E-06 | 9.92E-05 | 6 / 88 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5, CD74 |
| M39398 | Allograft Rejection | MSigDB C2 BIOCARTA (v7.3) | 4.35E-06 | 1.03E-04 | 6 / 89 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5, CXCL9 |
| M4741 | Systemic lupus erythematosus | $\begin{gathered} \text { MSigDB C2 } \\ \text { BIOCARTA } \\ \text { (v7.3) } \\ \hline \end{gathered}$ | 4.66E-06 | 1.07E-04 | 7 / 139 | HLA-DMA, FCGR2B, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5, C1S |
| M13103 | Autoimmune thyroid disease | MSigDB C2 <br> BIOCARTA <br> (v7.3) | 4.98E-06 | 1.11E-04 | 5 / 52 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5 |
| 1458885 | Th1 and Th2 cell differentiation | BioSystems: KEGG | 5.27E-06 | 1.14E-04 | 6 / 92 | JAK2, HLA-DMA, HLA-DOA, HLA-DPB1, HLADRA, HLA-DRB5 |
| 83121 | Autoimmune thyroid disease | BioSystems: KEGG | 5.48E-06 | 1.15E-04 | 5 / 53 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5 |
| 153910 | Phagosome | BioSystems: KEGG | 9.15E-06 | 1.85E-04 | 7 / 154 | HLA-DMA, FCGR2B, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5, CD209 |
| 125138 | Viral myocarditis | BioSystems: KEGG | 9.35E-06 | 1.85E-04 | 5 / 59 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5 |
| M6910 | Cytokines and Inflammatory Response | $\begin{gathered} \text { MSigDB C2 } \\ \text { BIOCARTA } \\ (v 7.3) \\ \hline \end{gathered}$ | 1.09E-05 | 2.10E-04 | 4 / 29 | CSF1, CSF3, HLA-DRA, HLA-DRB5 |
| M12294 | Viral myocarditis | $\begin{gathered} \hline \text { MSigDB C2 } \\ \text { BIOCARTA } \\ (\mathrm{v} 7.3) \\ \hline \end{gathered}$ | 2.17E-05 | 4.07E-04 | 5 / 70 | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, HLA-DRB5 |
| M39363 | Type II interferon signaling (IFNG) | $\begin{gathered} \text { MSigDB C2 } \\ \text { BIOCARTA } \\ (\text { v } 7.3) \end{gathered}$ | 2.94E-05 | 5.38E-04 | 4 / 37 | JAK2, CXCL10, CXCL9, IFIT2 |
| M1473 | B Lymphocyte Cell Surface Molecules | $\begin{gathered} \text { MSigDB C2 } \\ \text { BIOCARTA } \\ (\mathrm{v} 7.3) \\ \hline \end{gathered}$ | 3.80E-05 | 6.50E-04 | 3 / 14 | FCGR2B, HLA-DRA, HLA-DRB5 |


| M22023 | Antigen <br> Processing and <br> Presentation | MSigDB C2 <br> BIOCARTA <br> (v7.3) | $3.80 \mathrm{E}-05$ | $6.50 \mathrm{E}-04$ | $3 / 14$ | HLA-DRA, HLA-DRB5, CD74 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 193147 | Osteoclast <br> differentiation | BioSystems: <br> KEGG | $3.82 \mathrm{E}-05$ | $6.50 \mathrm{E}-04$ | $6 / 130$ | CSF1, CSF1R, FCGR2B, TNFRSF11A, LCP2, |
| 83122 | Systemic lupus <br> erythematosus | BioSystems: <br> KEGG | $4.34 \mathrm{E}-05$ | $7.22 \mathrm{E}-04$ | $6 / 133$ | HLA-DMA, HLA-DOA, HLA-DPB1, HLA-DRA, |
| M6856 | Hematopoietic cell <br> lineage | MSigDB C2 <br> BIOCARTA <br> (v7.3) | $6.22 \mathrm{E}-05$ | $1.01 \mathrm{E}-03$ | $5 / 87$ | CSF1, CSF1R, CSF3, HLA-DRA, HLA-DRB5 |
| 1269547 | Chemokine <br> receptors bind <br> chemokines | BioSystems: <br> REACTOME | $8.32 \mathrm{E}-05$ | $1.31 \mathrm{E}-03$ | $4 / 48$ | CCR2, CXCL10, CXCL9, CCR5 |


| 1268690 | Eukaryotic Translation Elongation | BioSystems: REACTOME | 2.17E-19 | 3.95E-16 | 23 / 98 | RPL30, RPL29, RPL32, RPL37A, RPLPO, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, RPSA, EEF1B2, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| M39495 | Cytoplasmic Ribosomal Proteins | MSigDB C2 <br> BIOCARTA <br> (v7.3) | 5.21E-19 | 4.74E-16 | 22 / 90 | RPL30, RPL29, RPL32, RPL37A, RPLP0, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1268691 | Peptide chain elongation | BioSystems: REACTOME | 1.13E-18 | 5.12E-16 | 22 / 93 | RPL30, RPL29, RPL32, RPL37A, RPLPO, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1269120 | Viral mRNA Translation | BioSystems: REACTOME | 1.13E-18 | 5.12E-16 | 22 / 93 | RPL30, RPL29, RPL32, RPL37A, RPLP0, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1268681 | Formation of a pool of free 40S subunits | BioSystems: REACTOME | 1.83E-18 | 6.65E-16 | 23/107 | RPL30, RPL29, RPL32, RPL37A, RPLPO, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, EIF3F, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1339156 | Selenocysteine synthesis | BioSystems: REACTOME | $2.36 \mathrm{E}-18$ | 7.16E-16 | 22 / 96 | RPL30, RPL29, RPL32, RPL37A, RPLPO, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1268692 | Eukaryotic <br> Translation Termination | BioSystems: REACTOME | 3.01E-18 | 7.81E-16 | 22 / 97 | RPL30, RPL29, RPL32, RPL37A, RPLPO, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| M189 | Ribosome | $\begin{gathered} \text { MSigDB C2 } \\ \text { BIOCARTA } \\ (\mathrm{v} 7.3) \end{gathered}$ | 5.81E-18 | 1.23E-15 | 21 / 88 | RPL30, RPL29, RPL32, RPL37A, RPLPO, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS20, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1269718 | Nonsense Mediated Decay (NMD) independent of the Exon Junction Complex (EJC) | BioSystems: REACTOME | 6.09E-18 | 1.23E-15 | 22 / 100 | RPL30, RPL29, RPL32, RPL37A, RPLPO, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |


| 1268686 | GTP hydrolysis and joining of the 60S ribosomal subunit | BioSystems: REACTOME | $2.28 \mathrm{E}-17$ | $3.77 \mathrm{E}-15$ | 23/119 | RPL30, RPL29, RPL32, RPL37A, RPLP0, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, EIF3F, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1268688 | L13a-mediated translational silencing of Ceruloplasmin expression | BioSystems: REACTOME | $2.28 \mathrm{E}-17$ | 3.77E-15 | 23/119 | RPL30, RPL29, RPL32, RPL37A, RPLP0, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, EIF3F, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1339149 | Selenoamino acid metabolism | BioSystems: REACTOME | 4.95E-17 | 7.50E-15 | $23 / 123$ | RPL30, RPL29, RPL32, RPL37A, RPLP0, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, RARS1, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1268680 | Cap-dependent Translation Initiation | BioSystems: REACTOME | 1.04E-16 | 1.35E-14 | 23 / 127 | RPL30, RPL29, RPL32, RPL37A, RPLP0, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, EIF3F, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1268679 | Eukaryotic Translation Initiation | BioSystems: REACTOME | 1.04E-16 | 1.35E-14 | 23 / 127 | RPL30, RPL29, RPL32, RPL37A, RPLP0, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, EIF3F, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1268689 | SRP-dependent cotranslational protein targeting to membrane | BioSystems: REACTOME | 1.77E-16 | 2.15E-14 | 22 / 116 | RPL30, RPL29, RPL32, RPL37A, RPLP0, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1269716 | NonsenseMediated Decay (NMD) | BioSystems: REACTOME | 4.54E-16 | 4.86E-14 | 22 / 121 | RPL30, RPL29, RPL32, RPL37A, RPLPO, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1269717 | Nonsense Mediated Decay (NMD) enhanced by the Exon Junction Complex (EJC) | BioSystems: REACTOME | 4.54E-16 | 4.86E-14 | 22 / 121 | RPL30, RPL29, RPL32, RPL37A, RPLP0, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |


| 1268678 | Translation | BioSystems: REACTOME | 3.92E-15 | 3.96E-13 | 24 / 165 | RPL30, RPL29, RPL32, RPL37A, RPLP0, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, EIF3F, RPL10A, RPSA, EEF1B2, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1269115 | Influenza Viral RNA Transcription and Replication | BioSystems: REACTOME | 8.08E-15 | 7.60E-13 | 22 / 138 | RPL30, RPL29, RPL32, RPL37A, RPLP0, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 83036 | Ribosome | BioSystems: KEGG | 8.35E-15 | 7.60E-13 | 23/154 | RPL30, RPL29, RPL32, RPL37A, RPLPO, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, MRPL9, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1269109 | Influenza Life Cycle | BioSystems: REACTOME | $3.14 \mathrm{E}-14$ | 2.71E-12 | 22 / 147 | RPL30, RPL29, RPL32, RPL37A, RPLP0, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1383086 | Major pathway of rRNA processing in the nucleolus and cytosol | BioSystems: REACTOME | 3.28E-14 | 2.71E-12 | 24 / 181 | DDX47, RPL30, RPL29, RPL32, RPL37A, RPLPO, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, UTP20, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1427846 | rRNA processing in the nucleus and cytosol | BioSystems: REACTOME | 1.39E-13 | 1.10E-11 | 24 / 193 | DDX47, RPL30, RPL29, RPL32, RPL37A, RPLP0, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, UTP20, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1269108 | Influenza Infection | BioSystems: REACTOME | 1.45E-13 | 1.10E-11 | 22 / 158 | RPL30, RPL29, RPL32, RPL37A, RPLPO, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1383085 | rRNA processing | BioSystems: REACTOME | 4.30E-13 | 3.13E-11 | 24 / 203 | DDX47, RPL30, RPL29, RPL32, RPL37A, RPLP0, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, UTP20, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |


| 1270158 | Metabolism of amino acids and derivatives | BioSystems: REACTOME | 1.99E-10 | 1.40E-08 | 28 / 367 | RPL30, RPL29, RPL32, RPL37A, RPLP0, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, ARG2, ACAD8, AUH, MCCC2, RPL10A, CKMT1A, RARS1, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1269056 | Infectious disease | BioSystems: REACTOME | 7.51E-08 | 5.06E-06 | 25 / 393 | RPL30, RPL29, RPL32, RPL37A, RPLPO, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, RPL10A, TAF6, TAF7, TAF11, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1268682 | Formation of the ternary complex, and subsequently, the 43S complex | BioSystems: REACTOME | 6.64E-07 | 4.32E-05 | 9 / 55 | RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, EIF3F, RPSA |
| 1268685 | Ribosomal scanning and start codon recognition | BioSystems: REACTOME | 2.17E-06 | $1.36 \mathrm{E}-04$ | 9 / 63 | RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, EIF3F, RPSA |
| 1268684 | Translation initiation complex formation | BioSystems: REACTOME | 2.49E-06 | 1.51E-04 | 9 / 64 | RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, EIF3F, RPSA |
| 1268683 | Activation of the mRNA upon binding of the capbinding complex and elFs, and subsequent binding to 43S | BioSystems: REACTOME | 2.84E-06 | 1.67E-04 | 9 / 65 | RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, EIF3F, RPSA |
| 1268677 | Metabolism of proteins | BioSystems: REACTOME | $2.61 \mathrm{E}-05$ | $1.48 \mathrm{E}-03$ | $53 / 1631$ | RPL30, RPL29, RPL32, RPL37A, RPLP0, RPLP1, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, ARG2, ACAD8, AUH, MCCC2, RPL10A, CKMT1A, RARS1, RPSA, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |
| 1268854 | Disease | BioSystems: REACTOME | 6.14E-05 | 3.38E-03 | 33 / 867 | RPL30, RPL29, RPL32, RPL37A, RPLP0, RPLP1, PDGFA, RPS3, RPS5, RPS11, RPS12, RPS13, RPS14, RPS20, ZFYVE9, NEURL1B, MCCC2, GAB2, RPL10A, NFKB2, TAF6, TAF7, TAF11, RPSA, AKAP9, RNF5, RPL13A, RPL3, RPL6, RPL7A, RPL12, RPL13, RPL19 |


| 1269649 | Gene Expression | BioSystems: REACTOME | 9.79E-05 | 5.24E-03 | 56/1844 | DDX47, MTA2, RPL30, RPL29, RPL32, RPL37A, RPLPO, RPLP1, RPS3, RPS5, RPS11, RPS12, <br> RPS13, RPS14, RPS20, TSC1, TRMT9B, <br> MED30, KAT5, BIRC5, PPA1, NABP1, FARS2, <br> EIF3F, ZNF133, ZNF562, ZNF200, TNFRSF10C, <br> TNFRSF10B, PRDM1, RPL10A, AURKA, NFIC, <br> SUPT6H, HNRNPAO, UTP20, YWHAQ, TAF6, <br> TAF7, TAF11, RARS1, SAP30L, CASP6, RPSA, <br> CCNE1, TGIF1, EEF1B2, RPL13A, RPA3, RPL3, <br> RPL6, RPL7A, RPL12, RPL13, AURKB, RPL19 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| M180 | Hypoxic and oxygen homeostasis regulation of HIF-1-alpha | MSigDB C2 <br> BIOCARTA <br> (v7.3) | 3.56E-04 | 1.85E-02 | 4 / 19 | RACK1, COPS5, EGLN3, EGLN2 |
| M39529 | Photodynamic therapy-induced NF-kB survival signaling | MSigDB C2 <br> BIOCARTA <br> (v7.3) | 4.29E-04 | 2.17E-02 | 5 / 35 | BIRC5, PTGS2, NFKB2, EGLN2, RELB |
| 138033 | Signaling by Aurora kinases | BioSystems: <br> Pathway <br> Interaction <br> Database | 1.01E-03 | 4.96E-02 | $2 / 3$ | AURKA, AURKB |

Table S4. Maternal asthma-discriminatory pathways of the black, turquoise, and yellow modules. For each of the 11, 36, and 33 enriched pathways of the black, turquoise, and yellow modules, respectively, a per-individual median normalized and covariateadjusted expression level was calculated for all expressed genes (CPM > 1 in at least $25 \%$ of subjects) in the pathway ( 3 to 1,844 per pathway. Random Forests selected pathways that discriminated between asthma cases with (MA; $\mathrm{N}=27$ ) and without an asthmatic mother (NMA; N=48); 3 (black), 16 (turquoise), and 18 (yellow) discriminatory pathways were identified as having Mean Decrease in Accuracy > 1. There were 231, 535, and 675 unique genes in total for the black, turquoise, and yellow modules, respectively; a median score of the normalized and covariate-adjusted expression for each set of module-specific genes were compared between MA and NMA cases (Fig. 2 in the main paper; Fig. S8).

| Black module (231 expressed genes) |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| ID | Name | Source | Mean Decrease in Accuracy | Median Expression (NMA) | Median Expression (MA) | \# of expressed genes / \# of total genes | Expressed genes |
| M9904 | T cell receptor signaling pathway | MSigDB C2 BIOCARTA (v7.3) | 2.10 | 5.06 | 5.00 | 95 / 108 | KRAS, RAF1, PDCD1, AKT3, NRAS, MAP3K8, FOS, PDPK1, PIK3R3, PAK4, MAP3K14, GRB2, CBLC, NCK1, RELA, PAK6, LCK, TEC, LCP2, CBL, CBLB, ZAP70, GSK3B, ITK, CHUK, RHOA, <br> MALT1, JUN, RASGRP1, PTPN6, CD3D, CD3E, CD3G, CD247, CD4, MAPK14, PPP3CA, ICOS, MAPK12, PTPRC, PPP3CB, PPP3CC, CD8A, PPP3R1, CD8B, LAT, NFATC1, NFATC2, NFATC3, NFATC4, PIK3CA, PIK3CB, CD28, PIK3CD, PIK3CG, PIK3R1, NFKB1, CHP2, NFKBIA, NFKBIB, NFKBIE, GRAP2, HRAS, PAK1, PAK2, DLG1, AKT1, AKT2, CARD11, BCL10, VAV3, TNF, PRKCQ, PLCG1, MAPK1, MAPK3, IKBKB, <br> MAPK11, MAPK9, MAPK13, MAP2K1, MAP3K7, NFAT5, MAP2K2, CDC42, FYN, MAP2K7, PIK3R5, VAV1, VAV2, NCK2, CDK4, CHP1, SOS1, SOS2 |


| 83059 | mTOR signaling pathway | BioSystems: KEGG | 2.21 | 5.30 | 5.35 | 131 / 151 | FNIP2, MAPKAP1, KRAS, RAF1, CAB39, SEH1L, ATP6V1A, ATP6V1B1, DDIT4, ATP6V1B2, ATP6V1C1, AKT3, <br> ATP6V1E1, RNF152, LAMTOR2, NRAS, DEPTOR, FZD3, SLC38A9, SGK1, LAMTOR5, WNT3, PDPK1, RPS6, <br> WNT5A, RPS6KA1, RPS6KA2, WNT7B, ATP6V1E2, RPS6KB1, RPS6KB2, <br> PIK3R3, WNT10B, TBC1D7, WNT2B, INSR, WNT9A, PRR5, DVL1, DVL2, DVL3, GRB2, GRB10, ATP6V1G1, RICTOR, TSC1, ATP6V1F, TSC2, IRS1, WNT4, PTEN, SKP2, CLIP1, AKT1S1, WDR59, GSK3B, SLC3A2, RHEB, RRAGC, CHUK, FZD1, FZD4, RHOA, FZD6, FZD7, WNT5B, FZD8, FZD9, LPIN1, STK11, STRADB, NPRL2, ATP6V1H, IGF1R, LAMTOR4, RRAGD, BRAF, TELO2, PIK3CA, MTOR, PIK3CB, PIK3CD, STRADA, RRAGA, PIK3R1, FZD5, RPTOR, WNT3A, ATP6V1D, EIF4B, WNT16, EIF4E, PRKAA1, EIF4EBP1, PRKAA2, HRAS, NPRL3, MIOS, WNT10A, FLCN, LRP6, LRP5, LAMTOR3, PRKCA, FNIP1, PRKCB, TTI1, SLC7A5, AKT1, CAB39L, AKT2, DEPDC5, SESN2, TNF, ATP6V1C2, ULK1, MAPK1, MAPK3, TNFRSF1A, LAMTOR1, IKBKB, MLST8, MAP2K1, MAP2K2, FZD2, ULK2, WDR24, SEC13, SOS1, EIF4E2, SOS2 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| M39602 | Neurodegeneration with brain iron accumulation (NBIA) subtypes pathway | MSigDB C2 BIOCARTA (v7.3) | 4.07 | 5.83 | 5.85 | 41 / 44 | ATG101, PIK3R4, DCAF17, ATG5, SCP2, <br> STK11, CP, MAP1LC3A, PLA2G6, ATG16L1, ATG14, TSC1, FTL, TSC2, GTPBP2, WIPI1, ATG10, PANK2, ULK1, COASY, ATG2A, RB1CC1, DEPTOR, ACACA, MLST8, ATG7, BECN1, ATG3, ATP13A2, PIK3C3, MTOR, AKT1S1, ATG13, FA2H, RPTOR, ATG12, WIPI2, C19orf12, RHEB, PRKAA1, SPTLC1 |
| Turquoise module (535 expressed genes) |  |  |  |  |  |  |  |


| ID | Name | Source | Mean Decrease in Accuracy | Median Expression (NMA) | Median Expression (MA) | \# of expressed genes / \# of total genes | Expressed genes |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 172846 | Staphylococcus aureus infection | BioSystems: KEGG | 1.16 | 5.03 | 4.67 | 44 / 56 | CFH, SELPLG, CFD, KRT10, FCGR1A, HLA-DMA, FCGR2A, HLA-DMB, FCGR2B, FCGR3A, FCGR3B, HLA-DOA, HLADPA1, HLA-DPB1, HLA-DQA1, HLADQA2, HLA-DQB1, HLA-DRA, HLA-DRB1, FPR1, FPR2, HLA-DRB5, ICAM1, FPR3, C1QA, C1QB, C1QC, C1R, C1S, C2, C3, C3AR1, C4A, C4B, C5, C5AR1, PLG, PTAFR, CFI, ITGAL, ITGAM, ITGB2, CFB, MASP2 |
| 200309 | Rheumatoid arthritis | BioSystems: KEGG | 1.87 | 5.20 | 5.05 | 67 / 90 | TGFB1, TGFB2, TGFB3, ATP6V1A, ATP6V1B1, JUN, ATP6V1B2, ATP6V1C1, IL15, ATP6V1E1, ATP6V0B, ATP6V1H, IL18, ATP6V0A1, ATP6V0D1, CSF1, ANGPT1, ATP6V0E1, HLA-DMA, HLADMB, TNFSF13, HLA-DOA, HLA-DPA1, HLA-DPB1, CD28, HLA-DQA1, CD86, HLA-DQA2, HLA-DQB1, TNFSF13B, FOS, HLA-DRA, HLA-DRB1, ATP6V1D, ATP6V1E2, ACP5, HLA-DRB5, ICAM1, ATP6V0E2, TLR2, ATP6V0A4, TLR4, TCIRG1, CCL2, CCL3, CCL3L1, <br> ATP6V1G1, CCL5, ATP6V1F, LTB, TNF, ATP6V1C2, MMP1, TNFRSF11A, CCL20, IL1B, ITGAL, CXCL6, CXCL5, CXCL1, CTSK, ITGB2, IL23A, CTSL, CXCL12, CXCL8, ATP6V0A2 |


| 213780 | Tuberculosis | BioSystems: KEGG | 1.00 | 5.28 | 5.21 | 142 / 179 | HSPD1, IL10RA, IL10RB, RAF1, MYD88, IL12A, MRC1, TRADD, AKT3, SPHK2, ATP6V0B, TRAF6, IL18, ATP6V0A1, CEBPB, CEBPG, CALM1, HLA-DMA, HLA-DMB, CALM2, HLA-DOA, CALM3, HLA-DPA1, CALML3, HLA-DPB1, HLADQA1, HLA-DQA2, HLA-DQB1, CAMK2B, CAMK2D, HLA-DRA, CAMK2G, HLADRB1, CAMP, HLA-DRB5, SRC, CYP27B1, BAD, APAF1, RIPK2, FADD, CASP3, ARHGEF12, CLEC7A, BAX, IRAK2, TCIRG1, CASP8, CASP9, CASP10, LAMP1, RELA, BCL2, TLR6, CALML5, CR1, ITGAM, ITGAX, RFX5, CREB1, ITGB2, IL23A, RFXAP, CALML6, CREBBP, CD209, STAT1, BID, NOD2, TGFB1, TGFB2, RHOA, IFNGR1, TGFB3, IFNGR2, JAK1, JAK2, KSR1, MALT1, PLA2R1, CLEC4E, CORO1A, ATP6V1H, MAPK14, ATP6V0D1, PPP3CA, PPP3CB, MAPK12, PPP3CC, PPP3R1, FCER1G, FCGR1A, CD14, FCGR2A, CIITA, FCGR2B, FCGR3A, FCGR3B, PIK3C3, CARD9, SPHK1, MRC2, TIRAP, RFXANK, NFKB1, TLR1, TLR2, ATP6VOA4, TLR4, CYCS, NFYA, NFYB, NFYC, SYK, RAB7A, CD74, C3, LSP1, AKT1, AKT2, BCL10, TNF, CALML4, MAPK1, MAPK3, EEA1, TNFRSF1A, MAPK8, MAPK11, IL1B, MAPK9, MAPK13, CTSD, NOS2, RAB5A, RAB5B, PLK3, CTSS, EP300, HSPA9, RAB5C, ATP6V0A2 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |


| 83078 | Hematopoietic cell lineage | BioSystems: KEGG | 3.38 | 4.37 | 3.97 | 70 / 97 | IL11RA, EPOR, ANPEP, HLA-DMA, HLADMB, HLA-DOA, HLA-DPA1, HLA-DPB1, HLA-DQA1, HLA-DQA2, HLA-DQB1, HLADRA, HLA-DRB1, HLA-DRB5, CD55, ITGA6, ITGA1, ITGA2, ITGA3, ITGA4, ITGA5, CR1, ITGAM, TFRC, CD1A, CD1C, CD1D, CD1E, CD2, CD3D, CD3E, CD3G, CD4, CD5, CSF1, CD7, CSF1R, CD8A, CD8B, KITLG, CD9, CSF3, CSF3R, FCGR1A, CD14, CD19, MS4A1, CD22, IL1R2, CD33, CD34, CD36, CD37, CD38, CD44, CD24, CD59, TNF, MME, IL1B, IL1R1, KIT, IL4R, IL5RA, IL6R, IL7, IL7R, GP1BA, GP5, GP9 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| M13950 | Asthma | MSigDB C2 BIOCARTA (v7.3) | 1.74 | 6.62 | 6.27 | 17 / 30 | TNF, FCER1A, MS4A2, FCER1G, EPX, HLA-DMA, HLA-DMB, HLA-DOA, HLADPA1, HLA-DPB1, HLA-DQA1, HLADQA2, HLA-DQB1, HLA-DRA, HLA-DRB1, HLA-DRB5, CD40 |
| 1469482 | Th17 cell differentiation | BioSystems: KEGG | 1.49 | 5.53 | 5.48 | 87 / 107 | IL12RB1, HIF1A, RARA, HLA-DMA, HLADMB, HLA-DOA, HLA-DPA1, HLA-DPB1, HLA-DQA1, HLA-DQA2, HLA-DQB1, FOS, HLA-DRA, HLA-DRB1, HLA-DRB5, TBX21, GATA3, IRF4, RELA, LCK, <br> RUNX1, IL23A, ZAP70, RXRA, RXRB, STAT1, STAT3, STAT5A, STAT5B, STAT6, CHUK, TGFB1, TYK2, IFNGR1, IFNGR2, JAK1, JAK2, JAK3, TGFBR1, TGFBR2, JUN, CD3D, CD3E, CD3G, CD247, CD4, MAPK14, PPP3CA, <br> PPP3CB, MAPK12, PPP3CC, PPP3R1, LAT, NFATC1, NFATC2, NFATC3, MTOR, NFKB1, IL21R, NFKBIA, NFKBIB, NFKBIE, AHR, RORA, RORC, PRKCQ, PLCG1, MAPK1, MAPK3, IKBKB, MAPK8, MAPK11, IL1B, MAPK9, IL1R1, MAPK13, IL1RAP, IL2RB, IL4R, IL6R, IL6ST, SMAD2, SMAD3, HSP90AA1, SMAD4, IL27RA, HSP90AB1 |


| 169642 | Toxoplasmosis | BioSystems: KEGG | 4.44 | 5.71 | 5.63 | 96 / 113 | IL10RA, IL10RB, MYD88, IL12A, AKT3, TRAF6, HLA-DMA, HLA-DMB, HLA-DOA, HLA-DPA1, HLA-DPB1, HLA-DQA1, HLADQA2, HLA-DQB1, HLA-DRA, PDPK1, HLA-DRB1, HLA-DRB5, BAD, LAMA1, CASP3, LAMA2, LAMA3, LAMA4, ITGA6, LAMA5, LAMB1, LAMB2, BIRC2, CASP8, BIRC3, CASP9, LAMB3, LAMC1, LAMC2, <br> TAB2, RELA, BCL2, BCL2L1, LY96, <br> ITGB1, LDLR, STAT1, STAT3, PPIF, CHUK, TGFB1, TYK2, TGFB2, IFNGR1, TGFB3, IFNGR2, JAK1, JAK2, MAPK14, MAPK12, PIK3R6, CIITA, PIK3CG, NFKB1, NFKBIA, TLR2, NFKBIB, TLR4, CYCS, CD40, SOCS1, AKT1, AKT2, CCR5, GNAI1, GNAI2, TNF, GNAI3, <br> TAB1, GNAO1, MAPK1, MAPK3, TNFRSF1A, IKBKB, MAPK8, MAPK11, MAPK9, MAPK13, MAP3K7, MAP2K3, HSPA1A, MAP2K6, HSPA1B, HSPA1L, HSPA2, NOS2, PIK3R5, HSPA6, HSPA8, ALOX5 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |


| 217173 | Influenza A | BioSystems: KEGG | 1.86 | 5.74 | 5.69 | 137 / 173 | RAF1, MYD88, IL12A, DNAJB1, AK MAP2K4, IL18, TRIM25, RAE1, <br> TMPRSS13, DDX58, HLA-DMA, HLADMB, TNFSF10, HLA-DOA, HLA-DPA1, <br> HLA-DPB1, CXCL10, HLA-DQA1, HLADQA2, HLA-DQB1, HLA-DRA, HLA-DRB1, <br> TICAM1, NLRX1, HLA-DRB5, ICAM1, PIK3R3, ACTB, SOCS3, NUP98, CASP1, ACTG1, TMPRSS4, CASP9, OAS1, OAS2, OAS3, IRF3, IRF7, RELA, TNFRSF10D, XPO1, TNFRSF10C, TNFRSF10B, HNRNPUL1, TNFRSF10A, FAS, FASLG, ADAR, ATF2, CREBBP, GSK3B, STAT1, STAT2, NLRP3, IFNAR1, IFNAR2, IVNS1ABP, TYK2, IFNGR1, IFNGR2, JAK1, JAK2, IFIH1, IRF9, JUN, CPSF4, RSAD2, MAPK14, RNASEL, MAPK12, MAVS, CIITA, IKBKE, PIK3CA, PABPN1, PIK3CB, EIF2S1, PIK3CD, PIK3R1, FDPS, NXT1, PYCARD, FURIN, NFKB1, TBK1, NFKBIA, NFKBIB, TLR3, TLR4, CYCS, AGFG1, TMPRSS2, PRKCA, PRKCB, CCL2, AKT1, CCL5, AKT2, TNF, EIF2AK4, MAPK1, MAPK3, TNFRSF1A, PLG, EIF2AK1, IKBKB, MAPK8, MAPK11, IL1B, MAPK9, MAPK13, MAP2K1, MAP2K2, MAP2K3, HSPA1A, MAP2K6, HSPA1B, HSPA1L, MAP2K7, HSPA2, EIF2AK2, EIF2AK3, DNAJC3, HSPA6, DDX39B, HSPA8, IL33, EP300, NXF1, MX1, CXCL8, VDAC1, PML, KPNA1, KPNA2 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| M615 | Intestinal immune network for IgA production | MSigDB C2 BIOCARTA (v7.3) | 2.18 | 4.74 | 4.53 | $30 / 48$ | TGFB1, TNFRSF13C, IL15, IL15RA, LTBR, ICOS, ITGA4, CCL28, HLA-DMA, PIGR, TNFSF13, HLA-DMB, HLA-DOA, HLA-DPA1, HLA-DPB1, CD28, CXCR4, <br> HLA-DQA1, CD86, HLA-DQA2, <br> MADCAM1, HLA-DQB1, ITGB7, <br> TNFSF13B, HLA-DRA, CXCL12, HLADRB1, HLA-DRB5, MAP3K14, CD40 |


| 1470924 | Interleukin-10 <br> signaling <br> BioSystems: <br> REACTOME |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |


| 193147 | Osteoclast differentiation | BioSystems: KEGG | 1.09 | 4.78 | 4.64 | 110 / 130 | CYLD, AKT3, LILRB3, TRAF2, LILRA2, TRAF6, SPI1, CAMK4, LILRB2, FOS, FOSB, FOSL2, ACP5, PIK3R3, NCF1, MAP3K14, SOCS3, OSCAR, BLNK, LILRA6, GRB2, TAB2, NCF2, NCF4, RELA, RELB, SIRPB1, SIRPA, TNFRSF11A, LCK, PPARG, TEC, LCP2, CREB1, LILRB1, STAT1, STAT2, <br> TNFRSF11B, GAB2, CHUK, IFNAR1, IFNAR2, SIRPG, TGFB1, TYK2, TGFB2, IFNGR1, IFNGR2, JAK1, TGFBR1, TGFBR2, TYROBP, IRF9, JUN, JUNB, JUND, MAPK14, PPP3CA, CSF1, <br> PPP3CB, MAPK12, CSF1R, PPP3CC, PPP3R1, FCGR1A, NFATC1, FCGR2A, NFATC2, FCGR2B, FCGR3A, FCGR3B, PIK3CA, PIK3CB, PIK3CD, SQSTM1, PIK3R1, NFKB1, NFKB2, NFKBIA, MITF, TREM2, SYK, SOCS1, AKT1, AKT2, TNF, <br> TAB1, PLCG2, MAPK1, MAPK3, <br> TNFRSF1A, IKBKB, MAPK8, MAPK11, <br> IL1B, MAPK9, FHL2, IL1R1, MAPK13, <br> MAP2K1, MAP3K7, FYN, MAP2K6, CTSK, MAP2K7, LILRB5, LILRA5, RAC1, LILRB4, CYBA |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1269330 | TNF receptor superfamily (TNFSF) members mediating noncanonical NF-kB pathway | BioSystems: REACTOME | 2.16 | 4.05 | 3.99 | 14 / 18 | TNFRSF12A, TNFRSF13C, TNFSF14, TNFSF12, BIRC2, BIRC3, TNFSF13B, LTB, TRAF2, TRAF3, LTBR, TNFRSF11A, MAP3K14, CD40 |
| 1269174 | Translocation of ZAP-70 to Immunological synapse | BioSystems: REACTOME | 1.64 | 5.13 | 4.57 | 17 / 22 | HLA-DPA1, HLA-DPB1, HLA-DQA1, HLADQA2, ZAP70, HLA-DQB1, PTPN22, HLADQB2, HLA-DRA, CD3D, HLA-DRB1, CD3E, CD3G, HLA-DRB5, CD247, CD4, LCK |


| 137998 | TCR signaling in naive CD4+ T cells | BioSystems: Pathway Interaction Database | 3.93 | 4.77 | 4.64 | 56 / 60 | STIM1, RASSF5, MALT1, RASGRP1, ORAI1, PAG1, PTPN6, RAP1A, CD3D, CD3E, CD3G, TRAF6, PTPN11, CD247, CD4, PTPRC, CSK, CD28, CD86, MAP3K8, MAP4K1, HLA-DRA, PDPK1, HLA-DRB1, SH3BP2, GRAP2, MAP3K14, DBNL, SHC1, GRB2, PRKCA, PRKCB, PRKCE, SLA2, AKT1, CARD11, NCK1, BCL10, STK39, PRKCQ, PLCG1, LCK, IKBKB, PTEN, LCP2, CBL, FYB1, CDC42, FYN, ZAP70, VAV1, ITK, GAB2, CHUK, RASGRP2, SOS1 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Yellow module (675 expressed genes) |  |  |  |  |  |  |  |
| ID | Name | Source | Mean Decrease in Accuracy | Median <br> Expression <br> (NMA) | Median Expression (MA) | \# of expressed genes / \# of total genes | Expressed genes |
| 1268690 | Eukaryotic Translation Elongation | BioSystems: REACTOME | 4.27 | 6.75 | 6.65 | 83 / 98 | RPL21, RPL22, RPL23A, RPL24, RPL26, <br> RPL27, RPL30, RPL27A, RPL28, RPL29, <br> RPL31, RPL36, RPL32, RPL34, RPL35A, <br> RPL36AL, RPL37, RPL37A, RPL38, <br> RPL41, RPLPO, RPLP1, RPLP2, RPS2, <br> RPS3, RPS3A, RPS5, RPS6, RPS7, <br> RPS8, RPS9, RPS11, RPS12, RPS13, RPS14, RPS15, RPS15A, RPS16, RPS17, <br> RPS18, RPS19, RPS20, RPSA, RPS21, <br> RPS23, RPS24, RPL14, RPS25, RPS26, <br> RPS27, RPS27A, RPS28, RPS29, <br> RPL39L, RPS27L, EEF1A1, RPL10A, <br> RPL23, EEF1B2, UBA52, EEF1D, EEF2, <br> FAU, RPL26L1, RPL22L1, RPL35, <br> RPL13A, RPL3, RPL4, RPL5, RPL6, <br> RPL7, RPL7A, RPL8, RPL9, RPL11, <br> RPL12, RPL13, RPL15, RPL17, RPL18, <br> RPL18A, RPL19 |


| M39495 | Cytoplasmic Ribosomal Proteins | MSigDB C2 BIOCARTA (v7.3) | 4.3 | 6.78 | 6.69 | 79 / 90 | RPL10A, RPL21, RPL22, RPL23A, RPL23, RPL24, RPL26, RPL27, RPL30, RPL27A, RPL28, RPL29, UBA52, RPL31, RPL36, <br> RPL32, RPL34, FAU, RPL35A, RPL37, RPL37A, RPL38, RPL41, RPLP0, RPLP1, RPLP2, RPS2, RPS3, RPS3A, RPS5, RPS6, RPS6KA1, RPS6KA2, RPS6KB1, RPS6KB2, RPS7, RPS8, RPS9, RPS11, RPS12, RPS13, RPS14, RPS15, RPS15A, MRPL19, RPS16, RPS17, RPS18, RPS19, RPS20, RPSA, RPS21, RPS23, RPS24, RPL14, RPS25, RPS26, RPS27, RPL35, RPS27A, RPS28, RPS29, RPL13A, RPL3, RPL4, RPL5, RPL6, RPL7, RPL7A, RPL8, RPL9, RPL11, RPL12, RPL13, RPL15, RPL17, RPL18, RPL18A, RPL19 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1268691 | Peptide chain elongation | BioSystems: REACTOME | 2.31 | 6.79 | 6.69 | 81 / 93 | RPL10A, RPL21, RPL22, RPL23A, RPL23, RPL24, RPL26, RPL27, RPL30, RPL27A, RPL28, RPL29, UBA52, RPL31, RPL36, RPL32, EEF2, RPL34, FAU, RPL35A, RPL36AL, RPL37, RPL37A, RPL38, RPL41, RPLP0, RPLP1, RPLP2, RPS2, RPS3, RPS3A, RPS5, RPL26L1, RPS6, RPS7, RPS8, RPS9, RPS11, RPS12, RPS13, RPS14, RPS15, RPS15A, RPS16, RPS17, RPS18, RPS19, RPS20, RPSA, RPS21, RPS23, RPS24, RPL14, RPS25, RPL22L1, RPS26, RPL35, RPS27, RPS27A, RPS28, RPS29, RPL39L, RPL13A, RPL3, RPL4, RPL5, RPL6, RPL7, RPL7A, RPL8, RPL9, RPL11, RPL12, RPL13, RPS27L, RPL15, EEF1A1, RPL17, RPL18, RPL18A, RPL19 |


| 1268681 | Formation of a pool of free 40S subunits | BioSystems: REACTOME | 1.62 | 6.7 | 6.63 | 92 / 107 | RPL21, RPL22, RPL23A, RPL24, RPL26, <br> RPL27, RPL30, RPL27A, RPL28, RPL29, <br> RPL31, RPL36, RPL32, RPL34, RPL35A, <br> RPL36AL, RPL37, RPL37A, RPL38, <br> RPL41, RPLPO, RPLP1, RPLP2, RPS2, <br> RPS3, RPS3A, RPS5, RPS6, RPS7, <br> RPS8, RPS9, RPS11, EIF3E, RPS12, RPS13, RPS14, RPS15, RPS15A, RPS16, <br> RPS17, RPS18, RPS19, RPS20, RPSA, <br> RPS21, RPS23, RPS24, RPL14, RPS25, RPS26, RPS27, RPS27A, RPS28, RPS29, <br> RPL39L, RPS27L, RPL10A, RPL23, <br> UBA52, FAU, RPL26L1, EIF3L, EIF3K, <br> EIF3A, EIF3B, RPL22L1, EIF3C, EIF3D, RPL35, EIF3F, EIF3G, EIF3H, EIF3I, <br> EIF3J, RPL13A, RPL3, RPL4, RPL5, RPL6, EIF3M, RPL7, RPL7A, RPL8, RPL9, RPL11, RPL12, RPL13, RPL15, RPL17, RPL18, RPL18A, RPL19 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1268692 | Eukaryotic <br> Translation <br> Termination | BioSystems: REACTOME | 2.91 | 6.68 | 6.63 | 84 / 97 | RPL21, RPL22, RPL23A, RPL24, RPL26, <br> RPL27, RPL30, RPL27A, RPL28, RPL29, RPL31, RPL36, RPL32, RPL34, RPL35A, RPL36AL, RPL37, RPL37A, RPL38, <br> RPL41, RPLPO, RPLP1, RPLP2, RPS2, RPS3, RPS3A, TRMT112, RPS5, RPS6, RPS7, RPS8, ETF1, RPS9, RPS11, RPS12, RPS13, RPS14, RPS15, RPS15A, APEH, RPS16, RPS17, RPS18, RPS19, RPS20, RPSA, RPS21, RPS23, RPS24, RPL14, RPS25, RPS26, RPS27, RPS27A, RPS28, RPS29, RPL39L, GSPT1, RPS27L, RPL10A, RPL23, UBA52, FAU, N6AMT1, RPL26L1, RPL22L1, RPL35, RPL13A, RPL3, RPL4, RPL5, RPL6, RPL7, RPL7A, RPL8, RPL9, RPL11, RPL12, RPL13, RPL15, RPL17, RPL18, RPL18A, RPL19 |


| M189 | Ribosome | MSigDB C2 BIOCARTA (v7.3) | 3.73 | 6.68 | 6.59 | 79 / 88 | RPL21, RPL10A, RPL22, RPL23A, RPL23, RPL24, RPL26, RPL27, RPL30, RPL27A, RPL28, RPL29, UBA52, RPL31, RPL32, RPL36, RPL34, RPL35A, FAU, RPL36AL, RPL37, RPL37A, RPL38, RPL41, RPLPO, RPLP1, RPLP2, RPS2, RPS3, RPS3A, RPS5, RPL26L1, RPS6, RPS7, RPS8, RPS9, RPS11, RPS12, RPS13, RPS15, RPS15A, MRPL13, RPS16, RPS17, RPS18, RPS19, RPS20, RPSA, RPS21, RPS23, RPL14, RPS24, RPS25, RPL22L1, RPS26, RPL35, RPS27, RPS27A, RPS28, RPS29, RPL13A, RPL3, RPL4, RPL5, RPL6, RPL7, RPL7A, RSL24D1, RPL8, RPL9, RPL11, RPL12, RPS27L, RPL13, RPL15, RPL17, RPL18, RPL18A, RPL19 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1269718 | Nonsense Mediated Decay (NMD) independent of the Exon Junction Complex (EJC) | BioSystems: REACTOME | 1.48 | 6.77 | 6.68 | 86 / 100 | RPL21, RPL22, RPL23A, RPL24, RPL26, <br> RPL27, RPL30, RPL27A, RPL28, RPL29, <br> RPL31, RPL36, RPL32, RPL34, RPL35A, <br> RPL36AL, RPL37, RPL37A, RPL38, <br> RPL41, RPLPO, RPLP1, RPLP2, RPS2, <br> RPS3, RPS3A, RPS5, RPS6, RPS7, <br> RPS8, ETF1, RPS9, RPS11, RPS12, <br> RPS13, RPS14, RPS15, RPS15A, RPS16, <br> RPS17, NCBP1, RPS18, RPS19, RPS20, <br> RPSA, RPS21, RPS23, RPS24, RPL14, <br> RPS25, RPS26, UPF1, RPS27, RPS27A, <br> RPS28, RPS29, RPL39L, PABPC1, <br> GSPT1, RPS27L, RPL10A, NCBP2, <br> RPL23, UBA52, FAU, RPL26L1, EIF4G1, RPL22L1, RPL35, RPL13A, RPL3, RPL4, RPL5, RPL6, RPL7, RPL7A, RPL8, RPL9, RPL11, RPL12, RPL13, RPL15, RPL17, RPL18, RPL18A, RPL19 |


| 1268686 | GTP hydrolysis and joining of the 60S ribosomal subunit | BioSystems: REACTOME | 2.78 | 6.63 | 6.60 | 102 / 119 | RPL21, RPL22, RPL23A, RPL24, RPL26, RPL27, RPL30, RPL27A, RPL28, RPL29, RPL31, RPL36, RPL32, RPL34, RPL35A, RPL36AL, RPL37, RPL37A, RPL38, RPL41, RPLPO, RPLP1, EIF4H, RPLP2, RPS2, RPS3, RPS3A, RPS5, RPS6, RPS7, RPS8, RPS9, RPS11, EIF3E, RPS12, RPS13, RPS14, RPS15, RPS15A, RPS16, RPS17, RPS18, RPS19, RPS20, RPSA, RPS21, RPS23, RPS24, RPL14, RPS25, RPS26, RPS27, RPS27A, RPS28, RPS29, RPL39L, RPS27L, RPL10A, RPL23, UBA52, FAU, EIF2S1, RPL26L1, EIF4A1, EIF4A2, EIF4B, EIF4E, EIF3L, EIF4G1, EIF2S2, EIF5, EIF5B, EIF3K, EIF3A, EIF3B, RPL22L1, EIF3C, EIF3D, RPL35, EIF3F, EIF3G, EIF3H, EIF3I, EIF3J, RPL13A, RPL3, RPL4, RPL5, RPL6, EIF3M, RPL7, RPL7A, RPL8, RPL9, RPL11, RPL12, RPL13, RPL15, RPL17, RPL18, RPL18A, RPL19 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |


| 1339149 | Selenoamino acid metabolism | BioSystems: REACTOME | 1.18 | 6.57 | 6.50 | 106 / 123 | RPL21, RPL22, RPL23A, EEFSEC, RPL24, RPL26, EPRS1, RPL27, RPL30, RPL27A, RPL28, RPL29, RPL31, RPL36, RPL32, RPL34, RPL35A, RPL36AL, RPL37, RPL37A, RPL38, RPL41, AIMP2, RARS1, RPLP0, RPLP1, RPLP2, AIMP1, RPS2, RPS3, MARS1, RPS3A, MAT1A, IARS1, RPS5, EEF1E1, RPS6, RPS7, RPS8, RPS9, RPS11, RPS12, RPS13, RPS14, LARS1, RPS15, RPS15A, RPS16, RPS17, RPS18, RPS19, DARS1, RPS20, RPSA, RPS21, RPS23, SCLY, RPS24, RPL14, RPS25, RPS26, RPS27, RPS27A, RPS28, RPS29, GNMT, RPL39L, PAPSS2, PAPSS1, HNMT, GSR, RPS27L, TXNRD1, RPL10A, RPL23, UBA52, PSTK, SEPSECS, FAU, KARS1, SARS1, RPL26L1, INMT, AHCY, SECISBP2, CTH, RPL22L1, RPL35, RPL13A, QARS1, RPL3, RPL4, RPL5, RPL6, RPL7, RPL7A, RPL8, RPL9, RPL11, RPL12, RPL13, RPL15, RPL17, RPL18, RPL18A, RPL19 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |


|  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |

RPL21, RPL22, RPL23A, RPL24, SMG1, RPL26, CASC3, RPL27, RPL30, RPL27A, RPL28, RPL29, RPL31, RPL36, RPL32, MAGOH, RPL34, RPL35A, RPL36AL, RPL37, RPL37A, RPL38, RPL41, RPLPO, RPLP1, RPLP2, RPS2, RPS3, RPS3A, EIF4A3, RPS5, RPS6, RPS7, RPS8, ETF1, RPS9, RPS11, RPS12, RPS13, RPS14, RPS 15, RPS15A, MAGOHB, RPS16, RPS17, PNRC2, NCBP1, RPS18, RPS19, RPS20, RPSA, RPS21, RPS23, RPS24, SMG5, RPL14, UPF3A, RPS25, RPS26, UPF1, RPS27, RPS27A, RPS28, RPS29, RPL39L, PABPC1, GSPT1, RPS27L, RPL10A, NCBP2, RPL23, PPP2CA, SMG8, PPP2R1A, UBA52, PPP2R2A, FAU, SMG7, UPF2, RNPS1, RPL26L1, EIF4G1, SMG9, RBM8A,
RPL22L1, RPL35, RPL13A, RPL3, RPL4, RPL5, RPL6, RPL7, RPL7A, RPL8, RPL9, RPL11, RPL12, RPL13, RPL15, DCP1A, RPL17, RPL18, SMG6, RPL18A, RPL19

| 1269115 | Influenza Viral RNA Transcription and Replication | BioSystems: REACTOME | 4.63 | 6.32 | 6.28 | 125 / 138 | RPL21, RPL22, IPO5, RPL23A, TPR, SEH1L, RPL24, RPL26, NUP50, RPL27, RPL30, RPL27A, RPL28, RPL29, RANBP2, RPL31, RPL36, RPL32, RPL34, RPL35A, RPL36AL, RPL37, RPL37A, RPL38, RPL41, RPLPO, NUP85, RPLP1, RAE1, NUP107, RPLP2, RPS2, RPS3, RPS3A, RPS5, RPS6, POLR2A, POLR2B, POLR2C, POLR2D, RPS7, RPS8, POLR2E, RPS9, POLR2G, POLR2H, RPS11, POLR2I, RPS12, NUP88, POLR2J, RPS13, NUP98, RPS14, POLR2K, RPS15, POLR2L, RPS15A, NUP43, RPS16, RPS17, RPS18, RPS19, RPS20, RPSA, RPS21, RPS23, NUP62, RPS24, NUP214, RPL14, RPS25, RPS26, RPS27, RPS27A, NUP42, RPS28, NUP58, RPS29, RPL39L, POM121C, GRSF1, NUP35, RPS27L, NUP54, NUP205, RPL10A, RPL23, UBA52, GTF2F1, GTF2F2, FAU, AAAS, NDC1, POM121, NUP155, NUP37, RPL26L1, NUP210, NUP133, RPL22L1, NUP188, NUP93, RPL35, RPL13A, RPL3, DNAJC3, RPL4, RPL5, NUP160, RPL6, RPL7, RPL7A, RPL8, NUP153, RPL9, RPL11, RPL12, HSP90AA1, RPL13, RPL15, RPL17, RPL18, RPL18A, RPL19 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |


|  |  |  |  |  |  |  |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| 1269109 | Influenza Life <br> Cycle | BioSystems: <br> REACTOME | 4.59 | 6.42 | 6.37 | $134 / 147$ |
|  |  |  |  |  |  |  |

RPL21, RPL22, IPO5, RPL23A, TPR, SEH1L, RPL24, RPL26, NUP50, RPL27, RPL30, RAN, RPL27A, RPL28, RPL29, RANBP2, RPL31, RPL36, RPL32, RPL34, RPL35A, RPL36AL, RPL37, RPL37A RPL38, RPL41, RPLP0, NUP85, RPLP1, RAE1, NUP107, RPLP2, CALR, RPS2, RPS3, RPS3A, RPS5, RPS6, CANX, POLR2A, POLR2B, POLR2C, POLR2D, RPS7, RPS8, POLR2E, RPS9, POLR2G, POLR2H, RPS11, POLR2I, RPS12, NUP88, POLR2J, RPS13, NUP98, RPS14, POLR2K, RPS15, POLR2L, RPS15A, NUP43, RPS16, RPS17, RPS18, RPS19, RPS20, RPSA, RPS21, RPS23, NUP62, RPS24, NUP214, RPL14, RPS25, RPS26, RPS27, RPS27A, NUP42, RPS28, NUP58, XPO1, RPS29, RPL39L, POM121C, GRSF1, NUP35, RPS27L, NUP54, NUP205, RPL10A, RPL23, UBA52, GTF2F1, GTF2F2, FAU, AAAS, NDC1, POM121, NUP155, NUP37, RPL26L1, NUP210, CLTA, CLTC, NUP133, RPL22L1, NUP188, NUP93, RPL35, RPL13A, HSPA1A, RPL3, DNAJC3, RPL4, RPL5, NUP160, RPL6, RPL7, RPL7A, RPL8, NUP153, RPL9, RPL11, RPL12, HSP90AA1, RPL13, RPL15, RPL17, KPNA1, RPL18, KPNB1, RPL18A, RPL19

| 1269108 | Influenza Infection | BioSystems: REACTOME | 5.35 | 6.30 | 6.27 | 144 / 158 | KPNA4, RPL21, KPNA5, RPL22, IPO5, RPL23A, TPR, SEH1L, RPL24, RPL26, NUP50, RPL27, RPL30, RAN, RPL27A, RPL28, RPL29, RANBP2, RPL31, RPL36, <br> RPL32, RPL34, RPL35A, RPL36AL, RPL37, RPL37A, RPL38, RPL41, RPLPO, NUP85, RPLP1, RAE1, NUP107, RPLP2, CALR, RPS2, RPS3, RPS3A, RPS5, RPS6, CANX, POLR2A, POLR2B, POLR2C, POLR2D, RPS7, RPS8, <br> POLR2E, RPS9, POLR2G, POLR2H, RPS11, POLR2I, RPS12, NUP88, POLR2J, RPS13, NUP98, RPS14, POLR2K, RPS15, POLR2L, RPS15A, NUP43, RPS16, RPS17, RPS18, RPS19, RPS20, RPSA, RPS21, RPS23, NUP62, RPS24, NUP214, RPL14, RPS25, RPS26, RPS27, RPS27A, NUP42, RPS28, NUP58, XPO1, RPS29, RPL39L, POM121C, GRSF1, NUP35, RPS27L, NUP54, NUP205, RPL10A, TGFB1, RPL23, KPNA7, UBA52, GTF2F1, CPSF4, GTF2F2, FAU, AAAS, NDC1, POM121, NUP155, ISG15, PABPN1, NUP37, RPL26L1, NUP210, CLTA, CLTC, NUP133, RPL22L1, NUP188, NUP93, RPL35, RPL13A, HSPA1A, EIF2AK2, RPL3, DNAJC3, RPL4, RPL5, NUP160, RPL6, RPL7, RPL7A, RPL8, NUP153, RPL9, RPL11, RPL12, HSP90AA1, RPL13, RPL15, RPL17, KPNA1, RPL18, KPNB1, KPNA2, RPL18A, KPNA3, RPL19 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |


| 1383085 | rRNA processing | BioSystems: REACTOME | 3.03 | 5.52 | 5.49 | 180 / 203 | RPL21, RPL22, DDX47, RPL23A, UTP6, RPL24, RPL26, RPL27, RPL30, RPL27A, RPL28, RPL29, RPL31, RPL32, RPL34, RPL35A, RPL36AL, RPL37, RPL37A, RPP21, RPL38, RPL41, RPLP0, RPLP1, RPLP2, FBL, RPS2, RPS3, RPS3A, RPP40, RPS5, RPS6, MRM1, SENP3, RPS7, RPS8, RPS9, NOL6, WDR46, RPS11, RPS12, BMS1, RPS13, RPS14, RPS15, RPS15A, PNO1, RPS16, RPS17, WDR18, RIOK3, EXOSC8, RPS18, RPS19, RPS20, RPS21, EXOSC5, <br> RPS23, RPS24, RPS25, RPS26, RPS27, RPS27A, RPS28, RPS29, RPL39L, <br> TEX10, ELAC2, WDR43, RPL10A, RPP25, RPL23, WDR3, UBA52, TRMT10C, FAU, DIMT1, WDR75, GAR1, UTP15, RRP36, IMP4, EXOSC6, PWP2, BYSL, EMG1, C1D, SNU13, NOC4L, UTP20, RRP7A, BOP1, NOP10, RPL22L1, UTP3, TSR3, NOP2, EXOSC4, MRM2, NSUN4, RPL36, EXOSC9, EXOSC10, XRN2, RIOK1, DCAF13, DDX49, WDR36, NOP56, DHX37, DDX52, TRMT112, NOB1, RPP30, RPP38, EXOSC3, EXOSC1, NOL11, THUMPD1, RPSA, RPL14, <br> HEATR1, NOL9, ERI1, RPP14, KRR1, NHP2, MTERF4, EXOSC2, TBL3, RPS27L, BUD23, FCF1, MRM3, NOP58, UTP18, NOP14, UTP25, TFB1M, PELP1, TSR1, CSNK1D, UTP11, CSNK1E, RRP9, RPL26L1, UTP4, PES1, NAT10, RCL1, PDCD11, WDR12, ISG20L2, LTV1, MPHOSPH10, MPHOSPH6, RPL35, PRORP, MTREX, RPL13A, RIOK2, EXOSC7, IMP3, RPL3, RPL4, RPL5, RPL6, RPL7, RPL7A, RPL8, RPL9, RPL11, RPL12, RPL13, RPL15, RPL17, UTP14C, RPL18, RPL18A, RPL19 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |


| 1270158 | Metabolism of amino acids and derivatives | BioSystems: REACTOME | 8.68 | 5.93 | 5.88 | 288 / 367 | RPL21, RPL22, RPL23A, TPO, RPL24 HGD, RPL26, EPRS1, RPL27, RPL30, RPL27A, RPL28, PHYKPL, RPL29, RPL31, RPL32, RPL34, RPL35A, RPL36AL, RPL37, RPL37A, RPL38, RPL41, GLS2, RPLPO, RPLP1, GAMT, ACADSB, IDO1, AUH, RPLP2, ACAT1, AIMP1, PDHB, RPS2, RPS3, MARS1, RPS3A, MAT1A, RPS5, PSMA1, RPS6, PSMA2, PSMA3, PSMA4, PSMA5, <br> PSMA6, BCKDK, PSMA7, RPS7, PSMB1, PSMB2, RPS8, RPS9, PSMB3, PSMB4, PSMB5, RPS11, PSMB6, RPS12, PSMB7, RPS13, NMRAL1, RPS14, PSMB8, RPS15, PSMB9, RPS15A, SRM, PSMB10, PSMC1, GATM, PIPOX, PSMC2, PSMC3, PSMC4, PSMC5, RPS16, BCAT1, PSMC6, RPS17, PSMD1, BCAT2, PSMD2, PSMD3, PSMD4, RPS18, GCDH, RPS19, DARS1, PSMD5, RPS20, PSMD7, BCKDHA, SERINC2, BCKDHB, PSMD8, PSMD9, RPS21, RPS23, RPS24, PSMD11, PSMD12, RPS25, PSMD13, MCCC2, UROC1, RPS26, PSME1, RPS27, RPS27A, PSME2, RPS28, MCCC1, PSPH, RPS29, DBT, TST, GNMT, RPL39L, NDUFAB1, DCT, HNMT, SQOR, DDC, ENOPH1, PHGDH, NAGS, CKB, TXNRD1, IVD, RPL10A, AFMID, PAOX, RPL23, PSMD6, CKMT1B, FAH, CKMT2, UBA52, DUOX1, FAU, KARS1, SARS1, PSME4, HIBCH, GRHPR, SUOX, GCLC, GCLM, HPD, SERINC1, CSAD, GLS, GLUD1, AHCY, GLUL, NQO1, DIO1, DIO2, PYCR1, ALDH18A1, DLAT, <br> SECISBP2, DLD, SERINC3, LIPT2, DLST, RPL22L1, ALDH9A1, PYCR2, SMOX, QARS1, CARNS1, GPT2, QDPR, DHTKD1, IL4I1, BBOX1, ALDH6A1, KYNU, PYCR3, TAT, GOT1, GOT2, |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |


|  |  |  |  |  |  |  | CKMT1A, EEFSEC, LIAS, PSAT1, RPL36, NAALAD2, PSMF1, MRI1, ASRGL1, <br> AIMP2, RARS1, HYKK, FOLH1, AZIN2, SEM1, ASPG, IARS1, EEF1E1, GPT, IYD, LARS1, SHMT1, DUOX2, OAT, DDO, RPSA, OAZ1, OAZ2, OCA2, SCLY, RPL14, ODC1, PPM1K, PAPSS2, <br> PAPSS1, OGDH, HIBADH, PDHX, KYAT1, KMO, GSR, SLC3A2, RPS27L, AZIN1, <br> ARG2, APIP, GSTZ1, SLC6A12, LIPT1, PSTK, SEPSECS, CRYM, ACAD8, MTAP, <br> ADO, GCAT, AASS, RPL26L1, INMT, <br> ETHE1, ASL, SLC25A15, ASNS, <br> SLC25A10, ASS1, CNDP2, MTR, AGMAT, <br> MTRR, HAAO, KYATЗ, SLC7A5, <br> ALDH4A1, CTH, PSME3, CARNMT1, <br> RPL35, ADI1, SERINC5, AADAT, RPL13A, <br> PCBD1, PSMD14, OAZ3, RPL3, RPL4, <br> RPL5, RPL6, RPL7, RPL7A, RPL8, <br> ALDH7A1, RPL9, RPL11, RPL12, RPL13, RPL15, HDC, RPL17, RPL18, RPL18A, <br> TPH1, RPL19 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |


| 1269056 | Infectious disease | BioSystems: REACTOME | 3.17 | 6.06 | 6.05 | 349 / 393 | RPL21, CBLL1, CDK9, RPL22, RPL23A, TPR, SEH1L, RPL24, RPL26, RPL27, <br> RPL30, EPS15, RPL27A, RPL28, STAM2, RPL29, RPL31, RPL32, ERCC2, RPL34, <br> RPL35A, RPL36AL, ERCC3, RPL37, <br> RPL37A, RPL38, RPL41, RPLPO, NUP85, RPLP1, HLA-A, RPLP2, RPS2, RPS3, RPS3A, RPS5, RPS6, RPS7, RPS8, RPS9, RPS11, RPS12, RPS13, RPS14, RPS15, RPS15A, RPS16, APOBEC3G, RPS17, RPS18, RPS19, RCC1, RPS20, RPS21, TSG101, RPS23, NUP62, RPS24, RPS25, HMGA1, RPS26, RPS27, RPS27A, RPS28, RPS29, RPL39L, NUP54, TXNRD1, RPL23, MET, KPNA7, UBA52, UBB, UBC, AP2M1, FAU, AP1S1, AP2S1, AP2A1, AP2A2, AP1B1, AP2B1, AP1G1, NUP37, NMT2, CLTA, FEN1, CLTC, GTF2H5, ANTXR1, CCR5, RPL22L1, HSPA1A, MNAT1, HSP90AA1, HSP90AB1, CHMP4C, MAP2K4, RPL36, PSMF1, VPS37A, RAE1, SLC25A4, AP1S3, NELFCD, NELFA, VPS4B, POLR2A, CHMP5, POLR2B, SH3GL1, POLR2C, POLR2D, SH3GL3, POLR2E, POLR2G, POLR2H, POLR2I, POLR2J, NELFB, POLR2K, POLR2L, VTA1, XPO1, XRCC4, XRCC5, SKP1, POM121C, PPIA, ARF1, NUP35, CHMP2B, NCBP2, PACS1, ATP6V1H, CHMP4A, NDC1, NUP155, ISG15, NUP133, CHMP3, NUP93, <br> CTNNB1, CTNND1, MAP2K1, MAP2K2, <br> FYN, MAP2K3, MAP2K6, MAP2K7, <br> EIF2AK2, DNAJC3, XRCC6, NUP50, <br> RNMT, RNGTT, VPS4A, PSMA1, PSMA2, PSMA3, PSMA4, PSMA5, B2M, PSMA6, PSMA7, PSMB1, PSMB2, SRC, PSMB3, PSMB4, PSMB5, PSMB6, PSMB7, PSMB8, PSMB9, PSMB10, PSMC1, PSMC2, PSMC3, PSMC4, PSMC5, |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |


|  |  |  |  |  |  |  | PSMC6, PSMD1, PSMD2, PSMD3, PSMD4, NCBP1, PSMD5, PSMD7, PSMD8, PSMD9, PSMD11, PSMD12, PSMD13, PSME1, PSME2, NUP58, SSRP1, CHMP2A, CCNK, BANF1, ELMO1, NUP205, RPL10A, PSMD6, CHMP7, CPSF4, STX1A, POM121, ANTXR2, PSME4, SV2B, SUPT4H1, CXCR4, SV2A, SUPT5H, NUP210, VAMP1, VAMP2, SYT1, AP1M1, TAF2, TAF4, TAF4B, TAF5, TAF6, TAF7, TAF9, TAF10, TAF11, TAF12, NMT1, TAF13, NUP160, BTRC, VPS37D, NUP153, RAC1, RBX1, KPNA1, TBP, KPNB1, KPNA2, KPNA3, KPNA4, NELFE, KPNA5, DOCK2, IPO5, NPM1, TCEA1, VPS37C, ELOC, ELOB, ELOA, RAN, RANBP1, RANBP2, RANGAP1, CHMP6, NEDD4L, PDCD6IP, CALM1, NUP107, CALM2, CALM3, SEM1, CALR, HBEGF, CANX, NUP88, NUP98, GRB2, AP1M2, NUP43, RPSA, NUP214, RPL14, SYT2, NUP42, STAM, LCK, CBL, CHMP4B, VPS37B, GRSF1, RPS27L, TGFB1, CUL5, CCNH, CCNT1, CCNT2, LIG1, GTF2A1, LIG4, GTF2A2, GTF2B, GTF2E1, GTF2E2, EEF2, GTF2F1, GTF2F2, GTF2H1, TAF3, GTF2H2, AAAS, CD247, GTF2H3, CD4, CD8B, CD9, PSIP1, GUCY2C, PABPN1, CD28, RPL26L1, FURIN, HGS, SUPT16H, CTDP1, PAK2, PSME3, NUP188, RPL35, VPS28, LTF, RPL13A, PSMD14, CDH1, PDZD3, RPL3, RPL4, RPL5, HCK, RPL6, RPL7, ELL, RPL7A, RPL8, RPL9, RPL11, RPL12, RPL13, RPL15, RPL17, RPL18, CDK7, RPL18A, RPL19 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |


| 1268685 | Ribosomal scanning and start codon recognition | BioSystems: REACTOME | 1.47 | 6.48 | 6.46 | $53 / 63$ | FAU, EIF4H, RPS2, RPS3, EIF2S1, RPS3A, RPS5, RPS6, EIF4A1, EIF4A2, EIF4B, EIF4E, RPS7, RPS8, EIF3L, RPS9, EIF4G1, RPS11, EIF3E, RPS12, EIF2S2, RPS13, EIF5, RPS14, RPS15, RPS15A, EIF3K, RPS16, RPS17, RPS18, RPS19, RPS20, RPSA, RPS21, RPS23, RPS24, EIF3A, EIF3B, RPS25, RPS26, EIF3C, EIF3D, RPS27, RPS27A, EIF3F, RPS28, EIF3G, EIF3H, RPS29, EIF3I, EIF3J, EIF3M, RPS27L |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1268683 | Activation of the mRNA upon binding of the capbinding complex and elFs, and subsequent binding to 43S | BioSystems: REACTOME | 2.01 | 6.44 | 6.40 | $54 / 65$ | FAU, EIF4H, RPS2, RPS3, EIF2S1, RPS3A, RPS5, RPS6, EIF4A1, EIF4A2, EIF4B, EIF4E, RPS7, RPS8, EIF3L, EIF4EBP1, RPS9, EIF4G1, RPS11, EIF3E, RPS12, EIF2S2, RPS13, RPS14, RPS15, RPS15A, EIF3K, RPS16, RPS17, RPS18, RPS19, RPS20, RPSA, RPS21, RPS23, RPS24, EIF3A, EIF3B, RPS25, RPS26, EIF3C, EIF3D, RPS27, RPS27A, EIF3F, RPS28, EIF3G, EIF3H, RPS29, EIF3I, EIF3J, PABPC1, EIF3M, RPS27L |

Table S5. Differentially expressed (DE) genes in the maternal asthma-discriminatory pathways of the turquoise module (see Figure 2 of the main paper) for bronchial (discovery) and nasal (replication) epithelial cells. Of 535 genes that were annotated in the 16 pathways and expressed in bronchial epithelial cells, one gene was DE between asthma cases with ( $N=27$ ) and without $(N=48)$ an asthmatic mother ( $\mathrm{FDR}<0.10$ ). Of 531 genes that were annotated in the 16 pathways and expressed in nasal epithelial cells, 27 genes were $D E$ between asthma cases with $(N=82)$ and without $(N=46)$ an asthmatic mother at ( $F D R<0.10$ ). These genes were among 13 of the 16 maternal asthma-discriminatory pathways.

| Bronchial epithelial cells (one DE gene) |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Gene | Name | Pathway(s) | logFC | P -value | FDR-adjusted P-value |
| ICOS | Inducible T Cell Costimulator | Intestinal immune network for IgA production | -0.97 | 1.61E-04 | 0.062 |
| Nasal epithelial cells (27 DE genes) |  |  |  |  |  |
| Gene | Name | Pathway(s) | logFC | P -value | FDR-adjusted P-value |
| IL2RB | Interleukin 2 Receptor Subunit Beta | Th17 cell differentiation | -0.54 | 1.05E-06 | 5.58E-04 |
| CCL5 | C-C Motif Chemokine Ligand 5 | Rheumatoid arthritis, Influenza A, Interleukin-10 signaling | -0.37 | 3.13E-06 | 8.32E-04 |
| CD3E | CD3e Molecule | Translocation of ZAP-70 to immunological synapse, TCR signaling in naïve CD4+ T cells | -0.35 | 4.52E-05 | 8.00E-03 |
| CD3D | CD3d Molecule | Hematopoietic cell lineage, Th17 cell differentiation, Translocation of ZAP-70 to immunological synapse, TCR signaling in naïve CD4+ T cells | -0.32 | 7.59E-05 | 0.010 |
| CD8A | CD8a Molecule | Hematopoietic cell lineage | -0.43 | 1.29E-04 | 0.012 |
| TCIRG1 | T Cell Immune Regulator 1, ATPase H+ Transporting V0 Subunit A3 | Rheumatoid arthritis | 0.11 | 1.36E-04 | 0.012 |
| C1QA | Complement C1q A Chain | Staphylococcus aureus infection | -0.30 | 4.04E-04 | 0.031 |
| C1QC | Complement C1q C Chain | Staphylococcus aureus infection | -0.43 | 6.10E-04 | 0.040 |
| CD2 | CD2 Molecule | Hematopoietic cell lineage | -0.28 | 7.28E-04 | 0.041 |
| FYN | FYN Proto-Oncogene, Src Family Tyrosine Kinase | Viral myocarditis, Osteoclast differentiation, TCR signaling in naïve CD4+ T cells | -0.29 | 7.85E-04 | 0.041 |
| C4B | Complement C4B (Chido Blood Group) | Staphylococcus aureus infection | -0.65 | 8.78E-04 | 0.041 |
| STAT1 | Signal Transducer And Activator Of Transcription 1 | Tuberculosis, Th17 cell differentiation, Toxoplasmosis, Influenza A, Osteoclast differentiation | -0.16 | 9.32E-04 | 0.041 |
| PLK3 | Polo Like Kinase 3 | Tuberculosis | 0.28 | 1.09E-03 | 0.044 |


| MAP4K1 | Mitogen-Activated Protein <br> Kinase Kinase Kinase <br> Kinase 1 | TCR signaling in naïve CD4+ T cells | -0.38 | $1.38 \mathrm{E}-03$ | 0.052 |
| :---: | :---: | :---: | :---: | :---: | :---: |
| PRF1 | Perforin 1 | Viral myocarditis | -0.54 | $1.63 \mathrm{E}-03$ | 0.058 |
| HIF1A | Hypoxia Inducible Factor 1 <br> Subunit Alpha | Th17 cell differentiation | 0.14 | $1.75 \mathrm{E}-03$ | 0.058 |
| PML | PML Nuclear Body Scaffold | Influenza A | -0.12 | $1.96 \mathrm{E}-03$ | 0.060 |
| RASSF5 | Ras Association Domain <br> Family Member 5 | TCR signaling in naïve CD4+ T cells | 0.15 | $2.02 \mathrm{E}-03$ | 0.060 |
| IL7 | Interleukin 7 | Hematopoietic cell lineage | -0.24 | $2.24 \mathrm{E}-03$ | 0.063 |
| LAMB2 | Laminin Subunit Beta 2 | Toxoplasmosis | 0.18 | $2.54 \mathrm{E}-03$ | 0.066 |
| FPR1 | Formyl Peptide Receptor 1 | Staphylococcus aureus infection, Interleukin-10 <br> signaling | 0.38 | $2.63 \mathrm{E}-03$ | 0.066 |
| CD9 | CD9 Molecule | Hematopoietic cell lineage | 0.05 | $3.08 \mathrm{E}-03$ |  |
| EP300 | E1A Binding Protein P300 | Tuberculosis, Influenza A | 0.10 | $3.09 \mathrm{E}-03$ |  |
| IL4R | Interleukin 4 Receptor | Hematopoietic cell lineage, Th17 cell <br> differentiation | 0.14 | $3.58 \mathrm{E}-03$ |  |
| PIK3R5 | Phosphoinositide-3-Kinase <br> Regulatory Subunit 5 | Toxoplasmosis | 0.36 | $4.19 \mathrm{E}-03$ | 0.079 |
| SIRPB1 | Signal Regulatory Protein <br> Beta 1 | Osteoclast differentiation | 0.56 | $4.39 \mathrm{E}-03$ |  |
| HSPA2 | Heat Shock Protein Family <br> A (Hsp70) Member 2 | Toxoplasmosis, Influenza A | 0.27 | $4.44 \mathrm{E}-03$ | 0.087 |

Table S6. Detailed functional information for each hub DMC.

| IImnID | Module | CHR | BP (Build 37) | UCSC_RefGene_Name | UCSC_RefGene_Accession | UCSC_RefGene_Group |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| cg05277504 | Green | 17 | 79961614 | ASPSCR1 | NM 024083 | Body |
| cg21531873 | Green | 12 | 110778616 | ATP2A2 | NM_001681 | Body |
| cg20436206 | Green | 5 | 146910671 |  |  |  |
| cg07317846 | Green | 2 | 42648344 |  |  |  |
| cg09459548 | Black | 5 | 119942454 | PRR16 | NM_016644 | Body |
| cg25921502 | Black | 10 | 50363896 | C10orf128 | NM_001010863 | 3'UTR |
| cg07270021 | Black | 8 | 60627336 |  |  |  |
| cg11964006 | Black | 10 | 53852689 | PRKG1 | NM_001098512 | Body |
| cg03296565 | Black | 17 | 51191056 |  |  |  |
| cg16284238 | Black | 6 | 87196595 |  |  |  |
| cg09175338 | Black | 17 | 10372273 | MYH4 | NM_017533 | 5'UTR |
| cg20699079 | Black | 2 | 137199850 |  |  |  |
| cg18437033 | Black | 12 | 130188284 | TMEM132D | NM_133448 | Body |
| cg03179496 | Blue | 15 | 29212340 | APBA2 | NM_005503 | TSS1500 |
| cg00030588 | Blue | 16 | 87100811 |  |  |  |
| cg15787712 | Blue | 19 | 13948243 | MIR24-2 | NR_029497 | TSS1500 |
| cg14236389 | Blue | 20 | 58631038 | C20orf197 | NM_173644 | 1stExon |
| cg04661929 | Blue | 17 | 75320035 | SEPT9 | NM_001113492 | 5'UTR |
| cg24292665 | Blue | 11 | 69065779 |  |  |  |
| cg11850468 | Blue | 5 | 180231185 | MGAT1 | NM_001114620 | TSS1500 |
| cg05007163 | Blue | 22 | 39266069 | CBX6 | NM_014292 | Body |
| cg14993283 | Red | 6 | 100906182 | SIM1 | NM_005068 | Body |
| cg17098147 | Red | 10 | 22634142 | SPAG6 | NM_172242 | TSS1500 |
| cg07416383 | Red | 7 | 155302899 | CNPY1 | NM_001103176 | 5'UTR |
| cg22717227 | Red | 10 | 50887568 | C10orf53 | NM_182554 | TSS200 |
| cg16066272 | Red | 14 | 52535949 | NID2 | NM_007361 | TSS200 |
| cg05323725 | Red | 17 | 35294713 | LHX1 | NM_005568 | 1stExon |
| cg00073837 | Red | 2 | 223177008 |  |  |  |
| cg04030584 | Turquoise | 16 | 67871218 | CENPT | NM_025082 | 5'UTR |
| cg22689324 | Turquoise | 1 | 1562397 | MIB2 | NM_001170687 | Body |
| cg10410146 | Turquoise | 7 | 44112411 | POLM | NM_013284 | 3'UTR |
| cg01259126 | Turquoise | 22 | 29705157 | GAS2L1 | NM_152237 | Body |
| cg09440150 | Turquoise | 5 | 6159585 |  |  |  |
| cg05991685 | Turquoise | 16 | 2818793 | SRRM2 | NM_016333 | Body |
| cg04662250 | Turquoise | 17 | 21157499 | C17orf103 | NM_152914 | TSS1500 |
| cg01005308 | Turquoise | 20 | 48768686 | TMEM189 | NM_001162505 | Body |
| cg12492380 | Turquoise | 7 | 98872169 | MYH16 | NR_002147 | Body |


| cg06082897 | Turquoise | 1 | 7767716 | CAMTA1 | NM_015215 | Body |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| cg19874640 | Turquoise | 5 | 134527449 |  |  |  |
| cg04841389 | Turquoise | 6 | 32015083 | TNXB | NM_019105 | Body |
| cg13120756 | Turquoise | 17 | 42386950 | RUNDC3A | NM_001144826 | Body |
| cg14444710 | Turquoise | 16 | 2587080 | PDPK1 | NM_031268 | TSS1500 |
| cg04662836 | Turquoise | 15 | 95334933 |  |  |  |
| cg01191154 | Turquoise | 22 | 37593672 |  |  |  |
| cg04329382 | Turquoise | 22 | 46480891 | LOC400931 | NR_027033 | TSS1500 |
| cg08758352 | Turquoise | 17 | 46205147 |  |  |  |
| cg24221742 | Turquoise | 17 | 74868220 | MGAT5B | NM_198955 | TSS1500 |
| cg09690040 | Yellow | 3 | 126569599 | CHCHD6 | NM_032343 | Body |
| cg05877397 | Yellow | 16 | 49672444 | ZNF423 | NM_015069 | Body |
| cg19807237 | Yellow | 1 | 218470982 | RRP15 | NM_016052 | Body |
| cg19077494 | Yellow | 2 | 158723291 | ACVR1 | NM_001105 | 5'UTR |
| cg00251716 | Yellow | 1 | 243479948 | SDCCAG8 | NM_006642 | Body |
| cg22060153 | Yellow | 2 | 101889653 | SNORD89 | NR_003070 | TSS200 |
| cg07016095 | Yellow | 15 | 71519190 | THSD4 | NM_024817 | Body |
| cg25061701 | Yellow | 17 | 62608856 | SMURF2 | NM_022739 | Body |
| cg25697442 | Yellow | 6 | 1708713 | GMDS | NM_001500 | Body |

Table S7. Associations between co-methylation modules and clinical measures after correcting for hub DMCs. (A) For each module, the number of correlated genes and hub DMCs are shown. No genes were associated with any of the modules after correcting for all of the module's respective hub DMCs (FDR-corrected $P>0.999$ ). (B) Correlation coefficients and $p$-values are shown between clinical measures and module eigenvectors. Significant $p$-values after Bonferroni-correcting for nine tests (clinical phenotypes; $p<5.56 \times 10^{-3}$ ) are shown in bold. The numbers of subjects with measurements for each variable are shown. Asthma severity is determined by STEP classification (1). ns, not significant ( $p>0.05$ ).

| Variables | WGCNA co-methylation modules (number of DMCs) |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | NMA-associated |  | MA-associated |  |  |  |  |
|  | Orange (31) | Green (35) | Black (15) | Blue $(74)$ | Red (17) | Turquoise (73) | Yellow (24) |
| A. Correlated Genes and hub DMCs |  |  |  |  |  |  |  |
| Number of correlated genes |  | 0 | 0 | 0 | 0 | 0 | 0 |
| Number of hub DMCs | 0 | 4 | 9 | 8 | 7 | 19 | 9 |
| B. Clinical Measures |  |  |  |  |  |  |  |
| Asthma Severity $(N=142)$ |  | $\begin{gathered} -4.21 \\ 4.6 \times 10^{-5} \\ \hline \end{gathered}$ | $\begin{aligned} & -2.34 \\ & 0.021 \end{aligned}$ | $\begin{gathered} -3.07 \\ 2.5 \times 10^{-3} \\ \hline \end{gathered}$ | ns | $\begin{gathered} -3.04 \\ 2.8 \times 10^{-3} \\ \hline \end{gathered}$ | $\begin{gathered} 3.07 \\ 2.6 \times 10^{-3} \\ \hline \end{gathered}$ |
| $\begin{gathered} \mathrm{FEV}_{1} \% \text { Predicted } \\ (\mathrm{N}=142) \\ \hline \end{gathered}$ |  | ns | ns | ns | ns | ns | ns |
| $\begin{aligned} & \mathrm{FEV}_{1} / \mathrm{FVC} \\ & (\mathrm{~N}=142) \end{aligned}$ |  | ns | ns | $\begin{gathered} 0.20 \\ 0.018 \end{gathered}$ | ns | ns | ns |
| Total Serum IgE $(\mathrm{N}=141)$ |  | ns | $\begin{gathered} \hline 0.21 \\ 0.013 \\ \hline \end{gathered}$ | ns | ns | ns | $\begin{aligned} & \hline-0.18 \\ & 0.037 \\ & \hline \end{aligned}$ |
| $\begin{gathered} \text { FeNO } \\ (\mathrm{N}=135) \end{gathered}$ |  | ns | ns | ns | ns | ns | ns |
| BAL Eosinophilia $(\mathrm{N}=140)$ |  | ns | ns | ns | ns | ns | ns |
| BAL Neutrophilia $(\mathrm{N}=140)$ |  | ns | ns | ns | ns | ns | ns |
| Blood Eosinophilia $(N=141)$ |  | ns | ns | ns | ns | ns | ns |
| Body Mass Index $(N=142)$ |  | ns | ns | ns | ns | ns | ns |

Table S8. Covariate selection. Principal Components Analysis (PCA) of the 398,186 CpGs that passed quality control was performed using the R function, prcomp, to identify potential technical or biological confounders. The DNA methylation PC1-PC10 and the \% variance explained by each PC are shown in the first column, followed by four potential technical (platform, batch, chip, recruitment source), seven biological (age, sex, self-reported race/ethnicity, current smoking, and ancestry PCs 1-3) covariates. The p-values for tests of association between each PC and each variable are shown. Associations were tested in a step-wise fashion, regressing out the effects of covariates in succession until there were no significant correlations with any of the top 10 DNA methylation PCs. Pvalues for the associations are shown for the (A) raw data, (B) after removing "chip effect" using ComBat (24), and (C) after further regressing out the effects of sex, age, current smoking status, and the first three ancestry PCs. The latter six variables, along with cigarette smoking at time of bronchoscopy, were included as covariates in all analyses. Although the smoking variable was not associated with any of the 10 DNA methylation PCs, we included it as a covariate because smoking is known to have strong effects on global DNA methylation levels. Significant p-values after correcting for 11 tests (potential confounders) using a Bonferroni correction ( $\mathrm{p}<4.55 \times 10^{-3}$ ) are shown in bold font. The effects of asthma and maternal asthma on each of the 10 DNA methylation PCs are also shown.

| A. Raw |  |  |  |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Technical Variables |  |  |  | Biological Variables |  |  |  |  |  |  | Asthma | Maternal Asthma |
| k | Platform | Batch | Chip | Recruitment Source | Age | Sex | Self-reported Race/Ethnicity | Current Smoking | Ancestry PC1 | Ancestry PC2 | Ancestry PC3 |  |  |
| $\begin{gathered} \hline \text { PC1 } \\ (19 \%) \end{gathered}$ | 5.11E-109 | 3.95E-107 | 2.81E-111 | 1.46E-04 | 0.356 | 0.504 | 0.722 | 0.117 | 0.058 | 0.573 | 0.223 | 0.019 | 0.966 |
| $\begin{gathered} \text { PC2 } \\ (12 \%) \\ \hline \end{gathered}$ | 0.817 | 0.010 | 4.67E-07 | 1.60E-03 | 0.287 | 0.182 | 2.72E-03 | 0.385 | 0.027 | 0.668 | 0.730 | 0.315 | 0.870 |
| $\begin{aligned} & \mathrm{PC} 3 \\ & \text { (9\%) } \end{aligned}$ | 0.621 | 2.20E-18 | 1.73E-25 | 2.35E-17 | 0.647 | 0.936 | 0.034 | 0.186 | 0.006 | 0.176 | 0.134 | 0.432 | 0.367 |
| $\begin{aligned} & \text { PC4 } \\ & (5 \%) \\ & \hline \end{aligned}$ | 0.171 | 5.43E-04 | 4.37E-07 | 0.011 | 3.35E-03 | 0.182 | 0.608 | 0.120 | 0.588 | 0.544 | 0.133 | 0.168 | 0.230 |
| $\begin{aligned} & \text { PC5 } \\ & (4 \%) \end{aligned}$ | 0.669 | 0.093 | 0.577 | 5.26E-05 | 2.15E-17 | 0.409 | 0.022 | 0.501 | 0.696 | 0.069 | 0.327 | $\begin{gathered} \hline 7.98 \mathrm{E}- \\ 07 \\ \hline \end{gathered}$ | 0.303 |
| $\begin{aligned} & \text { PC6 } \\ & (2 \%) \\ & \hline \end{aligned}$ | 0.415 | 1.75E-03 | 6.23E-14 | 4.80E-04 | 0.542 | 0.249 | 5.40E-04 | 0.694 | 2.46E-04 | 0.029 | 0.365 | 0.045 | 0.014 |
| $\begin{aligned} & \hline \text { PC7 } \\ & (2 \%) \\ & \hline \end{aligned}$ | 0.747 | 0.288 | 0.082 | 7.46E-06 | 0.065 | 0.889 | 4.19E-14 | 0.564 | 3.78E-20 | 1.24E-06 | 0.198 | 0.855 | 0.874 |
| $\begin{aligned} & \hline \text { PC8 } \\ & (2 \%) \\ & \hline \end{aligned}$ | 0.454 | 0.609 | 4.92E-08 | 0.195 | 0.088 | 0.941 | 0.571 | 0.735 | 0.911 | 0.984 | 0.663 | 0.096 | 0.309 |
| $\begin{aligned} & \text { PC9 } \\ & (1 \%) \\ & \hline \end{aligned}$ | 0.757 | 0.741 | 0.308 | 0.113 | 9.05E-05 | 0.080 | 2.18E-07 | 0.417 | 3.95E-09 | 0.002 | 0.478 | $\begin{gathered} 9.67 \mathrm{E}- \\ 05 \\ \hline \end{gathered}$ | 0.021 |
| $\begin{aligned} & \text { PC10 } \\ & (1 \%) \end{aligned}$ | 0.935 | 2.98E-03 | $3.78 \mathrm{E}-11$ | 0.005 | 0.810 | 0.071 | 0.560 | 0.104 | 0.589 | 0.933 | 0.557 | 0.089 | 0.764 |
| B. After ComBat |  |  |  |  |  |  |  |  |  |  |  |  |  |
|  | Technical Variables |  |  |  | Biological Variables |  |  |  |  |  |  |  |  |
| k | Platform | Batch | Chip | Recruitment Source | Age | Sex | Self-reported Race/Ethnicity | Current Smoking | Ancestry PC1 | Ancestry PC2 | Ancestry PC3 | Asthma | Maternal Asthma |
| $\begin{gathered} \text { PC1 } \\ (14 \%) \\ \hline \end{gathered}$ | 0.791 | 0.937 | 1.000 | 0.208 | 0.328 | 0.322 | 0.036 | 0.604 | 0.479 | 0.131 | 0.294 | 0.052 | 0.401 |
| $\begin{aligned} & \mathrm{PC2} \\ & (7 \%) \end{aligned}$ | 0.507 | 0.859 | 0.999 | 0.549 | 0.020 | 0.107 | 0.130 | 0.395 | 0.641 | 0.662 | 0.891 | 0.020 | 0.089 |
| $\begin{aligned} & \text { PC3 } \\ & (6 \%) \\ & \hline \end{aligned}$ | 0.969 | 0.777 | 1.000 | 8.93E-04 | 1.70E-18 | 0.485 | 0.112 | 0.114 | 0.435 | 0.241 | 0.093 | $\begin{gathered} 8.09 \mathrm{E}- \\ 05 \end{gathered}$ | 0.716 |
| $\begin{aligned} & \hline \text { PC4 } \\ & \text { (3\%) } \end{aligned}$ | 0.858 | 0.329 | 0.999 | 8.85E-05 | 0.658 | 0.155 | 6.27E-16 | 0.584 | 7.93E-24 | 2.60E-08 | 0.505 | 0.404 | 0.425 |
| $\begin{aligned} & \text { PC5 } \\ & (2 \%) \end{aligned}$ | 0.812 | 0.920 | 1.000 | 0.288 | 2.73E-03 | 0.273 | 0.720 | 0.571 | 0.622 | 0.777 | 0.103 | $\begin{gathered} 2.64 \mathrm{E}- \\ 05 \\ \hline \end{gathered}$ | 1.08E-03 |
| $\begin{aligned} & \text { PC6 } \\ & (2 \%) \end{aligned}$ | 0.688 | 0.976 | 1.000 | 0.737 | 0.003 | 0.076 | 2.46E-07 | 0.871 | 2.19E-08 | 1.03E-03 | 0.734 | 0.040 | 0.461 |
| $\begin{aligned} & \text { PC7 } \\ & (2 \%) \\ & \hline \end{aligned}$ | 0.690 | 0.889 | 1.000 | 0.015 | 0.005 | $\begin{gathered} \text { 9.27E- } \\ 06 \\ \hline \end{gathered}$ | 0.013 | 0.408 | 0.023 | 0.017 | 0.021 | 0.779 | 0.333 |
| $\begin{aligned} & \text { PC8 } \\ & (1 \%) \\ & \hline \end{aligned}$ | 0.893 | 0.873 | 1.000 | 0.555 | 0.990 | 0.292 | 0.080 | 0.698 | 0.138 | 0.461 | 0.602 | 0.207 | 0.405 |
| $\begin{aligned} & \hline \text { PC9 } \\ & (1 \%) \\ & \hline \end{aligned}$ | 0.929 | 0.969 | 1.000 | 0.215 | 0.203 | 0.046 | 0.061 | 0.241 | 0.232 | 0.694 | 0.612 | 0.099 | 0.511 |
| $\begin{aligned} & \text { PC10 } \\ & (1 \%) \end{aligned}$ | 0.252 | 0.690 | 1.000 | 0.422 | 0.038 | 0.823 | 0.256 | 0.791 | 0.078 | 0.083 | 0.264 | 0.154 | 0.636 |
| C. After ComBat, ancestry PCs 1-3, age, sex, and current smoking status |  |  |  |  |  |  |  |  |  |  |  |  |  |


|  | Technical Variables |  |  |  | Biological Variables |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| k | Platform | Batch | Chip | Recruitment Source | Age | Sex | Self-reported Race/Ethnicity | Current Smoking | $\begin{gathered} \text { Ancestry } \\ \text { PC1 } \end{gathered}$ | Ancestry PC2 | $\begin{gathered} \text { Ancestry } \\ \text { PC3 } \end{gathered}$ | Asthma | Maternal Asthma |
| $\begin{gathered} \hline \text { PC1 } \\ (15 \%) \end{gathered}$ | 0.569 | 0.849 | 1.000 | 0.148 | 0.887 | 1.000 | 0.729 | 0.570 | 0.944 | 0.996 | 0.878 | 0.095 | 0.660 |
| $\begin{aligned} & \mathrm{PC2} \\ & (8 \%) \end{aligned}$ | 0.404 | 0.749 | 0.998 | 0.903 | 0.834 | 1.000 | 0.406 | 0.832 | 0.917 | 0.994 | 0.822 | 0.172 | 0.156 |
| $\begin{aligned} & \text { PC3 } \\ & (4 \%) \\ & \hline \end{aligned}$ | 0.932 | 0.998 | 1.000 | 0.099 | 0.854 | 1.000 | 0.614 | 0.206 | 0.927 | 0.995 | 0.843 | $\begin{gathered} 1.51 \mathrm{E}- \\ 08 \\ \hline \end{gathered}$ | 0.010 |
| $\begin{aligned} & \text { PC4 } \\ & (3 \%) \end{aligned}$ | 0.982 | 0.817 | 1.000 | 0.112 | 0.994 | 1.000 | 0.984 | 0.567 | 0.997 | 1.000 | 0.994 | 0.061 | 0.094 |
| $\begin{aligned} & \text { PC5 } \\ & (2 \%) \end{aligned}$ | 0.438 | 0.520 | 0.999 | 0.601 | 0.908 | 1.000 | 0.846 | 0.730 | 0.954 | 0.997 | 0.901 | 0.103 | 0.643 |
| $\begin{aligned} & \mathrm{PC6} \\ & (2 \%) \end{aligned}$ | 0.466 | 0.897 | 1.000 | 0.817 | 0.971 | 1.000 | 0.653 | 0.345 | 0.986 | 0.999 | 0.969 | 0.440 | 0.422 |
| $\begin{aligned} & \text { PC7 } \\ & (2 \%) \end{aligned}$ | 0.270 | 0.426 | 0.999 | 0.465 | 0.838 | 1.000 | 0.987 | 0.072 | 0.919 | 0.994 | 0.826 | 0.793 | 0.808 |
| $\begin{aligned} & \text { PC8 } \\ & \text { (1\%) } \end{aligned}$ | 0.696 | 0.955 | 1.000 | 0.701 | 0.912 | 1.000 | 0.706 | 0.325 | 0.956 | 0.997 | 0.906 | 0.623 | 0.882 |
| $\begin{aligned} & \text { PC9 } \\ & \text { (1\%) } \end{aligned}$ | 0.486 | 0.747 | 1.000 | 0.010 | 0.909 | 1.000 | 0.610 | 0.133 | 0.955 | 0.997 | 0.902 | 0.023 | 0.996 |
| $\begin{aligned} & \text { PC10 } \\ & (1 \%) \end{aligned}$ | 0.629 | 0.875 | 1.000 | 0.865 | 0.946 | 1.000 | 0.894 | 0.962 | 0.973 | 0.998 | 0.942 | 0.066 | 0.748 |

## Supplementary References

1. A. Tandon, N. Patterson, D. Reich, Ancestry informative marker panels for African Americans based on subsets of commercially available SNP arrays. Genet Epidemiol 35, 80-83 (2011).
2. G. Jun et al., Detecting and estimating contamination of human DNA samples in sequencing and array-based genotype data. Am J Hum Genet 91, 839-848 (2012).
3. S. Andrews (2010) FastQC: a quality control tool for high throughput sequence data.
4. A. Dobin, T. R. Gingeras, Mapping RNA-seq Reads with STAR. Curr Protoc Bioinformatics 51, 11.14.11-11.14.19 (2015).
5. M. D. Robinson, A. Oshlack, A scaling normalization method for differential expression analysis of RNA-seq data. Genome Biol 11, R25 (2010).
6. C. W. Law, Y. Chen, W. Shi, G. K. Smyth, voom: Precision weights unlock linear model analysis tools for RNA-seq read counts. Genome Biol 15, R29 (2014).
7. P. Langfelder, S. Horvath, WGCNA: an R package for weighted correlation network analysis. BMC Bioinformatics 9, 559 (2008).
8. N. A. E. a. P. Program, Expert Panel Report 3 (EPR-3): Guidelines for the Diagnosis and Management of Asthma-Summary Report 2007. J Allergy Clin Immunol 120, S94-138 (2007).
9. J. Chen, E. E. Bardes, B. J. Aronow, A. G. Jegga, ToppGene Suite for gene list enrichment analysis and candidate gene prioritization. Nucleic Acids Res 37, W305-311 (2009).
10. A. J. Coyle, J. C. Gutierrez-Ramos, The role of ICOS and other costimulatory molecules in allergy and asthma. Springer Semin Immunopathol 25, 349-359 (2004).
11. H. Maazi, O. Akbari, ICOS regulates ILC2s in asthma. Oncotarget 6, 24584-24585 (2015).
12. K. H. Shalaby et al., ICOS-expressing CD4 T cells induced via TLR4 in the nasal mucosa are capable of inhibiting experimental allergic asthma. J Immunol 189, 2793-2804 (2012).
13. A. Rossnerova et al., Factors affecting the 27 K DNA methylation pattern in asthmatic and healthy children from locations with various environments. Mutat Res 741-742, 18-26 (2013).
14. K. K. Belanger, B. T. Ameredes, I. Boldogh, L. Aguilera-Aguirre, The Potential Role of 8Oxoguanine DNA Glycosylase-Driven DNA Base Excision Repair in Exercise-Induced Asthma. Mediators Inflamm 2016, 3762561 (2016).
15. S. Liong, R. Lim, G. Barker, M. Lappas, Hepatitis A virus cellular receptor 2 (HAVCR2) is decreased with viral infection and regulates pro-labour mediators OA. Am J Reprod Immunol 78 (2017).
16. E. Noguchi et al., Positional identification of an asthma susceptibility gene on human chromosome 5q33. Am J Respir Crit Care Med 172, 183-188 (2005).
17. H. Z. Shi, Z. F. Xie, J. M. Deng, Y. Q. Chen, C. Q. Xiao, Soluble CD86 protein in serum samples of patients with asthma. Thorax 59, 870-875 (2004).
18. P. Balbo, M. Silvestri, G. A. Rossi, E. Crimi, S. E. Burastero, Differential role of CD80 and CD86 on alveolar macrophages in the presentation of allergen to T lymphocytes in asthma. Clin Exp Allergy 31, 625-636 (2001).
19. Z. Zhang, J. Wang, O. Chen, Identification of biomarkers and pathogenesis in severe asthma by coexpression network analysis. BMC Med Genomics 14, 51 (2021).
20. K. Amin, C. Janson, I. Harvima, P. Venge, G. Nilsson, CC chemokine receptors CCR1 and CCR4 are expressed on airway mast cells in allergic asthma. J Allergy Clin Immunol 116, 1383-1386 (2005).
21. K. J. Carpenter et al., Therapeutic targeting of CCR1 attenuates established chronic fungal asthma in mice. Br J Pharmacol 145, 1160-1172 (2005).
22. P. Joubert et al., Expression and regulation of CCR1 by airway smooth muscle cells in asthma. $J$ Immunol 180, 1268-1275 (2008).
23. M. Gruchała-Niedoszytko et al., Analysis of the differences in whole-genome expression related to asthma and obesity. Pol Arch Med Wewn 125, 722-730 (2015).
24. J. T. Leek, J. D. Storey, Capturing heterogeneity in gene expression studies by surrogate variable analysis. PLoS Genet 3, 1724-1735 (2007).
